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THE EXTERNAL MALE GENITALIA AND THE PHYLOGENY OF BLATTARIA AND MANTODEA

by

KLAUS-DIETER KLASS



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ABSTRACT

The external male genitalia of Blattaria and Mantodea (phallomeres, phallomere complex) are highly complicated structures, which are always extremely asymmetrical. They are provided with many sclerites and muscles. Their cuticular surface is complexly folded, and there are many distinct in- and evaginations (the formative elements), which may have the shape of spines, lobes, bulges, pouches, apodemes, tendons, etc.. The knowledge of phallomere morphology is extremely incomplete, and the potential for phylogenetic research inherent in these structures has so far hardly been used.

In 4 species of Mantodea and 10 species of Blattaria the sclerites, muscles, and formative elements of the phallomere complex and some other parts of the male postabdomen have been investigated in detail. Most of the subgroups of Blattaria (subfamilies in the system of McKittrick 1964) and four families of Mantodea (of the system of Beier 1968) are represented in this sample. Certain parts of the phallomeres are described for some further species of Blattaria.

A detailed homology hypothesis is presented for the sclerites, muscles, and formative elements of the phallomeres, which includes the homologies between Blattaria and Mantodea. The common ground-plan of Blattaria and Mantodea has been reconstructed. Phallomere characters have been evaluated in terms of phylogeny.

The resulting phylogenetic hypothesis is roughly as follows: In Mantodea, the basal dichotomy is between Mantoididae and the other families; the second one is between Chaeteessidae and the remaining families. In Blattaria, the basal dichotomy is between Blattinae + Polyzosteriinae and the remainder. These remaining Blattaria can be divided into three groups: The first consists of Tryonicinae only. The second contains Cryptocercidae as well as Lamproblattinae and Polyphaginae, the two latter taxa being especially closely related. The third group comprises Blattellidae and Blaberidae. Blattellidae are clearly paraphyletic, with Blaberidae as a rather subordinate subgroup. The first offshoot within Blattellidae (+ Blaberidae) are the Anaplectinae. The subsequent offshoots are various species of Plectopterinae, which is a paraphyletic taxon, too. Blaberidae, Nyctiborinae, Blattellinae and Ectobinae together form a holophyletic group. Nyctiborinae and Blaberidae are possibly sister-groups.

Some other important results are:

(1) The asymmetry of the phallomere complex is homologous in Blattaria and Mantodea, and the morphology of each side is quite similar in the two groups. In Mantodea the hook-process hla (sclerite L3 of McKittrick 1964) is missing; this might be the consequence of a derived copulation procedure.

(2) In the common ground-plan of Blattaria and Mantodea asymmetry is already as extreme as in the extant species. The opinion of Mizukubo & Hirashima (1987) that the stemspecies of Blattaria still had symmetrical phallomeres is refuted.

(3) The ground-plan morphology is most extensively retained in the Mantodea Mantoididae (left side) and Chaeteessidae (right side). In Blattaria, Blattinae have retained many ground-plan features, but in some other phallomere characters they are rather derived. The phallomeres of Cryptocercidae are not close to the Blattarian ground-plan as it is the opinion of McKittrick (1964).

(4) The hypothesis of Bohn (1987) that the side-reversed similarities of the phallomeres of Blaberidae on the one hand and of some subgroups of Blattellidae on the other are due to homology is highly supported. They are not due to parallel evolution as it is the view of Mizukubo & Hirashima (1987).

(5) Homologies between the left and the right side of the phallomere complex can be recognised in only very few respects. Probably, most of the complex morphology of the phallomeres has evolved when asymmetry had already been established. The concept of side-homologous subregions in Mizukubo & Hirashima (1987) and the assumptions of side-homologies in Grandcolas (1994) are refuted.

ZUSAMMENFASSUNG

Die äußeren männlichen Genitalien der Blattaria und Mantodea (Phallomeren, Phallomerenkomplex) sind äußerst komplizierte Strukturen. Sie sind mit vielen Skleriten und Muskeln ausgestattet und sind immer völlig asymmetrisch. Ihre Cuticula ist stark in sich verfaltet, und es finden sich viele markante Ein- und Ausstülpungen (formative Elemente), die die Form von Stacheln, Lappen, Beulen, Taschen, Apodemen, Sehnen o.ä. haben können. Die Phallomeren waren bislang noch kaum vergleichend-morphologisch bearbeitet, und die Möglichkeiten, die sie der Phylogenie-Forschung bieten könnten, wurden bislang noch kaum genutzt.

An 4 Arten der Mantodea und 10 Arten der Blattaria wurden die Sklerite, Muskeln und formativen Elemente des Phallomerenkomplexes sowie einige weitere Teile des Postabdomens eingehend untersucht. Die meisten höherrangigen Teilgruppen der Blattaria (Unterfamilien im System von McKittrick 1964) und vier Familien der Mantodea (nach dem System von Beier 1968) sind in dieser Auswahl vertreten. Teilbereiche der Phallomeren wurden an weiteren Arten der Blattaria untersucht.

Für die Sklerite, Muskeln und formativen Elemente der Phallomeren wird eine detaillierte Homologiehypothese vorgestellt, die auch die Homologiebeziehungen zwischen Blattaria und Mantodea einschließt. Der gemeinsame Grundbauplan der Blattaria und Mantodea konnte weitestgehend rekonstruiert werden. Die Phallomeren-Merkmale wurden in Hinblick auf die Phylogenie ausgewertet.

Aus den Merkmalsverteilungen ergibt sich die folgende Phylogenie-Hypothese: Innerhalb der Mantodea besteht die basalste Dichotomie zwischen Mantoididae und den restlichen Familien, die nächstfolgende zwischen Chaeteessidae und den verbleibenden Familien. Innerhalb der Blattaria ist eine basale Schwestergruppenbeziehung zwischen Blattinae + Polyzosteriinae und den ganzen restlichen Gruppen anzunehmen. Diese restlichen Blattaria lassen sich in drei Gruppen gliedern: Der ersten gehören nur die Tryonicinae an. Die zweite Gruppe umfaßt die Cryptocercidae, Polyphaginae und Lamproblattinae, wobei die beiden letzteren besonders enge Beziehungen zeigen. Der dritten Gruppe sind die Blattellidae und die Blaberidae zuzurechnen. Die Blattellidae sind eindeutig paraphyletisch: die Blaberidae sind eine untergeordnete Teilgruppe dieser Familie. Innerhalb der Blattellidae (+ Blaberidae) sind die Anaplectinae der basalste Seitenzweig. Die nachfolgenden Abzweigungen werden von verschiedenen Vertretern der Plectopterinae repräsentiert, womit auch dieses Taxon als paraphyletisch anzusehen ist. Blaberidae, Nyctiborinae, Blattellinae und Ectobiinae bilden gemeinsam eine holophyletische Gruppierung. Nyctiborinae und Blaberidae sind möglicherweise Schwestergruppen.

Weitere bedeutsame Ergebnisse sind:

(1) Die Asymmetrie des Phallomerenkomplexes ist bei Blattaria und Mantodea homolog, und die Morphologie jeder Seite ist ziemlich ähnlich. Bei Mantodea fehlt der Hakenfortsatz hla (L3-Sklerit in McKittrick 1964), was sich vielleicht als Folge eines apomorphen Kopulationsverhaltens interpretieren läßt.

(2) Im gemeinsamen Grundbauplan der Blattaria und Mantodea ist der Phallomerenkomplex bereits im selben Ausmaß (und in derselben Weise) asymmetrisch wie bei den rezenten Vertretern. Die Ansicht von Mizukubo & Hirashima (1987), die Phallomeren seien bei der letzten gemeinsamen Stammart der Blattaria noch symmetrisch gewesen, wird zurückgewiesen.

(3) Die Morphologie des Grundbauplans wird am umfangreichsten bei den Mantodea Mantoididae (linker Teil) und Chaeteessidae (rechter Teil) beibehalten. Innerhalb der Blattaria sind besonders bei den Blattinae viele Merkmale des Grundbauplans erhalten, in manchen Merkmalen der Phallomeren ist diese Gruppe allerdings bereits stark abgeleitet. Die Meinung von McKittrick (1964), daß die Phallomeren der Cryptocercidae dem Grundbauplan der Blattaria besonders nahestünden, ist abzulehnen.

(4) Die Hypothese von Bohn (1987), daß die seitenverkehrten Ähnlichkeiten der Phallomeren der Blaberidae und mancher Teilgruppen der Blattellidae als Homologien anzusehen sind, wird umfassend bestätigt. Eine Entstehung dieser Ähnlichkeiten durch parallele Evolution, wie sie Mizukubo & Hirashima (1987) annehmen, erscheint äußerst unwahrscheinlich.

(5) Homologien zwischen linker und rechter Seite des Phallomerenkomplexes lassen sich nur bezüglich sehr weniger Elemente begründen. Es ist zu vermuten, daß ein großer Teil der komplexen Phallomeren-Morphologie erst ausgebildet wurde, als bereits eine deutliche Asymmetrie etabliert war. Das Konzept seitenhomologer Subregionen von Mizukubo & Hirashima (1987) und auch die Annahmen von Seitenhomologien in Grandcolas (1994) lassen sich widerlegen.

1. INTRODUCTION

Blattaria, Mantodea, and Isoptera form a holophyletic group called Dictyoptera (s. lat.; Kristensen 1995) or Blattopteroidea (Hennig 1969). The relationships between the three groups are unresolved. Hennig (1969) regards Mantodea as the sister-group of Blattaria + Isoptera, and he points out the possibility that Isoptera might be a subgroup of Blattaria – closely related to the Blattarian family Cryptocercidae. These assumptions probably reflect the most parsimonious solution, but the arguments are scarce. Thorne & Carpenter (1992) assume that Isoptera are the sister-group of Blattaria + Mantodea. However, their results are not very convincing, since for many characters the assumed polarities are questionable (Kristensen 1995; Klass 1995).

Concerning the internal phylogeny of Blattaria and of Mantodea, the current ideas are based on the extensive investigations of McKittrick (1964) and McKittrick & Mackerras (1965) (Blattaria) and on the survey in Beier (1968) (Mantodea). In terms of systematics, these authors will be followed in this paper. In some aspects the ideas of these authors are well-founded, but many points are still debatable.

Beier (1968) divides the Mantodea into 8 families: Chaeteessidae, Metallyticidae, Mantoididae, Amorphoscelididae, Eremiaphilidae, Hymenopodidae, Mantidae, and Empusidae. These are not grouped into higher-ranked categories. Chaeteessidae are more primitive than all other families in that their hind-wings have a complete second anal-vein and in that their fore-legs are beset with stout setae rather than thorns. In Metallyticidae the second anal vein is vestigial, and in the other families the vein is completely missing. Hence, Chaeteessidae seem to be the first offshoot and Metallyticidae the second.

McKittrick (1964) divides the Blattaria into two sister-groups, Blattoidea and Blaberoidea. The Blattoidea, comprising Cryptocercidae and Blattidae, do not reveal a single feature that could be unambiguously regarded as a synapomorphy of the two families. The phylogenetic position of Cryptocercidae has been reanalysed by Grandcolas (1994), who assumes that they are a subgroup of Polyphaginae. However, this assignment is also not very convincing, since many of the homology assumptions upon which this assignment is based are debatable. The Blattidae of McKittrick, comprising Blattinae, Polyzosteriinae, Tryonicinae, and Lamproblattinae, are based on features most of which can be suspected to be plesiomorphic for Blattaria, and the family might be para- or even polyphyletic. The Blaberoidea of McKittrick, including Polyphagidae, Blattellidae, and Blaberidae are founded on the presence of a pair of special compound sclerites in the ovipositor, the crosspieces. However, crosspieces are simple gonangula strictly homologous with those of the other Blattaria (Klass 1995, in press), and the holophyly of Blaberoidea is thus highly questionable. McKittrick's assumption that Blattellidae and Blaberidae are closely related is well-founded. However, the exact relations between the two families are uncertain. Interpreting the morphological results of McKittrick (1964) concerning the male and female genitalia from the viewpoint of phylogenetic systematics and parsimony, the Blaberidae would have to be regarded as a rather subordinate subgroup of Blattellidae; however, not all features are consistently supporting this view. The relations between the various subgroups of Blattellidae, which are Anaplectinae, Plectopterinae, Blattellinae, Ectobiinae, and Nyctiborinae, are also rather unclear.

The external male genitalia of Blattaria and Mantodea (the phallomere complex composed of the phallomeres) have a highly complicated morphology. The knowledge of these structures is extremely scarce. However, a large potential for phylogenetic research can be supposed to be inherent in them, and this will be used in this paper to contribute to the solution of the basic problems of Blattarian and Mantodean phylogeny.

The phallomere complex, or at least its anterior part, is concealed within a genital pouch. Abdominal sternite 9 is a saucer-shaped subgenital plate, and the pouch is mainly formed from the intersternal membrane between the sternites 9 and 10, which is deeply invaginated anteriad. The ejaculatory duct opens into this pouch, and the phallomeres are evaginations surrounding the genital opening. The phallomere complex is provided with many sclerites

and muscles, and with many distinct in- and evaginations of the cuticle (formative elements such as processes, lobes, pouches, apodemes, or tendons). The structure as a whole is always completely asymmetrical. Its morphology is quite variable within Mantodea (LaGreca 1954) and extremely so within Blattaria (McKittrick 1964). In Isoptera the phallomeres are said to be missing (Weidner 1970), a situation which is, according to Matsuda (1976), an element of the neotenic traits generally observable within this taxon.

The phallomere morphology of Mantodea has been studied by Snodgrass (1937) in *Tenodera sinensis*, by Levereault (1936, 1938) in *Stagmomantis carolina*, and by LaGreca & Rainone (1949) in *Mantis religiosa*. In each of these studies the cuticular elements and the musculature are described. The three species are closely related (Mantidae, Mantinae, Mantini in Beier 1968), and their phallomeres are rather similar. LaGreca (1954) compared the cuticular elements of the phallomeres of several Mantodea. In this selection all families of Beier (1968) are represented, except for those regarded as most primitive: Chaeteessidae, Metallyticidae, and Mantoididae.

Regarding the phallomeres of Blattaria, the paper of Snodgrass (1937) is the most important of the earlier contributions. The sclerotisations and – in part – the musculature of *Periplaneta americana*, *Blatta orientalis* (both: Blattidae, Blattinae), *Blattella germanica* (Blattellidae, Blattellinae), and *Ectobius lapponicus* (Blattellidae, Ectobiinae) are described. The phallomeres of Blattidae and Blattellidae are very different from each other, and assumptions on homology relations are made only to a very small extent.

McKittrick (1964) investigated the phallomere sclerites in 24 genera of Blattaria and gives a homology hypothesis. However, the descriptions are not very detailed, and the musculature has not been studied. Thus, this homology hypothesis is not very convincing in many points. McKittrick introduced a new terminology for the phallomere sclerites: The terms are composed of several letters and numbers, each giving some information about the position and the homology relations of the respective sclerite. McKittrick regards the phallomeres of *Cryptocercus* (Cryptocercidae) as primitive within Blattaria.

Grandcolas (1994) studied the phallomere sclerites of *Cryptocercus* and some Polyphaginae and Blattinae. He finds many synapomorphies for *Cryptocercus* and (subgroups of) Polyphaginae and assigns *Cryptocercus* to Polyphaginae. However, the homology relations assumed for the sclerites are disputable in many cases.

Mizukubo & Hirashima (1987) studied the phallomere sclerites and muscles of *Periplaneta* (various species; Blattinae), *Blattella* (various species; Blattellinae), and *Opisthoplatia orientalis* (Blaberidae). They use – with some modifications – the terminology of McKittrick. The authors homologise the phallomere sclerites according to their relative positions and their mutual relations. Furthermore, they introduce a new topic into the discussion: Homologies between elements of the left and of the right half of the phallomere complex are considered. In their analysis they dismiss the musculature as a valuable reference system for homologising. The phallomeres of the stem-species of Blattaria (excluding Mantodea) are supposed to be still symmetrical. Accepting this view, the asymmetry of the phallomere complex would have to be regarded as non-homologous in Blattaria and Mantodea. In the case of Blattellidae and Blaberidae, which families show obvious but side-reversed similarities in their phallomere morphology, Mizukubo &

Hirashima assume a parallel evolution of these similarities, and they also assume a still symmetrical morphology in the common stem-species of these two families. In McKittrick (1964), the same view is indicated through the designation of the sclerites. In contrast, Bohn (1987) supposes that the phallomeres of Blaberidae have undergone a change of their left-right-asymmetry and that the similarities concerned are homologous. The point of discussion is the same in the case of Plectopterinae, whose phallomeres also show side-reversed similarities with the other subfamilies of Blattellidae.

The knowledge of the morphology of the phallomeres and the other parts of the male postabdomen of Blattaria and Mantodea is extremely incomplete. The few hypotheses concerning homology relations (between species and between the left and right halves of the phallomere complex), the ground-plan, and the evolution of the phallomeres are not very convincing. Furthermore, nothing is known about homology relations between the phallomeres of Blattaria and Mantodea. Thus, there is compelling need for a large-scaled morphological investigation of the phallomere complex.

To do this is the intention of this paper. Phallomere morphology will be analysed in Blattarian and Mantodean species representing the various subgroups. This investigation should be as detailed as possible in order to avoid misinterpretations due to superficial observation and in order to get as many arguments as possible for assumptions and conclusions. The homology relations between the various species will be worked out in detail, and possible homologies between the left and the right side of the phallomere complex will be considered. The ground-plan features of the phallomeres will be reconstructed – focused on the common ground-plan of Blattaria and Mantodea, if there is one. The special condition of the phallomere elements in the various species and their evolution will be discussed in detail. The characters of the phallomeres will be evaluated in order to establish a phylogenetic hypothesis for Blattaria and Mantodea. The terminology for the phallomere elements will be based on the common ground-plan of Blattaria and Mantodea. Mantodea – in accordance with the homology relations assumed. A standardised, wellfounded, and well-defined use of the terminology might also be valuable for taxonomic research and the description of species.

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2. MATERIAL AND METHODS

Species investigated

Mantodea

Mantoididae Mantoida schraderi Rehn, 1951

Chaeteessidae Chaeteessa caudata Saussure, 1871

Metallyticidae Metallyticus violaceus Burmeister, 1838

Mantidae Sphodromantis Stal, 1871 (sp. indet.); Mantis religiosa (Linné, 1758)

Blattaria

Blattoidea

Cryptocercidae Cryptocercus punctulatus Scudder, 1862

Blattidae

Blattinae Archiblatta hoeveni Sn. v. Vollenhoven, 1862; Blatta orientalis Linné, 1758; Deropeltis Burmeister, 1838 (sp. indet.); Periplaneta americana (Linné, 1758)

Polyzosteriinae Eurycotis floridana (Walker, 1868)

Tryonicinae Tryonicus parvus (Tepper, 1895); Tryonicus angustus (Chopard, 1924)

Lamproblattinae Lamproblatta albipalpus Hebard, 1919

Blaberoidea

Polyphagidae

Polyphaginae *Polyphaga aegyptiaca* (Linné, 1758); *Ergaula capensis* (Saussure, 1893); *Ergaula capucina* (Brunner v. W., 1893)

Blattellidae

Anaplectinae Anaplecta Burmeister, 1838 (sp. indet.) Plectopterinae Nahublattella Bruijning, 1959 (sp. indet.); Euphyllodromia angustata (Latreille, 1811); Supella longipalpa (Fabricius, 1798)

Blattellinae *Parcoblatta lata* (Brunner v. W., 1865); *Loboptera decipiens* (Germar, 1817)

Ectobiinae Ectobius sylvestris (Poda, 1761)

Nyctiborinae Nyctibora Burmeister, 1838 (sp. indet.)

Blaberidae Blaberus craniifer Burmeister, 1838; Byrsotria fumigata (Guérin-Méneville, 1857); Blaptica Stal, 1874 (sp. indet.); Nauphoeta cinerea (Olivier, 1789)

The assignment of the respective genera to the various taxa is adopted from McKittrick (1964) and Beier (1968). The only exceptions are: The assignment of *Tryonicus* is taken from McKittrick & Mackerras (1965). *Polyphaga* and *Ergaula* are assigned to Polyphaginae according to Grandcolas (1994). *Euphyllodromia* is assigned to Plectopterinae by Roth (1967). *Archiblatta* is in its phallomere morphology rather close to *Deropeltis* (McKittrick 1964, fig.108), which is assigned to Blattinae by McKittrick (1964). *Nahublattella* is in its phallomere morphology very close to *Lophoblatta* (McKittrick 1964, fig.113), which is assigned to Plectopterinae by McKittrick (1964).

In the subsequent text these species will be named by their generic name alone. (Tryonicus

is always *T. parvus*, *Ergaula* is always *E. capensis*. *T. angustus* and *E. capucina* will be addressed by their complete names).

Sphodromantis, Mantis, Deropeltis, Periplaneta, Blatta, Eurycotis, Supella, Parcoblatta, Loboptera, Blaberus, Nauphoeta, Blaptica, Byrsotria, Ergaula capucina, and Polyphaga have been reared in the laboratory and were available as freshly killed specimens. Lamproblatta, Anaplecta, Nahublattella, Euphyllodromia, Nyctibora, and Mantoida have been stored in 4% formaldehyde, Cryptocercus and Ectobius in 70% isopropanol. Chaeteessa, Metallyticus, Archiblatta, the two Tryonicus-species, and Ergaula capensis were dried specimens.

Preparation

For the examination of the musculature the abdomina were cut off and stored in 75% isopropanol for at least three days. For the study of cuticular elements the soft tissues were removed by treating the abdomina with 10% KOH for 4-20 hrs. at 40°C. The remaining cuticular structures were then washed in distilled water and stored in 75% isopropanol. Descriptions of morphological structures are always based on preparations of several specimens, with the exception of some species of which only one or two specimens were available (*Mantoida, Chaeteessa, Metallyticus*, both species of *Tryonicus, Ergaula capensis, Archiblatta*). Preparation was performed with sharp forceps and iris scissors. In the observation of small and weakly sclerotised structures it was sometimes useful to underlay the object with a piece of aluminium foil.

Remarks on the figures 1-319

- In all figures anterior is towards the top and posterior is towards the bottom of the sheet.
- The cuticle generally has an internal surface, which is in contact with the epidermis, and an external surface. In all figures the cuticle is partly seen from internally and partly from externally.
- Dark areas are sclerotised; white areas are membranous.
- Muscles are hatched longitudinally, in correspondence with the course of their fibers.
- In each figure only those structures are shown which can be directly seen by the observer and which are not covered by other structures. Thus, e.g. sclerites covered by membrane are not shown, even if they can be easily seen through the membrane in an original preparation.
- Mostly the cuticle is very thin, and in drawing it is regarded as a convoluted plane without thickness. Only in some cases when the cuticle is strongly thickened its thickness is considered in drawing.
- In drawing, the geometry of the cuticular foldings and of the other elements is strictly held to. Continuous black lines represent edges. Edges are understood throughout as lines along which the cuticle or the surface of a muscle curves beneath itself and thus vanishes from the observer's view. What appears as an edge is dependent on the angle of view. Edges of the cuticle can be external or internal: External edges are directed to the exterior, and along them the external surface of the cuticle is visible; internal edges are directed to the interior of the body, and along them the internal surface of the cuticle is visible. Edges beneath the visible surface are sometimes drawn as broken lines.

- The boundaries of the insertion areas of muscles are also shown by continuous black lines if they are directly visible. Parts of these boundary lines which are covered by the muscle itself or by other structures are drawn as broken lines. In some cases the insertion areas alone are drawn without the respective muscles (mainly in the figures showing the subgenital plates, e.g. fig.5); the boundary of the insertion area is again shown by a continuous black line.
- Undulate lines are cutting lines (through cuticle of normal thickness) or bound cut surfaces (of muscles or strongly thickened cuticle cut through).
- The series of figures pertaining to a certain species has as a whole been worked out with the intention of showing all relative positions of all elements of the phallomeres, including all the membranous foldings.
- The series of figures for the various species are designed for the best possible comparability. For example, in the overall views of the postabdomina (compare fig.1, 2, 3 and fig.58, 59, 60) the cutting lines have the same course in each species (their courses are, so to speak, homologous), and, if present, the same muscles are shown. Or, the right phallomeres of Blattaria are always shown in the same four aspects.

Remarks on homology

As a principle, elements regarded as homologous will be given the same name, and elements given the same name are regarded as homologous. Minor exceptions to this rule, mainly due to a not very high probability of homology, will be explicitly mentioned in the text.

Mainly the criteria of the relative position and of the special structure (1. and 2. major criteria of Remane 1952) will be used in this paper. These will be applied to the following kinds of structures, whose relative position and special structure will be comprehensively discussed in the homology analysis:

- Sclerotised areas of the cuticle (sclerites).
- Articulations or other special relations between sclerites.
- Formative elements: more or less distinct evaginations or invaginations of the cuticular surface of the phallomere complex (processes, ridges, pouches, tendons, apodemes, lobes, etc.).
- Insertion areas of muscles.
- The opening of the ejaculatory duct or genital opening.
- The opening of the phallomere-gland (a gland within the left part of the phallomere complex).

The sclerotisations of the phallomeres will be divided into areas which are strictly homologous in the different species. These areas are the main sclerites (or sclerite groups, if these main sclerites have split into several isolated sclerites) and – as their subunits – the sclerite regions. Some difficulties arise in this demarcation of homologous areas within the cuticular surface of different species and in the standardisation of this procedure. The following example shall illustrate these problems: Provided: In two compared species A and B homology is certain (as much as it can be) for a sclerite as well as for a muscle. In species A the muscle inserts on the sclerite, in species B the muscle inserts on the

membrane next to the sclerite. The situation in A is primitive, the situation in B is derived. The derivation which B shows as compared with A can be interpreted in two different ways: (1) In B the insertion of the muscle has shifted from the sclerite to the membrane. (2) In B the sclerite has diminished and has "lost" the insertion of the muscle. According to (1) the sclerites of the two species are homologous in a strict sense. According to (2) they are homologous only in part, since in species B part of the sclerotisation has been lost. In one peculiar case the special circumstances can suggest an interpretation according to either (1) or (2). In many cases, however, an objective decision in favor of one of the two alternatives is hardly possible, and it is debatable whether a discussion of such a case is of importance at all. The interpretation will then be done in that way which seems to be more suitable for the explanation of homology relations.

3. GENERAL DESCRIPTION AND MORPHOLOGICAL DISCUSSION OF THE POSTABDOMEN AND OF THE MALE GENITAL REGION OF DICTYOPTERA

The postabdomen of male Dictyoptera comprises the abdominal segments 9-11 and the telson, which contains the anus (Snodgrass 1937). Matsuda (1976) postulates a twelfth segment for the ground-plan of insects, and this would also affect the interpretation of the Dictyopteran postabdomen. According to Matsuda himself, p.52, however, this "segment" contains neither mesoderm rudiments, nor ganglion rudiments, nor appendage rudiments. Thus, it does not fulfil either criterion to be regarded as a segment. This "twelfth segment" could be regarded as a (highly reduced) segment, if it is demonstrated to be homologous with a true segment (containing mesoderm) of another group of Arthropoda, having lost its segmental organs secondarily. This, however, has not been shown. Therefore, the twelve-segment-theory of Matsuda is not followed here.

Subsequently the general morphology of the postabdomen and the phallomeres of Blattaria and Mantodea will be described. This will essentially be a description of the common ground-plan of Blattaria and Mantodea, whose reconstruction will be substantiated step by step later on in this paper.

3.1. The cuticular elements

Abdominal segment 9

The sternite of segment 9 (subgenital plate, **S9** in fig.320, 321) always forms a large lobelike extension to the posterior, which reaches or even exceeds the morphological posterior end of the body (with the anus **Af** in fig.320, 321c). Most species have a large membranous or only weakly sclerotised area in the anterior half of the subgenital plate (**M** in fig.320, 321b,d,k). The (heavier) sclerotisation is continuous in antero-posterior direction only in the lateralmost parts, to the left and to the right of area **M** (fig.321k). The lateral parts of the subgenital plate (**S91** in fig.321k) curve upwards. The posterior edge of the subgenital plate bears styli (**S9s** in fig.320, 321b,d,k).

Along the anterior margin of the subgenital plate the intersternal membrane connecting sternites 8 and 9 adjoins and bends back sharply to the posterior margin of sternite 8

(compare fig.320). A more or less extensive anterior part of the subgenital plate is thus concealed by the posterior part of sternite 8 from ventrally and can be regarded as a broad apodeme or internal apophysis. The paired parts which project especially deeply anteriad (**S9a** in fig.321b,d,k) are the lateral sternal apodemes or apophyses; these paired parts will be designated as the apophyses of the subgenital plate subsequently. The summits of what I call apophyses can reasonably be regarded as homologous in the various species though they can take various positions from far lateral (like in fig.265) to far medial (like in fig.22). However, strict homology is certainly not true for the whole apophyses (= paired parts): The apophyses can be separated from each other to the far posterior (like in fig.265), the paired parts being very long, or the whole median part of the subgenital plate is produced far anteriad and only the anteriormost parts show the paired condition (like in fig.296). The apophyses present in the latter situation seem to correspond only to the anteriormost parts of the apophyses present in the former situation, and probably some median fusion has taken place in the posterior part. Hence, the term "apophysis" as used here is not intended to claim strict homology.

According to Walker (1922) and other authors, the subgenital plate is not the sternite of segment 9 only but is probably composed of: (1) the true sternite 9 (the part anterior to **M**); (2) the paired but medially fused coxites of segment 9 – probably serially homologous with the thoracic coxae or with more extensive basal parts of the thoracic appendages. If this composition is true, the subgenital plate is a coxosternite. The styli **S9s** sitting upon the coxites are probably serially homologous with distal parts of the thoracic legs.

The tergite of segment 9 (**T9** in fig.320, 321a) resembles the more anterior tergites, but like tergite 8 it is rather short. Its lateral parts (= paratergites, **T9p** in fig.321b) incline ventrad from the dorsal main part – often along a distinct edge. The ventral margins of the paratergites overlap in most cases, and to a varied extent, the lateral parts of **S9**.

The position of the phallomere complex

The intersternal membrane between sternites 9 and 10 is deeply invaginated anteriad to form the walls of the funnel-shaped genital pouch: The ventral wall of this pouch extends anteriad from the lateral and posterior edges of the subgenital plate (Vw in fig.321k, left half, fig.320). The dorsal wall extends anteriad from the anterior margins of the paraprocts **Pp** and **Pv**-sclerites (**Dw** in fig.320, 321b). The lateral walls extend anteriad from the posterior edge of the pleural membrane between tergite 9 and subgenital plate (**Sw** in fig.321b,d).

Deep in the genital pouch the cuticle turns posteriad again and forms the walls of the phallomere complex. The edge or line of turning, along which the walls of the genital pouch meet the walls of the phallomere complex, will be called the basal line (**Bl** in fig.320, 321b,d). Hence, the phallomere complex seems to be exclusively an elaboration of the intersternal area between sternites 9 and 10. The ejaculatory duct (**D** in fig.321b,d,e,g) opens on the phallomere complex.

In many Blattaria and Mantodea, the ventral wall of the genital pouch (Vw in fig.320, 321k), which covers the posterior part of the subgenital plate from dorsally, contains a sclerotisation (**S9d** in fig.320, 321b,d,k). **S9d** is regarded as a dorsal sclerotisation of the

subgenital plate and is possibly the sclerotisation of the dorsal walls of the fused coxites and hence a part of the appendages of segment 9. This **S9d** can be either separated from or connected with the ventral main sclerotisation of the subgenital plate (around the lateral and posterior edges of the plate, as in fig.321k), and it may either occupy an extensive part of the ventral wall of the genital pouch or is restricted to the marginal areas close to the edges of the subgenital plate. The sclerotisations comprised in **S9d** are certainly not homologous in a strict sense throughout the species.

The phallomere complex

The phallomere complex will be divided into two main parts belonging to the left and to the right half of the body: left complex and right phallomere. This major division is shown in fig.321e and f, where the two parts are separated (compare fig.321d). Both are complicated structures with intensively folded cuticle and with sclerotised and membranous areas. Left complex and right phallomere are extremely asymmetrical in all Blattaria and Mantodea. The phallomere-gland (penis-gland, conglobate gland; **P** in fig.321e) opens on the left complex; at least its outlet channel is cuticulised. Since the morphology of the phallomere complex is highly variable within Blattaria and Mantodea, a description valid for all subgroups is impossible. The following description corresponds to the common ground-plan of Blattaria and Mantodea. In addition, some important derived states will be mentioned.

Left complex

Several left-lateral and ventral sclerites are designated L4 (fig.321e,g,i): A large crescentshaped L4-sclerite occupies the left edge of the left complex, including the adjacent margins of the dorsal and ventral walls, and the anteriormost ventral wall. Along most of this sclerite there runs an apodeme (swe in fig.321e,g), which is groove-like posteriorly but solid and beam-like anteriorly (the groove is filled in by the cuticle becoming thickened). The posteriormost part of the sclerite occupies a short process (pda in fig.321e,g). As a derived condition, the dorsal part of the sclerite can be strongly expanded to the right, and the dorsal and the ventral parts of the sclerite can be separated. A second, plate-like L4-sclerite lies in the right ventral wall. Another L4-sclerite in the anterior left ventral wall bears a node-like process (nla in fig.321i; present in Blattaria only). These three L4-sclerites can be separated from or connected with each other in the anterior ventral wall.

In the central and right parts of the left complex there are two pouches invaginated anteriad, which lie one above the other. The walls of the dorsal pouch (**pne** in fig.321e,n) are largely occupied by the hood-shaped sclerite L1. The phallomere-gland (**P** in fig.321e) opens into this **pne**-pouch. The ventral pouch (**lve** in fig.321e,g) contains the L2-sclerite, which is often restricted to the dorsal wall of the pouch and extends like an arch along its anterior and lateral margins. The left posterior part of sclerite L2 leaves the **lve**-pouch and extends onto a process (**paa** in fig.321e,g) immediately to the right of the **pda**-process. The sclerotisations of L4 and L2 are connected in between the processes **pda** and **paa**. The right end of sclerite L2 articulates with sclerite L1 (articulation A2 in fig.321e,n).

The ventral wall of the **lve**-pouch is almost completely membranous, and it is at the same time the left-anterior part of the dorsal wall of a large ventral lobe (**vla** in fig.321e,g,i). The ejaculatory duct (**D** in fig.320, 321g) opens into this wall. In some species this wall contains a small sclerite **L5** (fig.321e,g). The ventral wall of the **vla**-lobe is part of the ventral wall of the whole left complex and is largely occupied by the right-ventral **L4**-sclerite (fig.321g,i).

All Blattaria, but not Mantodea, have a large hook-like evagination from the left ventral wall of the left complex (**hla** in fig.321i). The **hla**-hook is largely occupied by sclerite **L3**, but a more or less extensive basal part is membranous (**30** in fig.321i). This membrane can be introverted, which procedure results in a more or less deep retraction of the hook into the left complex.

Right phallomere

The anteriormost ventral wall is occupied by the plate-like sclerite **R3** (fig.321f,h). Along the lateral and anterior margins of **R3** (parts of the basal line **BI**) the sclerotisation of **R3** extends somewhat into the wall of the genital pouch, and these margins of sclerite **R3** form a groove- (as seen from externally) or fold-like (as seen from internally) apodeme **age** (fig.321f,h). Like in the **swe**-apodeme, parts of this **age**-apodeme can be filled in by the cuticle becoming thickened, and the respective parts of **age** are beam-like.

Behind the central part of **R3** the ventral wall is extensively invaginated dorsad and anteriad (**cbe** in fig.321f,h), and this invagination is partly sclerotised in its dorsal wall (anterior part of sclerite **R1** in fig.321f). Blattaria, but not Mantodea, have a sclerite **R2** left-ventral to the **cbe**-invagination (fig.321f,h), which articulates with the left posterior end of **R3** (articulation **A7** in fig.321f) and with the left end of **R1** (articulation **A6** in fig.321f). Sclerite **R2** and the posterior margin of the anterior part of **R1** often form tooth-or ridge-like cuticular evaginations (on **R1**: **pva** in fig.321h).

The part of the right phallomere posterodorsal to the **cbe**-invagination is composed of a large dorsal lobe (**fda** in fig.321f) and a ventral tooth (**pia** in fig.321h, which is in most species much smaller than the **fda**-lobe). The **fda**-lobe and the **pia**-tooth are confluent along the right edge of the right phallomere, and they diverge to the left. The dorsal wall of the **fda**-lobe – and often parts of its ventral wall, too – as well as the dorsal and ventral walls of the **pia**-tooth are occupied by the posterior part of sclerite **R1**. The sclerites **R1** and **R3** articulate with each other at the anterior right edge of the right phallomere (articulation **A3** in fig.321f,h). In the anteriormost dorsal wall of the **fda**-lobe, part of the cuticle is invaginated to form a hollow, long and narrow, membranous tendon (**tre** in e.g. fig.74; not shown in fig.321), which is present in some Blattaria only.

Discussion of the basic division of the phallomere complex

I propose this division of the phallomere complex into a left complex and a right phallomere. However, earlier suggestions for a basic division differ from this hypothesis: Snodgrass (1936, 1937) divides the phallomere complex of Blattinae into a ventral, a right, and a left phallomere. Beier (1970) follows Snodgrass regarding Blattaria as well as Mantodea, and he terms these main divisions hypophallus, right epiphallus, and left

epiphallus. The ventral phallomere (= vla-lobe in the previous description) lies ventral to the genital opening (in a strict morphological sense anterior to it); i.e. the genital opening is in its anteriormost dorsal wall. The right phallomere (= right phallomere in the previous description) and the left phallomere (= left complex minus the vla-lobe) have their bases in the areas dorsal (in a strict sense: posterior) and lateral to the genital opening. Snodgrass (1937) deduces this basic division from his investigations of the ontogenetic stages of the phallomere complex in *Periplaneta americana* and *Blatta orientalis* (both: Blattinae): In medium-sized nymphs the phallomere complex consists of three distinct lobes, which hardly reveal any further elaborations. One lobe is situated medioventral to the prospective genital opening (prospective ventral phallomere), the other two take positions dorsolateral to the genital opening (prospective right and left phallomeres). Thus, a composition of a medioventral, a right-dorsal, and a left-dorsal basic element seems plausible, and according to Snodgrass (1937) the ventral phallomere is an unpaired medioventral element.

Quadri (1940) studied the ontogeny of the phallomere complex of *Blatta orientalis* in more detail. In first instar nymphs he finds one pair of lobes with an invagination between them (rudiment of ejaculatory duct). In the third instar each lobe is divided into a dorsal and a ventral secondary lobe, and thus four lobes surround the prospective genital opening. Later, the two left lobes form the left phallomere (more or less by fusion, without any clear border remaining). The ventral right lobe shifts to the left, into a position beneath the genital opening, and becomes the ventral phallomere. The dorsal right lobe maintains its position and becomes the right phallomere. Thus, according to Quadri, the ventral phallomere is a ventral part of the right half of the phallomere complex. Later Snodgrass (1957) took over the opinion of Quadri but still used the tripartite division in his terminology.

Concerning the assignment of the ventral phallomere, or ventral lobe **vla**, my own observations as well as the mode of innervation (Pipa 1988) are in conflict with the views of both Snodgrass (1937) and Quadri (1940):

In some aberrant specimens of Blattaria the phallomere complex is completely symmetrical. I could find two such specimens: (1) *Polyphaga aegyptiaca* (Polyphaginae) with two "right" phallomeres being mirror-images of each other; there was no trace of a ventral phallomere, which is present in normal specimens. Unfortunately, the specimen had been dried and macerated, and the relations to the internal genitalia and the presence of an ejaculatory duct could not be investigated. (2) *Deropeltis* sp. (Blattinae) with two "left" phallomeres and two complete ventral phallomeres, both pairs being mirror-images of each other. The phallomere-gland is paired. The ejaculatory duct is, as usual, unpaired. It opens in the median plane – in that area where the dorsal walls of the left and the right ventral phallomeres are confluent with each other. Thus, the location of the genital opening – in relation to the ventral phallomeres – is the same as in normal specimens, and the two ventral phallomeres are arranged in a way that this relative position is true of both of them.

Pipa (1988) studied the innervation of the male postabdomen in *Periplaneta americana*: From the posterior part of the last abdominal ganglion – a compound ganglion formed from the ganglion rudiments of abdominal segments 7 to 11 – one pair of nerves runs to the phallomeres (phallic nerve = nerve 5a in Pipa). Their basal branchings are symmetrical. After entering the phallomeres, where the branches innervate the phallomere muscles, the branching pattern becomes completely asymmetrical. The ventral phallomere gets its innervations exclusively from branches coming from the left nerve.

The morphology of the two symmetrical specimens and the innervation pattern suggest that the ventral phallomere is neither an unpaired ventromedian element of the phallomere complex nor a part of its right half but a lobe-like part of its left half. I term it the **vla**-lobe, and the left and ventral phallomeres together I term the left complex. There is another, more practical (though not decisive) reason for this concept: The morphological relations between the left phallomere and the ventral phallomere are often very close, and the border between them is in many cases not very distinct. And this is with high probability the ground-plan situation (like in fig.321e,i). However, the question of the correct assignment of the ventral phallomere or **vla**-lobe is certainly not finally settled.

The homology relations between the phallomere elements of Blattaria and Mantodea on the one hand and the elements of the external genitalia of other Ectognathan taxa on the other are completely unclear. Only the earliest rudiments or primary phallic lobes can be reliably regarded as homologous.

The abdominal segments 10 and 11 and the telson

This morphologically posteriormost part of the body lies dorsal to the phallomere complex and covers it completely (Blattaria) or partly (Mantodea) (fig.320, 321a,b). For many sclerotisations of this region it is unclear whether they belong to abdominal segment 10 or 11 or to the telson, or to the segment 12 proposed by Matsuda (1976).

Description of morphology

The principal morphology of this area is in Blattaria and Mantodea always quite similar: Tergite 10 (**T10** in fig.320, 321a,b) is somewhat triangular by a more or less pronounced expansion of its median part to the posterior. Along the posterior edge of tergite 10 (**X** in fig.320, 321a,b) the cuticle bends sharply ventrad and anteriad, and the sclerotisation of **T10** often – and to a varied extent – follows this bend and forms the ventral sclerotisation of tergite 10 (**T10v** in fig.320, 321b). In some species tergite 10 is longitudinally divided along its midline by a stripe of membrane (a derived condition).

The cerci (E11 in fig.321a,b) are the appendages of abdominal segment 11. They have their bases laterally beneath the posterior edge of tergite 10. The basal article of each cercus has at its dorsal basal margin a distinct articulation with a node-like thickening on the posterior margin of tergite 10 (articulations A98 in fig.321b and e.g. fig.58). Median to the cercal bases there may be some further sclerotisations (three pairs at most; not shown in fig.321): The crescent-shaped Ca-sclerites (e.g. in fig.263) are close to the cercal bases and often lie upon distinct bulge-like evaginations. The Cb- and Cc-sclerites take more median positions (e.g. in fig.169, 170).

The anterolateral parts of tergite 10 curve ventrad and then mesad; these parts are the paratergites (T10p in fig.321b,c), which take a position posterolateral to the phallomere

complex. The ventromedian ends of the paratergites **T10p** are in most species distinctly articulated with the lateral ends of the paraprocts **Pp** (articulations **A99** in fig.321b,c).

Median to the paratergites **T10p** and beneath the **T10v**-sclerotisation there is on each side a transverse (or oblique) bulge, the subanal lobe (**sbl** in fig.321b,c). The paraprocts **Pp** are always present as one pair of sclerites (fig.321b,c). From their lateral ends at the articulations **A99** they extend mesad and sclerotise more or less extensive parts of the subanal lobes **sbl**. Laterally the paraprocts are restricted to the ventral **sbl**-walls, medially they curve more and more into their dorsal walls (fig.321b,c). Consequently, the posterior parts of the paraprocts are curved upwards and back anteriad, and the paraprocts have a ventral anterior margin and a dorsal anterior margin. (The latter will subsequently be designated as the posterior margin, which is true in a strict morphological sense). The median tips of the subanal lobes (**Y** in fig.321b,c) lie on both sides of the anus (**Af** in fig.321c) and are either membranous or also sclerotised by the paraprocts. The median walls of the subanal lobes continue anteriad into the lateral walls of the rectum (**Re** in fig.321b,c).

In front of the (ventral) anterior margins of the paraprocts there is often another pair of ribbon-shaped sclerites (Pv in fig.320, 321b,c). These Pv-sclerites are either completely free (like in fig.321) or connected with the paraprocts laterally. In some species separate Pv-sclerites are missing, and in these cases they seem to have fused to the anterior margins of the paraprocts.

The membrane anterior to the ventral sclerotisation of tergite 10 (**T10v** in fig.321b) is evaginated to form an unpaired supraanal lobe (**spl** in fig.320, 321b,c), whose ventral wall continues anteriad into the dorsal wall of the rectum (**Re** in fig.320, 321b,c). In Mantodea the supraanal lobe bears a sclerotisation in its dorsal wall, the epiproct (**Ep** in fig.320, 321b,c). In many Blattaria the supraanal lobe is still distinct but never has a sclerotisation. In other Blattaria the supraanal lobe is no longer distinct from other small membranous foldings in the anal region, and its presence is questionable.

Discussion of morphology

Concerning all these elements, only the assignment of the anterior part of tergite 10 (**T10** including **T10p**) to abdominal segment 10 and the consideration of the cerci as the appendages of segment 11 is generally accepted. Regarding the other elements there are various opinions. Snodgrass (1933, 1936, 1937) regards the cerci and the subanal and supraanal lobes as elements of segment 11, the paraprocts being the medially divided sternite and the epiproct being the tergite of segment 11. Sternite 10 is assumed to be missing, tergite 10 is the true tergite 10, with no other elements incorporated, and the telson is only a small membranous ring surrounding the anus. Chopard (1917), Walker (1922), and Ford (1923) differ from Snodgrass only in assuming a participation of sternite 10 within the anterior margins of the paraprocts. This is said to be indicated by the articulations between the paraprocts and the paratergites of segment 10 (**A99** in fig.321b,c) and Matsuda (1976) consider segment 11 as strongly reduced – the cerci being its only persisting products – and regard the subanal lobes and the paraprocts as well as the

supraanal lobe and the epiproct as elements of the telson (Heymons) or of a twelfth segment (Matsuda; the only difference to Heymons is that Matsuda regards this posteriormost part of the body as a segment). Matsuda regards tergite 10 of Mantodea as a proper one, but tergite 10 of Blattaria is supposed to contain the epiproct.

These differences in the interpretation of the terminal elements are accompanied by a confused situation in the terminology for these structures. This concerns the usage of e.g. the terms subanal lobe, supraanal lobe, epiproct, paraproct, tergite 11, sternite 11, and telson. The comparison of the results of the various authors is thus rather difficult. For a correct interpretation of the elements concerned some clarifying investigations of ontogeny and morphology would be necessary. To do this is not the purpose of this paper, and the terminology for the respective elements subsequently practised is a descriptive one, not the morphologically correct one.

These controversial opinions, however, have to be discussed as far as homology relations within Dictyoptera are involved. This concerns the elements called tergite 10 T10 and supraanal lobe spl in the above description (fig.320, 321), which are, according to Matsuda (1976), both not homologous in Mantodea on the one hand and in Blattaria and Isoptera on the other. (This difference is the same for females). Matsuda's opinion is as follows: Blattaria and Isoptera show in their ontogeny a very early differentiation of a supraanal lobe (meaning of supraanal lobe here: the dorsal part of the embryonic telson - Heymons - or segment 12 - Matsuda; not the structure called **spl**-lobe above!). By the extensive reduction of segment 11 during embryonic development this supraanal lobe comes into a position immediately behind abdominal tergum 10. The dorsal segmental border between supraanal lobe and tergum 10 then vanishes and these regions become fused. Thus, in the imago the sclerite called "tergite 10" T10 above is regarded as a compound sclerite containing the true tergite 10 and the epiproct (the latter considered as the tergite of the telson or of the twelfth segment, respectively). In Mantodea, however, the differentiation of this supraanal lobe is delayed until postembryonic development. No fusion of supraanal lobe and tergum 10 takes place. Thus, in the Mantodean imago "tergite 10" T10 is the true tergite 10, and the epiproct $\mathbf{E}\mathbf{p}$ is still situated behind it as a separate sclerite on an independent supraanal lobe.

If this is true, the element I call supraanal lobe **spl** (fig.321b) would be: (1) the supraanal lobe sensu Heymons and Matsuda in Mantodea (dorsum of telson or segment 12); (2) a posterior part of the supraanal lobe sensu Heymons and Matsuda in Blattaria / Isoptera (a lobe-like posterior part of the dorsum of the telson or segment 12). The condition in Mantodea, if not regarded as a neotenic trait, would be more primitive than the situation in Blattaria and Isoptera.

Matsuda (1976) refers to the results of earlier workers: Heymons (1895), Wheeler (1889), and Cholodkowsky (1891) for Blattaria; Graber (1890), Hagan (1917), Görg (1959), and Walker (1919, 1922) for Mantodea. From the data contained in these papers the following view results:

- Looking at the descriptions in Heymons, the fusion between tergum 10 and the dorsal part of the telson (or segment 12) really seems to take place in Blattaria.

- In Graber, Hagan, and Görg, however, I could not find any observation contradicting the same fusion in Mantodea: None of these authors treats the development of the region concerned in sufficient detail.
- Matsuda agrees with Walker and also Snodgrass (1933), p.73, and (1936), p.42, about Mantodea: Supraanal lobe and tergite 10 are not fused, and tergite 10 of the adults is a proper one. The two latter authors (the only ones from whom Matsuda could have adopted his assumption for Mantodea), however, make the same assumption for Blattaria, too. They regard as I did in the above description the membranous lobe (spl in fig.320, 321b) of Blattaria as homologous with the spl-lobe of Mantodea. Thus, the opinions of Walker and Snodgrass for Mantodea cannot in a comparison with the results of Heymons for Blattaria serve to state a difference between Blattaria / Isoptera and Mantodea.
- Accepting Heymons' results, in Blattaria the supraanal lobe sensu Walker and Snodgrass (= spl-lobe in my terminology) is posterior to or a posterior part of the supraanal lobe sensu Heymons. According to Matsuda, in Mantodea the former (spl-)lobe is differentiated in a postembryonic stage. Such a late elaboration of the spl-lobe is possibly also true of Blattaria (and Isoptera?); at least, to my knowledge, an embryonic rudiment of this lobe is not mentioned in the literature.
- Thus, no argument comes from the data used by Matsuda to contradict the homology of the spl-lobes of Blattaria and Mantodea. The assumption of a difference between Blattaria and Mantodea is based upon a comparison of non-comparable data.

Hence, the elements I call supraanal lobe **spl** and tergite of segment 10 **T10** might both be regarded as homologous in Blattaria and Mantodea – whatever structures these may be in a strict morphological sense. Moreover, there are some arguments supporting these homologies: (1) The supraanal lobe of Mantodea and the membranous lobe found in many Blattaria (**spl**) show exactly the same relations to surrounding elements – namely those shown in fig.320, 321b,c. (2) My own investigations of the musculature of the respective region in *Sphodromantis* (Mantodea), *Lamproblatta, Eurycotis*, and *Cryptocercus* (Blattaria) had the result that muscle insertions are present neither on the lobe of *Sphodromantis* nor on that of the Blattarian species, and the relations of these lobes to the surrounding muscles are the same in both groups. (3) Investigations in the same species show that the tergites 10 are provided with the same sets of muscle insertions. An unpaired muscle running from the posterior part of tergite 10 to the rectum (present in all these species) could be the musculus epiprocto-analis (Weidner 1982). The position of its dorsal insertion might support the view that the true epiproct has been incorporated into tergite 10 in both Blattaria and Mantodea.

3.2. The musculature

Most muscles are compact, and the insertion areas are well-defined. Some others, however, are rather diffuse, and it is not possible to exactly demarcate their insertion areas. (Such a diffuse condition will be mentioned in the muscle lists in chapter 5.). The data given in the figures must be considered with these reservations.

According to their morphological arrangement, the following groups of muscles can be distinguished:

Phallomere muscles: Intrinsic muscles of the phallomere complex. All muscles of this group will be studied. Three subgroups will be distinguished:

a) Intrinsic muscles of the left complex.

b) Intrinsic muscles of the right phallomere.

c) Muscles connecting the left complex and the right phallomere.

Phallomero-sternal muscles: Muscles connecting parts of the phallomere complex or the ventral and lateral walls of the genital pouch with the subgenital plate. All muscles of this group will be studied.

Ventral muscles: Muscles connecting successive sternites (mainly the respective anterior margins). These muscles are, compared with the more anterior segments, quite reduced in the postabdomen. Only the muscles belonging to abdominal segment 9 (running from sternite 9 to the – possibly obsolete – sternite 10) will be studied.

Dorsal muscles: Muscles connecting successive tergites. In some species lateral parts of the dorsal muscles of segment 9 have shifted in a way that they can hardly be recognised as dorsal muscles but seem to be muscles from the tergite to the phallomere complex. Only these parts of the dorsal muscles will be studied.

Dorsoventral muscles: Muscles connecting tergite and sternite of the same segment. If there really are vestiges of appendages contained in the subgenital plate, some of the muscles included here might be muscles from the tergite to the appendages. These muscles will be considered only in part.

Rectal muscles: Muscles from the ectodermal rectum to e.g. the anterior margin of the subgenital plate, the tergite 10, or the paraprocts, which function as dilators or suspensors of the rectum. These have in most cases clearly demarcated insertion areas on the respective parts of the exoskeleton, but the fibers diverge like a fan on their way to the rectum, and the rectal insertions are composed of many small insertion areas, which are often widely separated from each other. Only those muscles inserting on the subgenital plate will be considered.

Muscles of the ejaculatory duct: The ejaculatory duct is covered by a mat of fibers showing a ring-like, spiral, or longitudinal arrangement. This musculature will not be investigated in detail.

4. TERMINOLOGIES AND ABBREVIATIONS

4.1. The terminology for the sclerites of the phallomere complex

Mantodea

The most elaborate terminology is that of LaGreca (1954). It is the only one that is based on quite detailed investigations of phallomere morphology and that has already been applied to a broader sample of species. However, some disadvantage lies in the fact that LaGreca uses some names for sclerites as well as for the formative elements of the cuticle

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(e.g. processes or pouches) on or in which these sclerites are situated. For example, lamina ventrale (= lv) designates the sclerite I call L2 and, at the same time, the pouch lve, which contains sclerite L2 (fig.321g). This ambiguity makes LaGreca's terminology rather impractical. The other terminologies put forward (e.g. Beier 1968) are not very handy because of their long terms, and they are by far not detailed enough for my purposes.

Blattaria

McKittrick (1964) has developed a new, very simple and handy terminology, which has been adopted by most of the subsequent workers. Mizukubo & Hirashima (1987) also employ it but propose some changes. In both terminologies, the names for sclerites are short sequences of letters and numbers, each position containing certain information. Some of these terms have already been applied in the description of the phallomeres in 3.1., e.g. **L2**, **R3**.

McKittrick regards the phallomere complex of *Cryptocercus punctulatus* as the most primitive and takes it as the reference type for her terminology. She adopts the tripartition of Snodgrass (1936) into left, right and ventral phallomeres, and according to this major division McKittrick basically distinguishes left, right and ventral sclerites, which get L, **R**, or **V**, respectively, in the first position of their names. Then, on the left and on the right phallomere, the sclerites are numbered separately. The ventral phallomere has only one sclerite. In this way seven main sclerites are distinguished (L1, L2, L3, V, R1, R2, R3). No assumptions concerning side-homologies are intended in this terminology. What McKittrick – starting fom this situation in *Cryptocercus* – regards as a product of a secondary "subdivision" or as a special region or "elaboration" of a main sclerite is expressed by the addition of one or two small letters (**d** = dorsal, **v** = ventral, **l** = lateral, **m** = median, **vm** = ventromedian). Sclerites of certain species regarded as completely new elements not present in *Cryptocercus* are given the next free number of the respective phallomere. This terminology is very handy and clear and contains a lot of information.

Mizukubo & Hirashima state side-homologies for the elements of the left and of the right half of the phallomere complex and integrate these assumptions in their terminology. For that purpose, they modify the terminology of McKittrick in two ways:

(1) According to the assumption of a plane of symmetry, they basically distinguish right and left elements (**R** or **L** in first position); then both **R** and **L** are grouped into dorsal and ventral elements (**D** or **V** in second position). The left-dorsal elements **LD** and the left-ventral elements **LV** compose the left phallomere. The right-ventral elements **RV** correspond to the ventral phallomere (**vla**-lobe in my terminology). The right-dorsal elements **RD** correspond to the right phallomere. Thus, the basic division into **LD**, **LV**, **RV**, and **RD** essentially conforms with the division of the phallomere-complex proposed by Quadri (1940).

(2) As regards the numbers and small letters, Mizukubo & Hirashima adopt the terminology of McKittrick, but changes are made in order to get side-homologous elements provided with the same names – except for \mathbf{R} or \mathbf{L} in the first position. These changes are, compared with McKittrick, not very extensive.

The terminology for Blattaria and Mantodea used in this paper

The terminology of Mizukubo & Hirashima will not be employed since I do not agree with the assumptions of these authors (discussion in chapter 8.). I will use a modified version of McKittrick's terminology and apply it to both Blattaria and Mantodea. There are three reasons why the terminology of McKittrick is not adopted unchanged, and I will proceed in the following way:

1. Reason: The tripartition in McKittrick's terminology (L, V, R) reflects the earlier view of Snodgrass (1936) that the ventral phallomere is a medioventral basic element of the phallomere complex. In my view the ventral phallomere = ventral lobe vla is a ventral part of the left half of the phallomere complex (= left complex).

1. Procedure: All sclerites of the left complex will get L, all sclerites of the right phallomere will get R in the first position of their names.

2. Reason: Like Mizukubo & Hirashima (1987), I cannot accept the view of McKittrick that the phallomeres of *Cryptocercus* are closest to the primitive Blattarian type and should be used as a reference type. I have taken the common ground-plan of the phallomeres of Blattaria and Mantodea (compare in 3.1.) as the basis of my terminology. The ground-plan pattern I assume for the phallomere sclerites is rather different from that proposed by McKittrick (compare fig.321e-i and McKittrick 1964, fig.106).

2. Procedure: Each sclerotisation that is assumed to be present as one isolated and undivided sclerite in its most primitive condition within the taxon comprising all Blattaria and Mantodea and their last common stem-species is designated as a main sclerite. Hence, these main sclerites can be (1) sclerites of the common ground-plan of Blattaria and Mantodea or (2) sclerites formed de novo (not by the division of sclerites already present before) in any subgroup of Blattaria or Mantodea. Each main sclerite will get its own number in the second position of its name. In the description in 3.1. these are the sclerites L1, L2, L3, L4, L5, R1, R2, and R3. Numbering is arbitrary. If any of these main sclerites becomes divided, the whole of its descendants is called sclerite group L1, L2, etc..

Unfortunately, for many sclerotisations the most primitive condition and the evolution are not completely clear, and there is in many cases, and to various extents, some uncertainty about whether a certain sclerotisation fulfils the definition of a main sclerite. As regards the sclerotisations shown in fig.321, L1, R1, and R3 are assumed to be isolated and undivided sclerites in the common ground-plan of Blattaria and Mantodea (fig.321e-i). L3, L5, and R2 are also isolated and undivided in their most primitive states, but they are possibly not yet present in this ground-plan; however, if a later origin is really true for them, they can be at least assumed to be new sclerotisations, not split off descendants of ground-plan sclerites. To regard L2 and L4 as two main sclerites is somewhat

subjective: (1) L2 and L4 are primitively connected in between the processes **paa** and **pda** (fig.321e,g); the interspace between **paa** and **pda** is here defined as the border between L2 and L4. (2) For L4 it is not clear whether it has been present as one, two, or three sclerites in the ground-plan (the latter alternative is shown in fig.321e,g,i). Apart from these early evolved sclerites, there are several main sclerites which are undoubtedly apomorphic for certain subgroups (L6..., R4...).

3. Reason: McKittrick names certain parts of her main sclerites with small letters in the third position, no matter whether these parts are (1) products of a complete division or (2) only certain regions of a main sclerite. However, these two situations represent two different aspects of a sclerotisation and its evolution: (1) On the one hand, a main sclerite is composed of one or more separate individual sclerites. Since divisions or fusions of sclerites can take place, the state of subdivision of a main sclerite is subjected to evolutionary changes. (2) On the other hand, a main sclerite, irrespective of its state of subdivision, consists of several regions each of which is characterised by taking a certain position, by having a certain shape, by occupying certain in- or evaginations of the cuticle, or by bearing certain muscle insertions or articulations. The properties of these regions undergo evolutionary changes, too. The special state of subdivision (1) and the special properties of the regions (2) of a main sclerite are largely independent of each other, and evolution works on both these aspects and has to be considered from both viewpoints.

3. Procedure: I strictly separate these two aspects (1) and (2) in my terminology, and for the designation of different parts of main sclerites two terminologies completely independent of each other will be used:

The **first terminology** serves to designate **individual sclerites** having originated by a division of a main sclerite: If any main sclerite is divided into two or more sclerites completely separated from each other by membrane, each of the sclerites will get one **capital letter** in the third position of its name (e.g. **L2D**, **R1C**). (The main sclerite has become a sclerite group. An individual sclerite as defined here may, however, be connected with parts of another main sclerite). If any of these individual sclerites undergoes a further division or a fusion, all resulting sclerites involved in this process will get a new capital letter. Equal designation of sclerites of different species means the assumption of homology. Different designation, however, does not always mean complete non-homology: Some small sclerites of one species can as a whole be homologous with one large sclerite of another species, and none of the sclerites will have the same letter. In the third position of these terms, I, O, and Q will not be used (danger of confusion with "1" and "0" in the figures); R will not be used on the right side, L on the left side. Among other things, this terminology serves for a clear reference between the text and the figures.

The meaning of a term designating a certain individual sclerite, e.g. L2D, is hence as follows:

L	2	D
a sclerotisation	belonging to	being the indi-
of the Left side,	main sclerite 2,	vidual sclerite D

The second terminology serves to designate certain regions of the main sclerites or sclerite groups: This regioning is essentially independent of any natural sub-division of the respective sclerotisations. To name these regions one small letter will be added in the third position (e.g. L2d, R1c). Equal designation of sclerotisations of different species means the assumption of homology. Different designation means complete non-homology. These names mainly serve for the demarcation of homologous areas on the main sclerites / sclerite groups of different species and for a description of their evolution as regards the properties listed above in **3. problem** (2). Therefore, this dividing into regions, or

regioning, will be performed with practicability as the main point of interest (the best possible way to explain homology relations). It is in general arbitrary. (The definition of the regions, however, will in some cases be done in correspondence with a concrete division of the respective main sclerite into smaller sclerites in a certain, arbitrarily selected species). Only the more complicated main sclerites will be divided into regions. The small letters are in most cases abbreviations of typical attitudes (e.g. position) of the regions. These abbreviations and the definition of the various regions will be given in the homology discussion of the respective main sclerites (chapter 6.).

The meaning of a term designating a certain sclerite region, e.g. R1c, is hence as follows:R1ca sclerotisation ofbelonging tobeing the region c, which

main sclerite 1.

the **R**ight side,

In Blattellidae Plectopterinae, some other Blattellidae, and Blaberidae the phallomere complex is side-reversed (Bohn 1987), being essentially a mirror-image of the phallomere complex of the other Blattarian subgroups. In the terminology for the sclerites, this fact will be taken into account by adding ' at the end of the term. This will be done in the terms for individual sclerites as well as in the terms for sclerite regions. (For example, L4U' is a L4U on a side-reversed phallomere complex, and it is on the right side). This procedure is different from Bohn (1987), who adds ' after the first letter (L4U' would be L'4U), but the meaning is the same. (Of course, since homology should be the basis of the terminology, the left complex will still have this name, if it is, after a reversal of the phallomere complex, on the right side of the body; the same practice will be applied to the right phallomere.)

4.2. Abbreviations for other sclerites of the postabdomen

The terminology for the sclerites of the postabdomen is largely pre-set by the earlier literature. Abbreviating is done according to the same principle as in the phallomere sclerites. A capital letter in the first position designates the category (\mathbf{S} = sternite; \mathbf{T} = tergite; \mathbf{E} = sclerotisation of an appendage = extremity). A number in the second position designates the abdominal segment the sclerite belongs to. A small letter in the third position (not obligatory) serves to designate a special region. As discussed in 3.1., there are many problems concerning the correct morphological assignment and designation of postabdominal sclerotisations. In such problematical cases neutral abbreviations will be used (e.g. \mathbf{Pp} = paraproct; \mathbf{Ep} = epiproct; all elements of the subgenital plate are called $\mathbf{S9}$ despite the possibility of true appendages being involved).

4.3. The terminology for the formative elements

The phallomere complex contains many areas where the cuticular body wall forms pouches, apodemes, tendons, hook-, spine-, tooth-, or lobe-like processes, or comparable

takes a rather central position

structures. These more or less distinct invaginations (or infoldings) and evaginations (or outfoldings) of the cuticle are responsible for the shape of the phallomere complex, and I will designate them as the formative elements. These will receive special names serving for a clear and practicable reference and for a clear presentation of homology relations. The terms are composed of three small letters. The first two letters are the individual name of the respective formative element, the third letter stands for one of the two possible categories (\mathbf{a} = evagination, German "Ausstülpung"; \mathbf{e} = invagination, German "Einstülpung"). A certain area of the cuticle can belong to two or more neighboring formative elements, and, in such a case, the respective terms "overlap" each other. For example, a certain area of the cuticle can be e.g. the dorsal wall of a specific evagination and the ventral wall of an immediately adjacent invagination at the same time, or a large evagination can fork into smaller ones in its distal part.

The first two letters are essentially selected arbitrarily but are mostly abbreviations for characteristic attitudes (e.g. vla = ventral lobe, evagination). In other cases the abbreviations of the terms of LaGreca (1954) have been used (e.g. lve = lamina ventrale, invagination; LaGreca's name for sclerite L2 and for the pouch containing L2; compare fig.321g). The formative elements of the phallomere complex termed in this paper are listed in 4.7.

Similar terms are used for formative elements outside the phallomere complex (e.g. **sbl** = subanal lobe), but the letter in third position is part of the abbreviation and does not stand for a category.

4.4. The terminology for the muscles

Muscles are named by a small letter determining the category, followed by a number for the individual muscle. Numbering is arbitrary. The categories are as follows:

- IIntrinsic muscles of the left complex(left)rIntrinsic muscles of the right phallomere(right)bMuscles connecting the left complex and the right phallomere(between)sPhallomero-sternal muscles(sternal)
- **p** Muscles not inserted on the phallomere complex but taken

into account (some ventral, dorsal, dorsoventral, and rectal muscles) (peripheral)

In some cases it can be reliably assumed that a muscle has divided. In other cases homology can be assumed for groups of muscles of different species, without the possibility to ascertain the homology relations for the single muscles of the group. In both these situations all muscles of the group get the same number, and the single muscles are distinguished by adding a small letter in the third position. In these cases equal letters do not mean homology of the respective muscles by principle; which homologies are assumed will be explicitly mentioned in the text.

For the muscles of the categories l and r it is not clear which of them have originally been pairs or if there are pairs at all. The muscles of category **b** are probably unpaired transverse muscles. The phallomero-sternal muscles **s** will be separately numbered on the left and on the right side, though pairs can be identified in many cases; those inserting on the left half of the subgenital plate are given odd numbers, those inserting on the right half are given even numbers. (In species having side-reversed phallomeres the reverse is true). Only the "peripheral" muscles \mathbf{p} will be designated pairwise.

4.5. The terminology for the articulations

Articulations between sclerites are named by A ("Articulation") and an added number (e.g. **A5**, **A98**). The numbers 1-79 are reserved for the articulations within the phallomere complex, the numbers 80-99 for the articulations outside the phallomere complex. Numbering is arbitrary. The term articulation is taken in its widest sense: each case of close contact between two sclerites. Even if the contact has become looser by evolutionary changes, the remaining vicinity of the sclerites will in many cases still be designated as an articulation (in order to designate the assumed homology of the respective vicinity). Places where articulations have been lost by a fusion of the sclerites concerned are often given the name of the respective articulation, but * is added (e.g. **A99***).

4.6. Survey of the terminologies used

	Main sclerites / sclerite groups of the phallomere complex:				
L	2				
Capital letter	Number				
(side: L, R)	(main sclerite)				
Regions of the main sclerites	s / sclerite groups of the phall	omere complex:			
L	2	d			
Capital letter	Number	Small letter			
(side: L, R)	(main sclerite)	(specific region)			
Individual sclerites of the ma	ain sclerites / sclerite groups o	of the phallomere complex:			
L	2	D			
Capital letter	Number	Capital letter			
(side: L, R)	(main sclerite)	(individual sclerite)			
Postabdominal sclerites outsi	de the phallomere complex:				
S	9	d			
Capital letter	Number	Small letter			
(category: S, T, E)	(abdominal segment)	(specific region)			
Formative elements of the phallomere complex:					
р	n	e			
Small letter	Small letter	Small letter			
(part of individual name)	(part of individual name)	(category: a, e)			
Muscles:					
r	4	b			
Small letter	Number	Small letter			
(category: l, r, b, s, p)	(individual muscle)	(separate part of the muscle)			

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Articulations:	
А	6
Capital letter A	Number
(articulation)	(individual articulation)

4.7. Abbreviations in the figures 1-321

CL = Capital letter; SL = Small letter; NR = Number; facultative parts of the terms are put in brackets; the abbreviations for the formative elements are listed separately.

A + NR	Articulation
Af	Anus
\mathbf{b} + NR (+ SL)	Muscle between left complex and right phallomere (or part of it)
Bl	Basal line
D	Ejaculatory duct
Dw	Dorsal wall of genital pouch
\mathbf{E} + NR (+ SL)	Sclerotisation of an appendage (or part of it)
E11	Sclerotisation of cerci
Ер	Epiproct
L + NR	Main sclerite / sclerite group of left complex
L + NR + CL	Separate sclerite of main sclerite / sclerite group of left complex
L + NR + SL	Region of main sclerite / sclerite group of left complex
l + NR (+ SL)	Muscle of left complex (or part of it)
Μ	Membranous or weaker sclerotised field of subgenital plate
Р	Phallomere-gland
p + NR (+ SL)	Muscles not inserted on phallomere complex
Рр	Paraproct
Pv	Ribbon-like sclerotisation anterior to paraproct
$\mathbf{R} + \mathbf{NR}$	Main sclerite / sclerite group of right phallomere
\mathbf{R} + NR + CL	Separate sclerite of main sclerite / sclerite group of right phallomere
$\mathbf{R} + \mathbf{NR} + \mathbf{SL}$	Region of main sclerite / sclerite group of right phallomere
$\mathbf{r} + \mathbf{NR} (+ \mathbf{SL})$	Muscle of right phallomere (or part of it)
Re	Rectum
S + NR (+ SL)	(Coxo-) Sternite (or part of it)
S9a	Apophysis of subgenital plate
S9d	Dorsal sclerotisation of subgenital plate
S91	Anterolateral part of subgenital plate
S9s	Stylus on subgenital plate
s + NR (+ SL)	Phallomero-sternal muscle (or part of it)
sbl	Subanal lobe
spl	Supraanal lobe
Sw	Lateral wall of genital pouch
$\mathbf{T} + \mathbf{NR} (+ \mathbf{SL})$	Tergite (or part of it)
Т9р	Paratergite of abdominal segment 9

Т10р	Paratergite of abdominal segment 10
T10v	Ventral sclerotisation of abdominal tergite 10
U	Pleura between abdominal tergite 9 and sternite 9
Vw	Ventral wall of genital pouch
X	Posterior edge of abdominal tergite 10
Y	Median part of subanal lobe

Formative elements of the phallomere complex

Column 2: LG = term derived from LaGreca (1954). Column 3: left = left complex; right = right phallomere. Column 4: If the formative element is sclerotised entirely or partly, the respective sclerotisation is given; if it is membranous, memb is listed; if both situations occur in different species, the more primitive situation is listed first.

Term	Meaning of term	Position	Sclerotisation	Shape
afa	a pofisi f alloide = af (LG)	left	memb, L1m	lobe, process
age	anterior groove	right	R3	fold/beam-like
	-			apodeme
are	apodeme, right	right	R3	fold-like apodeme
ate	anterior tendon	left	memb, L4n?	tendon
boe	bulb, opposite	left	L2d	hood-shaped apodeme
cbe	central bulge	right	R1t	large, shallow
	-	-		invagination
cla	central lobe	left	memb	lobe
cwe	cleft "Wulst"	right	R1t	curved cuticular
				thickening
dca	dorsal cushion	left	memb, L1	1-more cushions,
				processes
dla	dorsal lobe	right	memb, R4	lobe
dte	dorsal trough	left	L2d + L4l	shallow invagination
fda	fallomero dorsale	right	R1	lobe
	di destra = fd (LG)			
fpe	fence, posterior	left	memb	fold-like invagination
goa	genital opening	left	memb	1-2 cushions, lobes
gta	-	left	L2p	process
hge	hook groove	left	L3	groove
hla	hook, left	left	L3	hook
ipe	inter, phallomere-	left	memb	fold-like invagination
	glands			
lba	lobe, between	left	L7	lobe
loa	lobo membranoso =	left	L1m	slender process
	lo (LG)			
lve	lamina ventrale = $lv (LG)$	left	L2	flat pouch
mla	median lobe	left	memb	lobe
nla	node, left	left	L4n	node, bulge
paa	p rocesso a picale = pa (LG)	left	L2d	process
	-			

	manale leature at	left		flat manual
pbe	pouch, between		memb	flat pouch
pda	processo distale = pd (LG)	left	L4l	process
pia	piastra ventrale = pi (LG)	right	R1v	tooth, ridge
pne	p rocesso anteriore =	left	Lla	pouch
	pn (LG)			
pra	prong, right	right	R1d	process
psa	posterior sting	left	L2m	bifid process
pva	processo ventrale	right	R1t	tooth, ridge
	sclerificato = pv (LG)			
rge	right groove	right	R1c	groove
sbe	sting bulb	left	L4l	bulb + channel
sla	sting, left	left	L4d	sting-like process
sra	sting, right	right	R1d	sting-like process
swe	sclerotised "Wulst"	left	L4l	fold/beam-like
				apodeme
tre	tendon, right	right	memb	long, narrow tendon
tve	tendon, virga	left	L4l	tendon, apodeme
vfa	ventral fold	left	memb	broad lobe
vge	virga groove	left	L4l	groove
via	virga	left	L2d + L4l	process
vla	ventral lobe	left	L4v	lobe
vpe	ventral pouch	left	memb	broad, flat pouch
vsa	ventral sting	left	L4l	sting-like process
vte	ventral tendon	left	memb	broad, flat tendon
xla		left	L2m	short process

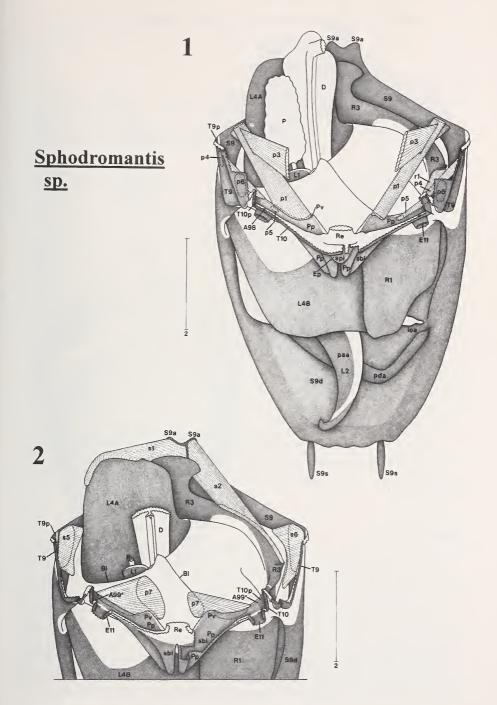
5. DESCRIPTIONS OF PHALLOMERES AND POSTABDOMINA

The complicated morphology of the phallomere complex and the courses of all investigated muscles will be described in detail. The morphology of the subgenital plate and of the posteriormost part of the abdomen can be largely seen from the figures and from the general descriptions in 3.1.. Only the condition of the following elements will be mentioned: **Pv**-sclerites; sclerites **Ca**, **Cb**, and **Cc** median to the cercal base; ventral sclerotisation of tergite 10 **T10v**; articulations **A98** (cercal base – posterior margin of tergite 10) and **A99** (paratergite 10 – paraproct); some peculiarities will also be pointed out.

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Figs.1,2: *Sphodromantis* sp. (Mantodea, Mantidae) – 1: Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, supraanal lobe, epiproct, subanal lobes, paraprocts, distal part of rectum, basal parts of cerci, and part of musculature. – 2: Same as in fig.1, after removal of further parts of abdominal tergites 9 and 10, parts of right paraproct, and supraanal lobe with epiproct. Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. Posterior to transverse line: like in fig.1. – Scale: 2mm.





5.1. Sphodromantis sp. (Mantodea, Mantidae)

Left complex

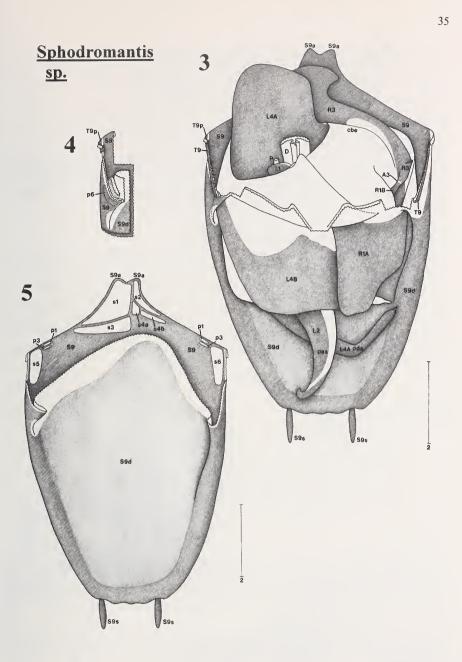
The dorsal wall is largely occupied by sclerite L4B (fig.9), the ventral wall by the large plate-like sclerite L4A (fig.6, 9-11). L4A extends posteriorly onto the process pda (fig.6, 12). On the left edge of the left complex L4A and L4B articulate (A1 in fig.6, 10). L4B has a solid keel-like apodeme (swe in fig.10; cross-section in fig.11) at its anterior left margin.

From beneath the posterior and right margins of L4B, two deep pouches invaginate to the left and anteriad and lie one above the other (pne and lve in fig.10, 11). The dorsal pouch pne (fig.10) is largely occupied by sclerite L1A (fig.10), which lies mainly in the ventral wall but bends anteriorly into the dorsal wall around the left edge of the pouch. The phallomere-gland (P in fig.10) opens into the membranous anteriormost ventral wall of the pne-pouch. The ventral pouch lve (fig.10-12) deepens extensively to the left. (The edge along the bottom of the lve-pouch is labelled 7 in fig.11). Its dorsal wall is occupied by sclerite L2. The ejaculatory duct (D in fig.11, 12) opens into its membranous ventral wall. The edge between the pouches **pne** and **lve** (1 in fig.10, 11) bears three processes: most anteriorly a hammer-shaped one (afa in fig.9-11); far behind this a long and narrow lobe (loa in fig.10); immediately behind the latter a stout hook (paa in fig.10). The anterior part of this edge 1 is occupied by sclerite L1B (fig.10, 11), which is separated from both L1A and L2 by narrow stripes of membrane (2 in fig.10, A2 in fig.11). L1B also sclerotises the afa-process and has a strip-like posterior extension into the dorsal wall of the loaprocess. The posterior part of sclerite L2 extends onto the paa-process (fig.11) and occupies most of its surface.

The membranous ventral wall of the **lve**-pouch with the genital opening is at the same time the dorsal wall of the large ventral lobe **vla** (= ventral phallomere; fig.10-12). Close to the genital opening there are two small membranous lobes (**goa** in fig.12). The ventral wall of the **vla**-lobe is part of the ventral wall of the left complex and is largely sclerotised by **L4A** (fig.6).

Right phallomere

The anterior ventral wall is occupied by sclerite **R3** (fig.6, 13): Its left part is expanded, its narrow right part curves posteriad and articulates (**A3** in fig.6, 8, 14) with the posteroventral sclerite **R1B**. The ventral wall behind the left and central parts of **R3** is broadly invaginated dorsad and anteriad (**cbe** in fig.13). The ventral wall behind the right part of **R3** bears two teeth, which are both sclerotised by **R1B**: the anterior **pva** and the posterior **pia** (fig.6, 14). **pia** and **pva** are both on the crest of a leftward projecting evagination of the ventral wall (fig.8, 14). The posterior part of the right phallomere is a large lobe (**fda** in fig.6, 13), with its dorsal wall completely occupied by sclerite **R1A**. Around the anterior right edge of the right phallomere **R1A** curves into the ventral wall, where it is connected with **R1B** by a stripe of very weak sclerotisation (**4** in fig.6, 14).



Figs.3-5: *Sphodromantis* sp. (Mantodea, Mantidae) – 3: Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – 4: Left margin of subgenital plate (compare fig.3); with insertion of muscle p6. – 5: Subgenital plate in dorsal view; with insertion areas of muscles (except p6). – Scale: 2mm.

The whole anterior and right margins of **R3** form a groove-like apodeme **age** (fig.6; crosssection in fig.8, 14). The cuticle along the bottom of the **age**-groove is thickened. In the left part of **R3** the **age**-groove abruptly deepens and, in addition, curves posteriad and finally back to the right (fig.6, 8, 13, 14). By this deepening and curving the left part of the **age**-apodeme is a crescent-shaped plate, which is sclerotised dorsally and ventrally (fig.6-8, 13, 14). The ventral wall of the genital pouch becomes evaginated by the posterior part of the crescent and forms a membranous pouch (**5** in fig.6, 7, 8, 13, 14). In the right posterior part of **R3** the **age**-apodeme bears an additional apodeme fold **are** (fig.6; crosssection in fig.8), which bears another small keel-like apodeme (**3** in fig.6, 13). The groovelike shape of the sclerotisation extends posteriad beyond articulation **A3** (where **R3** ends and the groove is no longer called **age**) onto sclerite **R1B** (fig.6, 8, 13).

Subgenital plate and posterior abdominal segments

Fig.1,2 (posterior segments); fig.5 (subgenital plate **S9**). The paraprocts **Pp** and the epiproct **Ep** are just weakly sclerotised and do not have distinct boundaries to the surrounding membranes; it is therefore not possible to determine the exact outlines of these sclerotisations, the presence of articulations **A99** (**A99*** in fig.2), and the presence of separate **Pv**-sclerites (compare fig.321b,c). The data given in fig.1, 2 must be considered with these reservations. A ventral sclerotisation of tergite 10 **T10v** was not found. The sclerites **Ca**, **Cb**, and **Cc** are missing (or strongly obsolete?). The articulations **A98** are well-developed.

Musculature

Muscles	Positions of insertions	in fig.
11	L1A (anteriorly on pne-pouch) – right anterior dorsal wall of left	
	complex	17
12	L1A (dorsally on pne-pouch) – L4B (right part)	15
13	L1A (posteroventrally on pne- pouch) – L2 (left part)	16
14	L2 (left-posterior part) – L4B (central part)	15
15	L4A (left-anterior part) – L2 (left-anterior part)	15

 $[\]longrightarrow$ p.37

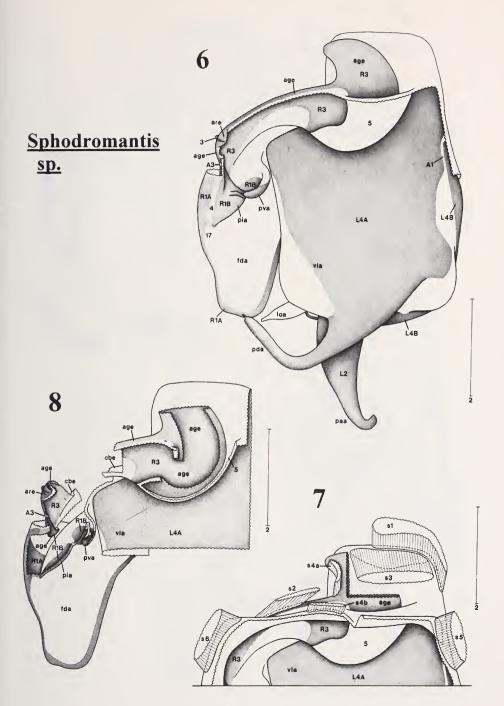
\longrightarrow p.38

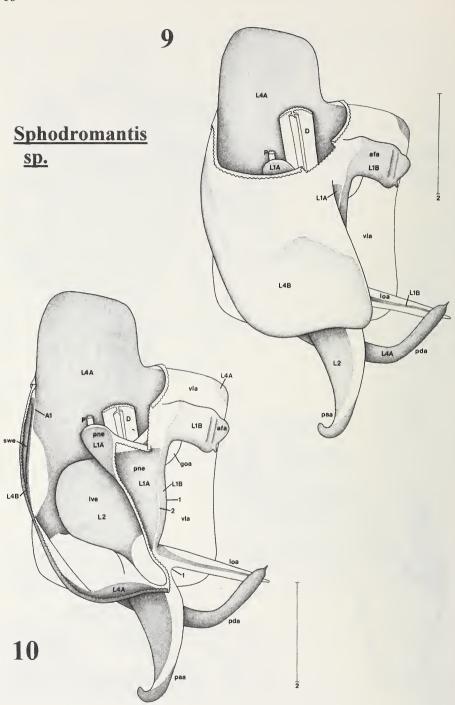
Figs.9,10: Sphodromantis sp. (Mantodea, Mantidae) – 9: Left complex in dorsal view. – 10: Left complex in dorsal view; some dorsal parts removed. – Scale: 2mm.

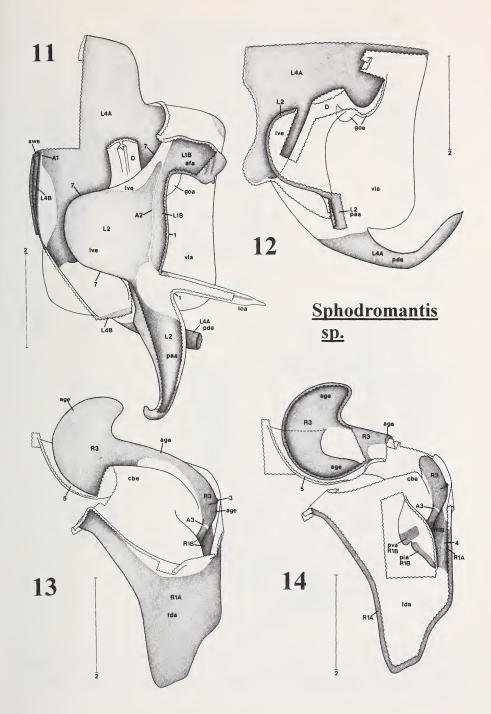
 \longrightarrow p.39

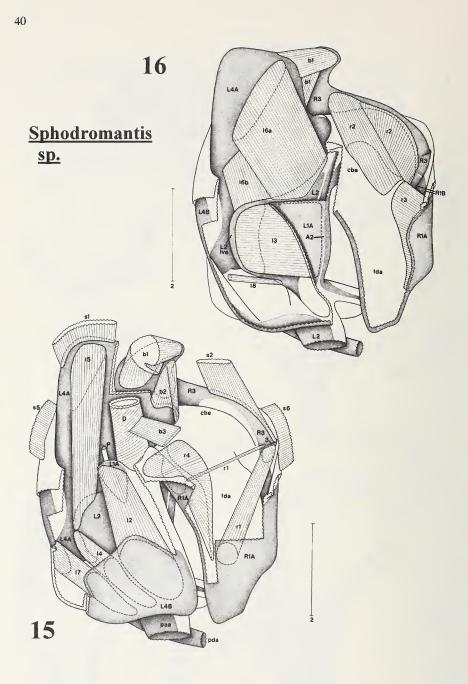
Figs.6-8: *Sphodromantis* sp. (Mantodea, Mantidae) – 6: Phallomere complex in ventral view. – 7: Anterior part of phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.6; left anterior part of sclerite R3 removed. – 8: Right phallomere with transition to left complex in ventral view; some ventral parts removed. – Scale: 2mm.

Figs.11-14: *Sphodromantis* sp. (Mantodea, Mantidae) – **11,12:** Left complex in dorsal view; with further removal of its parts (mainly of dorsal ones). – **13:** Right phallomere in dorsal view; – **14:** Right phallomere in dorsal view; some parts removed (mainly dorsal ones). – Scale: 2mm.

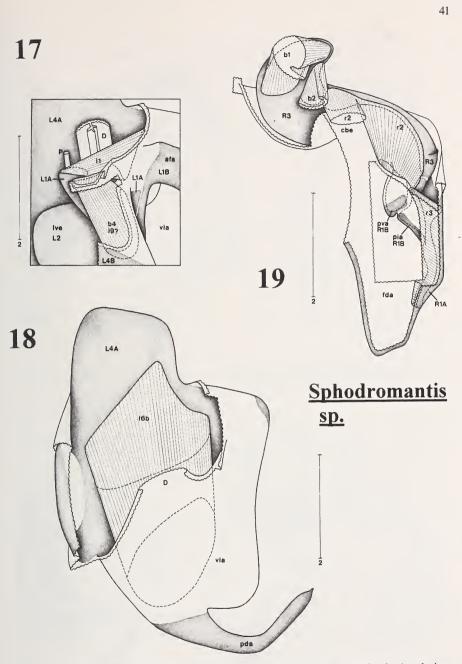








Figs.15,16: *Sphodromantis* sp. (Mantodea, Mantidae) – Left complex and right phallomere in dorsal view; each figure with some muscles; parts of phallomere complex removed to various extents. – Scale: 2mm.



Figs.17-19: *Sphodromantis* sp. (Mantodea, Mantidae) – 17: Detail of left complex in dorsal view (compare fig.10); with some muscles. – 18: Left complex in dorsal view; with muscle l6b; dorsal parts largely removed. – 19: Right phallomere in dorsal view; with some muscles; dorsal parts largely removed. – Scale: 2mm.

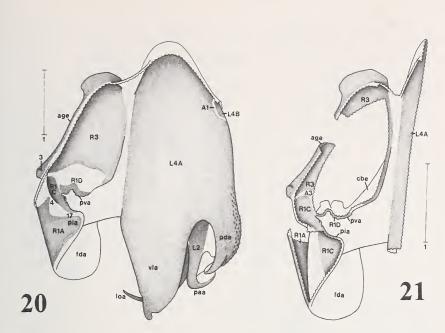
16a	L4A (anterior part) – L2 (right-anterior part)	16
16b	L4A (central part) – dorsal wall of vla-lobe behind genital opening	16, 18
17	L4A (left-posterior part) – L4B (left-posterior part)	15
18	Membrane behind left posterior end of L4A – L2 (posterior part);	
	only very few fibers; completely missing in some specimens	16
r1	R3 (right margin) – R1A (dorsal wall of fda-lobe)	1, 15
r2	R3 – R1B on pva-tooth and membrane of cbe-invagination	16, 19
r3 r4	R1A (right part) – R1B on pia -tooth and right ventral wall of fda -lobe R1A (left-anterior part) and membrane behind it – left ventral wall of	16, 19
	fda-lobe	15
b1	L4A (anterior margin) $-$ R3 (left anterior part)	15, 16, 19
b2	L4A (anterior margin) and vla-lobe (anteriormost part) – R3	
	(left-anterior part)	15, 19
b3	Ejaculatory duct \mathbf{D} – left ventral wall of right phallomere	15
b4 (19?)	Longitudinally within right dorsal wall of left complex	17
s1	S9 left side (medially and most anteriorly) – L4A (anterior margin)	2, 5, 7, 15
s2	S9 right side (medially and most anteriorly) $-$ R3 (lateral anterior	
	margin)	2, 5, 7, 15
s3	S9 left side (medially and anteriorly) – anteriormost left ventral wall	
	of genital pouch	5, 7
s4a	S9 right side (medially and anteriorly) $- R3$ (left ventral wall of	
	age-apodeme)	5, 7
s4b	S9 right side (medially and anteriorly) – anteriormost median ventral	
	wall of genital pouch	5, 7
s5	S9 left side (laterally and anteriorly) – anterior left wall of genital pouch	2, 5, 7, 15
s6	S9 right side (laterally and anteriorly) – anterior right wall of genital	
	pouch	2, 5, 7, 15
p1 (pair)	S9 – paraproct Pp (anterior margin, corresponding to Pv -sclerotisation)	1, 5
p3 (pair)	S9 – rectum (ventral wall); in most specimens p3 penetrates p1.	1, 5
p4 (pair)	T9 (lateralmost anterior margin, possibly corresponding to paratergite	
	T9p) – anterior margin of paratergite T10p	1
p5 (pair)	T10 (lateralmost anterior margin) – paraproct Pp (lateral anterior	
	margin, corresponding to Pv-sclerotisation)	1
p6 (pair)	T9 (lateralmost part) – S9 (lateral margin)	1, 4
p7 (pair)	Membrane anterior to paraproct Pp – anterior margin of paraproct Pp	
	and paratergite T10p (border between Pp and T10p not clear)	2

5.2. Metallyticus violaceus (Mantodea, Metallyticidae)

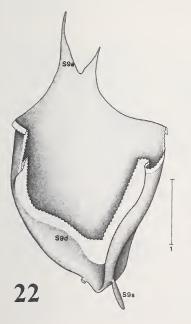
Left complex

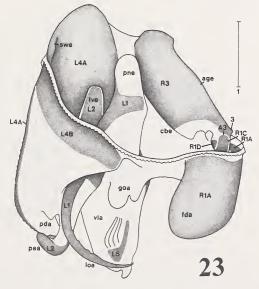
The left complex resembles that of *Sphodromantis*. However, sclerite L4B (fig.23) is ribbon-shaped and restricted to the anterior dorsal wall. The ventral sclerite L4A extends

Figs.20-23: *Metallyticus violaceus* (Mantodea, Metallyticidae) – **20:** Phallomere complex in ventral view. – **21:** Right phallomere with transition to left complex in ventral view; some ventral parts removed. – **22:** Subgenital plate in dorsal view; left stylus missing. – **23:** Left complex and right phallomere in dorsal view. – Scale: 1mm.



Metallyticus violaceus





posteriorly onto two processes (fig.20, 27): the left **pda**-process and the right **vla**-lobe. **L4A** and **L4B** articulate on the left edge of the left complex (A1 in fig.20, 24). **L4A** bears, anterior to A1, a small keel-like apodeme (swe in fig.23, 24; cross-section in fig.25).

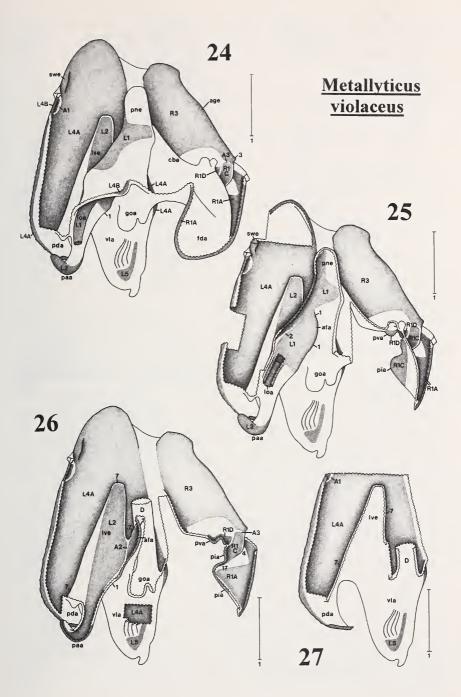
Like Sphodromantis, Metallyticus has a dorsal pouch **pne** and a ventral pouch **lve**, which contain the sclerites **L1** and **L2** (fig.23-25). Within the **pne**-pouch (fig.24, 25) **L1** occupies the ventral wall and parts of the dorsal wall. In the ventral wall **L1** is partly divided by a stripe of membrane (**2** in fig.25). A phallomere-gland, whose opening were to be expected within the **pne**-pouch, was not found. The ventral pouch **lve** (fig.26) deepens strongly anteriad. Its dorsal wall is occupied by the ribbon-shaped **L2**-sclerite. The edge between the pouches **pne** and **lve** (**1** in fig.25, 26) has its anterior starting point in the ventral wall of the **pne**-pouch. The anteriormost part of edge **1** bulges to the right (**afa** in fig.25, 26). Most posteriorly edge **1** bears a somewhat hook-like process **paa**, whose ventral wall is sclerotised by a posterior extension of **L2** (fig.20, 25, 26). Another sclerotised process **loa** (fig.23-25) arises from the posterior ventral wall of the **pne**-pouch (fig.25). Anterior to the **loa**-process, in the area of the bulge **afa**, **L1** bends around the edge **1** into the dorsal wall of the **lve**-pouch, where it has a hinge-like articulation with the right margin of **L2** (**A2** in fig.26).

The membranous ventral wall of the **lve**-pouch is at the same time the left part of the dorsal wall of the ventral lobe **vla** (= ventral phallomere; fig.27). The ejaculatory duct (**D** in fig.26, 27) opens into the anterior ventral wall of the **vla**-lobe. Dorsal to the genital opening there are two membranous lobes (**goa** in fig.25, 26). Posterior to the genital opening the dorsal **vla**-wall contains a small sclerite **L5**, with some small but distinct folds to the left of it. The ventral **vla**-wall is completely sclerotised by a part of **L4A** (fig.20).

Right phallomere

Sclerite **R3** occupies the anterior ventral wall (fig.20, 24). The left end of **R3** has a short extension to the posterior (fig.20, 26). The anterior and right margins of **R3** form a groove-like apodeme **age**, which deepens abruptly in its left part but does not curve like in *Sphodromantis* (fig.20, 24; cross-section in fig.21). Next to its right-posterior end the **age**-apodeme bears a small keel-like apodeme (**3** in fig.20, 24). The right posterior end of **R3** articulates (**A3** in fig.21, 24, 26) with the posteroventral sclerite **R1C**. Sclerite **R1D** is situated to the left of **R1C** and posterior to **R3**; it is in close contact with **R1C** and bears a dental ridge (**pva** in fig.20, 25, 26). The ventral wall of the right phallomere posterior to **R3** and **R1D** is extensively invaginated dorsad and anteriad (**cbe** in fig.21, 24). Sclerite **R1C** extends from articulation **A3** posteriad onto a large ventral tooth (**pia** in fig.20, 26), occupying mainly its dorsal wall. Like in *Sphodromantis*, the area of articulation **A3** and the adjacent part of **R1C** are somewhat groove-like – a posterior extension of the **age**-apodeme on **R3**. The posterior part of the right phallomere is composed of a large dorsal lobe (**fda** in fig.23, 24) and of the ventral **pia**-tooth. The dorsal wall of the **fda**-lobe is

Figs.24-27: *Metallyticus violaceus* (Mantodea, Metallyticidae) – Left complex and right phallomere in dorsal view; with successive removal of their parts (mainly of dorsal ones); fig.27: all parts of right phallomere removed. – Scale: 1mm.



completely occupied by sclerite **R1A**. Around the right edge of the right phallomere **R1A** curves ventrad and sclerotises the ventral wall of the **pia**-tooth (fig.20, 26). **R1A** is completely separated from **R1C**.

Subgenital plate

Fig.22. In the specimen examined only the right stylus S9s was preserved.

5.3. Chaeteessa caudata (Mantodea, Chaeteessidae)

Left complex

Sclerite L4 occupies the whole ventral and dorsal walls (fig.28, 31). The dorsal and ventral parts of L4 are firmly connected along the left edge of the left complex. Within the ventral part of L4, the left, right, and anterior margins as well as an anterior transverse bridge are distinctly heavier sclerotised (fig.28, 32).

A dorsal **pne**-pouch and a ventral **lve**-pouch invaginate from beneath the right margin of the dorsal L4 and take a position in the center of the left complex. These pouches contain the sclerites L1 and L2 (fig.32, 34). The **pne**-pouch has its anterior part deeply invaginated anteriad, its posterior part has the shape of a groove open to the right. L1 occupies the whole ventral wall of the pne-pouch; only within the anterior part of pne L1 curves around the left edge of the pouch into the dorsal wall; here it has a short extension directed rightposteriad (fig.32). The phallomere-gland (P in fig.32, 34) opens into the membranous right wall of the pne-pouch. The ventral pouch lve deepens strongly anteriad. The ribbon-shaped sclerite L2 occupies most of its dorsal wall (fig.34). The edge 1 (fig.34) between the pouches **pne** and **lve** bears in its posterior part a long process **paa**, whose ventral wall is sclerotised by a posterior extension of L2 (fig.28, 34). Immediately anterior to the paabase, L1 curves from the ventral wall of the pne-pouch into the dorsal wall of the lvepouch (around edge 1 in fig.34, 35), where it articulates with L2 (A2 in fig.34, 35). Anterior to this L1-curvature edge 1 bears a membranous process afa. Edge 1 forks immediately anterior to **afa**. Between the two branches the small membranous pouch **pbe** deepens to the left. pbe is situated between the pouches pne and lve.

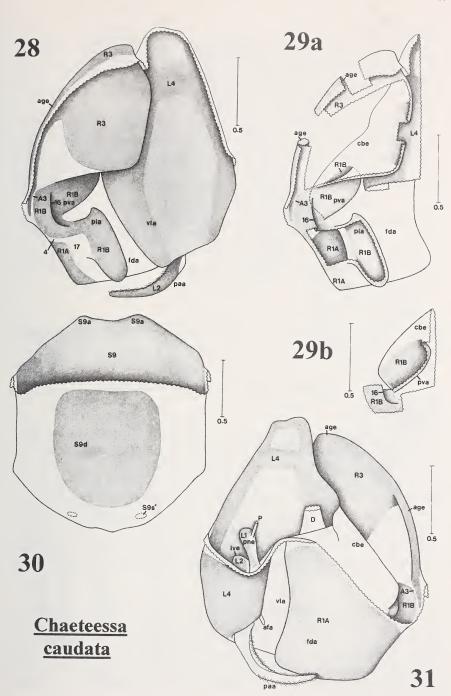
The membranous ventral wall of the **lve**-pouch is at the same time the left dorsal wall of a large ventral lobe **vla** (= ventral phallomere; fig.28, 32). The ejaculatory duct (**D** in fig.32, 35) opens into the anterior dorsal wall of the **vla**-lobe – far on the right side and outside the **lve**-pouch. The ventral wall of the **vla**-lobe is sclerotised by a part of the ventral **L4** (fig.28).

Figs.28-31: Chaeteessa caudata (Mantodea, Chaeteessidae) – 28: Phallomere complex in ventral view.

^{- 29}a: Right phallomere with transition to left complex in ventral view; some ventral parts removed.

^{- 29}b: Detail of right phallomere in ventral view (compare fig.29a); with sclerite R1B and tooth pva.

^{- 30:} Subgenital plate in dorsal view; styli missing. - 31: Left complex and right phallomere in dorsal view. - Scale: 0.5mm.



Right phallomere

Sclerite **R3** in the anterior ventral wall is hatchet-shaped (fig.28, 31). Its anterior and right margins form an apodeme **age** (fig.28, 32). The left part of **age** is groove-like, the right part is beam-like (the groove is filled in by the cuticle being thickened; cross-section through **age** in fig.29a, 33). The leftmost part of **R3** bends back to the right (fig.29a, 32, 33).

Posterior to the left part of **R3** the ventral wall of the right phallomere is invaginated dorsad and anteriad (**cbe** in fig.29a, 32). The right posterior end of **R3** has a broad articulation **A3** with the posteroventral sclerite **R1B**. From **A3** sclerite **R1B** extends to the left onto the anterior tooth **pva**, on which way it bends dorsad along the edge **16** (fig.28, 29a), as well as onto the posteroventral tooth **pia**. The dorsal and ventral walls of **pva** and **pia** are completely sclerotised by **R1B** (fig.28, 29a, 33).

The posterior part of the right phallomere is composed of the large dorsal lobe **fda** (fig.31, 32) and the ventral tooth **pia**. The dorsal wall of **fda** is occupied by sclerite **R1A**. Around the right edge of the right phallomere **R1A** curves into the ventral wall, where it is restricted to the right margin and completely separated from **R1B** by membrane (4 and 17 in fig.28, 32).

Subgenital plate

Fig.30. The styli **S9s** have been lost in the examined specimen; only their points of insertion are shown (**S9s***).

5.4. Mantoida schraderi (Mantodea, Mantoididae)

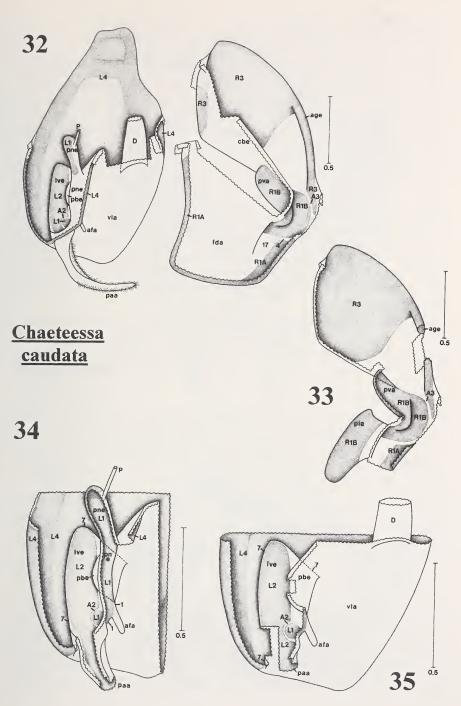
Left complex

Sclerite L4 extends along the left, anterior, and right margins of the ventral wall (fig.41, 44). Along the whole left edge of the left complex L4 also curves into the dorsal wall (fig.44, 45), where it is restricted to the left margin. Only a very distinct part of L4 in the anterior dorsal wall extends farther to the right (L4d in fig.44). An apodeme swe runs along the whole left arm of L4 (fig.44, 45). The anterior part of swe is beam-like by the cuticle being thickened (cross-section of swe in fig.46). To the posterior this thickening decreases, and swe is groove-shaped (cross-section in fig.45).

To the right of the dorsal part of L4, a dorsal pouch **pne** (fig.44, 45) and a ventral pouch **lve** (fig.45, 46) are invaginated anteriad, which contain the sclerites L1 and L2. Within

Figs.32-35: *Chaeteessa caudata* (Mantodea, Chaeteessidae) – **32:** Left complex and right phallomere (separated from each other) in dorsal view; some parts removed (mainly dorsal ones; compare fig.31).

^{- 33:} Right phallomere in dorsal view; further parts removed (mainly dorsal ones; compare fig.32).
- 34: Left posterior part of left complex in dorsal view; further parts removed (mainly dorsal ones; compare fig.32). - 35: Posterior part of left complex in dorsal view; right posterior part of left complex with genital opening retained; further parts removed in the left half (mainly dorsal ones; compare fig.34). - Scale: 0.5mm.



the pne-pouch, the hood-shaped L1 occupies most of the ventral wall and the left margin of the dorsal wall (fig.45). The left-dorsal posterior margin of L1 articulates with a small sclerite on the process loa (fig.45). The right posterior margin of L1 has a ribbon-like extension. The phallomere-gland (P in fig.44, 45) opens into the anteriormost membranous part of the **pne**-wall. The ventral pouch **lve** (fig.46) is more tranversely extended (and less antero-posteriorly as in Metallyticus and Chaeteessa). The edge along the bottom of the lve-pouch is labelled 7 in fig.46, 47. The ejaculatory duct (D in fig.46) opens into the right part of lve. Sclerite L2 extends like an arch along the margins of the dorsal wall of lve. In the anteriormost left edge of the pouch, however, it bends into the ventral wall of lve (fig.47), and from here it extends posteriad to join the left posterior end of L4. The area where these posterior ends of L2 and L4 are interconnected is as a whole upcurved (fig.45) and bears two short processes: the right, somewhat pointed paa, whose sclerotisation belongs to L2, and the left, bulge-like pda, whose sclerotisation is part of L4. The edge 1 (fig.45, 46) between the pouches pne and lve is transversely orientated, not longitudinally as in the previous species. Far to the right of paa the right posterior ends of L1 and L2 contact each other (articulation A2 in fig.45, 46) – exactly in the edge 1. Immediately to the right of A2 the invagination of the lye-pouch - and thus also the edge 1 -ends (fig.46), and immediately to the right of this point the membranous lobe afa has its base.

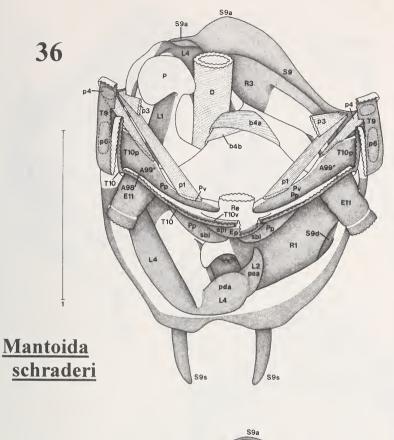
The membranous ventral wall of the **lve**-pouch is at the same time the anterior dorsal wall of the ventral lobe **vla** (= ventral phallomere; fig.46, 47). The ejaculatory duct (**D** in fig.46, 47) opens most anteriorly, and quite far to the right, into this membrane. The ventral wall of the **vla**-lobe is partly sclerotised by the right posterior part of **L4** (fig.41, 47).

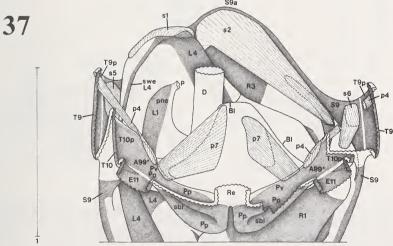
Right phallomere

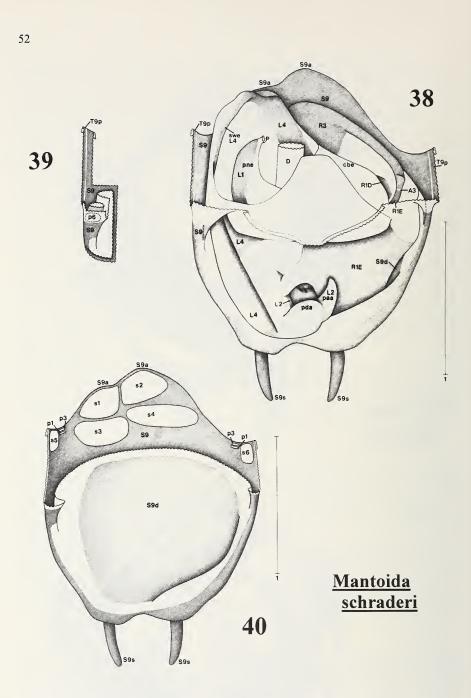
Sclerite **R3** in the anterior ventral wall is hatched-shaped (fig.41, 44). Its anterior and right margins form an **age**-apodeme (fig.41, 44, 45), which is distinctly groove-like in its left part but more beam-like in its right part (cross-section through **age** in fig.43, 45). In its left part **age** is deeper. On the utmost right posterior part of **R3** the **age**-apodeme bears a small keel-like apodeme (**3** in fig.43, 44).

The horseshoe-shaped sclerite **R1D** lies in the ventral wall posterior to the central part of **R3** and sclerotises the tooth **pva** (fig.41, 43, 45). Posterior to the left and central parts of **R3** and posterior to **R1D** the ventral wall of the right phallomere is invaginated dorsad and anteriad (**cbe** in fig.43-45). The right posterior end of **R3** articulates (**A3** in fig.41, 44) with the ventral part of sclerite **R1E**. The groove called **age** on **R3** extends beyond

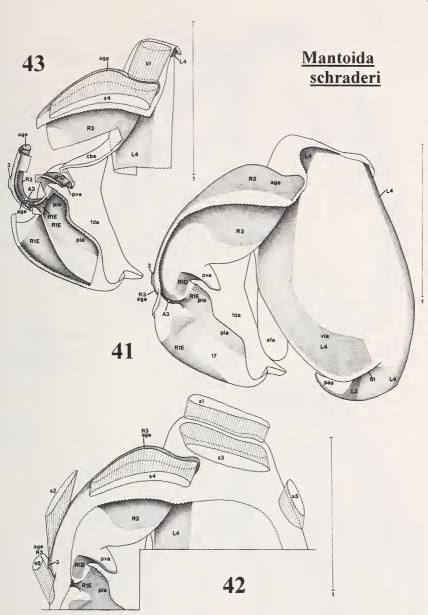
Figs.36,37: *Mantoida schraderi* (Mantodea, Mantoididae) – **36:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, supraanal lobe, epiproct, subanal lobes, paraprocts, distal part of rectum, basal parts of cerci, and part of musculature. – **37:** Same as in fig.36, after removal of further parts of abdominal tergites 9 and 10, parts of right paraproct, and supraanal lobe with epiproct. Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. Posterior to transverse line: like in fig.36. – Scale: 1mm.



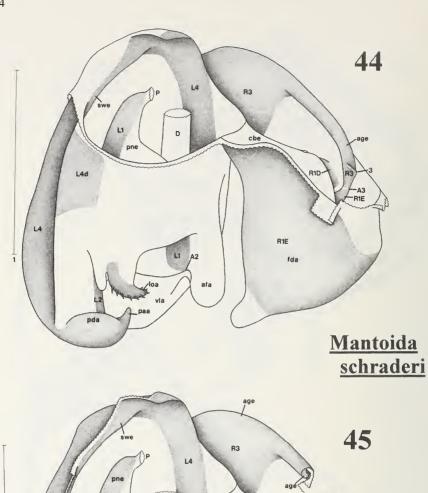


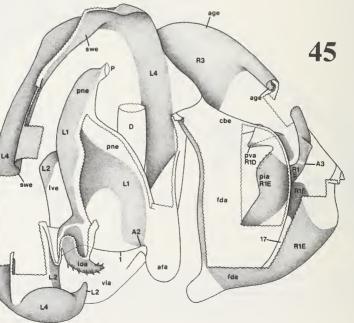


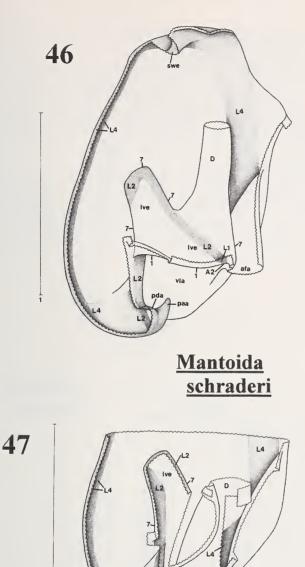
Figs.38-40: *Mantoida schraderi* (Mantodea, Mantoididae) – **38:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **39:** Left margin of subgenital plate (compare fig.38); with insertion of muscle p6. – **40:** Subgenital plate in dorsal view; with insertion areas of muscles (except p6). – Scale: 1mm.



Figs.41-43: *Mantoida schraderi* (Mantodea, Mantoididae) – **41:** Phallomere complex in ventral view. – **42:** Anterior part of phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.41. – **43:** Right phallomere with transition to left complex in ventral view; with some muscles; some ventral parts removed. – Scale: 1mm.







Figs.46,47: *Mantoida schraderi* (Mantodea, Mantoididae) – Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones). – Scale: 1mm.

vla

Figs.44,45: *Mantoida schraderi* (Mantodea, Mantoididae) – 44: Left complex and right phallomere in dorsal view; some parts removed (mainly dorsal ones). – Scale: 1mm.

articulation A3 (where it is no longer called **age**) onto **R1E** (fig.41). The anterior margin of **R1E** is in close contact with **R1D** (fig.41).

The posterior part of the right phallomere is composed of the large dorsal lobe **fda** (fig.44, 45), with **R1E** in its dorsal wall, and the ventral tooth **pia**. Sclerite **R1E** extends from articulation **A3** left-posteriad onto the **pia**-tooth and occupies most of its dorsal and ventral walls (fig.41, 43, 45). Another part of **R1E** extends narrowly to the right edge of the phallomere, curves dorsad, and, becoming broader again, occupies most of the dorsal wall of the **fda**-lobe (fig.41, 44).

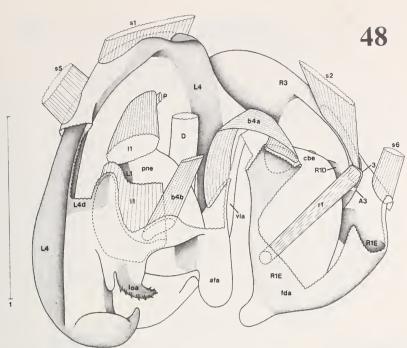
Subgenital plate and posterior abdominal segments

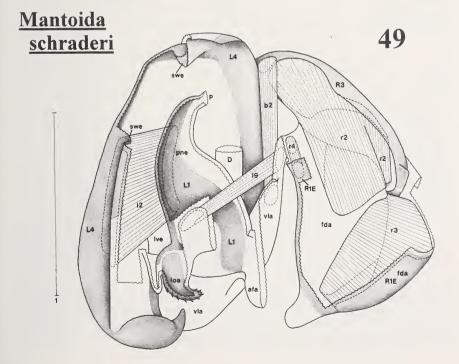
Fig.36, 37 (posterior segments); fig.40 (subgenital plate **S9**). The ventral part of tergite 10 **T10v** is very narrow but distinct. Separate **Pv**-sclerites are missing; they are assumed to have been incorporated into the anterior margins of the paraprocts **Pp**. The sclerites **Ca**, **Cb**, and **Cc** are missing. The articulations **A99** are missing (**A99***: paratergites **T10p** and paraprocts **Pp** have fused). The articulations **A98** are well-developed.

Musculature

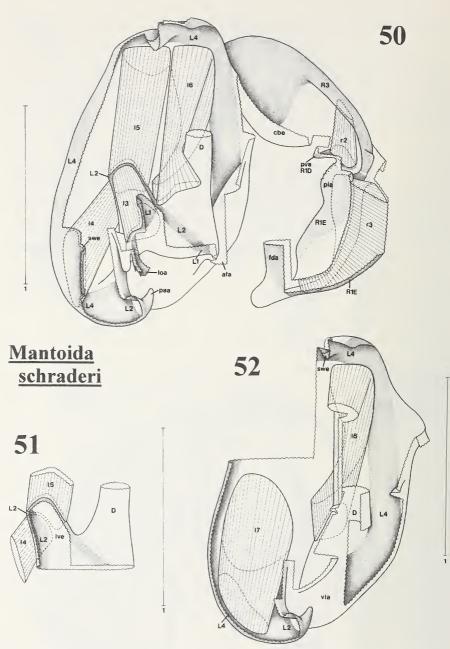
Muscles	Positions of insertions	in fig.
11	L1 (anteriorly on pne -pouch) – central dorsal wall of left complex near right end of L4d	48
12	L1 (left-ventrally on pne-pouch) – L4 (dorsally on left arm, on	
	swe-apodeme)	49
13	L1 (posteroventrally on pne-pouch) – L2 (left-anterior part)	50
14	L2 (left-anterior part) – L4 (dorsally on left arm, on swe-apodeme)	50, 51
15	Left anterior membranous ventral wall of left complex – L2	
	(left-anterior part)	50, 51
16	Right anterior membranous ventral wall of left complex – L2	
	(right part) and dorsal wall of vla-lobe around genital opening	50, 52
17	Left posterior membranous ventral wall of left complex – L4	
	(posterior ventral part of left arm)	52
19	Transversely within right dorsal wall of left complex	49
r1	R3 (right margin) – R1E (dorsal wall of fda-lobe)	48
r2	R3 – R1D on pva-tooth and membrane of cbe-invagination	49, 50
r3	R1E (right part) – R1E on pia-tooth and right ventral wall of fda-lobe	49, 50
r4	R1E (left-anterior part) – left ventral wall of fda-lobe	49
b1	Membrane behind right anterior margin of L4 – R3 (left anterior margin) 43
b2	Dorsal wall of vla-lobe (right-anterior part) – R3 (left anterior margin)	49
b4a	Dorsal wall of vla-lobe (right-anterior part) - R1E (left anterior margin)	36, 48
b4b	Central dorsal wall of left complex - membrane posterior to anterior	
	margin of R1E (in fig.36 the insertion is beneath muscle b4a)	36, 48

Figs.48,49: *Mantoida schraderi* (Mantodea, Mantoididae) – Left complex and right phallomere in dorsal view; each figure with some muscles; parts of phallomere complex removed to various extents. – Scale: 1 mm.









Figs.50-52: *Mantoida schraderi* (Mantodea, Mantoididae) – **50:** Left complex and right phallomere in dorsal view; each figure with some muscles; parts of phallomere complex removed. – **51:** Detail of left complex in dorsal view (compare fig.46); with some muscles. – **52:** Left complex in dorsal view; with some muscles; parts of left complex removed (mainly dorsal ones). – Scale: 1mm.

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s1	S9 left side (medially and most anteriorly) $-$ L4 (anterior margin)	37, 40, 42, 48
s2	S9 right side (medially and most anteriorly) – R3 (lateral anterior margin)	
s3	S9 left side (medially and anteriorly) – anteriormost left ventral wall of	
	genital pouch	40, 42
s4	S9 right side (medially and anteriorly) $- R3$ (left ventral wall of	
	age-apodeme)	40, 42, 43
s5	S9 left side (laterally and anteriorly) - anterior left wall of genital pouch	37, 40, 42, 48
s6	S9 right side (laterally and anteriorly) – anterior right wall of genital	
	pouch	37, 40, 42, 48
p1 (pair)	S9 – paraproct Pp (anterior margin, corresponding to Pv-sclerotisation)	36, 40
p3 (pair)	S9 – rectum (ventral wall)	36, 40
p4 (pair)	T9 (lateralmost anterior margin, also extending onto paratergite T9p) -	
	anterior margin of paratergite T10p (far medially); on both sides	
	completely divided into a dorsal (fig.36: on T9) and a ventral	
	(fig.37: on T9p) bundle	36, 37
p5 (pair)	Not investigated; presence highly probable	
p6 (pair)	T9 (lateralmost part) – membrane immediately above lateral margin	
	of S9	36, 39
p7 (pair)	Membrane anterior to paraproct \mathbf{Pp} – anterior margin of paraproct \mathbf{Pp}	
	and paratergite T10p (border between Pp and T10p not clear)	37

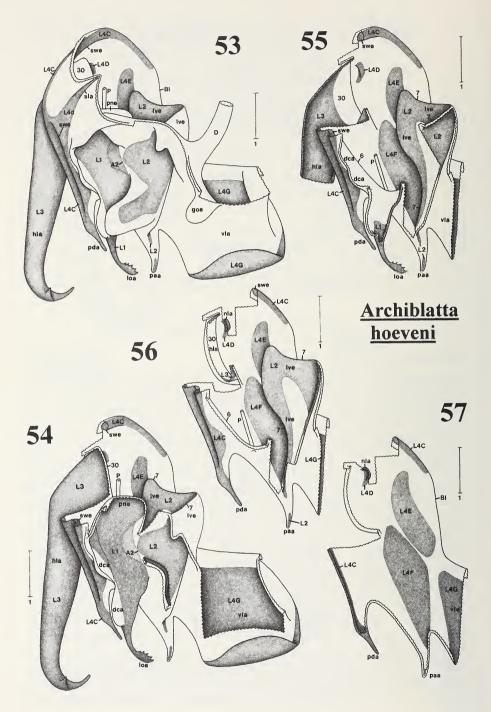
5.5. Archiblatta hoeveni (Blattaria, Blattidae, Blattinae)

Left complex

L4 is a group of five sclerites: The crescent-shaped L4C occupies the whole left edge and the anteriormost left ventral wall of the left complex (fig.53). That part along the left edge has a distinct dorsal extension to the right (L4d in fig.53), the end of which forms a spine sla. Along the whole of L4C there runs an apodeme swe (fig.53-56), whose anterior part is beam-like by the cuticle being thickened (anterior cross-section of swe in fig.54); to the posterior this thickening decreases, and swe becomes more and more groove-shaped (posterior cross-section of swe in fig.54). The sclerites L4D, L4E, L4F, and L4G lie in the ventral wall of the left complex. L4D bears a node-like process nla (fig.56, 57). L4E and L4F are simple plates. L4G occupies the ventral wall of a large ventral lobe vla (= ventral phallomere; fig.53, 54, 57), and its posterior part is upcurved.

In the center of the left complex there are two pouches **pne** and **lve** (fig.53-56), which contain the sclerites **L1** and **L2**. The anterior part of **L1** lies in the ventral wall of the small and flat **pne**-pouch (fig.53, 54). The posterior part of **L1** extends onto a broad lobe, which is divided into three processes posteriorly (fig.54, 55): the two short, membranous **dca** (left side) and the long, sclerotised **loa** (right side). Along the lateral and posterior edges of this lobe the cuticle bends ventrad and anteriad (fig.55) as far as to the edge **6**, where it turns posteriad again. The phallomere-gland (**P** in fig.55, 56) opens immediately posterior to edge **6**.

The large sclerite L2 adjoins to the right of L1 (fig.53, 54), and the two sclerites articulate (A2 in fig.53, 54). From A2 sclerite L2 extends to the right, curves ventrad and – becoming narrower – back to the left; then it bends posteriad and runs to the posterior edge of the



left complex; here it ends at the base of a small process **paa** (fig.56). This arch-like course of **L2** extends along the margins of a pouch **lve**, which is, like **L2** itself, curved dorsoventrally (in contrast to Mantodea, whose **lve**-pouches extend within one plane). Thus, the left parts of the **lve**-pouch and of **L2** lie beneath the **pne**-pouch as in Mantodea, but their right parts curve upwards into the same plane which also contains sclerite **L1**. The edge along the bottom of this **lve**-pouch is labelled **7** in fig.54, 55. The invagination of the **lve**-pouch starts immediately anterior to the **paa**-process (posterior end of **7** in fig.54), where **L2** leaves the pouch and approaches **L1**. According to the curvature of the **lve**-pouch, it is preferable to name that wall of **lve** containing sclerite **L2** the inner one (instead of dorsal) and the opposite wall the outer one (instead of ventral). **L2** is restricted to the inner **lve**-wall; only most posteriorly it bends into the outer (or ventral) **lve**-wall. Then it leaves the pouch and ends on the **paa**-process (fig.55, 56). **paa** is, except for another small distal sclerite (probably a split off part of **L2**), membranous (fig.55-57).

The membranous outer (or ventral) wall of the **lve**-pouch is at the same time the dorsal wall of the **vla**-lobe (fig.53-56; in the figures **vla** is pulled to the right). The ejaculatory duct (**D** in fig.53) opens into this wall. Dorsal to the genital opening there is a small membranous lobe **goa**. The ventral wall of the **vla**-lobe is sclerotised by **L4G**. The posterior edge of the **vla**-lobe continues leftward into the posterior edge of the remaining left complex, where the **paa**-process follows (fig.53, 57).

The large hook **hla** is evaginated from the left anterior ventral wall of the left complex. **hla** is, except for its basalmost walls (**30** in fig.54-56), completely sclerotised by **L3** (fig.53-55). **hla** is retractable for a very short distance, since the basal membranous walls **30** can be introverted (this state is shown in the figures).

Right phallomere

The right phallomere is only schematically shown in fig.330f. Differences to *Eurycotis* (following species) will be explained in 6.7.4., 6.7.5., and 6.7.6.

5.6. Eurycotis floridana (Blattaria, Blattidae, Polyzosteriinae)

Left complex

The left complex resembles that of *Archiblatta*. L4 is a group of three sclerites: The crescent-shaped L4H occupies the left edge and the left anterior ventral wall of the left complex (fig.65, 66). That part of L4H in the left edge broadens in its posterior half. A beam-like apodeme swe (fig.65, 66; anterior and posterior cross-section in fig.66) runs along L4H. At the anterior end of the swe-apodeme L4H bends abruptly posteriad and broadens into a plate bearing the node-like process nla (fig.68, 69) in its left half. The

Figs.53-57: Archiblatta hoeveni (Blattaria, Blattidae, Blattidae) – 53: Left complex in dorsal view. – 54-57: Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones). – Scale: 1mm.

plate L4F lies in the posterior ventral wall. Along its anterior margin the membrane is evaginated posteroventrad to form the broad lobe mla (fig.63, 69). L4G occupies most of the ventral and right walls of a large ventral lobe vla (= ventral phallomere; fig.63, 65, 66).

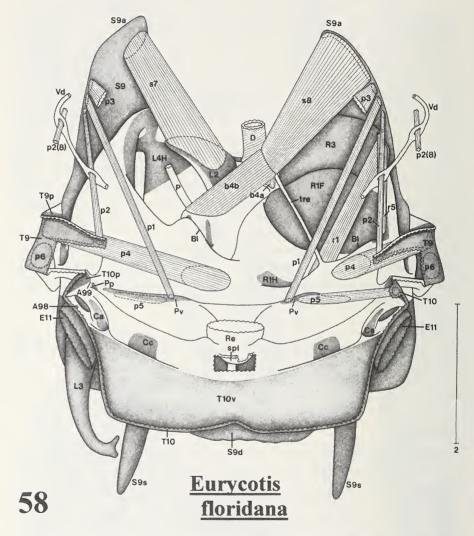


Fig.58: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, supraanal lobe, subanal lobes (covered), paraprocts (mostly covered), Pv-sclerites, distal part of rectum, basal parts of cerci, parts of vasa deferentia, and part of musculature. Supraanal lobe shown through a window cut into the membrane anterior to ventral sclerotisation of abdominal tergite 10 T10v. – Scale: 2mm.

Sclerite L1 lies in the central dorsal wall of the left complex (fig.65-67). Its anterior part occupies the ventral wall of a pouch-like invagination (**pne** in fig.65-67). Its posterior part has a longitudinal furrow (**8** in fig.67) and extends onto two processes (**dca** in fig.67, 68). The phallomere-gland (**P** in fig.68) opens into the ventral wall of the right **dca**-process. Two small sclerites L6A and L6B lie in the left-dorsal wall of the **pne**-pouch; each bears

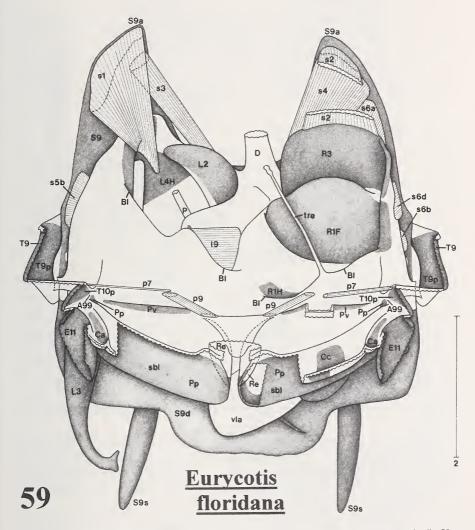


Fig.59: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – Male postabdomen as in fig.58, after removal of further parts of abdominal tergites 9 and 10 (especially T10v) and supraanal lobe. Distal part of rectum, basal parts of cerci, and dorsal wall of right subanal lobe cut open. Another part of musculature shown. – Scale: 2mm.

a spine (fig.65, 66). Sclerite L2 adjoins to the right of L1 (fig.67), and the two sclerites articulate (A2 in fig.67). Like in *Archiblatta*, L2 is dorsoventrally curved (fig.67, 68), and the anterior part of L2 lies in the inner wall of a large pouch (lve in fig.65-68; the edge along the bottom of the lve-pouch is labelled 7 in fig.66-68). The anteroventral parts of L2 and lve deepen strongly to the left to form a tongue-like apodeme (lve in fig.67). At the posterior margin of L2 there are three processes, the largest of which is the completely sclerotised **paa** (fig.67).

The outer wall of the **lve**-pouch is completely membranous and is at the same time the dorsal wall of the **vla**-lobe (fig.65-67; in the figures **vla** is pulled to the right). The ejaculatory duct (**D** in fig.65, 66) opens into the anterior part of this wall. Dorsal to the genital opening there is a membranous lobe **goa** (fig.66). In contrast to *Archiblatta*, the main part of the **vla**-lobe is separated by a deep notch (**9** in fig.63) from the remaining ventral parts of the left complex. The **hla**-hook (fig.65-67) is like in *Archiblatta*.

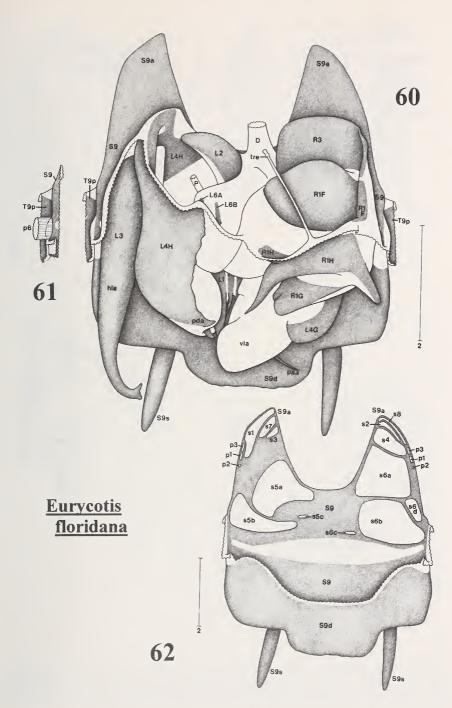
Right phallomere

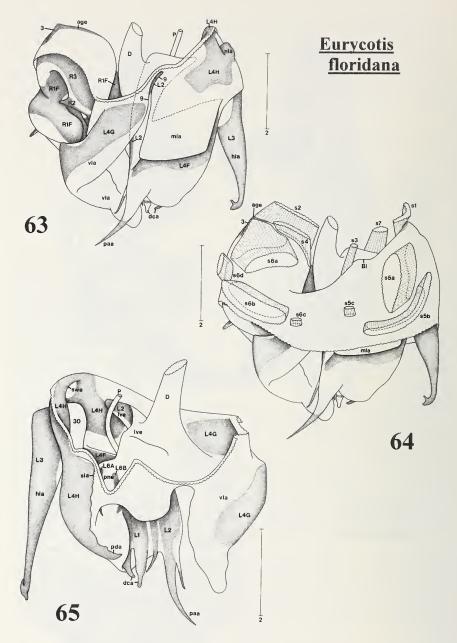
Sclerite **R3** in the anterior (right-)ventral wall is a curved plate (fig.74-77). The right and the right anterior margins of **R3** form a groove-like apodeme **age** (fig.74, 77; cross-section through **age** in fig.78). The right part of **age** bears a keel-like apodeme (**3** in fig.74, 77). Posterior to the left part of **R3** sclerite **R2** adjoins, and the two sclerites articulate (**A7** in fig.75-77). **R2** forms a dental ridge (fig.74-77). Posterior to the central part of **R3** the ventral wall of the right phallomere is extensively invaginated dorsad and anteriad (**cbe** in fig.74-76; compare fig.77 and 78). This **cbe**-invagination takes a position in the center of the right phallomere.

Posterior to the right end of **R3** sclerite **R1F** adjoins, and the two sclerites are articulated (**A3** in fig.74, 75, 77). **R1F** extends from its central part behind **A3** in three directions: The first arm bends left-dorsad (along edge **16**, fig.77, 78) and occupies the whole right-dorsal wall of the **cbe**-invagination (fig.74, 75, 78). This arm forms a dental ridge (**pva** in fig.75, 78, 80) at its posterior margin and articulates with the left-dorsal end of **R2** (**A6** in fig.75, 76) at its median end. The second arm of **R1F** extends posteriad and sclerotises the anterior part of a two-pointed ventral tooth (**pia** in fig.77). The third arm of **R1F** extends posterodorsad (fig.74); its dorsal margin folds back to the right to form a sclerotised groove (**rge** in fig.74); the **rge**-groove is a posterior continuation of the **age**-groove on **R3**.

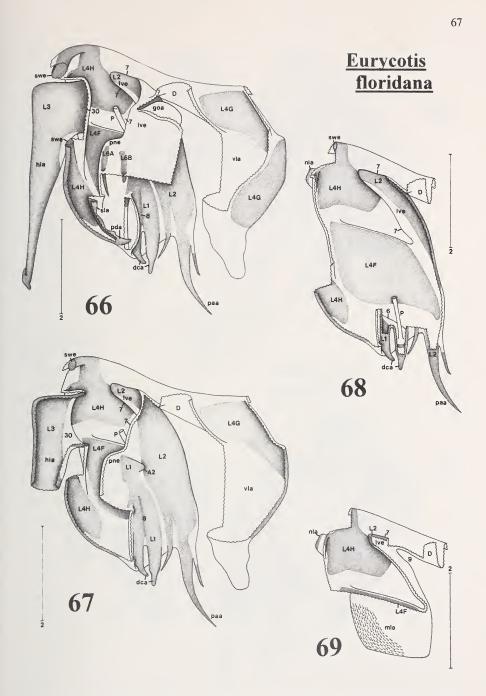
The posterior part of the right phallomere is composed of the dorsal lobe **fda** (fig.74) and the ventral tooth **pia** (fig.77, 78). **fda** and **pia** are confluent along the right edge of the right phallomere and diverge towards the left. The dorsal wall of the **fda**-lobe and parts of its ventral wall are occupied by sclerite **R1H** (fig.74, 76). The left part of **R1H** scle-

Figs.60-62: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – **60:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **61:** Left margin of subgenital plate (compare fig.60); with insertion of muscle p6. – **62:** Subgenital plate in dorsal view; with insertion areas of muscles (except p6). – Scale: 2mm.



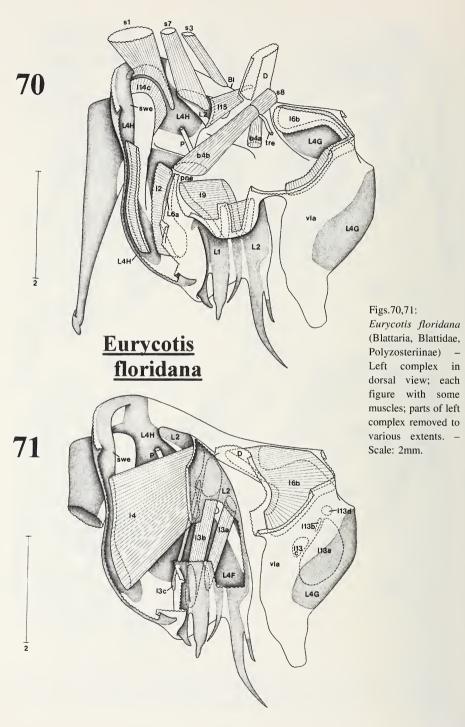


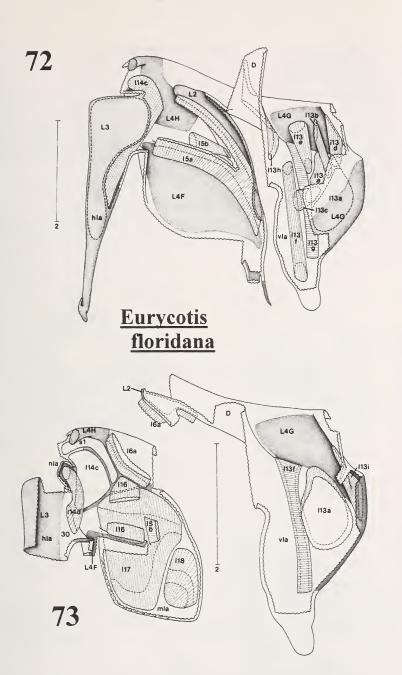
Figs.63-65: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – **63:** Phallomere complex in ventral view. – **64:** Phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.63. – **65:** Left complex in dorsal view. – Scale: 2mm.



Figs.66-69: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones). – Scale: 2mm.







Figs.72,73: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – Scale: 2mm.

rotises the spine **sra**; the left end of **R1H** is somewhat spoon-shaped. The right end of **R1H** extends posteriorly onto the process **pra** (fig.74) and articulates anteriorly with the dorsal arm of **R1F** (**A8** in fig.74). The tendon **tre** (fig.74), a very long and narrow invagination of the cuticle, has its origin in the anteriormost dorsal wall of the **fda**-lobe. The ventral tooth **pia** is anteriorly sclerotised by the second arm of **R1F** (fig.77, 78); posteriorly **pia** is sclerotised by **R1G**, which articulates with **R1F** in the dorsal wall of **pia** (**A9** in fig.78). **R1G** has a narrow extension to the right, which reaches the **pra**-process and maintains a narrow connection with the dorsal **R1H** (fig.74, 77, 78). Anterior to this sclerite bridge there is a large membranous area (**17** in fig.74, 77).

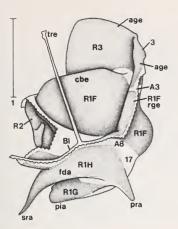
Subgenital plate and posterior abdominal segments

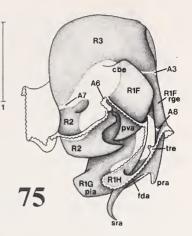
Fig.58, 59 (posterior segments); fig.62 (subgenital plate **S9**). The ventral part of tergite 10 **T10v** is very extensive. Strip-like **Pv**-sclerites are present; they are completely free from the paraprocts **Pp**. The **Ca**-sclerites lie on curved bulges immediately median to the cercal bases. The **Cc**-sclerites are dorsoventrally curved plates on paired shallow outfoldings beneath the anterior margin of **T10v** and above the subanal lobes **sbl**. **Cb**-sclerites are missing. The articulations **A98** and **A99** are well-developed.

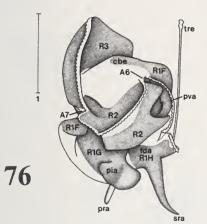
Musculature

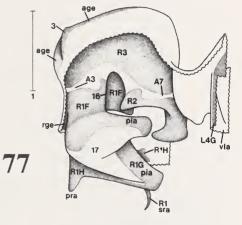
Muscles	Positions of insertions	in fig.
12	Membranous left wall of pne -pouch – L4H (left part, on swe -apodeme)	70
l3a,b,c	L1 and membrane to the left of L1 (posteroventrally on pne-pouch) – L	2
	(anterodorsal part)	71
14	L2 (anterodorsal part) – L4H (left part, on swe-apodeme)	71
15a	L4F (anterior margin) – L2 (ventral left margin)	72
15b	Membranous ventral wall of left complex anterior to L4F - L2 (ventral	
	left margin)	72, 73
16a	L4H (plate-like part in ventral wall of left complex) – anterior ventral	
	wall of lve-pouch	73
16b	L4G (anterior margin) and membrane anterior to L4G – dorsal wall	
	of vla-lobe around genital opening	70, 71
19	Transversely in central dorsal wall of left complex	59, 70
113a-i	Muscles within vla-lobe; mainly diffuse	71, 72, 73
114c	L4H (to the right of nla-process) – hla-hook (dorsally on L3)	70, 72, 73
l14d	L4H (on nla-process) – hla-hook (ventrally on membranous base 30)	73
115	Ejaculatory duct D next to its opening – L2 (anterior margin)	70
116	L4H (plate-like part in ventral wall of left complex) – L4F (anterior	
	margin)	73
117	L4H (plate-like part in ventral wall of left complex) and membrane	
	posterior to it – ventral wall of mla-lobe	73

Figs.74-78: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – **74:** Right phallomere in dorsal view. – **75:** Right phallomere in left-dorsal (somewhat anterior) view. – **76:** Right phallomere in left-ventral view; with transition to left complex. – **78:** Right phallomere in right-ventral view; most elements shown in fig.77 removed. – Scale: 1mm.



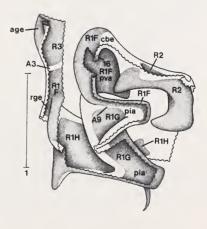




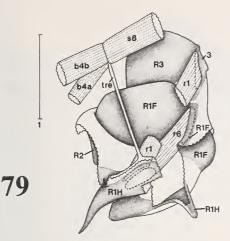


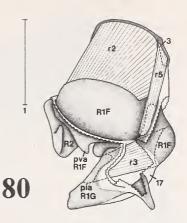
<u>Eurycotis</u> <u>floridana</u>

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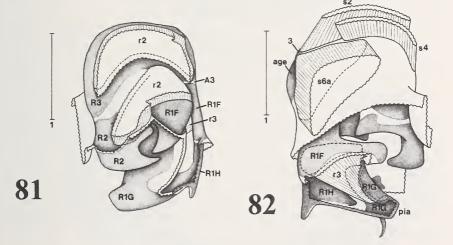


118	Path insertions on ventral well of mle lobe	72
	Both insertions on ventral wall of mla -lobe	73
r1	R3 (right margin) – R1H (anterior dorsal wall of fda-lobe)	58, 79
r2	R3 – cbe-invagination: R1F (anterior margin), membrane, and R2	00.01
	(dorsal margin)	80, 81
r3	R1F (central part, posterior to articulation A3) – R1G (on pia-tooth)	80, 81, 82
r5	R3 (right margin) – R1F (dorsal margin, on rge-apodeme)	58, 80
r6	R1F (dorsal margin, on rge -apodeme) – R1H (anterior dorsal wall	70
	of fda-lobe)	79
b4a	Outer wall of lve-pouch (anterior part next to genital opening) –	
	tre-tendon	58, 70, 79
b4b	pne-pouch (anterodorsal wall) – tre-tendon	58, 70, 79
s1	S9 left side (anteriorly on apophysis S9a) – L4H (anteriormost left	
	ventral wall of left complex)	59, 62, 64 70
s2	S9 right side (anteriorly on apophysis S9a) – R3 (anterior margin)	59, 62, 64, 82
s3	S9 left side (anteriorly on apophysis S9a) – anteriormost ventral wall	
	of left complex	59, 62, 64, 70
s4	S9 right side (anteriorly on apophysis S9a) – R3 (left anterior margin)	59, 62, 64, 82
s5a	S9 left side (posterior to apophysis S9a, medially) – anterior left	
	ventral wall of genital pouch; diffuse	62, 64
s5b	S9 left side (posteriorly and laterally) – posterior left ventral wall of	
	genital pouch	59, 62, 64
s5c	S9 left side (posteriorly and far medially) – median ventral wall of	
	genital pouch; small, diffuse	62, 64
s6a	S9 right side (posteriorly on apophysis S9a) – anteriormost right	
	ventral wall of genital pouch	59, 62, 64, 82
s6b	S9 right side (posteriorly and laterally) – posterior right ventral wall	
	of genital pouch	59, 62, 64
s6c	S9 right side (posterior to apophysis S9a) – median ventral wall of	(a) (1
0	genital pouch; small, diffuse	62, 64
s6d	S9 right side (posteriorly and far laterally) – right wall of genital pouch (20 ± 10^{-1})	59, 62, 64
s7	S9 left side (anteriorly on apophysis S9a) – L2 (anteriorly on lve-pouch)	58, 62, 64, 70
s8	S9 right side (most anteriorly on apophysis S9a) – tre-tendon	58, 62, 70, 79
p1 (pair)	S9 – Pv-sclerite (median part)	58, 62
p2 (pair)	S9 - T9 (lateral anterior margin); in most specimens the p2 pass	
	through eyelets of the vasa deferentia Vd. A pair of muscles having	
	the same course is also present in segment 8: p2(8).	58, 62
p3 (pair)	S9 – rectum (ventral wall)	58, 62
p4 (pair)	T9 (lateralmost anterior margin) – membrane far anterior to Pv -sclerite;	50
	ventral insertion of left p4 far to the right	58
p5 (pair)	T10 (lateralmost anterior margin) – Pv -sclerite	58
p6 (pair)	T9 (lateralmost part) – S9 (lateral margin) Membrane enterior to B ¹ calculateral margin	58, 61
p7 (pair)	Membrane anterior to $\mathbf{P}v$ -sclerite – anterior margin of paratergite T10p Membrane anterior to $\mathbf{P}v$ -sclerite (next to $\mathbf{r7}$), membrane median to	59
p9 (pair)	Membrane anterior to Pv -sclerite (next to $p7$) – membrane median to inner and of Pv sclerite beneath reatum	50
	inner end of Pv -sclerite, beneath rectum	59





Eurycotis floridana



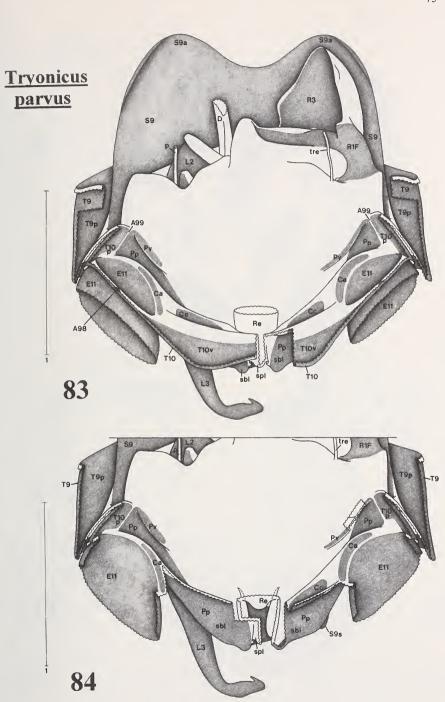
Figs.79-82: *Eurycotis floridana* (Blattaria, Blattidae, Polyzosteriinae) – **79:** Right phallomere in dorsal view; with some muscles; some dorsal elements removed. – **80:** Right phallomere in dorsal view; with some muscles; dorsal elements more extensively removed than in fig.79. – **81:** Right phallomere in left-dorsal (somewhat anterior) view; with some muscles; some dorsal elements removed. – **82:** Right phallomere in right-ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.77; pia-tooth cut open to show muscle r3. – Scale: 1mm.

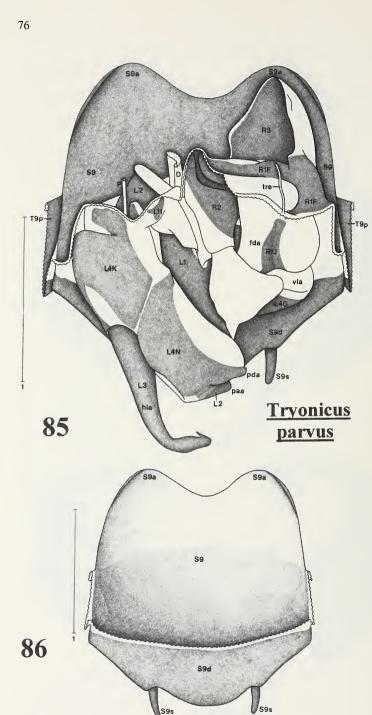
5.7. Tryonicus parvus (Blattaria, Blattidae, Tryonicinae)

Left complex

L4 is a group of three sclerites in the ventral wall and on the left edge of the left complex (fig.87, 88, 91): L4G occupies the ventral wall of a broad ventral lobe vla (= ventral phallomere; fig.87, 88, 98). L4K is a broadly horseshoe-shaped sclerite in the left anterior ventral wall and bears a bulge-like process (nla in fig.87-89, 97, 98). L4N occupies the posterior left edge of the left complex and has a hinge-like articulation with L4K (A5 in fig.88, 97, 98). Anterodorsally L4N has a long ribbon-like extension to the anterior (L4d in fig.88-97). The posterior end of L4N is on a short bulge-like process pda (fig.91, 94, 96) and is to the right connected with the L2-sclerotisation of another process paa (fig.94). The small sclerite L11 lies in the anteriormost left dorsal wall (fig.88, 91). The right anterior part of the dorsal wall has some complicated membranous foldings (fig.91-93). Beneath and posterior to these foldings there are the somewhat cup-shaped L1-sclerite and, to the right of L1, the dorsoventrally curved L2-sclerite. The plate-like dorsal part of L1 (fig.94) has a hinge-like articulation with L2 (A2 in fig.94). The posterior parts of L1 occupy two bulge-like processes (dca in fig.94, 95; only the ventral wall of the right dca-process is membranous). The ventral part of L1 sclerotises the dorsal wall of a broad pouch (pne in fig.95). This pne-pouch is strongly deepened in its rightmost part, where L1 has a ribbon-like extension to the anterior. At the left end of the pne-pouch L1 is in close contact with the anterior end of the L4d-ribbon (fig.94, 95). The ventral wall of the **pne**-pouch is membranous and contains the opening of the phallomere-gland (**P** in fig.96). L2 occupies the dorsal wall right-anterior to L1 (fig.94). From here it curves ventrad and back to the left (fig.95) into a position beneath the **pne**-pouch; then it extends, becoming narrower, to the posterior edge of the left complex (fig.97). The posterior end of L2 occupies the paa-process, which is upcurved and somewhat hook-like. For most of its length L2 extends broadly along the anterior edge of a pouch (lve in fig.94, 95, 97), which is dorsoventrally curved – in the same way as L2 itself and as it has been described for L2 and lve of Archiblatta. The edge along the bottom of the lve-pouch is labelled 7 in fig.92, 94, 96, 97. Ventrally the invagination of the lve-pouch begins at the posterior right margin of L4K (posterior end of edge 7 in fig.97). Immediately posterior to this point L2 and L4K are in close contact. L2 is restricted to the inner wall of the lve-pouch. The outer lve-wall is completely membranous and is at the same time the dorsal (or left) wall of the vla-lobe (fig.91, 92; in the figures vla is pulled to the right). The ejaculatory duct (D in fig.87, 91, 92) opens far anteriorly into this membrane. In the rightmost dorsal wall of the vla-lobe there lies a ribbon-like extension of the R2-sclerite of the right phallomere (**R2m** in fig.91, 101, 102).

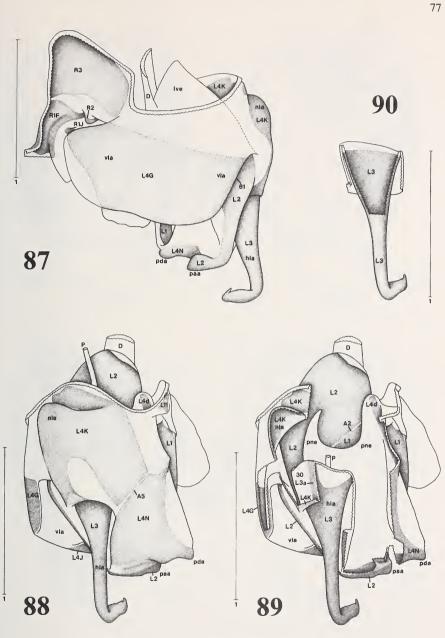
Figs.83,84: *Tryonicus parvus* (Blattaria, Blattidae, Tryonicinae) – **83:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, supraanal lobe, subanal lobes, paraprocts, Pv-sclerites, distal part of rectum, and basal parts of cerci. – **84:** Same as in fig.83, after removal of further parts of abdominal tergites 9 and 10 (especially T10v), parts of paraprocts, and supraanal lobe. Distal part of rectum, basal parts of cerci, and right subanal lobe cut open. Anterior to transverse line: like in fig.83. – Scale: 1mm.





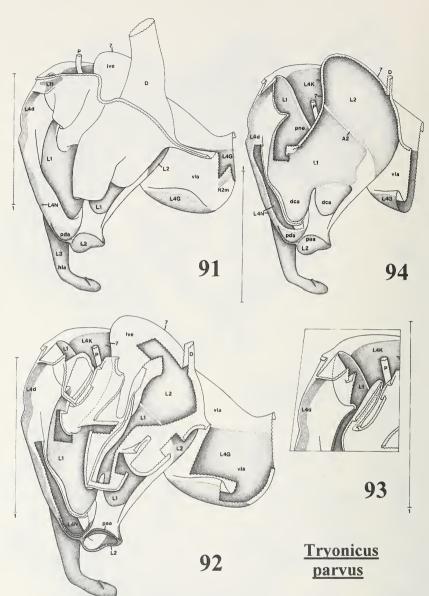
Figs.85,86:

Tryonicus parvus (Blattaria, Blattidae, Tryonicinae) – **85**: Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **86**: Subgenital plate in dorsal view. – Scale: 1mm.

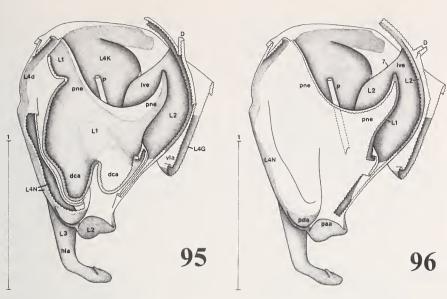


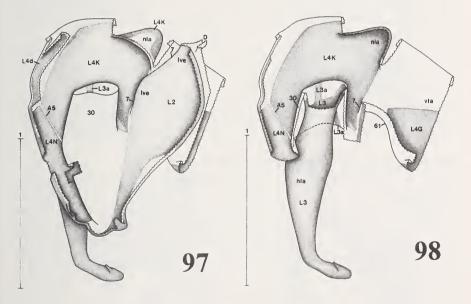
Tryonicus parvus

Figs.87-90: *Tryonicus parvus* (Blattaria, Blattidae, Tryonicinae) – 87: Phallomere complex in ventral view. – 88: Left complex in left-dorsal view. – 89: Left complex in left-dorsal view; some parts removed (compare fig.88). – 90: hla-hook and sclerite L3 in left-dorsal view. – Scale: 1mm.



Figs.91-94: *Tryonicus parvus* (Blattaria, Blattidae, Tryonicinae) – **91:** Left complex in right-dorsal view. (In the text this view is designated as dorsal). – **92-94:** Left complex in right-dorsal view; with successive removal of its parts (mainly of dorsal ones); figs.92,93: scale slightly larger; fig.93: detail from fig.92, some more parts removed. – Scale: 1mm.





Tryonicus parvus

Figs.95-98: *Tryonicus parvus* (Blattaria, Blattidae, Tryonicinae) – Left complex in right-dorsal view; with successive removal of its parts (mainly of dorsal ones). – Scale: 1mm.

The large hook hla is evaginated from the left ventral wall of the left complex. The distal part of hla is sclerotised by L3 (fig.87-90, 97, 98), a large basal part of hla is membranous (30 in fig.97, 98). By introverting this membrane 30, hla can be retracted rather far into the phallomere (the retracted state is shown in the figures). L3 is connected with L4K by a ribbon of weak sclerotisation (L3a in fig.89, 98), which crosses the membrane 30.

Right phallomere

Sclerite **R3** occupies the (right-)anterior ventral wall (fig.99-103). Sclerite **R2** articulates with the left posterior end of **R3** (A7 in fig.100-102). **R2** is a plate of irregular shape, with a ridge-like elaboration in its left-dorsal part (fig.99, 100, 103). The right-ventral part of **R2** has the extension **R2m** (fig.101, 102) mentioned above. Posterior to the central part of **R3** the ventral wall of the right phallomere curves dorsad and somewhat anteriad to form a central invagination (**cbe** in fig.99-101; compare fig.102 and 104).

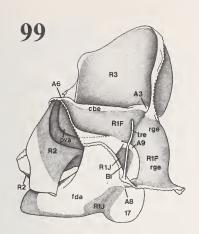
Posterior to the right part of **R3** sclerite **R1F** adjoins, and the two sclerites articulate (**A3** in fig.99, 100, 102). **R1F** extends from its central part behind **A3** in two directions: The ventral arm bends left-dorsad (along edge **16**, fig.102, 104) and largely occupies the right-dorsal wall of the **cbe**-invagination (fig.99, 104). The posterior margin of this arm forms a ridge (**pva** in fig.99, 103). The median end of this arm articulates with the left-dorsal end of **R2** (**A6** in fig.99, 100, 101). The dorsal arm of **R1F** extends posterodorsad (fig.99, 100, 102); its dorsal margin folds back to the right and ventrad to form a sclerotised groove **rge** (fig.99, 102). Behind **A3** and on the ventral arm the sclerotised cuticle is considerably thickened (cross-sections in fig.104).

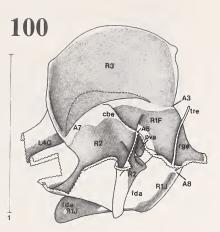
The posterior part of the right phallomere is a large dorsal lobe **fda** (fig.99, 103), whose dorsal and ventral walls are partly occupied by sclerite **R1J**. The dorsal anterior end of **R1J** articulates with the dorsal arm of **R1F** (**A8** in fig.99), its ventral anterior end is in close contact with the base of the ventral arm of **R1F** (**A9** in fig.102, 103). The tendon **tre** (fig.99) originates from the anteriormost dorsal wall of the **fda**-lobe. In its right wall the **fda**-lobe has a large membranous area (**17** in fig.99, 103).

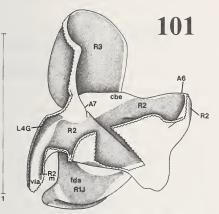
Subgenital plate and posterior abdominal segments

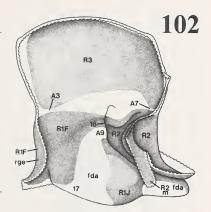
Fig.83, 84 (posterior segments); fig.86 (subgenital plate **S9**). The ventral part of tergite 10 **T10v** is moderately extensive. Strip-like **Pv**-sclerites are present; they are laterally connected with the paraprocts **Pp**. The **Ca**-sclerites are curved ribbons immediately median to the cercal bases (no distinct bulges present). The **Cc**-sclerites are dorsoventrally curved plates on paired shallow outfoldings beneath the anterior margin of **T10v** and above the subanal lobes **sbl**. **Cb**-sclerites are missing. The articulations **A98** and **A99** are well-developed.

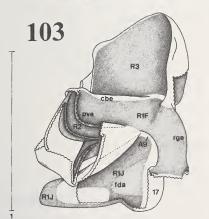
Figs.99-104: *Tryonicus parvus* (Blattaria, Blattidae, Tryonicinae) – **99:** Right phallomere in dorsal view. – **100:** Right phallomere in left-dorsal (somewhat anterior) view. – **101:** Right phallomere in left-ventral view. – **102:** Right phallomere in right-ventral view. – **103:** Right phallomere in dorsal view; some dorsal elements removed. – **104:** Right phallomere in right-ventral view; most elements shown in fig.102 removed. – Scale: 1mm.



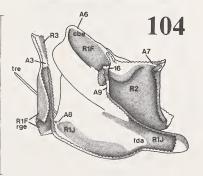


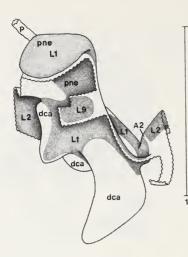


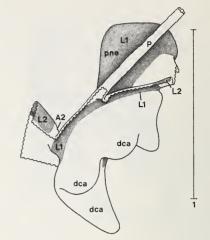




Tryonicus parvus

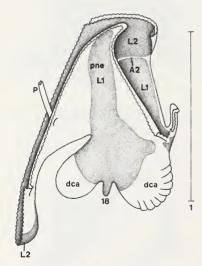




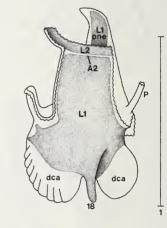




<u>Ergaula</u> <u>capensis</u>







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<u>Tryonicus</u> angustus

5.8. Polyphaga aegyptiaca (Blattaria, Polyphagidae, Polyphaginae)

Left complex

Sclerite L1 is situated in the central dorsal wall; its anterior part lies within a deep pouch (pne in fig.117). Beneath the pne-pouch there is another very deep and broad pouch (lve in fig.118, 122), with the arch-shaped L2-sclerite extending along its edges. The left posterior dorsal wall contains the sclerotisation of two processes (pda and paa in fig.117, 118) and of an intervening invagination (dte in fig.117, 118), which is firmly connected with the L2-sclerotisation in the lve-pouch around the posterior edge of the left complex (10 in fig.118, 122). This dorsal sclerotisation is composed of parts of L2 (right part) and of L4 (left part: L4N). Sclerite L8 lies in the right dorsal wall of the left complex (fig.117). From the left wall of the left complex there protrudes a large hook-process (hla in fig.117) with its L3-sclerite. The large L4M-sclerite occupies the ventral wall (fig.115, 117, 126). Another L4-sclerite lies within the ventral base of the hla-hook (L4K in fig.122-124). The anterior hood-shaped part of L1 largely occupies the walls of the **pne**-pouch (fig.111, 118, 119, 120). In the left-dorsal half of the pne-pouch the walls are membranous especially in the posterior part, where the phallomere-gland (P in fig.118, 120, 121) opens from ventrally. The anterior end of L1 is flat and plateau-like. Posteriorly L1 leaves the pouch and has an arm-like extension on each side (fig.118, 120, 121). The extensions curve ventrad and approach each other again (fig.121). The membranous cuticle enclosed by this (open) sclerite-ring forms two cushion-like evaginations (dca in fig.117, 120, 121). The lve-pouch (fig.118, 122) spans almost the whole breadth of the left complex. (The edge along the bottom of the lve-pouch is labelled 7 in fig.122, 123). L2 occupies the margins of the dorsal (fig.122) and ventral (fig.123) walls of the lve-pouch. The left part of L2 in the dorsal lve-wall broadens posteriorly and bends around the posterior edge of the left complex (along 10 in fig.118, 119a, 122) into the dorsal wall. Here it continues into the sclerotisation of the paa- and pda-processes and of the dte-invagination (fig.117-119b; L2 and L4N, with the border between them somewhere within dte). The pda-process is finger-like; the paa-process is saucer-shaped and partially encloses the dca-processes from left-ventrally. The pda-sclerotisation has a tongue-like extension to the left (L4d in fig.118, 123, 124). The paa-sclerotisation has an arm-like extension to the right (12 in fig.118, 119a). The right parts of L2 and of the lve-pouch curve dorsad and back to the

left (fig.118, 122, 123), and along this curvature the lve-pouch becomes rapidly less deep.

Figs.105-108: **105,106**: *Ergaula capensis* (Blattaria, Polyphagidae, Polyphaginae) – **105**: Sclerite L1 in dorsal pouch pne in dorsal view; with some surrounding elements and phallomere-gland P. – **106**: Sclerite L1 in dorsal pouch pne in ventral view; with some surrounding elements and phallomere-gland P. – **107,108**: *Tryonicus angustus* (Blattaria, Blattidae, Tryonicinae) – **107**: Sclerite L1 in dorsal pouch pne in left-dorsal view; with some surrounding elements and phallomere-gland P. – **108**: Sclerite L1 in dorsal pouch pne in right-ventral view; with some surrounding elements and phallomere-gland P. – **108**: Sclerite L1 in dorsal pouch pne in right-ventral view; with some surrounding elements and phallomere-gland P. – **108**: Sclerite L1 in dorsal pouch pne in right-ventral view; with some surrounding elements and phallomere-gland P. – **108**: Sclerite L1 in dorsal pouch pne in right-ventral view; with some surrounding elements and phallomere-gland P. – **108**: Sclerite L1 in dorsal pouch pne in right-ventral view; with some surrounding elements and phallomere-gland P. – **108**: Sclerite L1 in dorsal pouch pne in right-ventral view; with some surrounding elements and phallomere-gland P. – Scale: 1mm.

This recurved dorsal part of the **lve**-pouch, with its dorsal and ventral walls sclerotised by **L2**, approaches **L1** and articulates with it (**A2** in fig.118, 120, 121). The ventral wall of the **lve**-pouch also shows this dorsoventral curvature, but it is in its posterior part additionally invaginated to the right, and the invagination is strengthened by an arm-like extension of **L2** (**11** in fig.118, 122, 123). The posterior end of this **L2**-extension is, around an edge, in close contact with the posterior margin of sclerite **L8** (fig.117, 118).

The ventral wall of the **lve**-pouch is, except for the **L2**-sclerotisations along its margins, membranous; it is at the same time the anterior dorsal wall of the very broad ventral lobe **vla** (fig.123). The ejaculatory duct (**D** in fig.123, 124) opens into the right part of this membrane. Dorsal to the genital opening there are some membranous outfoldings (**goa** in fig.122-124). Sclerite **L5** lies in the left dorsal wall of the **vla**-lobe and is in close contact with the posterior margin of **L2** (fig.123, 124).

The **hla**-hook (with **L3**) is evaginated from the left wall of the left complex (fig.117). The base of **hla** is rather complicated (fig.122-125a) and contains sclerite **L4K** in its posteroventral part. **L4K** shows a horseshoe-like dorsoventral curvature, with a broad ventral and a pointed dorsal part. The rightmost part of the left complex is the lobe **lba** with sclerite **L7** in its ventral wall (fig.115, 117, 118). The **lba**-lobe is distinctly separated from the **vla**-lobe.

Right phallomere

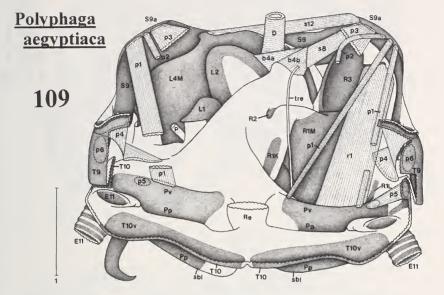
Sclerite **R3** occupies the anterior (right-)ventral wall (fig.134-137). The right margin of **R3** forms a groove-like apodeme **age** (fig.134, 137). Posterior to the left part of **R3** sclerite **R2** adjoins (fig.135-137), and the two sclerites are fused; a strip of weaker sclerotisation is probably the suture (A7* in fig.135-137). Along its ventral margin **R2** forms a ridge bearing several processes (fig.136, 137, 141), the largest of which is behind A7*. Posterior to the central part of **R3** the ventral wall of the right phallomere curves dorsad and somewhat anteriad to form a sclerotised central invagination (**cbe** in fig.134-136; compare fig.137 and 138).

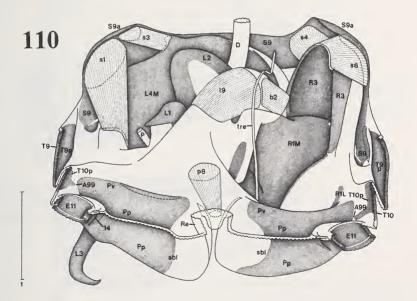
 $[\]longrightarrow$ p.85

Figs.109,110: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – **109:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, subanal lobes, paraprocts, distal part of rectum, basal parts of cerci, and part of musculature. – **110:** Same as in fig.109, after removal of further parts of abdominal tergites 9 and 10 (especially T10v). Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. – Scale: 1mm.

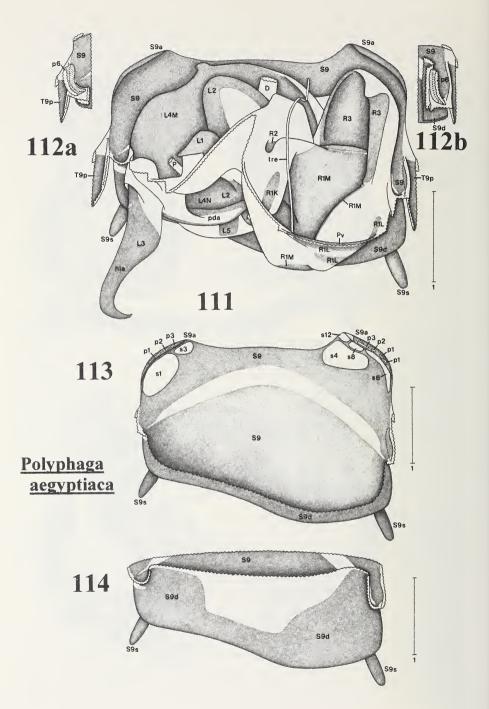
 $[\]longrightarrow$ p.86

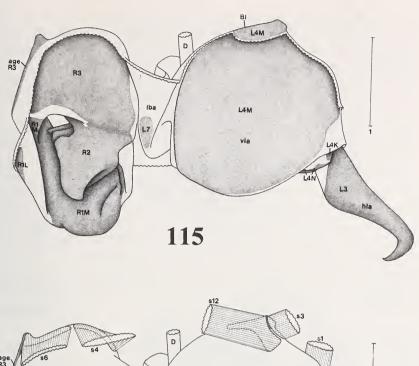
Figs.111-114: Polyphaga aegyptiaca (Blattaria, Polyphagidae, Polyphaginae) – 111: Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – 112a,b: Left (a) and right (b) margins of subgenital plate (compare fig.111); with insertions of muscles p6. – 113: Subgenital plate in dorsal view; with insertion areas of muscles (except p6); most of dorsal sclerotisation S9d of subgenital plate removed. – 114: Subgenital plate in dorsal view; anterior part of ventral sclerotisation removed, dorsal sclerotisation S9d complete. – Scale: 1mm.

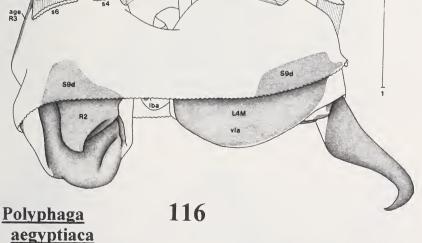












Figs.115,116: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – 115: Phallomere complex in ventral view. – 116: Phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.115, with parts of dorsal sclerotisation S9d of subgenital plate in its posterior part (compare fig.114). – Scale: 1mm.

Posterior to the right part of **R3** the large **R1M**-sclerite adjoins, and the two sclerites articulate (**A3** in fig.134, 135, 137). From its central part behind **A3** sclerite **R1M** extends to the left, where it bends left-dorsad (along edge **16**; fig.137, 138) and then occupies the right-dorsal wall of the **cbe**-invagination (fig.134). The left-ventral wall and the top of **cbe** are sclerotised by a plate-like part of **R2** (fig.134-137); the dorsal margin of this **R2**-part is fused to the anterior margin of **R1M**. A line of weaker sclerotisation (**13** in fig.134, 138) is probably the boundary between **R2** and **R1M**.

From its fusion line 13 with R2 and from articulation A3, R1M extends far posteriad as a dorsoventrally curved plate of irregular shape (fig.134, 137). It largely occupies the ventral and right walls of the posterior part of the right phallomere (labelled fda and pva in fig.134-138). The dorsal margin of R1M folds back to the right to form a sclerotised groove (rge in fig.134, 138, 140); rge is a posterior continuation of the age-groove on R3 and extends to the posterior edge of the fda-lobe. Dorsal to rge there is an outfolding to the right, which contains the very weak ribbon-like sclerites R1L (fig.134). At the left end of the fda-lobe there is a dorsal outfolding to the left, which contains sclerite R1K in its ventral wall (fig.134). Beneath R1K the left marginal part of R1M forms a longitudinal ridge projecting to the left (pva, compare fig.134 and 139). The dorsal wall of the fda-lobe is mostly membranous (fig.134), and most anteriorly the tendon tre has its origin (fig.134, 135, 139).

Subgenital plate and posterior abdominal segments

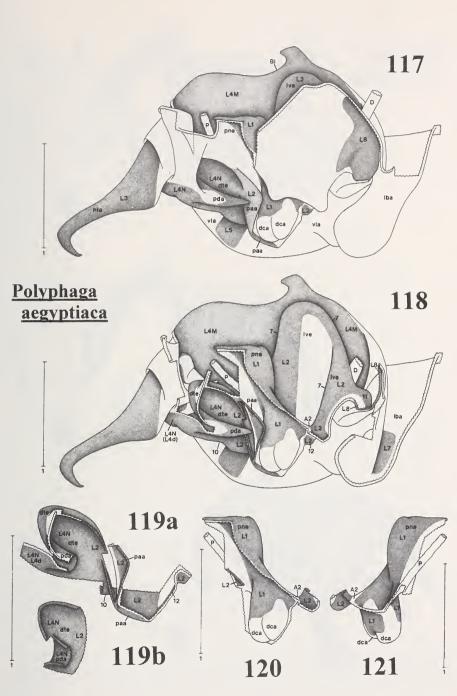
Fig.109, 110 (posterior segments); fig.113, 114 (subgenital plate **S9**). The entire tergite 10 **T10**, including its ventral part **T10v**, is divided along its midline. **T10v** is moderately extensive. Separate **Pv**-sclerites are missing; they are assumed to have been incorporated into the anterior margins of the paraprocts **Pp**. The sclerites **Ca**, **Cb**, and **Cc** are missing. The bulges next to the cercal bases the **Ca**-sclerites lie upon in other species, however, are present (compare fig.59). The articulations **A99** are well-developed. The articulations **A98** are missing: the sclerotisations **E11** and **T10** are far away from each other. Each subanal lobe **sbl** has a small groove (**14** in fig.110) beneath the cercal base.

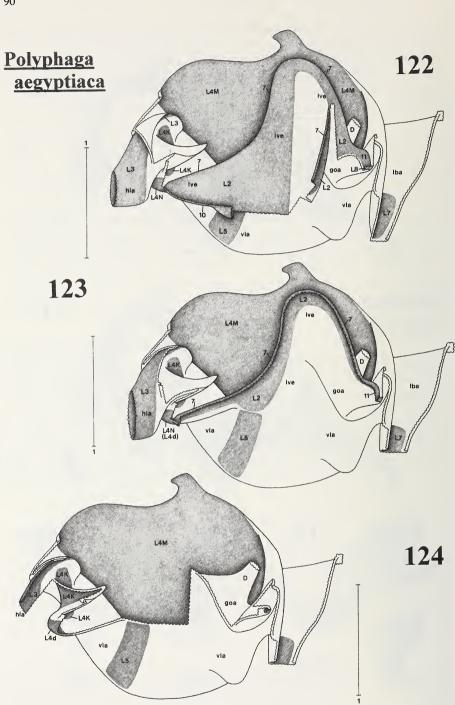
^{——→} p.89

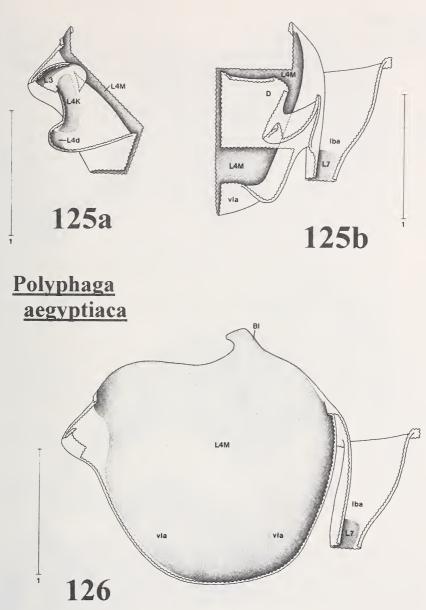
Figs.117-121: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – 117: Left complex in dorsal view. – 118: Left complex in dorsal view; some parts removed (mainly dorsal ones). – 119a,b: Dorsal parts of sclerites L4 and L2, separated from remainder of left complex, in dorsal view, – 120: Sclerite L1 in dorsal pouch pne, separated from remainder of left complex, in dorsal view; with some surrounding elements and phallomere- gland P. – 121: Sclerite L1 in dorsal pouch pne, separated from remainder of left complex, and phallomere-gland P. – Scale: 1mm.

[→] p.90

Figs.122-124: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – Left complex in dorsal view; with further successive removal of its parts (mainly of dorsal ones); fig.122: left complex after removal of the parts shown in fig.119-121. – Scale: 1mm.







Figs.125,126: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – **125a,b:** Left (a) and right (b) part of left complex, separated from remainder of left complex, in dorsal view; further parts removed (mainly dorsal ones; compare fig.124). – **126:** Ventral wall of left complex in dorsal view. – Scale: 1mm.

Musculature

l2 L1 (plateau-like anterior end of pne -pouch	n) – L4M (left anterior part) 1	27 129 120 121
· · · · ·		27, 128, 129, 131
13 L1 (right-ventrally on pne-pouch) – L2 (right anterior part) 1	28, 130, 131
14 L2 (left anterior part) – L4K (dorsal righ	t end, within base of	
hla-hook)		29, 130, 132
l5 $L4M$ (anterior part) – $L2$ (left part) and r	membrane to the left of it 1	30, 133
l6a L4M (anterior part) – L2 (right anterior p	part) 1	30, 133
16b L4M (posterior to 16a) – ejaculatory duct	D next to its opening 1	.32
19 Transversely within right dorsal wall of le	eft complex; ventral parts:	
L1 - L2 (areas next to articulation A2)	1	10, 127, 129, 130
l10 L2 (left posterior part) – L4N and L2 on	invagination dte 1	29, 130
l11 L4K (anteroventral part) – L4d (= left pa		28, 132
l12 L2 (rightmost part) – L8; very short and	stout 1	28, 129, 130
113 Ejaculatory duct D next to its opening (ve	entral wall) – dorsal wall	
of vla-lobe (rightmost part)	1.	32
r1 R3 (right margin) – R1M (dorsal margin,	on posterior part of	
rge-groove), R1L, and membrane in betw	veen these sclerotisations 1	09, 139
r2 R3 – cbe-invagination: R1M (anterior ma	rgin) and R2	
(right-dorsal part)	1.	40, 141
r6 R1M (dorsal margin, on rge-groove) – R	1K and surrounding	
membranes	1.	40
r9 R3 (left-posterior part) – R2 (left-ventral	part) 1-	41
b2 Right dorsal wall of left complex (next to	L8) – R3	
(left-posterior part)	1	10, 127, 141
b4a L2 (right anterior edge of lve-pouch) – tr	e-tendon 1	09, 127, 129,
	1:	30, 139
b4b L8 (anterior margin) – tre-tendon	1	09, 127, 139
s1 S9 left side (laterally and anteriorly) – me	embrane anterior to	
hla-hook	1	10, 113, 116, 127

\longrightarrow p.93

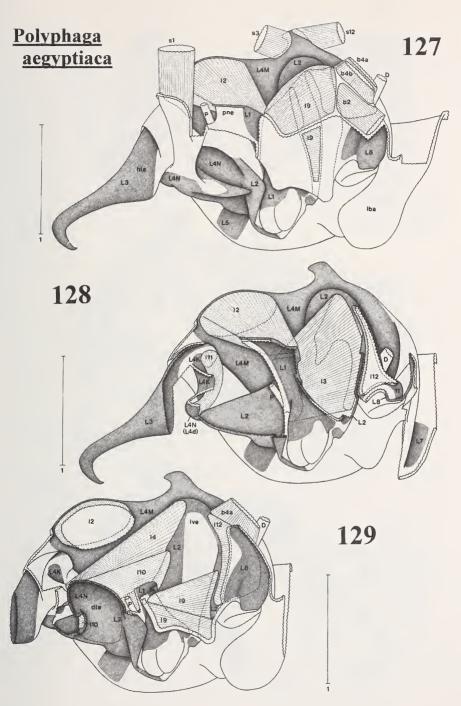
Figs.127-129: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – Scale: 1mm.

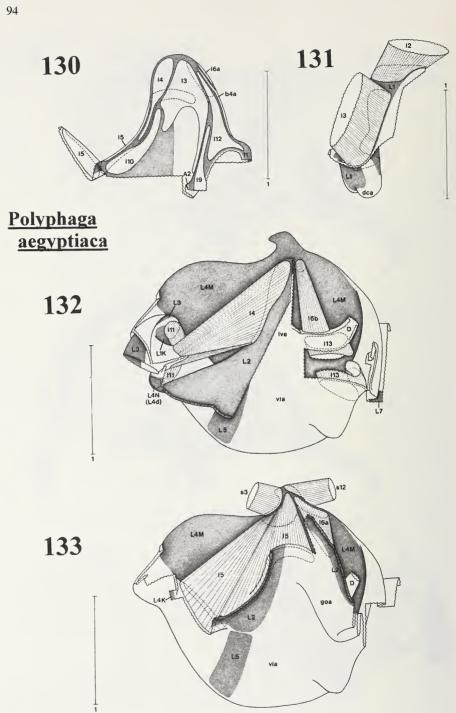
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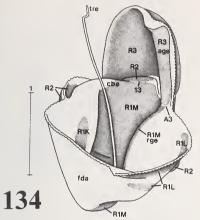
Figs.130-133: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – **130:** Sclerite L2 in ventral pouch lve in dorsal view; with insertion areas (white) of muscles (compare fig.122). – **131:** Sclerite L1 in dorsal pouch pne in ventral view; with muscles l2 and l3 (compare fig.121). – **132,133:** Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – Scale: 1mm.

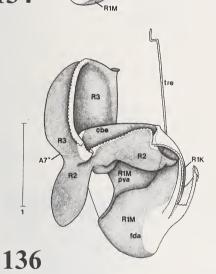
 \rightarrow p.95

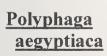
Figs.134-138: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – **134:** Right phallomere in dorsal view. – **135:** Right phallomere in left-dorsal (somewhat anterior) view. – **136:** Right phallomere in left-ventral view. – **137:** Right phallomere in right-ventral view. – **138:** Right phallomere in right-ventral view; most elements shown in fig.137 removed. – Scale: 1mm.

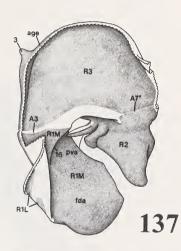


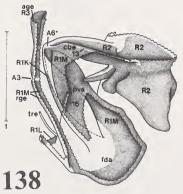












age tre

R3

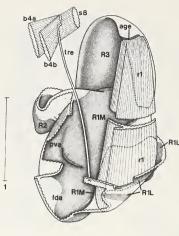
R1M

fda

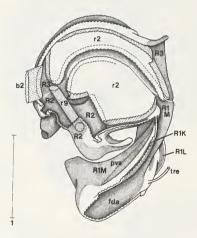
135

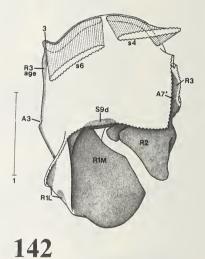
çbe

R2



139





141

Polyphaga aegyptiaca

Figs.139-142: *Polyphaga aegyptiaca* (Blattaria, Polyphagidae, Polyphaginae) – **139,140:** Right phallomere in dorsal view; each figure with some muscles; dorsal elements removed to various extents. – **141:** Right phallomere in left-dorsal (somewhat anterior) view; with some muscles; some left-dorsal elements removed. – **142:** Right phallomere in right-ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.137. – Scale: 1mm.

s3	S9 left side (most anteriorly, median to $s1$) – L4M (anterior margin)	110, 113, 116,
		127, 133
s4	S9 right side (anteriorly) – R3 (left anterior margin)	110, 113, 116, 142
s6	S9 right side (laterally and anteriorly) $- R3$ (right anterior margin)	110, 113, 116, 142
s8	S9 right side (anteriorly) – tre-tendon	109, 113, 139
s12	S9 right side (most anteriorly, median to s8) - L4M (anterior margin)	109, 113, 116,
		127, 133
p1 (pair)	S9 - paraproct Pp (anterior margin, corresponding to	
	Pv-sclerotisation); right muscle divided into two bundles	109, 113
p2 (pair)	S9 – T9 (lateral anterior margin)	109, 113
p3 (pair)	S9 – rectum (ventral wall)	109, 113
p4 (pair)	T9 (lateralmost anterior margin) – membrane anterior to paraproct	
	Pp (far laterally); right muscle inserting on lateral wall of genital	
	pouch, next to right phallomere; both muscles twisted	109
p5 (pair)	T10 (lateralmost anterior margin) - paraproct Pp (lateral anterior	
	margin, corresponding to the Pv-sclerotisation)	109
p6 (pair)	T9 (lateralmost part) - S9 (lateral margin) (left muscle only) and	
	adjacent membranes; ventral insertion of right muscle extending far	
	anteriad into right wall of genital pouch, lying immediately anterior	
	to insertion of p4	109, 112a,b
p8	Longitudinally within membrane between Pv-sclerotisations	110

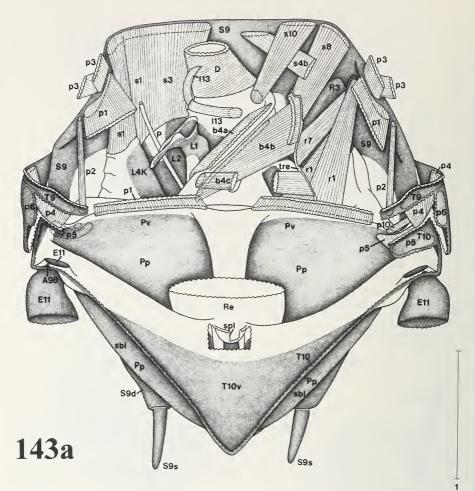
5.9. Cryptocercus punctulatus (Blattaria, Cryptocercidae)

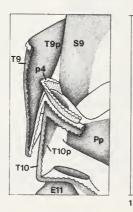
Left complex

L4 is a group of four sclerites: The ribbon-shaped L4N (fig.150; L4d is part of L4N) lies on a transverse outfolding of the dorsal wall, whose right end is somewhat lobe-like (**paa** in fig.150). The ventral wall of this outfolding extends far anteriad, where L4K takes its position. The small L4P, not present in all specimens, lies at the anteriormost left edge of the left complex. L4G is a plate in the posterior ventral wall of the ventral lobe **vla** (= ventral phallomere; fig.148, 152).

Sclerite L1 lies in the central dorsal wall. Its hood-shaped anterior part occupies most of the walls of a deep pouch (**pne** in fig.150-154). The anterior summits of L1 and **pne** are expanded and plateau-like, with upcurved margins. The posterior part of L1 leaves the **pne**-pouch and has an arm-like extension on each side. These extensions curve ventrad and join each other again to form a complete sclerite ring (fig.153, 154). The membranous cuticle enclosed by this ring forms two cushion-like bulges **dca**, with a small sclerotised peak between them (**18** in fig.153). The phallomere-gland (**P** in fig.152, 153) opens into the membrane to the left of L1.

The large sclerite L2 lies ventral to L1 (fig.152). To the posterior L2 becomes narrower, and then it curves around the posterior edge of the left complex into the dorsal wall (fig.151, 152). The area of this curvature forms a large bulge (paa in fig.150-152). In the dorsal wall L2 extends anteriad as far as to the opening of the phallomere-gland. The right anterior part of L2 lies in the dorsal wall of a pouch (lve in fig.150-152; the edge along





Cryptocercus punctulatus

143b

the bottom of the pouch is labelled 7). The broad ejaculatory duct (D in fig.150, 151) opens most anteriorly into this lve-pouch.

The ventral wall of the **lve**-pouch is at the same time an anterior part of the dorsal wall of the **vla**-lobe. The ribbon-shaped sclerite **L5** (fig.151, 152) lies more posteriorly in the dorsal **vla**-wall. The ventral **vla**-wall is membranous anteriorly and sclerotised by **L4G** posteriorly.

The large hook **hla** (fig.150, 151), whose distal part is sclerotised by L3, is evaginated from the left wall of the left complex – beneath and somewhat posterior to L4K.

Right phallomere

Sclerite **R3** occupies the anterior (right-)ventral wall (fig.160-163). The lateral and anterior margins of **R3** form a weakly sclerotised groove-like apodeme **age** (fig.160, 163). Along the right margin of **R3** the ventral sclerotisation of **age** folds to the left (**19** in fig.163, 164). Sclerite **R2** articulates with the left part of **R3** (**A7** in fig.161-163). **R2** has the shape of a ridge (fig.161, 162). Posterior to the central part of **R3** the ventral wall of the right phallomere curves dorsad and somewhat anteriad to form a central invagination (**cbe** in fig.160-162; compare fig.163 and 164).

Posterior to the right part of **R3** sclerite **R1F** adjoins, and the two sclerites articulate (**A3** in fig.160, 161, 163). From its central part behind the **A3**-articulation **R1F** extends in two directions: The ventral arm bends left-dorsad (behind the edge **16** in fig.163, 164) and extends into the right-dorsal wall of the **cbe**-invagination (fig.160, 164). This part of **R1F** bulges outwards (**pva** in fig.163, 164) by the cuticle being extensively thickened (cross-section in fig.164). The median end of this arm articulates with the left-dorsal end of **R2** (**A6** in fig.160, 164). The dorsal arm of **R1F** extends posterodorsad (fig.160, 163); anteriorly its dorsal margin folds back to the right to form a sclerotised groove (**rge** in fig.160, 163), which is a posterior continuation of the **age**-groove on **R3**.

The posterior part of the right phallomere is a large dorsal lobe **fda** (fig.160-163, 166), whose dorsal and ventral walls are partly occupied by sclerite **R1J**. The dorsal anterior tip of **R1J** approaches the dorsal arm of **R1F** (**A8** in fig.160), its ventral anterior tip approaches the base of the ventral arm of **R1F** (**A9** in fig.163, 166). The tendon **tre** has its origin in the anteriormost dorsal wall of **fda** (fig.160). In its right wall **fda** has a large membranous area (**17** in fig.160, 166). At the left end of **fda** there is another small sclerite **R1K** (fig.160).

Figs.143a,b: *Cryptocercus punctulatus* (Blattaria, Cryptocercidae) – **a:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, supraanal lobe, subanal lobes, paraprocts, distal part of rectum, basal parts of cerci, and part of musculature. Supraanal lobe shown through a window cut into the membrane anterior to ventral sclerotisation of abdominal tergite 10 T10v. – **b:** Posterior insertion of muscle p4 at anterior margin of abdominal tergite 10 T10. Enlarged detail from left part of fig.143a, further parts of abdominal tergite 9 T9 and anterior part of p4 removed. – Scale: 1mm.

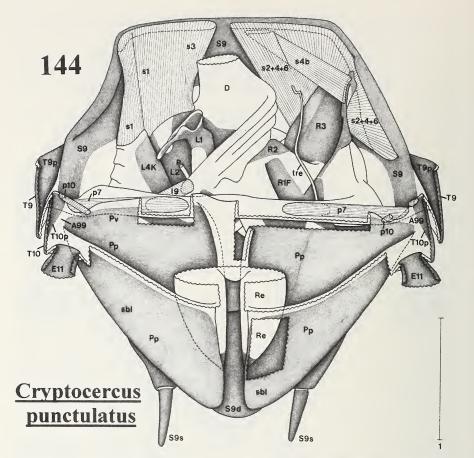


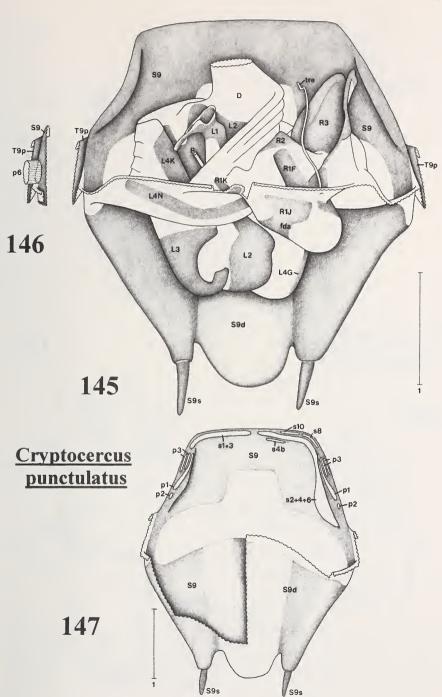
Fig.144: *Cryptocercus punctulatus* (Blattaria, Cryptocercidae) – Male postabdomen as in fig.143a, after removal of further parts of abdominal tergites 9 and 10 (especially T10v) and supraanal lobe. Distal part of rectum, basal parts of cerci, dorsal wall of right subanal lobe, and anterior margins of paraprocts cut open. – Scale: 1mm.

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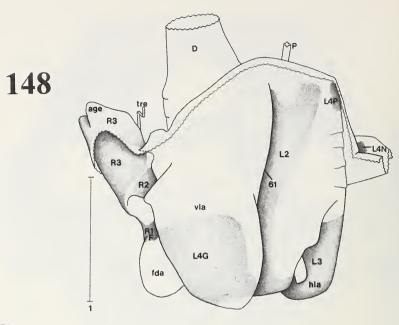
 \longrightarrow p.102

Figs.148,149: *Cryptocercus punctulatus* (Blattaria, Cryptocercidae) – **148:** Phallomere complex in ventral view. – **149:** Phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.148. – Scale: 1mm.

Figs.145-147: *Cryptocercus punctulatus* (Blattaria, Cryptocercidae) – **145:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **146:** Left margin of subgenital plate (compare fig.145); with insertion of muscle p6. - 147: Subgenital plate in dorsal view; with insertion areas of muscles (except p6); dorsal sclerotisation S9d of subgenital plate complete in the right part but largely removed in the left part. – Scale: 1mm.

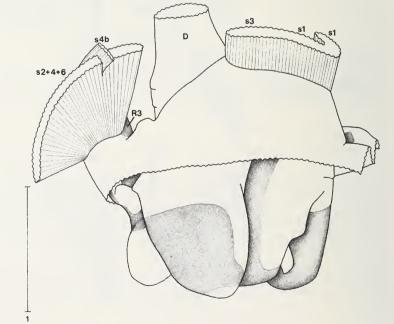


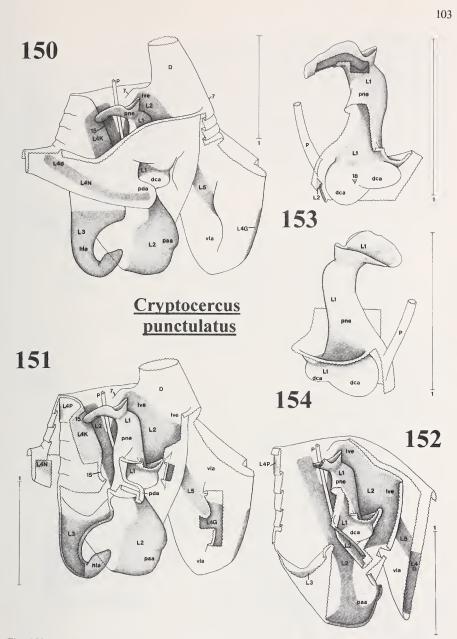




<u>Cryptocercus</u> <u>punctulatus</u>







Figs.150-154: Cryptocercus punctulatus (Blattaria, Cryptocercidae) – 150: Left complex in dorsal view. – 151, 152: Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones); fig.152: pne-pouch with some adjacent parts completely cut off from the other elements. – 153: Sclerite L1 in dorsal pouch pne in dorsal view; with some surrounding membranes, part of sclerite L2, and phallomere-gland P. – 154: Sclerite L1 in dorsal pouch pne in ventral view; with some surrounding membranes and phallomere-gland P. – Scale: 1mm.

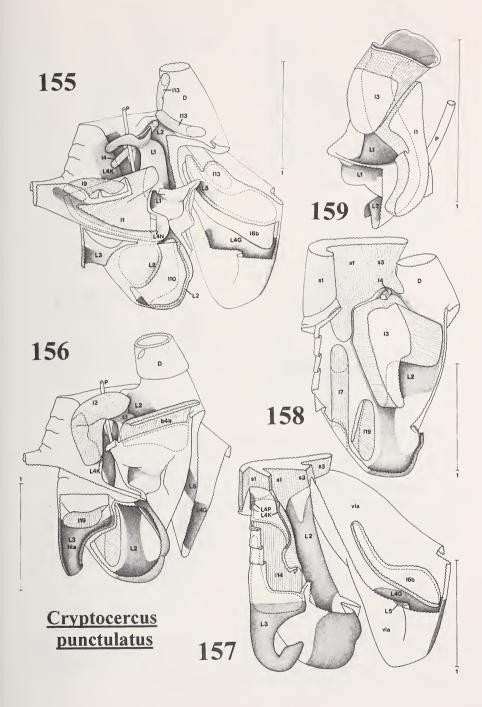
Subgenital plate and posterior abdominal segments

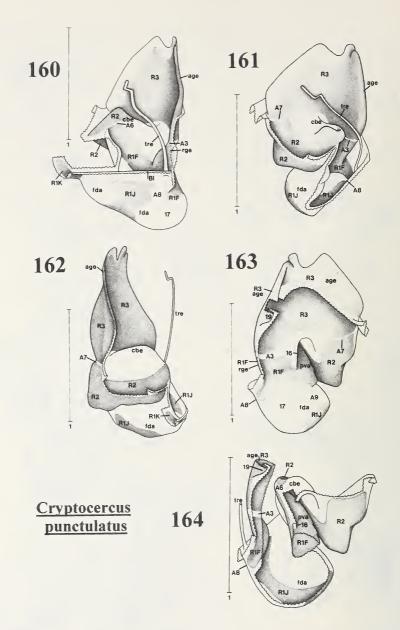
Fig.143a,b. 144 (posterior segments); fig.147 (subgenital plate **S9**). The whole postabdomen is retracted anteriad and completely covered by the heavily sclerotised tergite and sternite of abdominal segment 7. The tergal and sternal sclerotisations of the postabdomen are rather weak. The ventral part of tergite 10 **T10v** is rather extensive. Separate **Pv**sclerites are missing; they are assumed to have been incorporated into the anterior margins of the paraprocts **Pp**. The sclerites **Ca**, **Cb**, and **Cc** are missing, and there are no **Ca**bulges (compare fig.59). The articulations **A98** are well-developed. **A99** are not true articulations since the contact between paratergite **T10p** and paraproct **Pp** is not very close.

Musculature

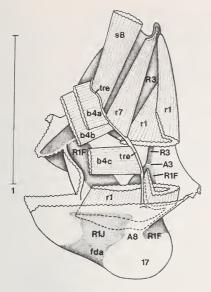
Muscles	Positions of insertions	in fig.
11	Membranous posterior dorsal wall of pne-pouch – L4N	
	(including L4d) and adjacent membranes	155, 159
12	L1 (plateau-like anterior end of pne-pouch) – L4K	156
13	L1 (right-ventrally on pne- pouch) – L2 (most of anterior half)	158, 159
14	L2 (right anterior part) – L4K; very delicate	155, 158
16b	L4G (anterior margin) and membrane anterior to L4G – dorsal wall	
	of vla-lobe posterior to genital opening (in part on L5)	155, 157
17	Left ventral wall of left complex - left posterior edge of left complex	
19	Transversely within anterior left dorsal wall of left complex	144, 155
110	L2 (posteriormost part, on paa-process) – membrane left-dorsal	
	to paa-process	155
113	Ejaculatory duct D next to its opening (dorsal wall) – anterior dorsal	140 155
11.4	wall of vla -lobe; anterior part of muscle divided into two bundles.	143a, 155
114	L4K and membrane anterior to base of hla-hook – hla-hook	157
119	(dorsal anterior margin of L3 and membrane anterior to it)	157
119	Left posterior ventral wall of left complex – hla -hook (ventral anterior margin of L3 and membrane anterior to it)	156, 158
		150, 158
r1	R3 (right margin) – anterior dorsal wall of fda-lobe, in part on	140 164 165
	anterior margin of R1J and on base of tre -tendon	143a, 164, 165
r2	R3 – cbe-invagination: R1F (anterior margin and left part),	1.66 1.67
	membrane, and R2 (dorsal margin)	166, 167
r3	R1F (dorsal and central parts, posterior to articulation A3) – R1J	166 167 169
	(right margin)	166, 167, 168
r7 	R3 (right anterior part) – tre-tendon	143a, 165 167
r8	Both insertions on central part of R3	
b4a	Right dorsal wall of left complex – tre -tendon	143a, 156, 165
b4b	Right dorsal wall of left complex – tre -tendon	143a, 165
b4c	Central dorsal wall of left complex – tre-tendon	143a, 165

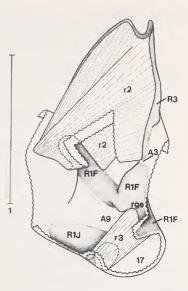
Figs.155-159: *Cryptocercus punctulatus* (Blattaria, Cryptocercidae) – **155-158:** Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – **159:** Sclerite L1 in dorsal pouch pne in ventral view; with muscles 11 and 13 (compare fig.154). – Scale: 1mm.



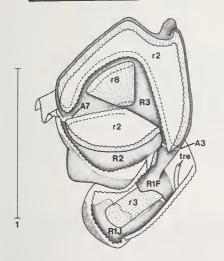


Figs.160-164: Cryptocercus punctulatus (Blattaria, Cryptocercidae) – 160: Right phallomere in dorsal view. – 161: Right phallomere in left-dorsal (somewhat anterior) view. – 162: Right phallomere in left-ventral view. – 163: Right phallomere in right-ventral view. – 164: Right phallomere in right-ventral view; most elements shown in fig.163 removed. – Scale: 1mm.





Cryptocercus punctulatus



s1+3(+7?)	S9 left side (most anteriorly; only medially) – ventral basal line B1 of left complex and membrane anterior to base of hla -hook; right	
	part of muscle inserting also on anterior margin of $L2 (= s7?)$	143a, 144, 147 149, 157, 158
s2+4+6	S9 right side (anteriorly; medially and laterally) $- R3$ (anterior	
	margin) and anterior ventral wall of genital pouch	144, 147, 149, 168
s4b	S9 right side (medially and anteriorly) $- R3$ (right anterior margin);	
	present in some specimens only	143a, 144, 147 149, 168
s8	S9 right side (most anteriorly) – tre-tendon	143a, 147, 165
s10	S9 right side (medially and most anteriorly) – ejaculatory duct D	
	next to its opening (right wall)	143a, 147
p1 (pair)	S9 – membrane anterior to paraproct Pp or Pv-sclerotisation;	
	very broad	143a, 147
p2 (pair)	S9 – T9 (lateral anterior margin)	143a, 147
p3 (pair)	S9 – rectum (ventral wall); divided into two groups of fibers on	
	both sides	143a, 147
p4 (pair)	T9 (lateralmost anterior margin, also extending onto paratergite	
	T9p) – T10 (lateralmost anterior margin, also extending onto	
	paratergite T10p)	143a,b
p5 (pair)	$T10\ (lateral\ anterior\ margin)$ – paraproct $Pp\ (lateral\ anterior\ margin,$	
	corresponding to Pv-sclerotisation)	143a
p6 (pair)	T9 (lateralmost part) – S9 (lateral margin)	143a, 146
p7 (pair)	Posteriad-directed outfolding anterior to paraproct Pp - membrane	
	anterior to "articulation" A99 (between paratergite T10p and	
	paraproct Pp)	144
p10 (pair)	Paratergite T10p (anterior margin) – paraproct Pp (lateral anterior	
	margin, corresponding to Pv-sclerotisation)	143a, 144

5.10. Lamproblatta albipalpus (Blattaria, Blattidae, Lamproblattinae)

Left complex

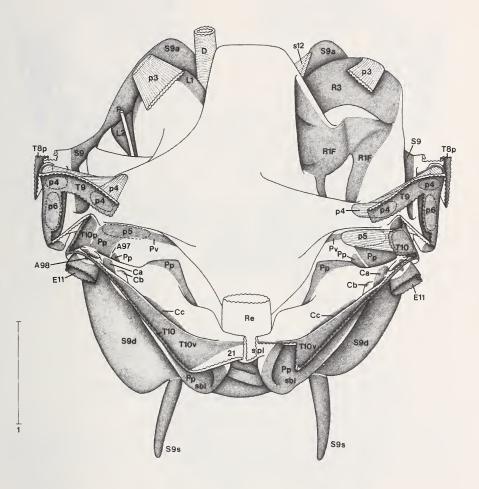
Sclerite L1 is situated in the right anterior dorsal wall; its anterior part lies within a large pouch **pne** (fig.177, 178). Posterior to L1 there is a complicated sclerotisation (L4T and L2C: fig.177-179, 182) bearing two processes **pda** and **paa**. Beneath these elements there is a large pouch **lve** (fig.180) containing the sclerites L2A and L2B in its dorsal wall. The

←____ p.107

Figs.165-168: *Cryptocercus punctulatus* (Blattaria, Cryptocercidae) – **165:** Right phallomere in dorsal view; with some muscles. – **166:** Right phallomere in dorsal view; with some muscles; some dorsal elements removed. – **167:** Right phallomere in left-dorsal (somewhat anterior) view; with some muscles; some left-dorsal elements removed. – **168:** Right phallomere in right-ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.163; fda-lobe cut open to show muscle r3. – Scale: 1mm.

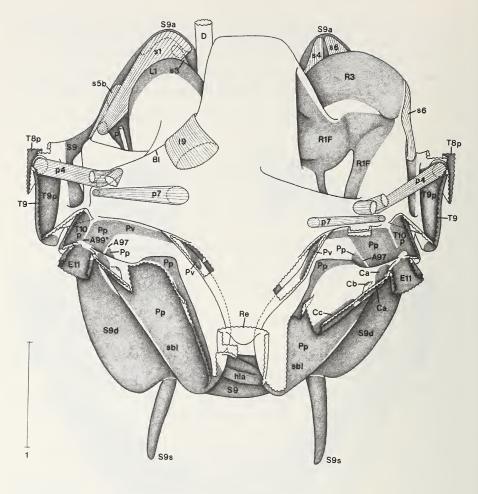
large hook-process **hla** (fig.177) with its sclerite **L3** protrudes from the left wall. Sclerite **L4K** takes a position dorsal to the base of **hla** (fig.177). **L4R** is the sclerotisation of the ventral wall of a broad ventral lobe **vla** (= ventral phallomere; fig.174, 181).

The **pne**-pouch (fig.177, 178) is deep but rather flat. The anterior part of L1 occupies the ventral wall of the **pne**-pouch and its anterior dorsal wall. The posterior part of L1



169 Lamproblatta albipalpus

Fig.169: *Lamproblatta albipalpus* (Blattaria, Blattidae, Lamproblattinae) – Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 8, 9, and 10, supraanal lobe, subanal lobes, paraprocts, Pv-sclerites, distal part of rectum, basal parts of cerci, and part of musculature. – Scale: 1mm.

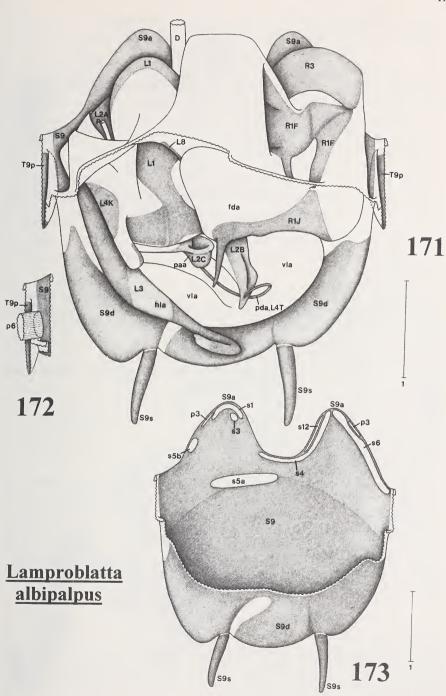


170 Lamproblatta albipalpus

Fig.170: *Lamproblatta albipalpus* (Blattaria, Blattidae, Lamproblattinae) – Male postabdomen as in fig.169, after removal of further parts of abdominal tergites 9 and 10 (especially T10v), parts of right paraproct, and supraanal lobe. Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. – Scale: 1mm.

 $[\]longrightarrow$ p.111

Figs.171-173: Lamproblatta albipalpus (Blattaria, Blattidae, Lamproblattinae) – **171:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **172:** Left margin of subgenital plate (compare fig.171); with insertion of muscle p6. – **173:** Subgenital plate in dorsal view; with insertion areas of muscles (except p6). – Scale: 1mm.



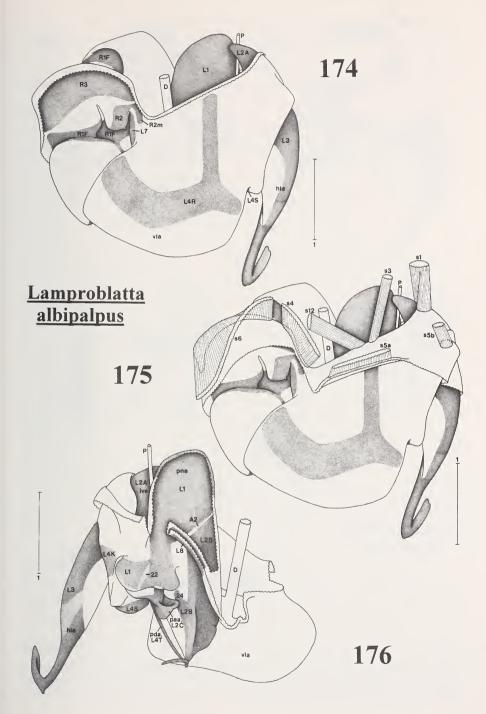
sclerotises a process (dca in fig.177, 178) and has a plate-like extension to the left, which is separated from the main part of L1 by a strip of weaker sclerotisation (22 in fig.176, 177).

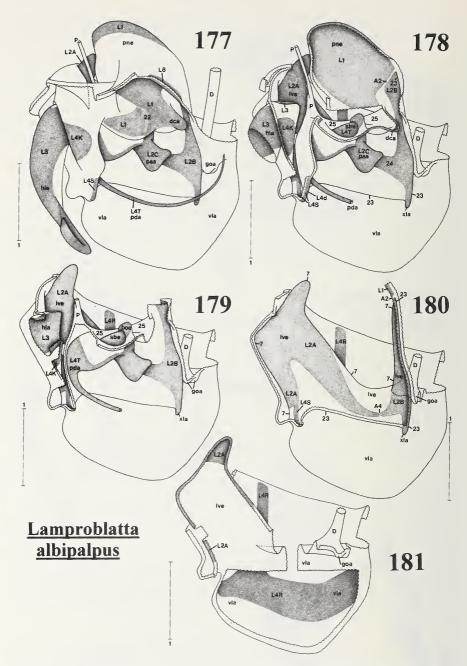
The complex sclerite posterior to L1 is composed of L4T – the sclerotisation of the spineshaped pda – and L2C – the sclerotisation of the cup-shaped **paa** (fig.176-179). **pda** is almost completely sclerotised in its dorsal wall but only basally in its ventral wall (fig.179, 182), and it resembles a hypodermic needle: At its pointed end (26 in fig.182) the cuticle is invaginated to form a very narrow channel (sbe in fig.182) which runs back through the whole spine and whose end is expanded and bulb-like (sbe in fig.182, 183; possibly the reservoir of a gland). The **paa**-process is completely sclerotised. At its right base the cuticle is deeply invaginated to form a heavily sclerotised hood-shaped apodeme (boe in fig.179, 182, 183) which caps the right end of the sbe-bulb (fig.182). The membrane (25 in fig.178, 179, 182) that adjoins this L4T+L2C-sclerite ventrally is somewhat invaginated anteriad, and here the phallomere-gland opens (P in fig.178, 179). Ventral to and to the left of this invagination the membrane extends posteriad towards the transverse edge 23, along which the cuticle bends ventrad and anteriad to continue into the dorsal wall of the **lve**-pouch (fig.180).

The lve-pouch spans almost the whole breadth of the left complex. (The edge along the bottom of the pouch is labelled 7 in fig.180). Its dorsal wall is largely occupied by the sclerites L2A (left part) and L2B (right part), which articulate with each other (A4 in fig.180). In the area around A4 the lve-pouch has a very deep recess from anteriorly. The right part of L2B bends dorsad and back to the left along the longitudinal part of edge 23 (compare fig.178 and 180). This dorsal part of L2B articulates with L1 anteriorly (A2 in fig.176, 178, 180); posteriorly it has an extension to the left (24 in fig.176, 178). L2A extends like an arch along the margins of the left dorsal lve-wall. Only in the anteriormost part of the lve-pouch the sclerite abruptly narrows, leaves the pouch (sclerotisation now designated L4S, fig.178, 180, with L4d as its distalmost part), and curves into the dorsal wall of the left complex (fig.177, 178).

The membranous ventral wall of the **lve**-pouch is at the same time the anterior dorsal wall of the **vla**-lobe (fig.174, 180, 181). The ejaculatory duct (**D** in fig.178-181) opens far on the right into this membrane. Dorsal to the genital opening there is a small membranous outfolding (**goa** in fig.177, 179-181). **L4R** in the ventral wall of the **vla**-lobe is a transverse plate with a ribbon-like anterior extension (fig.174, 181). The small sclerite **L7** lies in the anterior right edge of the **vla**-lobe and is in close contact with a ribbon-like extension of the **R2**-sclerite of the right phallomere (**R2m** in fig.174, 191-193). Another small sclerite **L8** lies in the right dorsal wall of the left complex (fig.176, 177).

Figs.174-176: Lamproblatta albipalpus (Blattaria, Blattidae, Lamproblattinae) – 174: Phallomere complex in ventral view. – 175: Phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.174. – 176: Left complex in right-dorsal view; dorsal wall of pne-pouch largely removed. – Scale: 1mm.





Figs.177-181: *Lamproblatta albipalpus* (Blattaria, Blattidae, Lamproblattinae) – **177:** Left complex in dorsal view. – **178-181:** Left complex in dorsal view; with successive removal of parts of left complex (mainly of dorsal ones). – Scale: 1mm.

The **hla**-hook (fig.174, 177) is evaginated from the left wall of the left complex and is largely sclerotised by L3. Around the base of **hla** the cuticle is circularly invaginated (fig.178, 179). Sclerite L4K shows a dorsoventral curvature: it lies mainly in the left dorsal wall of the left complex, above the base of **hla** (fig.177), but its left part bends like a horseshoe ventrad into the invagination around the **hla**-base (fig.178).

Right phallomere

Sclerite **R3** occupies the anterior ventral wall (fig.190-194). In the posterior part of **R3** the cuticle is considerably thickened (cross-section in fig.193). Sclerite **R2** articulates with the left posterior margin of **R3** (**A7** in fig.190-194). **R2** forms a large ridge (fig.191, 192, 194), whose left-dorsal part curves dorsad and slightly back to the right (fig.190). The right-ventral end of **R2** has the extension **R2m** (fig.191-195). Posterior to the central part of **R3** the ventral wall of the right phallomere curves dorsad to form a narrow, groove-like central invagination (**cbe** in fig.190, 191; compare fig.193 and 195).

Posterior to the right part of **R3** sclerite **R1F** adjoins, and the two sclerites articulate (**A3** in fig.190-194). From its central part behind the **A3**-articulation **R1F** extends in two directions: The ventral arm bends left-dorsad (at and behind edge **16** in fig.193, 195) and largely occupies the right-dorsal wall of the **cbe**-invagination (fig.190). The distal part of this arm forms a somewhat spoon-shaped process **pva** (fig.190-195). At its distal anterior margin this arm articulates with **R2** (**A6** in fig.190, 195); at its basal posterior margin it has a distinct extension (**20** in fig.190, 192-195). The dorsal arm of **R1F** extends posterodorsad (fig.190, 191) and forms a sclerotised groove (**rge** in fig.190, 195). The part of **R1F** posterior to **A3**, the extension **20**, and the dorsal arm show an extensive thickening of the cuticle directed to the interior of the phallomere (cross-sections in fig.195).

The posterior part of the right phallomere is a large dorsal lobe **fda** (fig.190, 194; in the figures **fda** is pulled to the right and to the posterior), whose dorsal and ventral walls are partly occupied by sclerite **R1J**. The dorsal anterior tip of **R1J** articulates with the dorsal arm of **R1F** (**A8** in fig.190), its ventral anterior margin articulates with the extension **20** of **R1F** (**A9** in fig.192-195, 197). Near the **A9**-articulation the cuticle of **R1J** is, like that of extension **20**, thickened to the interior, and the articulation is thus very stout and deeply immersed in the phallomere (fig.193, 195). The right wall of **fda** has a large membranous area (**17** in fig.190, 193, 194). The posterior edge of **fda** bears a sclerotised spine (**sra** in fig.190).

Subgenital plate and posterior abdominal segments

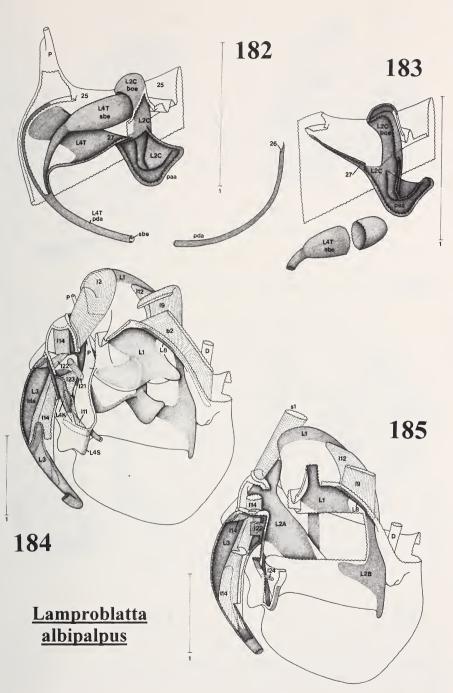
Fig.169, 170 (posterior segments); fig.173 (subgenital plate **S9**). Tergite 10 **T10** is not completely divided longitudinally, but around its posterior edge there is a median membranous field (**21** in fig.169). The ventral part of tergite 10 **T10**v is moderately extensive and is, except for an anterior transverse bridge, also divided by membrane **21**. The paraprocts **Pp** are divided (by the articulations **A97**) into a large median part sclerotising the dorsal wall of the subanal lobe **sb1** and a small lateral plate-like part. Along the anteriormost and medianmost dorsal wall of the subanal lobe **sb1** each paraproct forms a heavy groove-like apodeme (fig.170; cut through on the right side). The lateral plate of

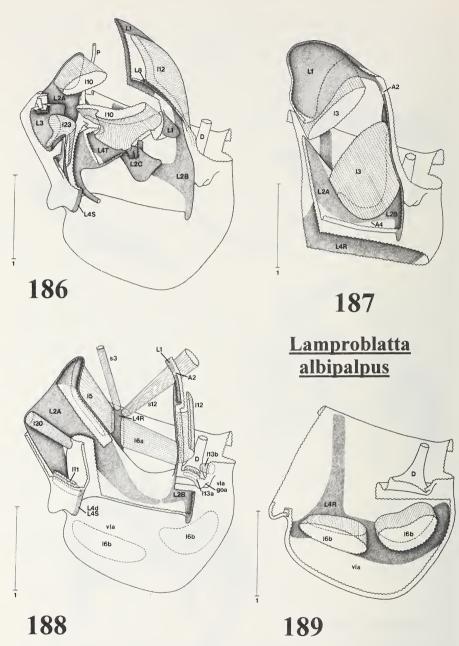
Pp is narrowly connected with the paratergite **T10p** laterally, and **A99*** is hence no longer a true articulation. Strip-like and twisted **Pv**-sclerites are present; they are laterally connected with the lateral plates of the paraprocts **Pp**. The **Ca**-sclerites are curved ribbons on rather indistinct bulges immediately median to the cercal bases. The very small **Cb**sclerites lie at the bottom of a small funnel-like invagination. The **Cc**-sclerites are dorsoventrally curved plates on a paired shallow outfolding beneath the anterior margin of **T10v** and above the subanal lobes **sbl**. The articulations **A98** are well-developed.

Musculature

Muscles	Positions of insertions	in fig.
12	L1 (left-anteriorly on pne-pouch) - membrane anterior to L4K and	
	to hla-base	184
13	L1 (ventrally on pne-pouch) – L2A and L2B (area of articulation A4)	187
15	L4R (anteriormost part) – L2A (anteriormost part)	188
16a	L4R (anteriormost part) – L2B (left posterior part)	188
16b	Left bundle: L4R (left-posterior part) – dorsal wall of vla-lobe, far	
	left-posterior to genital opening	188, 189
	Right bundle: L4R (right-posterior part) – dorsal wall of vla-lobe,	
	posterior to genital opening	188, 189
19	Anterior dorsal wall of genital pouch – L8	170, 184, 185
110	L2A (anteriormost part) – L4T and L2C between processes	
	paa and pda, membranous area 25 (compare fig.182)	186
111	L4K (posterodorsal part) – L4d (= dorsal part of L4S) and	
	membrane to the left of it	184, 188
112	Membrane next to L2B (right ventral wall of lve-pouch) – L8 and	
	membrane in dorsal wall of pne-pouch	184, 185, 186, 188
l13a	Ejaculatory duct D next to its opening – dorsal wall of goa-lobe	188
l13b	Ejaculatory duct D next to its opening – dorsal wall of vla-lobe	
	immediately to the right of genital opening	188
114	Membrane anterior to L4K and to hla-base – hla-hook (on L3);	
	muscle divided into an anterior and a posterior bundle inside hla.	184, 185
120	L2A (leftmost part) – membrane left-posterior to opening of	
	phallomere-gland P	188
121	Membrane anterior to L4K – membrane left-posterior to opening	
	of phallomere-gland P; very delicate	184
122	Membrane anterior to L4K – hla-hook (basally and dorsally on L3)	184, 185
123	L4K – hla-hook (basally and ventrally on L3)	184, 186
124	Membrane posterior to L4K – membrane left-posterior to opening	
	of phallomere-gland P; very few diffuse fibers	185

Figs.182-185: Lamproblatta albipalpus (Blattaria, Blattidae, Lamproblattinae) – 182: Sclerites L2C and L4T, processes paa and pda, and phallomere-gland P in dorsal view. – 183: Sclerites L2C and L4T and process paa in dorsal view; some further parts removed (compare fig.182). – 184,185: Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – Scale: 1mm.





Figs.186-189: *Lamproblatta albipalpus* (Blattaria, Blattidae, Lamproblattinae) – Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – Scale: 1mm.

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r2	$R3$ – $cbe\mbox{-invagination:}\ R1F$ (anterior margin), membrane, and $R2$	
	(dorsal margin)	197, 198
r3	R1F (dorsal and central parts, posterior to articulation A3) - R1J	
	(right margin)	197, 198, 199
r6	R1F (dorsal margin, on rge-groove) - dorsal wall of fda-lobe	
	(in part on R1J)	196
b2	L8 and membrane to the right of it (= right dorsal wall of	
	vla-lobe) – membrane ventral to R2	184, 198
s1	S9 left side (most anteriorly on apophysis S9a) – membrane anterior	
	to hla-base	170, 173, 175, 185
s3	S9 left side (anteriorly on apophysis S9a) – L4R (anteriormost part)	170, 173, 175, 188
s4	S9 right side (anteriorly on and median to apophysis S9a) - R3	
	(left anterior margin)	170, 173, 175, 199
s5a	S9 left side (posteriorly and medially) – left ventral wall of genital	
	pouch	173, 175
s5b	S9 left side (posteriorly and quite laterally) – left wall of genital	
	pouch	170, 173, 175
s6	S9 right side (anteriorly on apophysis S9a and laterally) – R3	
	(right anterior margin)	170, 173, 175, 199
s12	S9 right side (anteriorly on apophysis S9a) - L4R (anteriormost part)	169, 173, 175, 188
p3 (pair)	S9 – rectum (ventral wall)	169, 173
p4 (pair)	T9 (lateral anterior margin, also extending onto paratergite T9p) –	
	membrane far anterior to Pv-sclerite; muscles on both sides divided	
	into three bundles (except for their ventralmost parts)	169, 170
p5 (pair)	T10 (lateralmost anterior margin) – Pv-sclerite	169
p6 (pair)	T9 (lateralmost part) – S9 (lateral margin)	169, 172
p7 (pair)	Membrane anterior to Pv-sclerite - membrane (far) anterior to	
	contact A99* between paratergite T10p and paraproct Pp	170

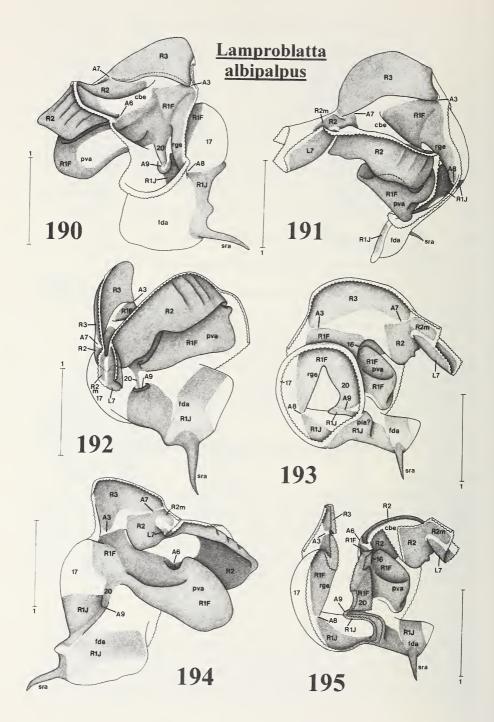
5.11. Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae)

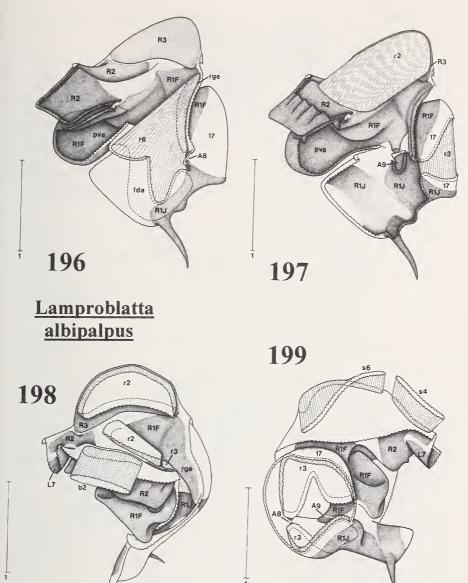
Left complex

Sclerite L4K (fig.205, 208-210) lies in the left wall. Its posterior part partly encloses the (retracted) hook hla (fig.209, 210) and its sclerite L3. The highly complicated L2-sclerite is in the center of the left complex (fig.210-215). Its anterior part forms a tube-like apodeme (lve-apodeme = anterior part of lve-pouch), on which the nla-bulge rests. At the left base of the lve-apodeme L2 forms, together with parts of L4 (L4N), a stout sclerite ring (fig.211, 212) bearing two processes: pda and paa (fig.209, 211, 214). From the right

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Figs.190-195: Lamproblatta albipalpus (Blattaria, Blattidae, Lamproblattinae) – 190: Right phallomere in dorsal view. – 191: Right phallomere in left-dorsal (somewhat anterior) view. – 192: Right phallomere in left-ventral view; membrane 17 largely removed. – 194: Right phallomere in ventral (somewhat posterior) view. – 195: Right phallomere in right-ventral view; most elements shown in fig.193 removed. – Scale: 1mm.





Figs.196-199: Lamproblatta albipalpus (Blattaria, Blattidae, Lamproblattinae) – 196: Right phallomere in dorsal view; with muscle r6; some dorsal elements removed. – 197: Right phallomere in dorsal view; with some muscles; dorsal elements more extensively removed than in fig.196. – 198: Right phallomere in left-dorsal (somewhat anterior) view; with some muscles. – 199: Right phallomere in right-ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.193; membrane 17 cut open to show muscle r3 (cut through). – Scale: 1mm.

base of the **lve**-apodeme **L2** extends to the right to form a curved plate-like sclerotisation (fig.212-214). The large lobe **vla**, with sclerite **L4G** in its ventral wall (fig.205), is the ventralmost part of the left complex.

L4K consists of a plate-like, dorsoventrally curved posterior and a bulge-like (nla in fig.206, 209; veiled by membrane in fig.208) anterior part (fig.209, 210), which are only narrowly connected (fig.205). The dorsal wall to the right of L4K is membranous; its central part is depressed ventrad and anteriad (pne in fig.208, 209). The hla-hook is evaginated from the left posterior wall of the left complex (fig.210, 212). The distal half of hla is sclerotised by L3, which is rather weak except for its distalmost part. The basal membranous half of the hla-hook (30 in fig.210-212) can be introverted and hla can thus be deeply retracted into the left complex (this state is shown in the figures). To the right of the hla-base the posterior wall of the left complex folds inwards (fpe in fig.210-213) – separating the area of hla from the area dominated by L2 (fig.211).

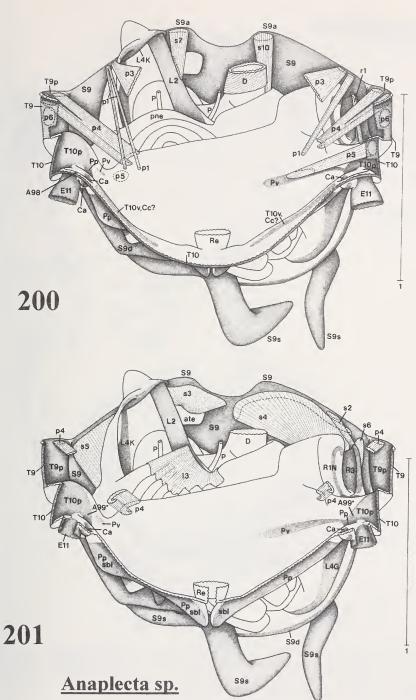
The **lve**-apodeme has its entire dorsal wall (fig.210, 211) and the margins of its ventral wall (fig.211, 225) sclerotised by **L2**. The sclerotised cuticle is considerably thickened (cross-sections in fig.211-215). The top of the **lve**-apodeme and the **nla**-process are firmly connected (two areas of the internal surface of the cuticle adhere to each other). The **lve**-apodeme is the narrow anterior part of a **lve**-pouch, which posteriorly expands to the right. At the base of the **lve**-apodeme, **L2** forks into a left and a right branch (immediately posterior to the cross-section through **lve** in fig.213).

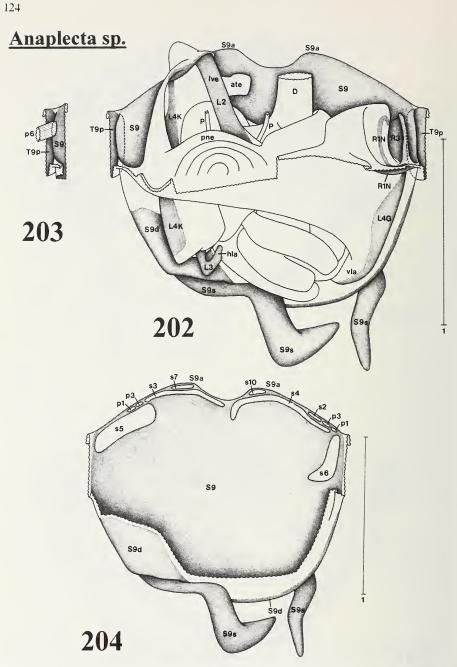
The left branch bears a node-like apodeme (**29** in fig.212, with a tuft of fine cuticular threads) and forms the sclerite-ring mentioned above (fig.211-213). This ring is slightly sunken anteriad into the left complex; the cuticle within the ring is evaginated to form the processes **paa** and **pda**, which are both sclerotised in their ventral walls only (fig.209, 214; the sclerotisations of both are connected with the basal ring). The membrane **31** in fig.211-214 is the area of contact between the bases of **paa** and **pda** (cut through in fig.214). Apart from **L2** (dorsal part of the ring, **paa**-sclerotisation), **L4** also contributes to this structure (**L4N**: ventral part of the ring, **pda**-sclerotisation). The left branch of **L2** has another posteriad-directed extension on its ventral side (**28** in fig.214, 215) which lies in the dorsal wall of another process (**gta** in fig.215, 216). The sclerotised cuticle of the left **L2**-branch is thickened in most of its parts (cross-sections in fig.212-215).

The right branch of L2 broadens and extends far to the right, where it curves dorsad (fig.212-214). Posteriorly this upcurved part extends somewhat back to the left and is involved in some complicated cuticular foldings (near 32 in fig.211, compare fig.209-213). Posterodorsal to the right L2-branch there are some additional membranous in- and outfoldings (fig.210-213).

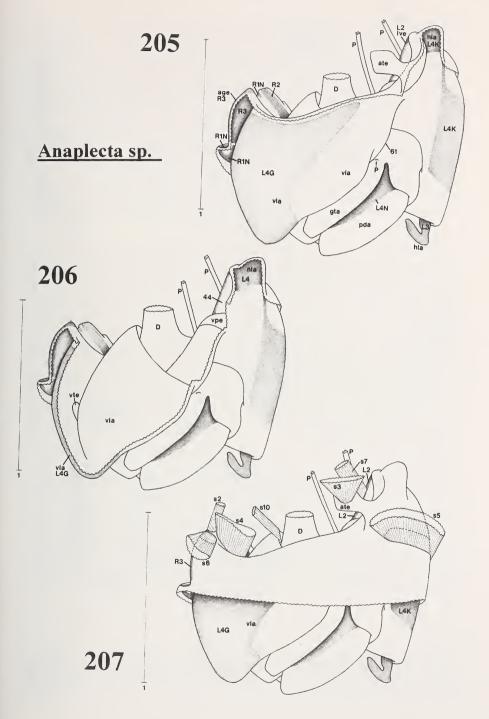
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Figs.200,201: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – **200:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, subanal lobes, paraprocts, Pv-sclerites, distal part of rectum, basal parts of cerci, and part of musculature. – **201:** Same as in fig.200, after removal of further parts of abdominal tergites 9 and 10 (especially T10v). Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. – Scale: 1mm.





Figs.202-204: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – **202:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **203:** Left margin of subgenital plate (compare fig.202); with insertion of muscle p6. – **204:** Subgenital plate in dorsal view; with insertion areas of muscles (except p6). – Scale: 1mm.



The ejaculatory duct **D** opens anterior to the right **L2**-branch (fig.210, 211). In its dorsal wall there is a small outfolding (**goa** in fig.212). The ventral wall of the duct continues posteriad into the dorsal wall of a broad membranous outfolding **vfa** (fig.211, 215). The **vla**-lobe, with sclerite **L4G** in its ventral wall (fig.205; cut open in fig.206; fig.214-220) is another broad outfolding ventral to **vfa**. The right part of the **vla**-lobe curves dorsad (fig.208). The right dorsal wall of the **vla**-lobe has a broad and flat invagination **vte** (fig.206, 208, 209, 219) functioning as a tendon (muscle **l6b** in fig.221).

Two phallomere-gland ducts (**P** in fig.215, 216) open anterior to the ventral wall of the **gta**-process. Between the orifices there is a small infolding (**ipe** in fig.215-217). Anterior to the orifices the ventral wall has a broad membranous pouch (**vpe** in fig.209-214, 216-218). Anterior to **vpe** the **nla**-bulge with its **L4K**-sclerotisation adjoins (fig.218, 219). Posterior to **nla** the anteriormost ventral wall of the genital pouch forms a membranous pouch **ate** (fig.205, 208), which functions as a tendon (muscles **s3** and **l6a** in fig.222).

Right phallomere

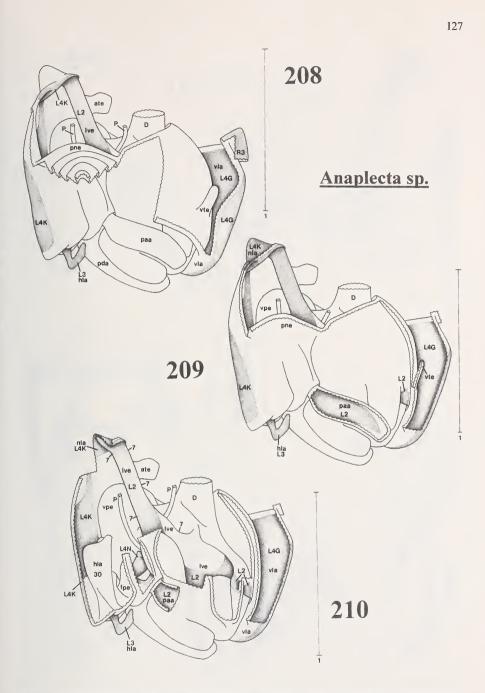
The triangular **R3**-sclerite occupies the anterior (right-)ventral wall (fig.226-229). The right and anterior margins of **R3** form a groove-like apodeme (**age** in fig.226, 229). Sclerite **R2** articulates with the left posterior margin of **R3** in two points (**A7** in fig.227-229). **R2** forms a ridge (fig.227, 228) with three bulges. Posterior to the central part of **R3** the ventral wall of the right phallomere curves dorsad and anteriad to form a central invagination (**cbe** in fig.226-228; compare fig.229 and 230). Posterior to the right part of **R3** there adjoins the large sclerite **R1N**, and the two sclerites articulate (**A3** in fig.226, 227, 229). **R1N** occupies most of the posterior part of the right phallomere, the broad dorsal lobe **fda** (fig.226, 231). From behind the **A3**-articulation **R1N** has a long extension to the left (**34** in fig.226), which lies in the dorsal wall of the **cbe**-invagination. The left end of extension **34** turns back to the right like a hook, and it articulates with the leftdorsal end of **R2** (**A6** in fig.226, 227, 230). The **fda**-lobe bears a sclerotised bulge in its ventral wall (**33** in fig.227, 228, 231).

Subgenital plate and posterior abdominal segments

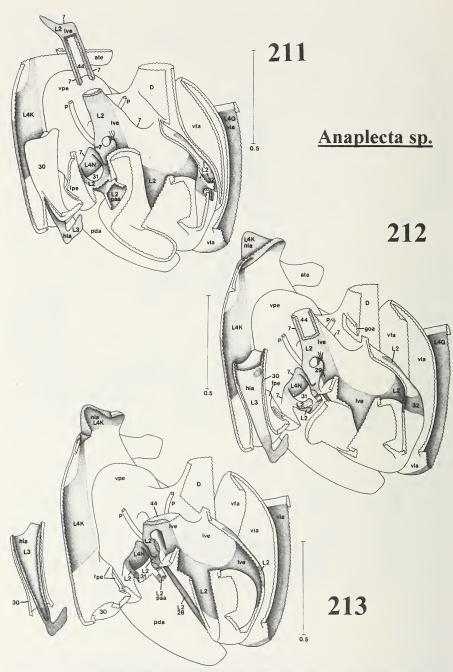
Fig.200, 201 (posterior segments); fig.204 (subgenital plate **S9**). Within the membrane extending anteroventrad from the posterior edge of tergite 10 **T10** there is a pair of ribbonlike sclerites (**T10v** or **Cc**?). Strip-like **Pv**-sclerites are present: The right one is well developed and connected with the paraproct **Pp** (or with paratergite 10 **T10p**?) laterally. The left one is very small and isolated. The articulations **A99** are missing (**A99*** in fig.201: paratergites **T10p** and paraprocts **Pp** have fused). The **Ca**-sclerites lie on curved bulges immediately median to the cercal bases. **Cb**- and **Cc**-sclerites are missing. The articulations **A98** are well-developed.

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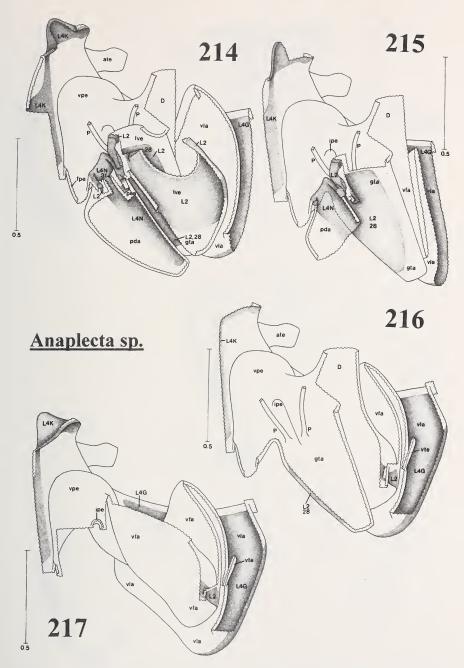
Figs.205-207: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – **205:** Phallomere complex in ventral view; some ventral parts removed (compare fig.205). – **206:** Phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.205. – Scale: 1mm.



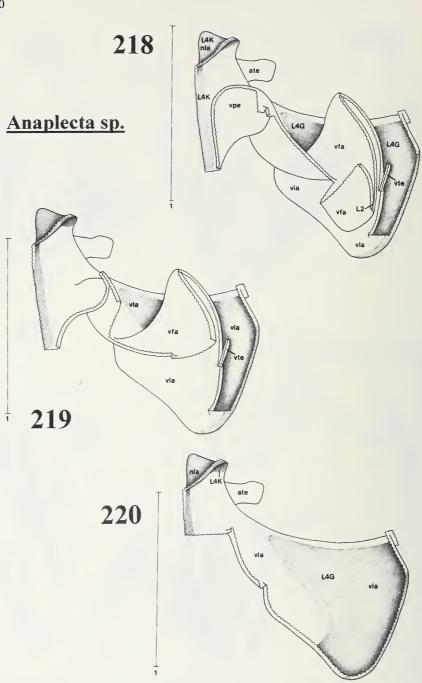
Figs.208-210: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – 208: Left complex in dorsal view.
209,210: Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones).
Scale: 1mm.



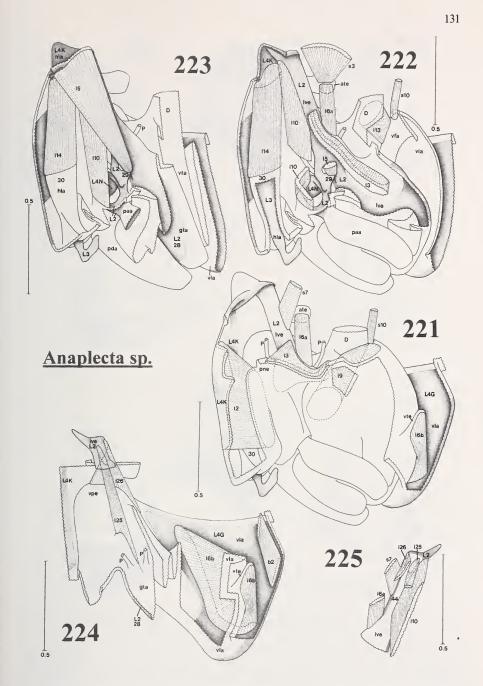
Figs.211-213: *Anaplecta* sp. (Blattaria, Blattellidae, Anaplectinae) – Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones); fig.213: hla-hook separated from remainder of left complex. – Scale: 0.5mm.



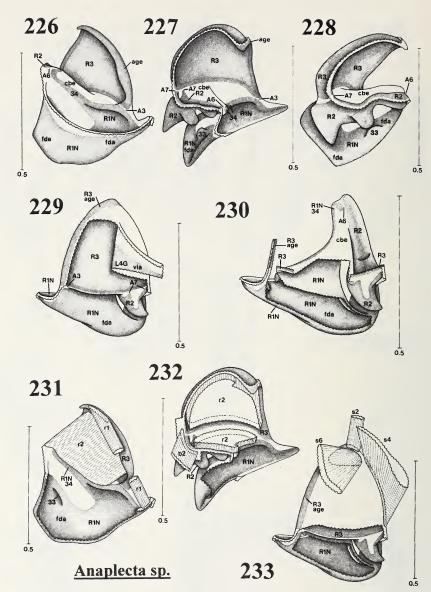
Figs.214-217: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones). – Scale: 0.5mm.



Figs.218-220: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones). – Scale: 1mm.



Figs.221-225: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – 221-224: Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – 225: Anterior part of lve-apodeme in ventral view; with the muscles inserting on it. – Scale: 0.5mm.



Figs.226-233: Anaplecta sp. (Blattaria, Blattellidae, Anaplectinae) – 226: Right phallomere in dorsal view. – 227: Right phallomere in left-dorsal (somewhat anterior) view. – 228: Right phallomere in left-ventral view; with transition to left complex. – 230: Right phallomere in right-ventral view; most elements shown in fig.229 removed. – 231: Right phallomere in dorsal view; with some muscles; some dorsal elements removed. – 232: Right phallomere in right-ventral view; with some muscles; some dorsal elements removed. – 232: Right phallomere in right-ventral view; with some muscles; some dorsal elements removed. – 233: Right phallomere in right-ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.229. – Scale: 0.5mm.

Musculature

Musslas	Desitions of incentions	1
Muscles	Positions of insertions	in fig.
12	Left dorsal wall of left complex (left part of pne-"pouch") – L4K	
	(posterior part)	221
13	Anterior central dorsal wall of left complex (anterior part of	
	pne -"pouch") – $L2$ and membrane to the right of $L2$ (right dorsal	
	wall of lve -pouch)	201, 221, 222
15	L4K (anterior part, dorsal base of nla-process) – L2 (on apodeme 29)	
16a	ate-tendon – L2 (right edge of lve-apodeme)	221, 222, 225
16b	L4G – vte-tendon in right dorsal wall of vla-lobe	221, 224
19	Transversely within right dorsal wall of left complex	221
110	L2 (anterior left edge of lve-apodeme) – sclerite ring with L4N and	
	L2 at common base of processes paa and pda	222, 223, 225
113	Ejaculatory duct D next to its opening (ventral wall) – bottom of	
	infolding between lobes vfa and vla	222
114	L4K (anterior part, ventral base of nla-process) – hla-hook	
	(anterior margin of L3)	222, 223
125	L2 (anterior ventral wall of lve-apodeme) – ipe-infolding between	
	openings of phallomere-gland ducts P	224, 225
126	L2 (anterior right edge of lve-apodeme) – anterior edge of vpe-	
	infolding	224, 225
r1	R3 (anterior right margin) – R1N (rightmost dorsal wall of	
	fda-lobe)	200, 231
r2	R3 – cbe-invagination: R1N-extension 34 (anterior margin),	
	membrane, and R2 (dorsal margin)	231, 232
b2	L4G (right margin) – membrane ventral to R2 and R3	224, 232
s2	S9 right side (laterally and anteriorly) – R3 (central anterior margin)	201, 204, 207, 233
s3	S9 left side (medially and anteriorly) – ate-tendon	201, 204, 207, 222
s4	S9 right side (entire anterior margin) $-$ R3 (left margin)	201, 204, 207, 233
s5	S9 left side (laterally and anteriorly) – left wall and left ventral	
	wall of genital pouch	201, 204, 207
s6	S9 right side (lateral margin) – anteriormost right ventral wall of	
	genital pouch	201, 204, 207, 233
s7	S9 left side (medially and most anteriorly) $-$ L2 (right edge	
	of lve-apodeme)	200, 204, 207,
		221, 225
s10	S9 right side (medially and most anteriorly) – bottom of infolding	
510	between lobes vfa and vla (ventral to ejaculatory duct)	200, 204, 207,
	between lobes via and via (vential to ejaculatory duct)	200, 204, 207, 221, 222
		221, 222
p1 (pair)	$\mathbf{S9}$ – membrane anterior to (right muscle) or median to	
	(left muscle) Pv-sclerite; very delicate; left muscle in most cases	
	divided into two bundles	200, 204
p3 (pair)	S9 – rectum (ventral wall)	200, 204
p4 (pair)	T9 (lateralmost anterior margin, also extending onto	
p. (puit)	paratergite $\mathbf{T9p}$) – membrane anterior to (right muscle) or	
	anteromedian to (left muscle) Pv -sclerite; muscles on both sides	
	divided into three bundles (except for their ventralmost parts)	200, 201
	in the second of the period and the second of pures	200, 201

p5 (pair)	T10 (lateralmost anterior margin) – on (right muscle) or median	
	to (left muscle) Pv-sclerite	200
p6 (pair)	T9 (lateralmost part) – S9 (lateral margin)	200, 203

5.12. Nahublattella sp. (Blattaria, Blattellidae, Plectopterinae)

All figures are side-reversed and show mirror-images of the original structures. In the subsequent descriptions and in the homology discussions (chapter 6.) the terms "left" and "right" will also be exchanged. (This will be practised in other Plectopterinae, too: *Euphyllodromia, Supella*). The natural orientation is shown in fig.236b and 239b.

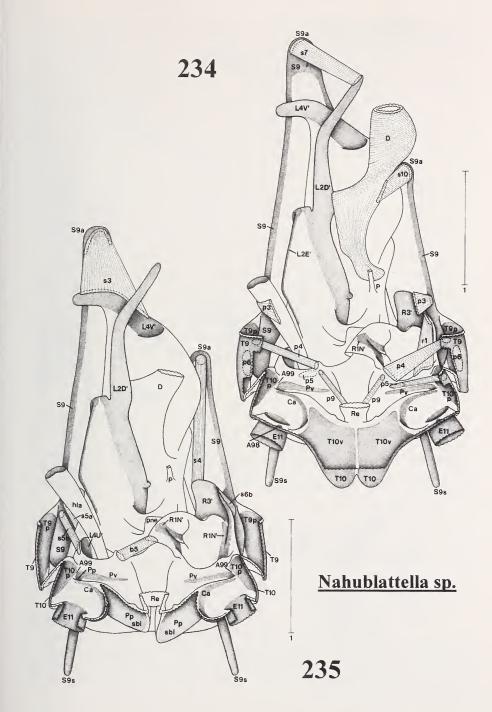
Left complex

Sclerite L1' lies in the posterodorsal wall (fig.243). The L2'-sclerotisations (L2D', L2E') are in the center of the left complex (fig.242-244). The anterior part of L2D' forms a tube-like apodeme (lve-apodeme = anterior part of lve-pouch), to the right of which the ejaculatory duct (D in fig.242) opens. At the left base of this apodeme, L2E' forms, together with a L4'-sclerotisation (L4N'), a large trifid process via (fig.244, 245). In the anteriormost ventral wall of the left complex lies sclerite L4V', which bears the whip-like process nla (fig.239a, 247). The left posterior edge of the left complex bears a long hook hla with its sclerite L3' (fig.242-244). The base of the hook is partly enclosed by the lateral sclerite L4U'.

The left posterior part of the left complex resembles a bulge whose left wall is sclerotised by L4U' (fig.242-244). The hla-hook is evaginated from the posterior wall of this bulge; its distal half is sclerotised by L3'. The membrane of the basal half (30 in fig.242-244) can be introverted, and hla can be retracted in the same way as in *Anaplecta* (all figures show hla in its retracted state). Right-ventral to the hla-base the posterior wall of the left complex folds inwards (fpe in fig.242-245): This fpe-fold separates the area of hla from the other parts of the left complex. The dorsal wall to the right of hla contains a bristle area (35 in fig.242). Ventral to this area there is a flat pouch invaginated to the anterior (pne in fig.242, 243). The ventral wall of the pne-pouch is sclerotised by the anterior part of L1'. The cup-shaped posterior part of L1' occupies a bulge-like process (dca in fig.243, 244) beset with setae. Beneath the dca-process there is a membranous lobe (cla in fig.244). Anteroventral to the cla-lobe the cuticle is again evaginated: The posterior end of sclerite L2D' forms a bifid process (psa in fig.244-246).

Figs.234,235: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures show mirrorimages of the original preparations. – **234:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, subanal lobes (covered), paraprocts (covered), Pv-sclerites, distal part of rectum, basal parts of cerci, and part of musculature. – **235:** Same as in fig.234, after removal of further parts of abdominal tergites 9 and 10 (especially T10v). Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. – Scale: Imm.





The whole central part of the left complex is invaginated anteriad to form a large pouch (lve in fig.242-246; the edges along the bottom of this lve-pouch are labelled 7 in fig.242). This invagination contains the lve-apodeme (middle part; with the L2D'-sclerotisation), the via-process (left side), and the terminal part of the ejaculatory duct **D** (right side). The lve-apodeme is completely sclerotised by L2D' – except for a membranous stripe (44 in fig.239a, 243, 245) in its ventral wall. At the base of the lve-apodeme the lve-pouch broadens. Here, L2D' has a short extension to the left (36 in fig.239a, 243-246) and a long ventral extension to the posterior (28 in fig.245, 246, which lies in the ventral wall of the lve-pouch). The main part of L2D', however, extends far posteriad within the dorsal wall of the lve-pouch (fig.242); it bears a small apodeme (37 in fig.242, 245), and its posteriormost part sclerotises the psa-process. Along its right margin this main part of L2D' folds ventrad and back to the left (towards edge 38 in fig.245; compare fig.244) to form a shallow sclerite groove. Along edge 38 the cuticle turns to the right again and continues into the dorsal wall of the ejaculatory duct (**D** in fig.244, 245). The ejaculatory duct opens from the right side into the lve-pouch (fig.242-246). The phallomere-gland (P in fig.242) opens posterior to the dorsal wall of the ejaculatory duct.

The via-process evaginates posteriad from the left wall of the lve-pouch (fig.244; in fig.241 via is isolated and shown in ventral view). Distally via branches into three spines paa, pda, and vsa. via has a basal and a distal sclerite separated by a membranous ring (39 in fig.241, 244, 245; sclerites not termed separately). The basal sclerite is roughly cylindrical (fig.244), with a deep recess at its ventral anterior margin (fig.241, 245). Ventrally it bears the vsa-spine. At its right anterior margin it articulates with extension 36 of sclerite L2D'

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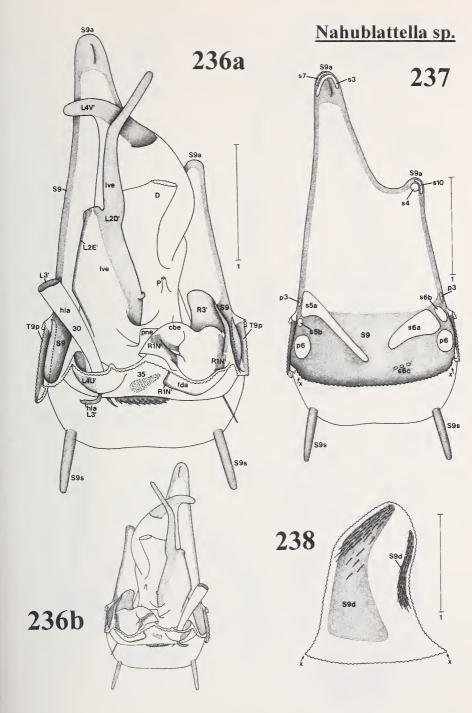
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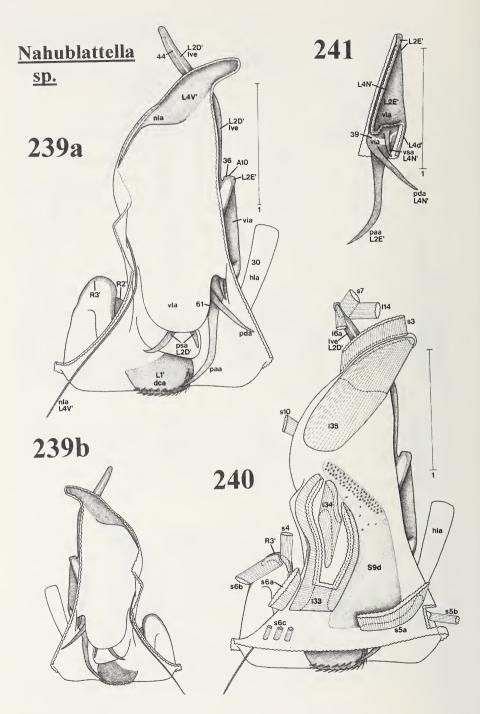
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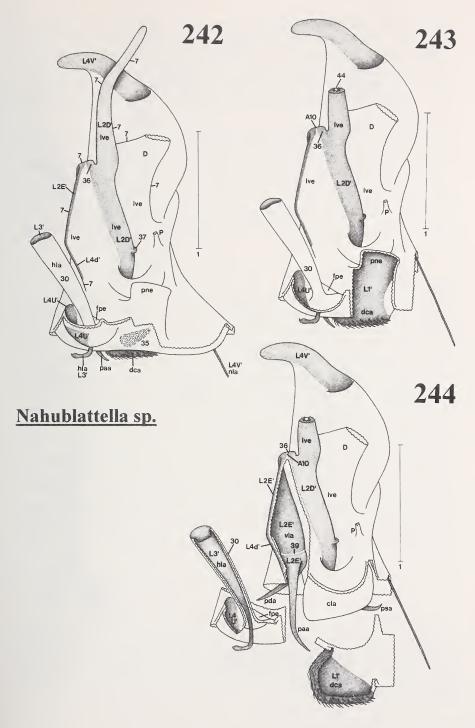
Figs.236-238: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures except 236b show mirror-images of the original preparations. – **236a:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **236b:** Same as in fig.236a but smaller scale and natural orientation. – **237:** Subgenital plate in dorsal view; with insertion areas of muscles (including p6). – **238:** Dorsal sclerotisation S9d of subgenital plate; the part of the cuticle shown has been cut off from the subgenital plate along the line between the points labelled x (compare x in fig.237). – Scale: 1mm.

Figs.239-241: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures except 239b show mirror-images of the original preparations. – **239a:** Phallomere complex in ventral view. – **239b:** Same as in fig.239a but smaller scale and natural orientation. – **240:** Phallomere complex in ventral view; with some muscles: ventral wall of genital pouch completely retained (compare fig.239a), including dorsal sclerotisation S9d of subgenital plate and its muscles. – **241:** via-process (including paa, pda, and vsa, and sclerotisations L2E' and L4N') in ventral view. – Scale: 1mm.

Figs.242-244: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures show mirrorimages of the original preparations. – **242:** Left complex in dorsal view. – **243,244:** Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones); fig.244: hook hla and adjacent areas as well as retained parts of process dca with sclerite L1' separated from remainder of left complex. – Scale: 1mm.







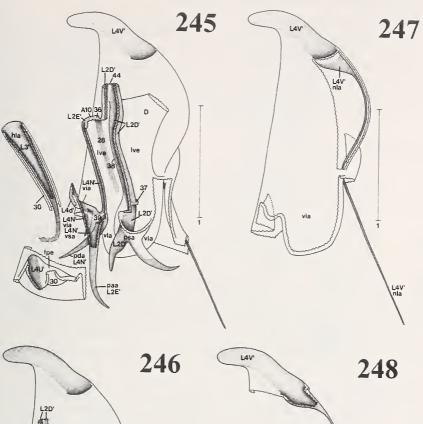
(A10 in fig.243-245). At its left anterior margin it has a ribbon-like extension (L4d' in fig.241, 242, 244, 245) running posteriad along the left edge of the lve-pouch (7 in fig.242). The distal sclerite branches into the sclerotisations of **paa** and **pda**. According to their assumed origin, the right-dorsal parts of the via-sclerotisation (including A10 and **paa**) are designated L2E', the left-ventral parts (including L4d' and **pda**) are designated L4N'. (The boundary between L2E' and L4N' is perpendicular to the division into a basal and a distal sclerite).

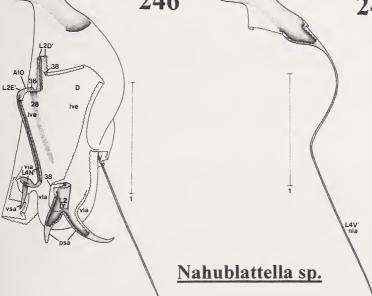
The ventral wall of the **lve**-pouch (fig.246, including the ventral wall of the ejaculatory duct **D**) is, except for the **L2D'**-extension **28** and the sclerotisations within the **lve**-apodeme, membranous. To the posterior it continues into the dorsal wall of a completely membranous lobe (**vla** in fig.245-247, 239a). The ventral wall of **vla** is part of the ventral wall of the left complex (fig.239a, 247). Sclerite **L4V'** (fig.239a, 247) occupies the anteriormost ventral wall; its right part extends onto and completely sclerotises the very long process **nla** (fig.239a, 247, 248). **nla** has a broad base but soon narrows to become whip-shaped.

Right phallomere

Sclerite **R3**' occupies the anterior (right-)ventral wall (fig.253-257); its lateral and anterior margins form a groove-like apodeme (age in fig.253, 256, 257). At its left and right ends **R3'** has extensions to the posterior (40 and 41 in fig.253-257). Posterior to the left part of R3' sclerite R2' adjoins; the two sclerites have a broad articulation (A7 in fig.255, 257). R2' is a plate of irregular shape, which as a whole slightly bulges posteriad (fig.254-257). It bears a tooth (42 in fig.254, 255, 260) projecting dorsad and a horseshoe-shaped bulge with small spines (43 in fig.254, 255, 260; seen from inside in fig.256, 258). Posterior to the central part of R3' the ventral wall of the right phallomere curves dorsad and slightly anteriad to form a narrow, groove-like central invagination (cbe in fig.253-256; compare fig.257 and 258), whose left-ventral wall is completely sclerotised by **R2**². Sclerite R1N' broadly articulates with the right posterior part of R3' (A3 in fig. 253, 255, 257). From the A3-articulation R1N' extends like an arch posteriad, leftward, and anteriad again; it occupies the margins of a broad dorsal lobe fda (fig.253), which is the posterior part of the right phallomere. The left anterior part of R1N' articulates with the left margin of R2' (A6 in fig.253-255, 258); at A6 R1N' turns sharply back to the right (34 in fig.253), and its bristled terminal part lies on a bulge in the posterior dorsal wall of the cbeinvagination (34 in fig.253, 255, 259).

Figs.245-248: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures show mirrorimages of the original preparations. – Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones); fig.245: hook hla and its base (with sclerite L4U') separated from remainder of left complex and from each other (compare fig.244); fig.248: only sclerite L4V' and process nla retained. – Scale: 1mm.





Subgenital plate and posterior abdominal segments

Fig.234, 235 (posterior segments); fig.237, 238 (subgenital plate **S9**). The dorsal sclerotisation **S9d** of the subgenital plate comprises two isolated sclerites (fig.238), which are asymmetrical and beset with stout setae. The entire tergite 10 **T10**, including its ventral part **T10v**, is divided along its midline. **T10v** is rather extensive. Strip-like **Pv**-sclerites are present; they are laterally connected with the paraprocts **Pp**. The **Ca**-sclerites are very long and lie on curved bulges immediately median to the cercal bases. **Cb**- and **Cc**-sclerites are missing. The articulations **A98** and **A99** are well-developed.

Musculature

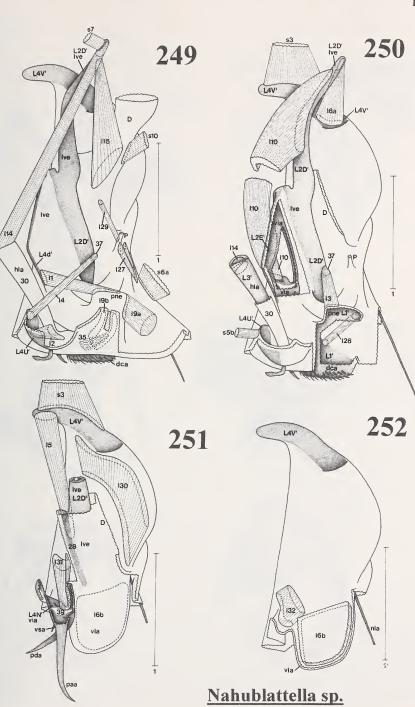
Muscle	Positions of insertions	in fig.
11	L1' (anteriorly on pne-pouch) – L4d' (= part of L4N')	249
12	Membranous basal part 30 of hla-hook – L4U' (dorsal part)	249
13	L1' (anteriorly on pne-pouch) – L2D' (posteriormost part, on	
	apodeme 37)	250
14	L2D' (posteriormost part, on apodeme 37) – L4U' (ventral part)	249
15	L4V' (left posterior margin) - L2D' (extension 28 in ventral wall	
	of lve-pouch)	251
16a	L4V' (right part) – L2D' (most anteriorly on lve-apodeme)	240, 250
16b	Ventral wall of vla-lobe – dorsal wall of vla-lobe	251, 252
19a	L1' (right wall of pne-pouch) – membrane to the right of pne-pouch	249
19b	Transversely in dorsal wall of left complex (including bristle area 35)	249, 261
110	L2D' (anterior left edge of lve-apodeme) – left base of via-process	
	(with L4N' and L2E')	250
114	L2D' (most anteriorly on lve-apodeme) – hla-hook (anterior margin	
	of L3')	240, 249, 250
115	L2D' (anteriorly on lve-apodeme) – ejaculatory duct D next to its	
	opening (dorsal wall)	249
127	Outlet channel of phallomere-gland \mathbf{P} – membrane to the right of \mathbf{P}	249
128	L1' (anterior right ventral wall of pne-pouch) – L1' (ventral anterior	
	margin)	250
129	Outlet channel of phallomere-gland P - ejaculatory duct D next to its	
	opening (dorsal wall)	249
130	Longitudinally in ventral wall of left complex	251

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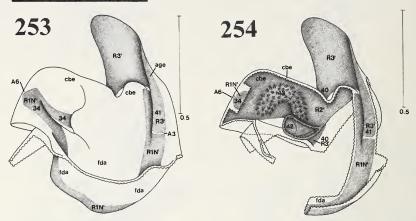
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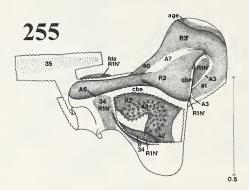
Figs.249-252: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures show mirrorimages of the original preparations. – Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents. – Scale: 1mm.

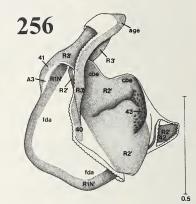
Figs.253-258: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures show mirrorimages of the original preparations. – **253**: Right phallomere in right-dorsal view. – **254**: Right phallomere in right-dorsal view; dorsal elements largely removed. – **255**: Right phallomere in dorsal (somewhat anterior) view; membrane of cbe-invagination cut open to show parts of R2' lying beneath it (42, 43). – **256**: Right phallomere in left view. – **257**: Right phallomere in ventral view. – **258**: Right phallomere in ventral view; most elements shown in fig.257 removed. – Scale: 0,5mm.

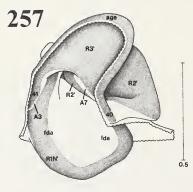


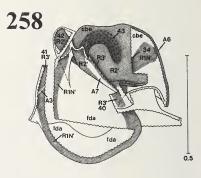
Nahublattella sp.

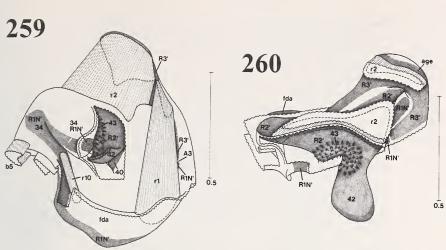


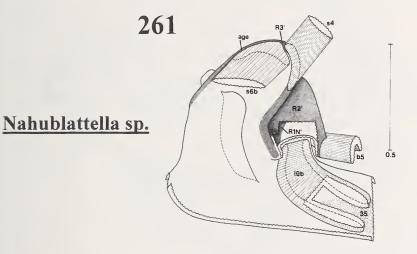












Figs.259-261: *Nahublattella* sp. (Blattaria, Blattellidae, Plectopterinae) – All figures show mirrorimages of the original preparations. – **259:** Right phallomere in right-dorsal view; with some muscles; some dorsal elements removed; membrane of cbe-invagination cut open to show parts of R2' lying beneath it (42, 43). – **260:** Right phallomere in dorsal (somewhat anterior) view; with muscle r2; dorsal elements removed. – **261:** Right phallomere in ventral view; with some muscles; ventral wall of genital pouch much more complete than in fig.257 (compare fig.239a). – Scale: 0,5mm.

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131	Left ventral wall of left complex – ventral base of via-process	
	(with L4N' and L2E')	251
132	Both insertions in left posterior ventral wall of left complex	252
133	S9d (right sclerite) – S9d (left sclerite)	240
134	Longitudinally in posterior ventral wall of genital pouch (membrane	
	in between the two S9d -sclerites)	240
135	Longitudinally in anterior ventral wall of genital pouch	240
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r2	R3' – cbe-invagination: membrane and R2' (dorsal margin)	259, 260
r10	R1N' (left ventral wall of fda-lobe) – left dorsal wall of fda- lobe	259
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s3	S9 left side (most anteriorly on apophysis S9a) – L4V' (anterior	, , ,
50	margin)	235, 237, 240,
	indi gitti)	250, 251
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	margin)	235, 237, 240, 261
s5a	S9 left side (far posteriorly; laterally and medially) – left ventral	,,,,
	wall of genital pouch	235, 237, 240
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p5 (pair)	T10 (lateralmost anterior margin) – membrane anterior to Pv-sclerite	
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p9 (pair)	Membrane anterior to Pv -sclerite – membrane beneath posterior	
	part of rectum	234

5.13. Parcoblatta lata (Blattaria, Blattellidae, Blattellinae)

Left complex

The left complex is not as complicated as in the previous species and contains only few sclerotisations. A deep infolding from the posterior (**fpe** in fig.268-271) divides the left complex into a left part with the retractable **hla**-hook and its **L3**-sclerite and a right part with the long **L2**-sclerite.

The left part has the shape of a bulge. The **hla**-hook evaginates from the posterior wall of the bulge; when retracted, **hla** lies in the center of the bulge (fig.268, 269; all figures show **hla** in the retracted state). The distal half of **hla** is sclerotised by **L3**, which becomes

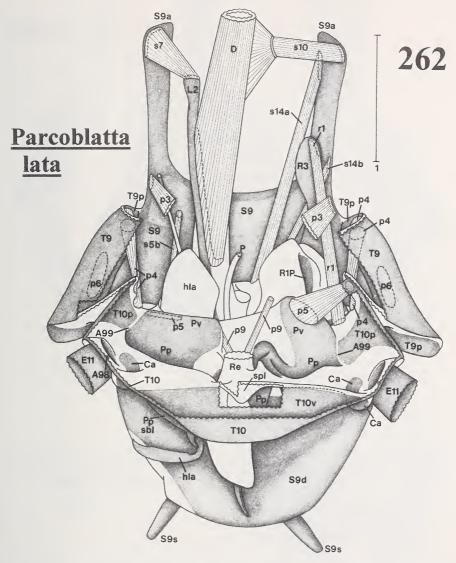


Fig.262: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, supraanal lobe, subanal lobes, paraprocts, distal part of rectum, basal parts of cerci, and part of musculature.
Supraanal lobe shown through a window cut into ventral sclerotisation of abdominal tergite 10 T10v.
Scale: 1mm.

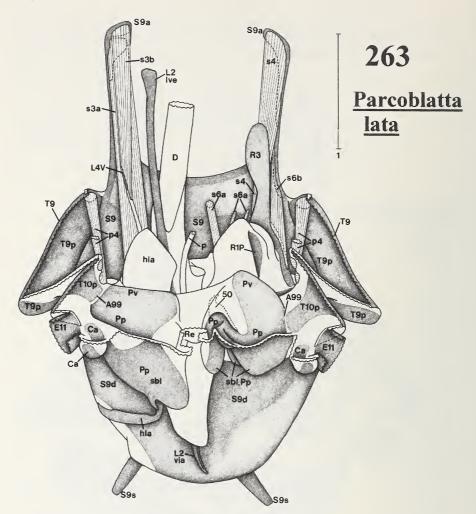
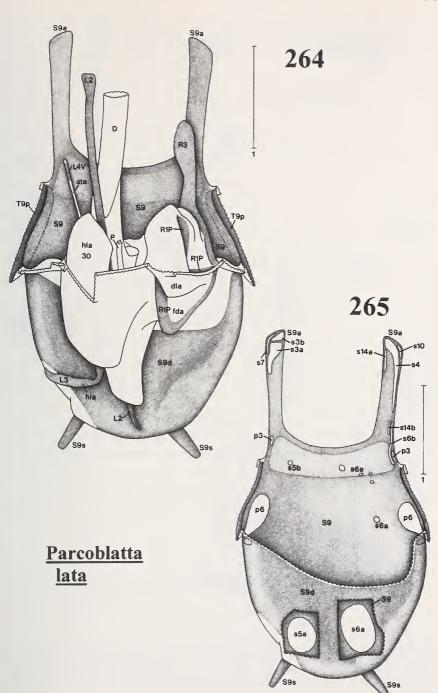
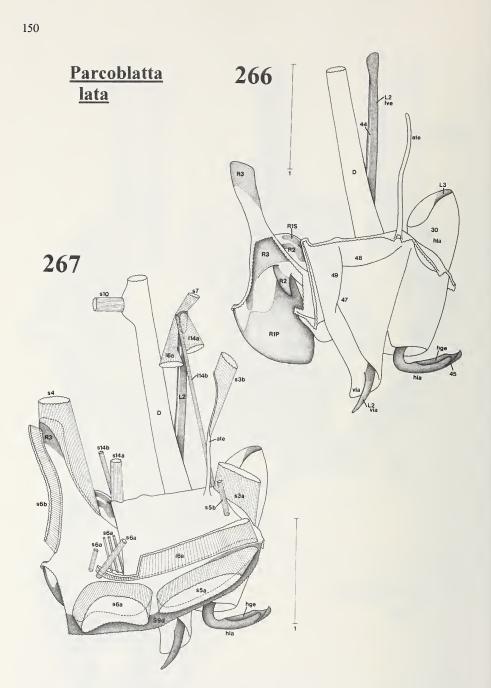


Fig.263: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – Male postabdomen as in fig.262, after removal of further parts of abdominal tergites 9 and 10 (especially T10v) and supraanal lobe. Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. – Scale: 1mm.

[→] p.149

Figs.264,265: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – **264:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **265:** Subgenital plate in dorsal view; with insertion areas of muscles (including p6). Insertion areas of s5a and s6a shown through two windows cut into dorsal sclerotisation S9d of subgenital plate. – Scale: 1mm.





Figs.266,267: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – **266**: Phallomere complex in ventral view. – **267**: Phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.266, with parts of dorsal sclerotisation S9d of subgenital plate in its posterior part (compare fig.265). – Scale: 1mm.

heavier distally. The membranous basal half (**30** in fig.268-270) of **hla** becomes introverted in the retraction of **hla** (fig.270, 271). The terminal leftward-bent part of **hla** has a groove along its anterior surface (**hge** in fig.266), whose ventral wall has a distinct notch (**45** in fig.266).

The part to the right of the **fpe**-infolding has in its center a deep invagination to the anterior (pouch lve in fig.268, 269) and a spine to the posterior (via in fig.268); both contain parts of sclerite L2. The anterior part of the lve-pouch is a narrow tube-like apodeme (lveapodeme) with a flattened and broadened top. It is completely sclerotised by L_2 – except for a membranous stripe (44 in fig.266) in its right-ventral (more anteriorly) or right (more posteriorly) wall, which does not reach the top of the apodeme. Roughly in the middle of L2, the ejaculatory duct joins the lve-pouch from the right (fig.268), and lve becomes much broader. At this point, the right edge of the lve-apodeme (with the membranous stripe 44) bends anteriad to continue into a dorsal fold of the ejaculatory duct D (fig.268, 269). The ventral main part of the ejaculatory duct extends to beneath the lve-pouch (fig.268, 269; cross-section in fig.270) and wraps partly around it from ventrally (fig.270-272). In the area posterior to the confluence of the lve-apodeme and the dorsal part of the ejaculatory duct, L2 is a groove-shaped sclerotisation in the left edge of the lve-pouch (cross-sections in fig.270-272). This groove-shape of L2 extends posteriad as far as to the posterior end of edge 7 (fig.270, 273), where L2 becomes completely restricted to the dorsal wall of the lve-pouch.

Posterior to this point L2 forms the sclerotisation of the via-spine (fig.272-275). via has a longitudinal groove in its right-dorsal wall (vge in fig.272-275), whose anterior end deepens to form a small, bulb-like, and heavily sclerotised apodeme (vge, tve in fig.273). The phallomere-gland P opens to the right of the tve-apodeme. The ventral wall of the left complex has some outfoldings in the area beneath the ejaculatory duct (47, 48, 49 in fig.266, 271, 272). In between these outfoldings and posterior to them the ejaculatory duct opens to the outside, and this area can be regarded as the genital opening. Dorsal to and to the left of the via-process there is a dorsoventrally curved membranous lobe (vla in fig.266, 268, 270).

The tendon **ate** (fig.266, 268, 271) has its origin in the anteriormost ventral wall of the genital pouch; it is a long and thin invagination of the cuticle with sclerite **L4V** in its anterior dorsal wall.

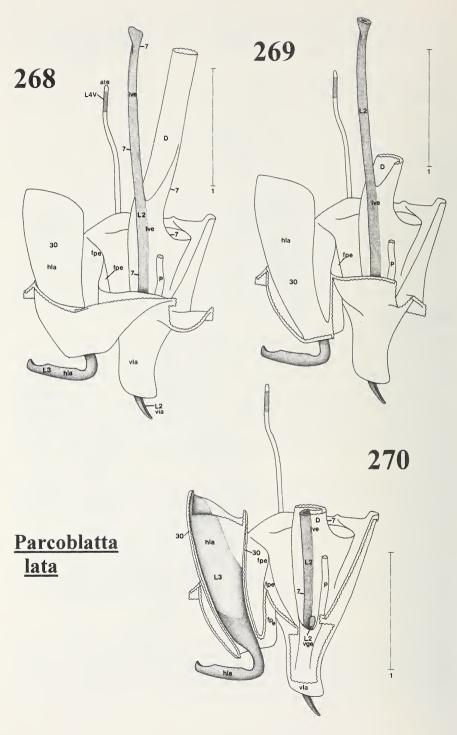
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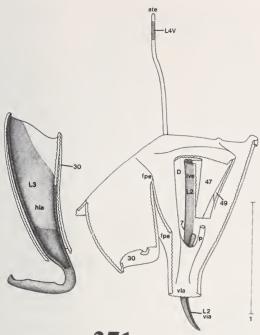
Figs.268-270: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – **268:** Left complex in dorsal view. – **269,270:** Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones). – Scale: 1mm.

 $[\]longrightarrow$ p.153

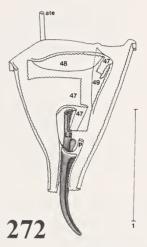
Figs.271-275: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – **271,272:** Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones); fig.271: hla-hook separated from remainder of left complex (compare fig.270). – **273-275:** Posteriormost part of sclerite L2 on process via, phallomere-gland P, and surrounding membranes in dorsal view (scale larger than in fig.272); with successive removal of parts of the cuticle (mainly of dorsal ones). – Scale: 1mm.

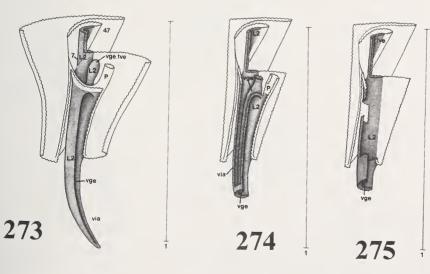






<u>Parcoblatta</u> <u>lata</u>





Right phallomere

The long, spatulate **R3**-sclerite occupies the anterior ventral wall (fig.280-284). Its right part has a long extension to the posterior (fig.280, 284). The **age**-groove or -apodeme is very broad at the anteriormost margins of **R3**; to the posterior it soon decreases and ends on both sides (fig.266, 284). Sclerite **R2** adjoins posterior to the left part of **R3**; the two sclerites are broadly separated by membrane (at **A7** in fig.282-284). From posterior to **A7**, **R2** extends anteriad and then curves to the left (fig.281, 283, 285). Most ventrally **R2** has a strong tooth (fig.283, 284); in its other parts it forms a very low ridge (fig.285). At its left end **R2** is fused to sclerite **R1S** (at **A6*** in fig.281, 283, 285). **R1S** likewise forms a low ridge (**pva** in fig.281, 282), and next to its fusion with **R2** it has a bulge-like cuticular thickening to the interior (**cwe** in fig.282, 283, 285). Posterior to the central part of **R3** the ventral wall of the right phallomere curves dorsad and anteriad to form a central invagination (**cbe** in fig.280, 281, 283; compare fig.284 and 285) with **R2** and **R1S** in its left-ventral wall.

Sclerite **R1P** adjoins posterior to the right part of **R3** (fig.281, 284), and the two sclerites articulate (**A3** in fig.281, 282, 284). **R1P** occupies the ventral wall and the margins of the dorsal wall of a large lobe **fda** (fig.281, 284). The left anterior tip of **R1P** closely approaches the free end of **R1S** (fig.281, 282). Above the **fda**-lobe there is another, smaller and membranous lobe (**dla** in fig.280).

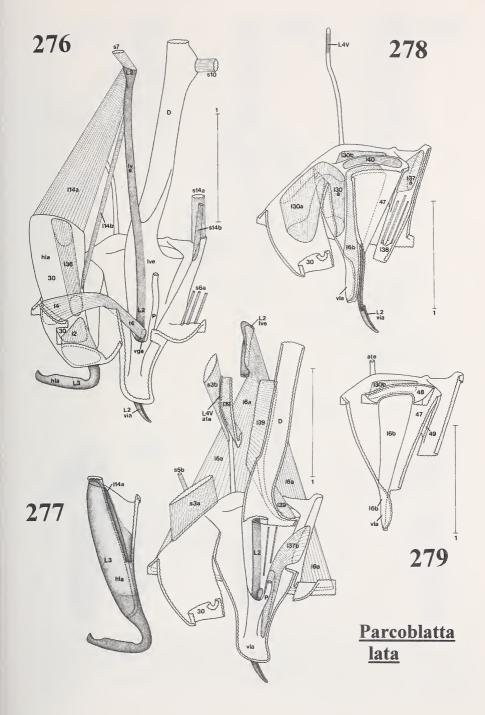
Subgenital plate and posterior abdominal segments

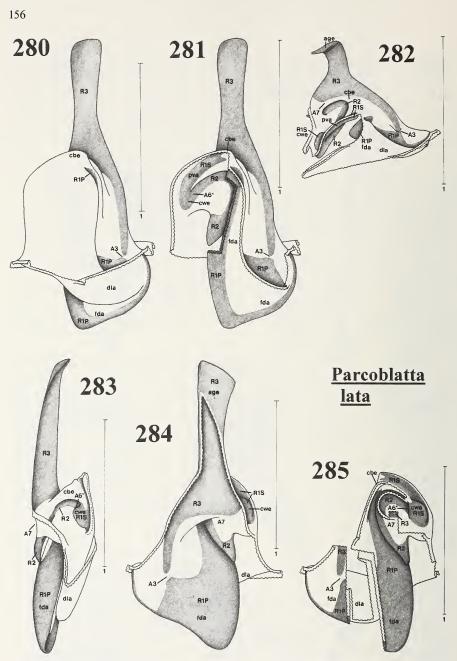
Fig.262, 263 (posterior segments); fig.265 (subgenital plate **S9**). The ventral part of tergite 10 **T10v** is rather extensive. There are no separate **Pv**-sclerites; the **Pv**-sclerotisations are assumed to be incorporated into the anterior parts of the paraprocts **Pp** (fig.263; a deep indentation at the median margin of each paraproct possibly marks the border between **Pp** and **Pv**). The **Ca**-sclerites lie on curved bulges immediately median to the cercal bases. **Cb**- and **Cc**-sclerites are missing. The articulations **A98** and **A99** are well-developed. The asymmetrical subanal lobes **sbl** are highly elaborated (fig.263): The left **sbl** bears a small spine on its posterior edge. The right **sbl** bears some posteriad-directed bulge-like processes and an anteriad-directed small hook (**50** in fig.263, veiled by membrane).

Musculature

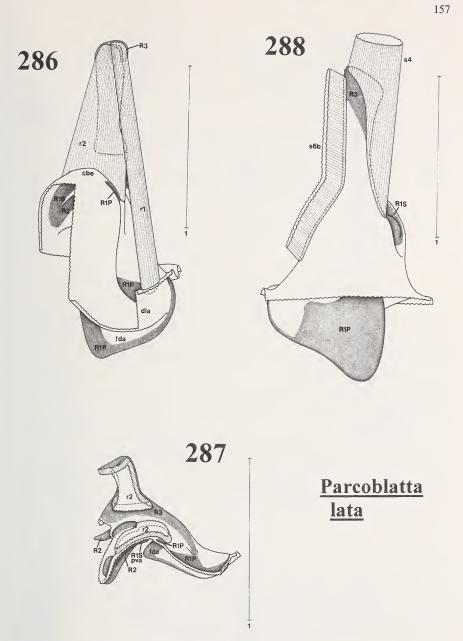
in fig.
276
276
267, 277

Figs.276-279: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents; fig.279: of muscle l6b only ventral insertion area shown. – Scale: 1mm.





Figs.280-285: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – **280**: Right phallomere in dorsal view; some dorsal elements removed. – **282**: Right phallomere in left-dorsal (somewhat anterior) view. – **283**: Right phallomere in left-ventral view. – **284**: Right phallomere in right-ventral view. – **285**: Right phallomere in right-ventral view; most elements shown in fig.284 removed. – Scale: 1mm.

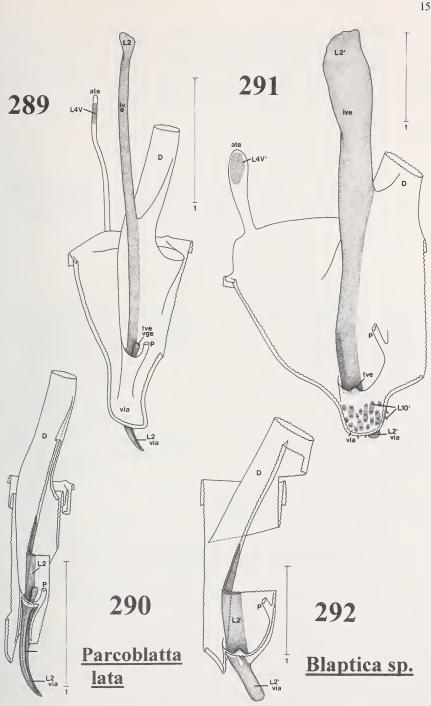


Figs.286-288: *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – 286: Right phallomere in dorsal view; with some muscles; some dorsal elements removed. – 287: Right phallomere in left-dorsal (somewhat anterior) view; with muscle r2; some dorsal elements removed. – 288: Right phallomere in right-ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.284.
Scale: 1mm.

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16b	Central ventral wall of left complex – posterior ventral wall of	278 270
11.4.	lve-pouch; diffuse	278, 279
114a	L2 (most anteriorly on lve-apodeme) – hla-hook (left part of L3)	267, 276, 277
114b	L2 (most anteriorly on lve-apodeme) – right part of membranous	2(7.27)
	basal part 30 of hla -hook	267, 276
130a	Longitudinally in ventral wall of left complex (on both sides of	
	fpe-infolding); diffuse	278
130b	Ventral wall of outfolding 48 - dorsal wall of outfolding 48; diffuse	278, 279
136	Longitudinally in membranous basal part 30 of hla-hook	276
137a	Transversely in anterior right ventral wall of left complex	278
137b	Longitudinally in posterior right ventral wall of left complex	277
138	Some isolated fibers in right ventral wall of left complex	278
139	ate-tendon (anterior part with $L4V$) – ejaculatory duct D	
	(ventral wall)	277
140	Transversely in anterior ventral wall of left complex; diffuse	278
r1	R3 (right-anteriormost part) – membrane in right anterior dorsal	
	wall of dla -lobe	262, 286
r2	R3 – cbe-invagination: R1S (right part), R1P (leftmost anterior	
	part), and membrane	286, 287
s3a	S9 left side (far anteriorly on apophysis S9a) – left ventral basal	
	line Bl of left complex	263, 265, 267, 277
s3b	S9 left side (most anteriorly on apophysis S9a) – ate-tendon	263, 265, 267,
		277
s4	S9 right side (anteriorly and posteriorly on apophysis S9a) – R3	
5.	(anterior and left margin)	263, 265, 267, 288
s5a	S9 left side (very far posteriorly) – left ventral wall of genital	200, 200, 201, 200
304	pouch (very far posteriorly, in part on S9d)	265, 267
s5b	S9 left side (medially) – anterior left ventral wall of genital pouch;	200, 207
300	very delicate	262, 265, 267, 277
s6a	S9 right side (main part inserting very far posteriorly, some smaller	202, 203, 207, 277
50a	bundles more anteriorly) – right ventral wall of genital pouch (very	
	far posteriorly, in part on anterior margin of S9d)	263, 265, 267, 276
s6b	S9 right side (laterally) – $\mathbf{R3}$ (entire right margin)	263, 265, 267, 288
		203, 203, 207, 200
s7	S9 left side (most anteriorly on apophysis S9a) – L2 (most anteriorly on luo apodemo).	262, 265, 267, 276
	anteriorly on lve-apodeme)	202, 203, 207, 270

Figs.289-292: **289,290:** *Parcoblatta lata* (Blattaria, Blattellidae, Blattellinae) – **289:** Central part of left complex in dorsal view; with sclerites L2 and L4V, tendon ate, process via, phallomere-gland P, and ejaculatory duct D; stippled area: insertion area of muscle 14. – **290:** Central part of left complex in dorsal view; with posterior part of sclerite L2, process via, phallomere-gland P, and ejaculatory duct D; many parts removed compared with fig.289. – **291,292:** *Blaptica* sp. (Blattaria, Blaberidae) – Mirror-images of the original preparations. – **291:** Central part of left complex in dorsal view; with sclerites L2' and L4V', sclerite-group L10', tendon ate, process via, phallomere-gland P, and ejaculatory duct D; stippled area: insertion area of muscle 14. – **292:** Central part of left complex in dorsal view; with posterior part of sclerite L2, process via, phallomere-gland P, and ejaculatory duct D; stippled area: insertion area of muscle 14. – **292:** Central part of left complex in dorsal view; with posterior part of sclerite L2, process via, phallomere-gland P, and ejaculatory duct D; stippled area: insertion area of muscle 14. – **292:** Central part of left complex in dorsal view; with posterior part of sclerite L2, process via, phallomere-gland P, and ejaculatory duct D; many parts removed compared with fig.291. – Scale: 1mm.



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s10	S9 right side (most anteriorly on apophysis S9a) - ejaculatory	
	duct D (right wall)	262, 265, 267, 276
s14a	S9 right side (far anteriorly on apophysis S9a) - right ventral	
	basal line BI of left complex	262, 265, 267, 276
s14b	S9 right side (far anteriorly on apophysis S9a) - right ventral	
	basal line Bl of left complex	262, 265, 267, 276
p3 (pair)	S9 – rectum (ventral wall)	262, 265
p4 (pair)	T9 (lateralmost anterior margin, also extending onto paratergite	
	T9p) - paratergite T10p (anterior margin); muscles on both sides	
	completely divided into two bundles	262, 263
p5 (pair)	T10 (lateralmost anterior margin) – anterior margin of	
	Pv-sclerotisation	262
p6 (pair)	T9 (lateralmost part) – S9 (lateral part, also extending onto dorsal	
	sclerotisation S9d of subgenital plate)	262, 265
p9 (pair)	Membrane anterior to Pv-sclerotisation - membrane median to	
	paraproct Pp , beneath rectum	262

5.14. Blaberus craniifer (Blattaria, Blaberidae)

All figures are side-reversed and show mirror-images of the original structures. In the subsequent descriptions and in the homology discussions (chapter 6.) the terms "left" and "right" will also be exchanged. (This will be practised in other Blaberidae, too: *Blaptica, Byrsotria, Nauphoeta*). The natural orientation is shown in fig.295b and 297b.

Left complex

The left complex resembles that of *Parcoblatta*. Again, a deep infolding from the posterior (**fpe** in fig.299-302) divides the left complex into a left part with the retractable **hla**-hook and its **L3**'-sclerite and a right part with the long **L2**'-sclerite.

The left part has the shape of a bulge whose left and ventral walls are occupied by sclerite L4U'. The hla-hook evaginates from the posterior wall of the bulge; when retracted, hla lies in the center of the bulge (fig.295a, 299; all figures show hla in a more or less retracted state). Only a small distal part of hla is sclerotised by L3'. Most of the extensive membranous basal part (30 in fig.299-302) of hla becomes introverted when hla becomes fully retracted. hla can be retracted more deeply than in the previous species. (Full retraction is shown in fig.295a, with the sclerotised part of hla completely veiled by membrane; in the other figures hla is only partly retracted – to an extent corresponding to the maximal retraction in *Parcoblatta*). The leftward-bent terminal part of hla has a groove along its anterior surface (hge in fig.297a), whose ventral wall has a distinct notch (45 in fig.297a).

The part to the right of the **fpe**-infolding has in its center a deep invagination to the anterior (pouch **lve** in fig.299, 300) and a sclerotised process to the posterior (**via** in fig.299); both contain parts of sclerite **L2'**. The anterior part of the **lve**-pouch is a short tube-like apodeme (**lve**-apodeme) with a flattened and broadened top. Most anteriorly the **lve**-apodeme is sclerotised all around, more posteriorly the right wall is membranous (**44** in fig.297a, 299, 300). Roughly one third down from the top of **L2'**, the ejaculatory duct **D** joins the **lve**-

pouch from the right (fig.299), and **lve** becomes broader. In the area posterior to this confluence, **L2'** is a groove-shaped sclerotisation in the left edge of the **lve**-pouch (cross-section in fig.301). This groove-shape of **L2'** extends posteriad as far as to the posterior end of edge 7 (fig.301), where **L2'** becomes completely restricted to the ventral wall of the **lve**-pouch. The phallomere-gland (**P** in fig.299-301) opens next to this point. The posteriormost part of **L2'** sclerotises the **via**-process (fig.299-302).

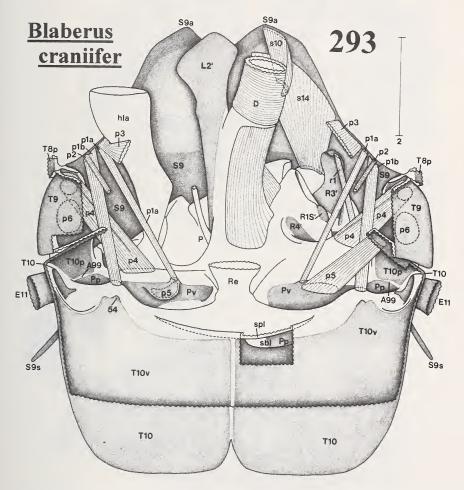
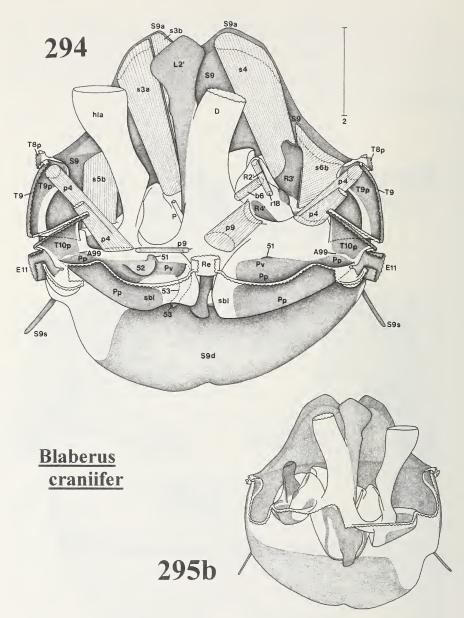
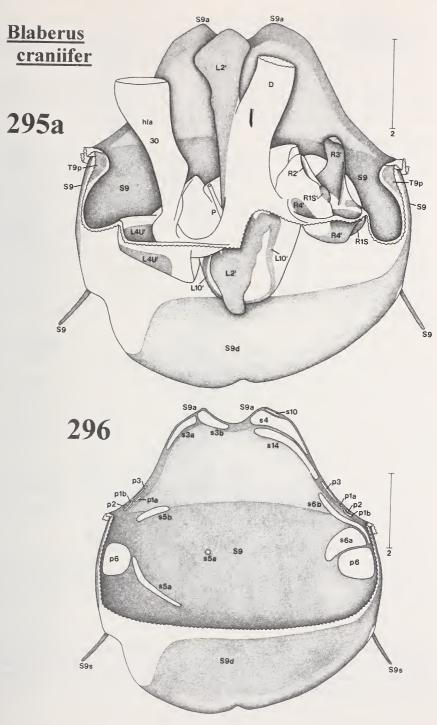


Fig.293: *Blaberus craniifer* (Blattaria, Blaberidae) – Mirror-image of the original preparations. – Male postabdomen in dorsal view; with phallomere complex, subgenital plate, marginal parts of abdominal tergites 9 and 10, supraanal lobe, subanal lobes (covered), paraprocts, distal part of rectum, basal parts of cerci, and part of musculature. Right part of supraanal lobe shown through a window cut into ventral sclerotisation of abdominal tergite 10 T10v. – Scale: 2mm.



Figs.294,295b: *Blaberus craniifer* (Blattaria, Blaberidae) – **294:** Male postabdomen as in fig.293, after removal of further parts of abdominal tergites 9 and 10 (especially T10 and T10v) and supraanal lobe. Distal part of rectum and basal parts of cerci cut open. Another part of musculature shown. – Mirror-image of the original preparations. – Scale: 2mm. – **295b:** Same as in fig.295a (next plate) but smaller scale and natural orientation (no mirror-image).



Beneath via, the ventralmost part of the left complex forms a broad ventral lobe vla, whose edges are sclerotised by the tuberculate L10'. The right anterior end of L10' is connected with L2'. The genital opening is more or less right-dorsal to the middle part of sclerite L2', though it is hardly possible to define its exact position.

The membranous tendon **ate** has its origin in the ventral basal line of the left complex (**Bl** in fig.297a, 302); it is a short and broad invagination of the cuticle. To the left of **ate** there is another small membranous invagination (**55** in fig.297a, 302).

Right phallomere

Sclerite **R3'** occupies the anterior (right-)ventral wall (fig.308-312a). Its right part has a short extension to the posterior (fig.308, 312a). The **age**-groove or -apodeme is very broad at the anteriormost margins of **R3'**; to the posterior it soon decreases and ends on both sides (fig.297a, 312a). Sclerite **R2'** adjoins posterior to the left part of **R3'**; the two sclerites are broadly separated by membrane (at **A7** in fig.312a). The ventral anterior tip of **R2'** lies in a small membranous pouch (**56** in fig.308, 312a,b, 313). From here **R2'** extends left-dorsad and forms a ridge (fig.310, 311). At its left end **R2'** is fused to the large sclerite **R1T'** (at **A6*** in fig.310, 313). Next to its fusion with **R2'**, **R1T'** has a bulge-like cuticular thickening to the interior (**cwe** in fig.308-310, 313). Posterior to the central part of **R3'** the ventral wall of the right phallomere curves dorsad and anteriad to form a rather indistinct central invagination (**cbe** in fig.308, 310, 313; compare fig.312a and 313).

The two large sclerites **R1T'** and **R4'** adjoin posterior to the right part of **R3'** (fig.308, 309, 312a). **R1T'** is loosely articulated with **R3'** (**A3** in fig.309, 310, 312a). **R1T'** and **R4'** are the sclerotisations of two lobes lying one above the other, which compose the posterior part of the right phallomere (**fda** and **dla** in fig.308, 309, 313). The left end of the **fda**-lobe is somewhat pointed (**58** in fig.309, 312a), and next to this point **R1T'** is fused to **R2'**. **R4'** mainly occupies the dorsal wall of the **dla**-lobe (fig.308); its right end curves into the ventral wall of the phallomere (**59** in fig.308, 309), where it closely approaches articulation **A3**.

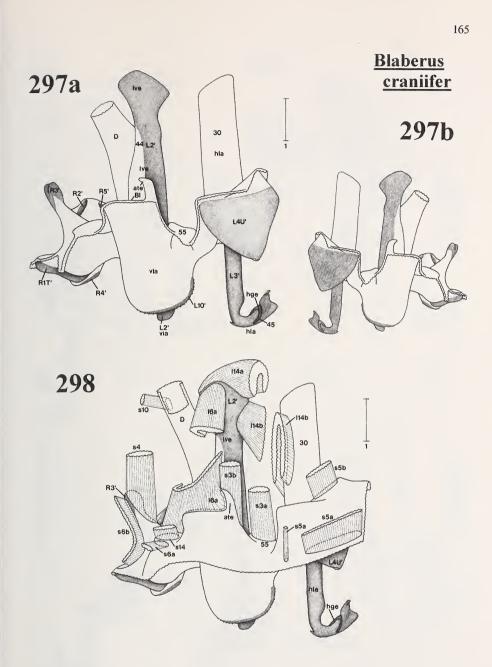
The bulged sclerite **R5'** lies in the left-ventral part of the right phallomere (fig.312a; removed from the other elements in fig.311; cut through in fig.309). The right phallomere can be retracted and protracted, and during this movement **R5'** flaps back and forth (compare fig.312a and 312b).

Subgenital plate and posterior abdominal segments

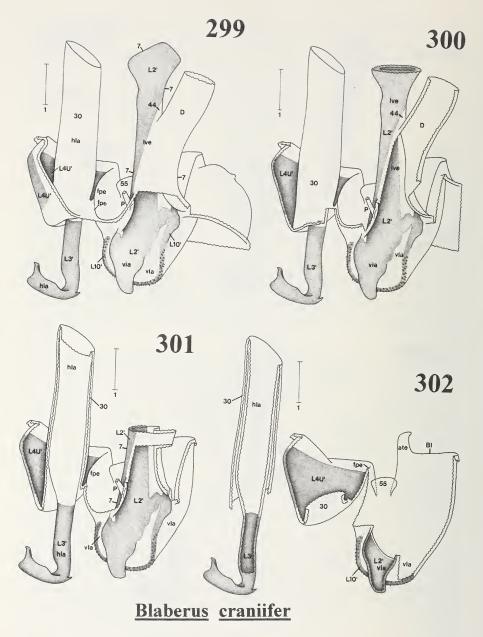
Fig.293, 294 (posterior segments); fig.296 (subgenital plate **S9**). The entire tergite 10 **T10**, including its ventral part **T10v**, is divided along its midline. **T10v** is very extensive; it has a pair of extensions to the anterior, which bear node-like apodemes (**54** in fig.293).

[←] ____ p.163

Figs.295a,296: *Blaberus craniifer* (Blattaria, Blaberidae) – All figures show mirror-images of the original preparations. – **295a:** Male postabdomen in dorsal view; with phallomere complex, subgenital plate, and lateral parts of abdominal tergite 9. – **296:** Subgenital plate in dorsal view; with insertion areas of muscles (including p6). – Scale: 2mm.

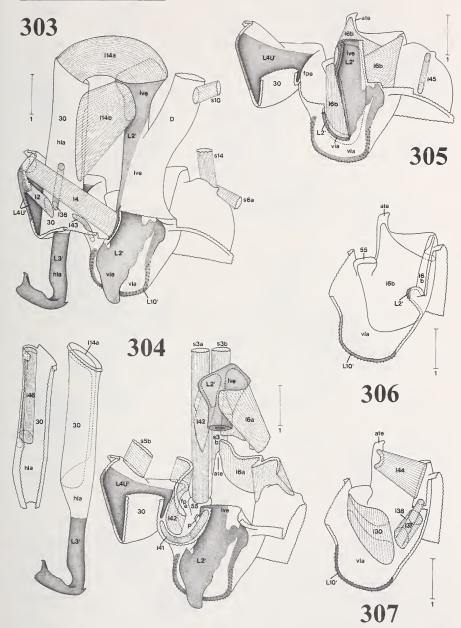


Figs.297,298: *Blaberus craniifer* (Blattaria, Blaberidae) – All figures except 297b show mirror-images of the original preparations. – **297a:** Phallomere complex in ventral view. – **297b:** Same as in fig.297a but smaller scale and natural orientation. – **298:** Phallomere complex in ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.297a. – Scale: 1mm.



Figs.299-302: *Blaberus craniifer* (Blattaria, Blaberidae) – All figures show mirror-images of the original preparations. – **299:** Left complex in dorsal view. – **300-302:** Left complex in dorsal view; with successive removal of its parts (mainly of dorsal ones); fig.302: hook hla separated from remainder of left complex (compare fig.301). – Scale: 1mm.

Blaberus craniifer



There are no separate **Pv**-sclerites; the **Pv**-sclerotisations are assumed to be incorporated into the anterior parts of the paraprocts **Pp** (fig.294; a deep indentation at the median margin of the left paraproct possibly marks the border between **Pp** and **Pv**). On their anterior margins the assumed **Pv**-sclerotisations bear the anteriad-directed node-like apodemes **51** (both sides, smaller on the right) and the posteriad-directed apodeme **52** (left side only). The sclerites **Ca**, **Cb**, and **Cc** are missing. The bulges next to the cercal bases the **Ca**-sclerites lie upon in the other species, however, are present (compare fig.263). The articulations **A99** are well-developed; the articulations **A98** are missing: The sclerotisations **E11** and **T10** are far away from each other. The left subanal lobe **sbl** bears a finger-like process (**53** in fig.294: mostly veiled by membrane) in its anterior ventral wall.

Musculature

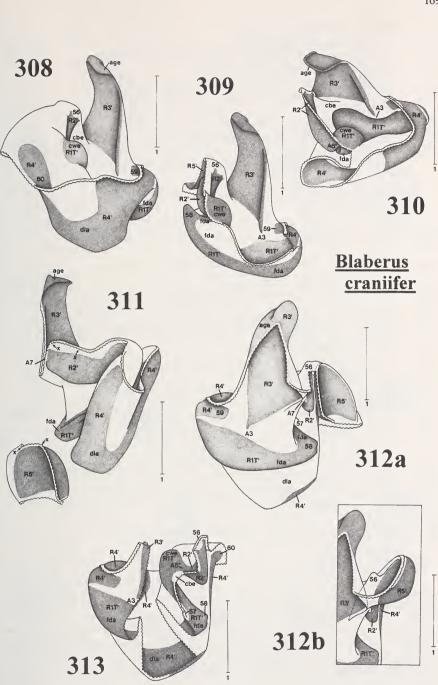
Muscles	Positions of insertions	in fig.
12	Membranous basal part 30 of hla-hook – L4U' (dorsal part)	303
14	L2' (left-posterior part) – L4U' (anterior part in left edge of left	
	complex) and membrane anterior to L4U'	303
16a	Anteriormost ventral wall of left complex and anteriormost ventral	
	wall of genital pouch - L2' (anteriormost right edge of	
	lve-apodeme)	298, 304
16b	Central ventral wall of left complex - L2' in posterior ventral wall	
	of lve-pouch; diffuse	305,306
l14a	L2' (most anteriorly on lve-apodeme) – hla-hook (left wall anterior	
	to L3')	298, 303, 304
l14b	L2' (anterior left wall of lve-apodeme) – right part of membranous	
	basal part 30 of hla-hook (insertion area horseshoe-shaped)	298, 303
130	Longitudinally in ventral wall of left complex (only to the right of	
	fpe-infolding); diffuse	307
136	Longitudinally in membranous basal part 30 of hla-hook	303

← ____ p.167

 \longrightarrow p.169

Figs.308-313: *Blaberus craniifer* (Blattaria, Blaberidae) – All figures show mirror-images of the original preparations. – **308:** Right phallomere in dorsal view. – **309:** Right phallomere in dorsal view; some dorsal elements removed (mainly lobe dla and sclerite R4'). – **310:** Right phallomere in left-dorsal (somewhat anterior) view; sclerite R5' removed. – **311:** Right phallomere in left-ventral view; sclerite R5' and surrounding membranes separated from remainder of right phallomere (along the undulate line between the points labelled x). – **312a:** Right phallomere in right-ventral view; – **312b:** Left part of right phallomere in right-ventral view; solerite R5' flapped to the anterior. – **313:** Right phallomere in right-ventral view; most elements shown in fig.312a removed. – Scale: 1mm.

Figs.303-307: *Blaberus craniifer* (Blattaria, Blaberidae) – All figures show mirror-images of the original preparations. – Left complex in dorsal view; each figure with some muscles; parts of left complex removed to various extents; fig.304: hook hla separated from remainder of left complex; of muscle 114a only posterior insertion area on hla shown; left picture shows part of the membranous base of hla (part of membrane 30) together with muscle 146; fig.306: of muscle 16b only ventral insertion area shown. – Scale: 1mm.



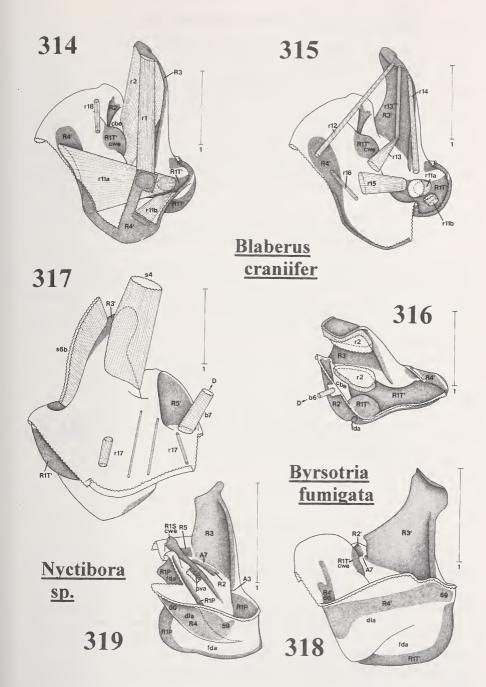
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137	In right ventral wall of left complex	307
138	In right ventral wall of left complex	307
141	Transversely in dorsal wall of left complex, between fpe-infolding	
	and opening of phallomere-gland P; diffuse	304
142	L2' (left anterior ventral wall of lve-apodeme) – fpe-infolding	304
143	Membranous basal part 30 of hla-hook – membrane to the right	
	of hla-base	303
144	ate-tendon – anterior right ventral wall of left complex; diffuse	307
145	Longitudinally in rightmost part of left complex	305
146	Longitudinally in membranous basal part 30 of hla-hook, distal	
	to 136; composed of diffuse fibers running within the insertion	
	area of 114a.	304 (left)
r1	R3' (right-anteriormost part) – R4' in right anterior dorsal wall	
	of dla-lobe	293, 314
r2	R3' – cbe-invagination: R1T' (left part) and membrane	314, 316
r11a	R4' (left edge of dla-lobe) – R1T' (right ventral wall of fda-lobe);	
	diffuse	314, 315
r11b	R4' (right dorsal wall of dla-lobe) – R1T' (rightmost ventral wall	
	of fda-lobe); diffuse	314, 315
r12	R3' (right-anteriormost part) – R4' (leftmost part); very delicate	315
r13	R3' (right-anteriormost part) – membrane posterior to	
	cwe-thickening; anterior part of r13 running within muscle r2	
	(compare fig.314), posterior part abruptly leaving r2 and running	
	to cwe; very delicate	315
r14	R3' (anterior right margin) – membrane to the right of right	
	posterior end of R3'; very delicate	315
r15	Membrane posterior to cwe -thickening – R1T' (right ventral wall	
	of fda -lobe); diffuse	315
r16	R4' (leftmost part) – membrane to the right of left part of R4';	
	very delicate, diffuse	315
r17	Longitudinally in ventral wall of genital pouch beneath right	
	phallomere; several delicate and diffuse bundles	317
r18	Membrane to the left of R2' - membrane to the left of cwe-thickening	294, 314
Fig.315 sh	ows the muscles r12 , r13 , r14 , r15 , and r16 . However, none of these	
19:010 011	one the massies rig, rie, rie, and rie is note of these	indoered n

Fig.315 shows the muscles **r12**, **r13**, **r14**, **r15**, and **r16**. However, none of these muscles was present in all of the investigated specimens, and in none of the specimens these muscles were present all together.

b6 Membrane to the left of **R2'** – ejaculatory duct **D** (posterior right dorsal wall) 294, 316

Figs.314-319: **314-317**: *Blaberus craniifer* (Blattaria, Blaberidae) – Figures 314-317 show mirrorimages of the original preparations. – **314,315**: Right phallomere in dorsal view; each figure with some muscles; dorsal elements removed to various extents. – **316**: Right phallomere in left-dorsal (somewhat anterior) view; with some muscles; some dorsal elements removed. – **317**: Right phallomere in right-ventral view; with some muscles; ventral wall of genital pouch more complete than in fig.312a. – **318**: *Byrsotria fumigata* (Blattaria, Blaberidae) – Mirror-image of the original preparations. – Right phallomere in dorsal view. – **319**: *Nyctibora* sp. (Blattaria, Blattellidae, Nyctiborinae) – Right phallomere in dorsal view. – Scale: 1mm.



172		
b7	Membrane posterior to $\mathbf{R5'}$ – ejaculatory duct \mathbf{D} (posterior right ventral wall)	317
s3a	S9 left side (far anteriorly on apophysis S9a) – ventral basal line of	
	left complex, on infolding 55	294, 296, 298,
		304
s3b	S9 left side (most anteriorly on apophysis S9a) – ate-tendon	294, 296, 298,
		304
s4	S9 right side (most anteriorly on apophysis S9a) – R3' (anterior	
	and left margin)	294, 296, 298, 317
s5a	S9 left side (far posteriorly) – left ventral wall of genital pouch	
	(far posteriorly); an additional delicate bundle runs more medially	296, 298
s5b	S9 left side (laterally) – left ventral basal line Bl of left complex	294, 296, 298, 304
s6a	S9 right side (far posteriorly and laterally) – right ventral wall of	
	genital pouch	296, 298, 303
s6b	S9 right side (laterally) – R3' (entire right margin)	294, 296, 298, 317
s10	S9 right side (on apophysis S9a) – ejaculatory duct D (right wall)	293, 296, 298, 303
s14	S9 right side (posterior to apophysis S9a) – right ventral basal line	
	Bl of left complex	293, 296, 298, 303
p1a (pair)	S9 – anterior margin of Pv-sclerotisation, on apodeme 51; very	
	delicate	293, 296
p1b (pair)	S9 – posterior (!) margin of paraproct Pp, in anteriormost dorsal	
	wall of subanal lobe sbl	293, 296
p2 (pair)	S9 – T9 (lateral anterior margin); very delicate	293, 296
p3 (pair)	S9 – rectum (ventral wall)	293, 296
p4 (pair)	T9 (lateralmost anterior margin, also extending onto paratergite	
	T9p) – membrane far anterior to Pv-sclerotisation; muscles on both	
	sides divided into two bundles in their dorsolateral parts	293, 294
p5 (pair)	T10 (lateralmost anterior margin) – anterior margin of	
	Pv-sclerotisation (left muscle on apodeme 52)	293
p6 (pair)	T9 (lateralmost part) – S9 (lateral part)	293, 296
p9 (pair)	Membrane anterior to Pv-sclerotisation – membrane median to	
	anterior margin of Pv-sclerotisation, beneath rectum; very	

5.15. Further species

asymmetrical

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In some further species only certain parts or elements of the phallomere complex have been investigated. These will be described within the respective sections of the following chapter 6.

For *Blatta orientalis*, *Deropeltis* sp., *Periplaneta americana* (Blattidae, Blattinae) and *Ergaula capucina* (Polyphagidae, Polyphaginae): Sclerites and most muscles of left complex (no figures for Blattinae; for *Ergaula capucina* the morphology of the leftmost part of the left complex is shown in fig.326d, 327d).

For *Tryonicus angustus* (Blattidae, Tryonicinae) and *Ergaula capensis* (Polyphagidae, Polyphaginae): Sclerite L1, pne-pouch (with opening of phallomere-gland), and dca-

processes (shown in fig.105-108). Only in *E. capensis*: morphology of right phallomere (shown schematically in fig.330m).

For Euphyllodromia angustata and Supella longipalpa (Blattellidae, Plectopterinae), Loboptera decipiens (Blattellidae, Blattellinae), Ectobius sylvestris (Blattellidae, Ectobiinae), Nyctibora sp. (Blattellidae, Nyctiborinae), and Byrsotria fumigata, Blaptica sp. and Nauphoeta cinerea (Blaberidae): Central part of left complex with sclerite L2, lvepouch, and via-process (all species; shown in fig.328c,d,f,g,h,i and 291, 292; no figures for Byrsotria). Morphology of right phallomere (all species except Ectobius, Loboptera, Nauphoeta; shown in fig.3300,r and 318, 319; no figures for Supella). Presence and special condition of some further elements of left complex: hge-groove, notch 45 (elements of hla-hook, compare fig.266), ate-tendon with its sclerite L4V (no figures).

6. HOMOLOGY RELATIONS AND CHARACTER STATES

In this chapter a homology hypothesis will be elaborated for the phallomere and postabdominal elements of the investigated species. This hypothesis should be as detailed as possible, and it should be provided with as many arguments as possible. The following list gives a survey which elements are discussed in which section. For the first five sections (left complex I-V) the discussed elements are listed. Which elements are contained in the

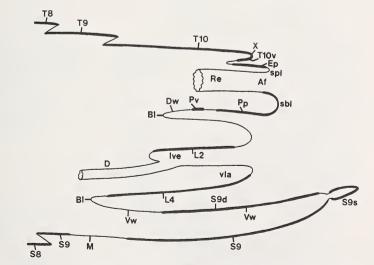


Fig.320: Male postabdomen and phallomere complex in median sagitto-longitudinal section. Left view; anterior \leftarrow , posterior \rightarrow . Only the cuticle is shown: Thin lines are membranous, thick lines are sclerotised cuticle. Styles and paraprocts are shown though they are not visible in a median section. Abbreviations in 4.7.

remaining sections is evident from the headlines. Some elements will be discussed in more than one section – according to the various aspects of their relative position.

- 6.1. Left complex I: Main sclerites L1 and L6 and associated elements (L1, L6 / pne, dca, loa, afa / l1, l2, l3, l9, l28, b4 / phallomere-gland P)
- 6.2. Left complex II: Main sclerite L2 and associated elements (L2 / lve, vla, pda, paa, via, gta, psa / l4, l5, l6, l8, l10, l12, s7 / ejaculatory duct D)
- 6.3. Left complex III: Main sclerites L4 and L10 and associated elements (L4, L10 / swe, pda, paa, vsa, via, nla, vla / 11, 12, 14, 15, 16, 17, 110, 111, 114, s1, s3, s12)
- 6.4. Left complex IV: Main sclerite L3 and associated elements (L3 / hla, hge, fpe / 114, 119, 122, 123, 136, 146)
- 6.5. Left complex V: Further main sclerites and muscles (L5, L6, L7, L8, L9, L10, L11 / lba / l7, l9, l12, l13, b2)
- 6.6. Left complex VI: The position of the phallomere-gland opening
- 6.7. The elements of the right phallomere

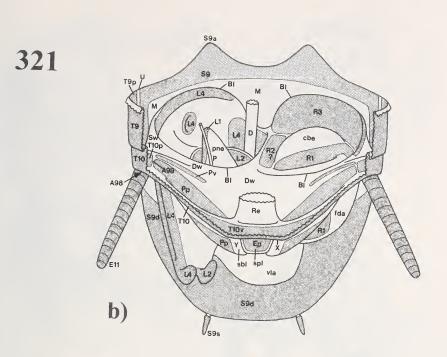
- c) Detail from left part of fig.321b after removal of some dorsal parts in dorsal view.
- d) Subgenital plate and phallomere complex in dorsal view.
- e) Left complex in dorsal view.
- f) Right phallomere in dorsal view.
- g) Left complex after removal of some dorsal parts in dorsal view.
- Right phallomere after removal of some dorsal parts in dorsal view. Ridges pia and pva shown through a window cut into ventral wall of lobe fda.
- i) Left complex in ventral view.
- Subgenital plate in dorsal view. Ventral wall of genital pouch with dorsal sclerotisation S9d of subgenital plate retained on left side but removed on right side.
- Postabdomen from segment 9 on after removal of rectum, supraanal lobe, epiproct, and of large parts of abdominal tergites 9 and 10, subanal lobes, and dorsal wall of genital pouch; dorsal view. With lateral parts of tergites 9 and 10, phallomere complex, ejaculatory duct, subgenital plate, paraprocts, ventral walls of subanal lobes, and basal parts of cerci. Dorsal lobe fda cut open lengthwise in its leftmost part. All ground-plan muscles of categories p (peripheral), s (phallomero-sternal), and b (between left complex and right phallomere) shown. p4 only shown on right side, p6 and p7 only on left side. Dorsal part of p3 (to rectum) removed.
- m)-o) Left complex after removal of some dorsal parts in dorsal view. Each figure with some groundplan muscles of category 1 (intrinsic muscles of left complex).
- p) Right phallomere after removal of some dorsal parts in dorsal view. With the ground-plan muscles of category r (intrinsic muscles of right phallomere). Ridge pia shown through a window cut into ventral wall of lobe fda.

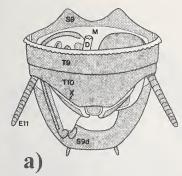
Stippled areas are sclerotised. Abbreviations in 4.7. Elements whose presence in the ground-plan of Blattaria and Mantodea is uncertain are provided with "?".

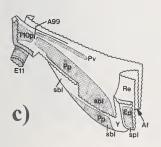
Fig.321: Male postabdomen and phallomere complex corresponding to the hypothetical ground-plan of Blattaria and Mantodea (on pages 175-177).

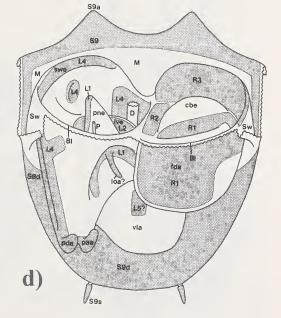
a) Postabdomen from segment 9 on in dorsal view.

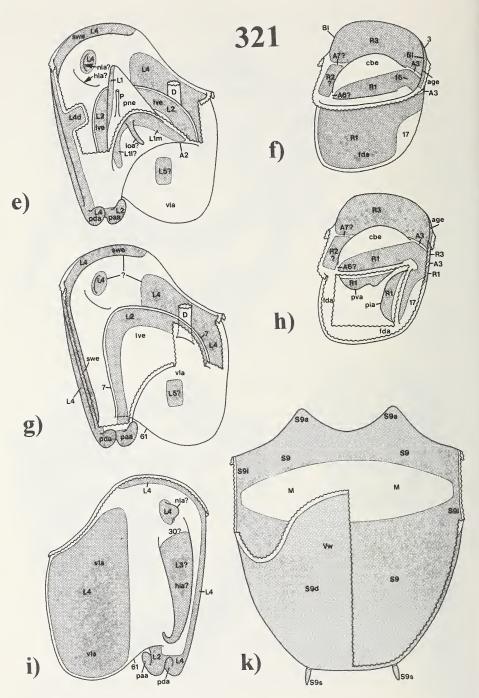
b) Postabdomen from segment 9 on after removal of central parts of abdominal tergites 9 and 10; dorsal view. With marginal parts of tergites 9 and 10, phallomere complex, ejaculatory duct, subgenital plate, paraprocts, epiproct, subanal lobes, supraanal lobe, cerci, and distal part of rectum.



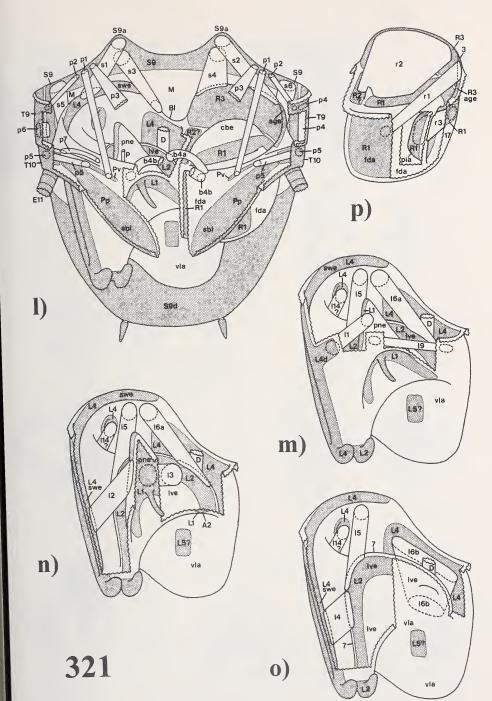


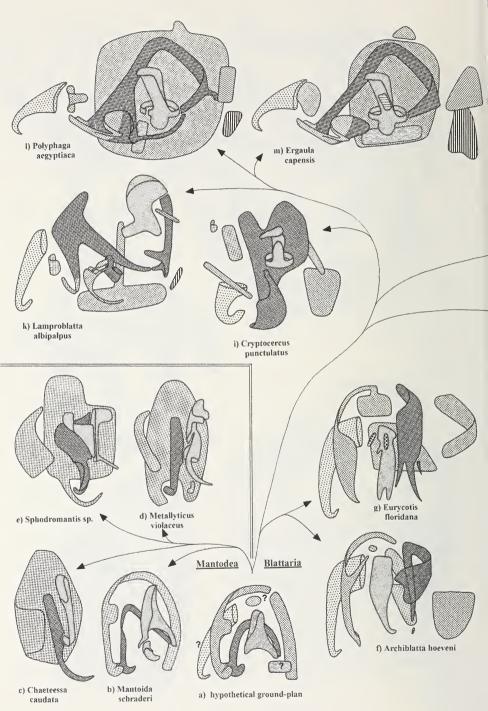












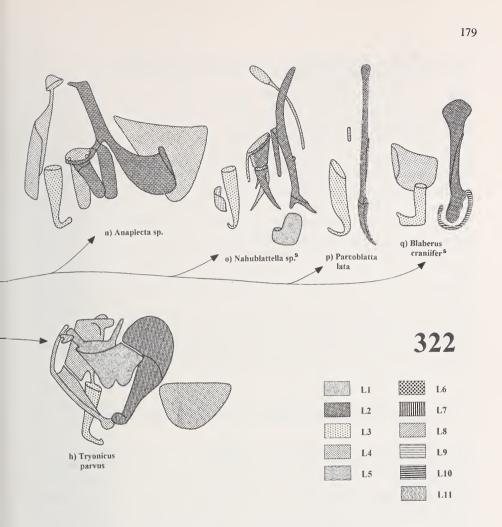


Fig.322: Left complex, evolution of main sclerites. – Only the sclerites of the left complex are shown – largely in their natural arrangement. Dorsal views. Species with "S" behind their names have side-reversed phallomeres, and a mirror-image of the original preparation is shown. The branching black lines represent the assumed phylogeny. The ground-plan is in some respects unclear (? in fig.322a, discussions in 6.3.1., 6.4., 6.5.): Presence or absence of sclerites L3 and L5; connection or separation of the L4-sclerites in the anterior ventral wall.

- 6.8. The muscles connecting the left complex and the right phallomere
- 6.9. The phallomero-sternal muscles
- 6.10. The subgenital plate and associated structures
- 6.11. The peripheral muscles
- 6.12. The terminal part of the abdomen
- 6.13. The asymmetry of the phallomere complex

In each of the sections 6.1.-6.4. and 6.7., as a first point, the homologies between Blattaria and Mantodea will be analysed. This will be done by a discussion of selected species of both groups which show similarities in the respective elements. Since Mantodea are not a subgroup of Blattaria, and vice versa, this comparison is an outgroup comparison for Blattaria as well as for Mantodea. Thus, it can serve (1) to reconstruct features of the common ground-plan of Blattaria and Mantodea and (2) to determine polarities of characters within Blattaria and within Mantodea. The ground-plan features will be shortly summarised within each section as a second point, and the complete ground-plan will be presented in chapter 7. As the third and fourth points of each section, the homologies, the special conditions, and, in part, the evolution of the respective elements will be discussed for Mantodea and then for Blattaria. The different states of the various characters, their distribution over the taxa, and their polarities will in most cases become clear from these discussions. An evaluation of the results in terms of evolution and phylogeny will be done in 7.2.-7.8.. Fig.322-333 show the homology relations of the elements discussed.

6.1. Left complex I: Main sclerites L1 and L6 and associated elements

6.1.1. Comparison between Blattaria and Mantodea

All Mantodea and most Blattaria have a pouch (**pne**) in the dorsal part of the left complex, the walls of which are largely sclerotised (sclerite **L1**). The membranous part of the **pne**-wall is on the right side in Mantodea (e.g. fig.44, 45) but left-dorsal or dorsal in Blattaria (e.g. fig.151). Arguments for the homology of **L1** and **pne** in Mantodea and Blattaria and indications for the ground-plan morphology of these elements can best be found by comparing *Mantoida* (fig.44, 45) with *Polyphaga* (fig.120, 121), *Ergaula* (fig.105, 106), and *Cryptocercus* (fig.153, 154); other Blattaria can also contribute.

In *Mantoida, Polyphaga, Ergaula* (both species), and *Cryptocercus* L1 and pne show several similarities:

- 1. The **pne**-pouch lies in the central dorsal wall of the left complex and is an invagination to the anterior.
- 2. The anterior part of L1 (region L1a in fig.323d,i,l,m) occupies most of the **pne**-wall and is hood-shaped.
- 3. The right posterior part of L1 is a distinct arm-like extension (region L1m in fig.323d,i,l,m).
- 4. A stout muscle runs from the posterior or central part of L1 to L2 in the dorsal wall of the lve-pouch: 13 (fig.50, 128, 158, 159; *Ergaula*: only *E. capucina* studied, no figure).

 Another stout muscle runs from the anterior or central part of L1 to L4-sclerotisations in or near the left edge of the left complex: l2 (fig.49, 128, 156; *Ergaula*: only *E. capucina* studied, no figure).

Mantoida, Polyphaga, and Cryptocercus have in common that:

6. The phallomere-gland **P** opens into the membranous part of the **pne**-wall. (In *Ergaula* the opening is beneath the **pne**-pouch and the **dca**-processes.)

Mantoida, Polyphaga, and Ergaula have in common that:

 The extension L1m (fig.323d,l,m) articulates with L2 (A2 in fig.45, 46, 105, 118). This articulation is rather narrow. (There is no contact between L1 and L2 in *Cryptocercus*: A2 is missing.)

Another feature is present only in Mantoida and Cryptocercus:

 A stout muscle runs from the pne-pouch to L4-sclerotisations in the dorsal wall of the left complex, the latter insertion being right-dorsal to the l2-insertion: l1 (fig.48, 155).

Regarding the **12**-insertion on **pne**, *Mantoida* is more similar to other Blattarian species (compare feature 5.):

9. In *Mantoida* (fig.49), *Eurycotis* (fig.70), and *Anaplecta* (fig.221) 12 has its right insertion in the left wall of the **pne**-pouch. (In *Cryptocercus, Ergaula*, and *Polyphaga* this insertion is on the anterior face of the **pne**-pouch.)

In *Cryptocercus, Polyphaga*, and *Ergaula* L1 has, apart from L1m on the right side, another distinct extension at its left posterior margin (region L1l in fig.323i,l,m). In *Cryptocercus* and *Ergaula* L1l joins L1m ventrally to form a complete sclerite-ring (region L1r in fig.323i,m). *Mantoida* has no L1l-extension, but other Mantodea have such an extension:

In *Metallyticus* (fig.323b) and *Chaeteessa* (fig.323c) L1 has an extension at its dorsal margin, which could well be homologous with the L1l of the respective Blattaria. (In *Mantoida*, fig.323d, the corresponding area of L1 is designated as a vestigial L1l.)

The formation of a sclerite-ring, however, does not seem to be a ground-plan element:

11. In *Mantoida*, *Chaeteessa*, *Metallyticus*, as well as in *Archiblatta* (fig.54, 55, 323f) and *Eurycotis* (fig.67, 68, 323e) the posterior part of L1 does not form a sclerite-ring.

Many Blattaria and Mantodea have distinct cuticular evaginations behind L1, which are either membranous or sclerotised by posterior parts of L1: dca (e.g. in fig.153) and loa (e.g. in fig.45, 54). The exact homology relations can hardly be determined for these formative elements. Only in some cases homology is evident, e.g. for the paired membranous cushions of *Polyphaga*, *Cryptocercus*, *Tryonicus angustus*, and probably *Archiblatta* (dca in fig.120, 153, 107, 54). It is unclear whether these dca-processes are elements of the common ground-plan of Blattaria and Mantodea and what their morphology was like in this ground-plan. As regards the process loa, *Mantoida* resembles *Archiblatta*:

12. At the posterior margin of L1 there is a completely sclerotised, curved and thorned process (loa in fig.45, 54). Its sclerotisation is connected with L1 in Archiblatta but articulated with L1 in Mantoida. However, the homology of these processes is not certain.



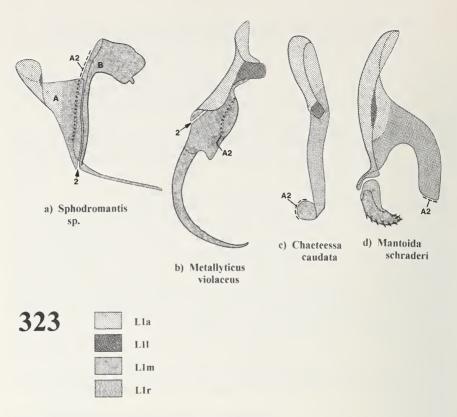
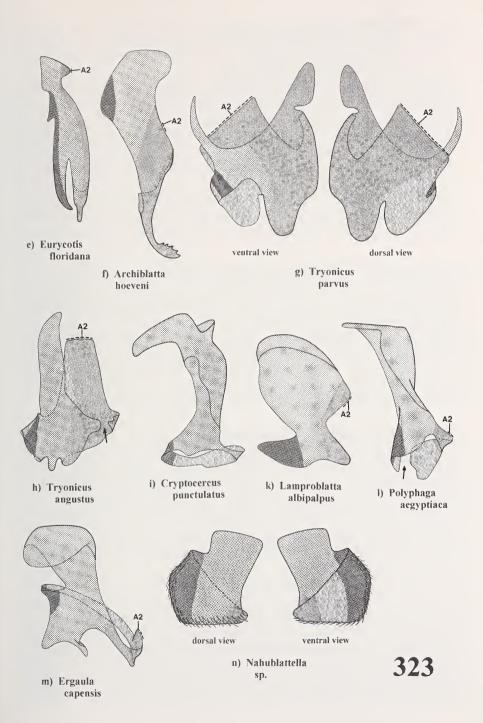


Fig.323: Left complex, homologous regions of main sclerite L1. – Only L1-sclerotisations are shown. Dorsal views, only left drawing of fig.323g and right drawing of fig.323n in ventral view. L1 is divided into the four regions L1a, L1l, L1m, and L1r (definition in 6.1.1.). If L1 is divided into several sclerites, these are labelled with the capital letters used in the text and in fig.1-319 (e.g. B = L1B). The part of the sclerite margin which forms articulation A2 with sclerite L2 is indicated by dashes; if part of this margin is covered by other parts of the sclerite, the dashes are shorter.



The membranous part of the pne-wall takes different positions within pne:

13. It is right-dorsal in *Mantoida* (fig.44, 45), dorsal in *Archiblatta* (fig.53, 54), left-dorsal in *Polyphaga*, *Ergaula*, and *Cryptocercus* (fig.117, 151). The ground-plan position can be assumed to be somewhere within this span.

This outgroup comparison suggests the similarities 1.-13. to be features of the common ground-plan of Blattaria and Mantodea (rather uncertain as regards 10.-12.).

Main sclerite L1 is divided into four regions (fig.323):

- L1a (anterior): The large anterior part of L1 within the pne-pouch (hood-shaped in most species).
- L1m (median): The extension at the median (right) posterior margin of L1a. L1m has an articulation A2 with L2.
- L11 (lateral): The extension at the lateral (left) posterior margin of L1a.
- L1r (ring): The sclerotisation connecting L1l and L1m ventrally and giving the posterior part of L1 the shape of a ring. (The ring can be complete or with a short gap.)

6.1.2. The elements in the common ground-plan of Blattaria and Mantodea

The features 1.-13. in 6.1.1. permit a reconstruction of the ground-plan morphology of **L1**, **pne**, and some adjacent elements (fig.321e,g): Region **L1a** is hood-shaped, is situated within a deep pouch **pne**, and has two posterior extensions: regions **L11** (not certain) and **L1m** (long and distinct). **L11** and **L1m** do not join each other ventrally to form a scleritering (no region **L1r**). **L1m** articulates with **L2** (A2). The membranous part of the **pne**wall is more or less dorsal and contains the opening of the phallomere-gland **P**. At the posterior margin of **L1** there is a thorned **loa**-processes (not certain). Muscles **11**, **12**, and **13** are present. **12** inserts on the left wall of the **pne**-pouch. **11** is dorsal to **12**.

6.1.3. Homology relations and character states of the elements in Mantodea

In *Chaeteessa* (fig.32, 34), *Metallyticus* (fig.24, 25), and *Sphodromantis* (fig.10) L1 and the **pne**-pouch are similar to *Mantoida* (fig.44, 45): The anterior part of L1 lies in the deep **pne**-pouch and is more or less hood-shaped. The phallomere-gland **P** (not found in *Metallyticus*) opens into the membranous part of **pne**. A large extension L1m (fig.323a-d) from the right posterior part of L1 articulates (A2) with the right part of L2.

In contrast to *Mantoida*, the membranous part of the **pne**-wall is not right-dorsal but on the right or right-ventral (*Sphodromantis*); this rotation (clockwise as viewed from behind) is regarded as derived.

The articulation area A2 is elaborated differently: In *Mantoida* A2 is exactly on the edge 1 between the pouches **pne** and **lve** (fig.45, 46). In the other species the L1m-extension bends around edge 1 into the dorsal wall of the **lve**-pouch, and here L1m and L2 lie in the same plane (fig.10, 11, 25, 26, 34, 35; fig.323a-c). This is assumed to be a derived condition. The sclerotisation bending around the edge is narrow in *Chaeteessa* but much broader in *Metallyticus* and *Sphodromantis*. In *Metallyticus*, the right, bending part of L1m is partly cut off from the basal part of L1m by the stripe of membrane 2 (fig.25, 323b). In *Sphodromantis* this separation is complete (2 in fig.10, 323a), and L1 has divided into

two sclerites **L1A** and **L1B**. This comparison with *Metallyticus* reveals that in *Sphodromantis* **L1B** is a right part of the **L1m**-region, that the articulation labelled **A2** in fig.11 is the true **A2**, and that the membranous stripe **2** is a derived feature.

The homology relations of the processes behind the **pne**-pouch and on the edge 1 between the pouches **pne** and **lve** (**loa**, **paa**, **afa** in fig.10, 25, 34, 45) are – with the exception of **paa**, which is discussed in 6.2.3. – somewhat difficult: The sclerotisation of **loa** originates in *Sphodromantis* (fig.10) and *Metallyticus* (fig.25) from that part of **L1** to the right of the membranous stripe **2** (fig.323a,b), and the base of **loa** is posterior to the bending part of **L1m**. Thus, homology is assumed for these **loa**-processes. The base of **loa** is far to the right in *Sphodromantis*, but more to the left, in the ventral wall of the **pne**-pouch, in *Metallyticus*. **loa** of *Mantoida* protrudes from the left-dorsal wall of the **pne**-pouch, but homology with the **loa** of the other species seems possible if a shift of **loa** is assumed – with the situation in *Metallyticus* being intermediate. *Chaeteessa* has no **loa**-processes. In *Sphodromantis* the sclerotisation of **loa** is reduced to a stripe in the dorsal wall (compare feature 12. in 6.1.1.).

The part of L1m bending ventrad around edge 1 sclerotises anteriorly the low bulge afa in *Metallyticus* (fig.25, 26) and the hammer-shaped afa in *Sphodromantis* (fig.10, 11); these afa might be homologous. The membranous lobes of *Mantoida* and *Chaeteessa* (afa in fig.34, 45) might be homologous with the afa of *Metallyticus* and *Sphodromantis* (not with loa), since their bases are anterior (not posterior) to the bending part of L1m. If this homology is true, in *Metallyticus* and *Sphodromantis* the L1m-region has, while becoming broader, additionally expanded onto the afa-processes.

6.1.4. Homology relations and character states of the elements in Blattaria

Ergaula, Polyphaga, and Cryptocercus

In Ergaula (fig.105, 106, 323m), Polyphaga (fig.120, 121, 323l), and Cryptocercus (fig.153, 154, 323i) L1 and pne are quite close to the ground-plan but also have some probably derived features: The anterior end of L1 is plateau-like, and the insertion of l2 has shifted to this plateau (fig.128, 156; compare 9. in 6.1.1.). The extensions L11 and L1m curve ventrad and approach each other. However, only in Ergaula and Cryptocercus the extensions unite to form a complete ring; in Polyphaga the ring is open (arrow in fig.323l). The dca-processes – with their bases encircled by the L1-ring – are very similar in Cryptocercus has a sclerotised peak (18 in fig.153) in between the dca, and the close contact between L1m and L2 (A2-articulation) has been lost (fig.151).

Tryonicus angustus and T. parvus

In *Tryonicus angustus* (fig.107, 108, 323h) L1, pne, and dca are similar to the previous species: The pne-pouch is very distinct and deep. The opening of the phallomere-gland has the same position as in *Polyphaga* and *Cryptocercus* (compare fig.107 and 120, 153). L1 articulates with L2 (A2 in fig.107, 108). The shape of the dca resembles *Polyphaga* and *Cryptocercus* (fig.107, 120, 153). The extensions L11 and L1m are distinct (fig.323h) and form a (open) sclerite ring encircling the dca-processes. The sclerotised peak 18

(fig.107, 108) resembles that of *Cryptocercus* (18 in fig.153), but its sclerotisation is connected with the L1-ring dorsally and ventrally.

Some features are certainly derived (compared with the previous species and with the ground-plan): L1a (fig.323h) and pne are flat (not hood-shaped). The L1-ring is not complete since L1m has a gap between its base on L1a and A2 (arrow in fig.323h; this situation differs from *Polyphaga* where the ring has a gap ventrally between L11 and A2: arrow in fig.323l). A2 has become a broad hinge-like articulation, and the part of L1m next to A2 is strongly enlarged (compare fig.323h and i,l,m). L1, pne, and dca are – compared with *Ergaula*, *Polyphaga*, and *Cryptocercus* – rotated 40° (counterclockwise as seen from behind): The membranous part of the pne-wall (removed in fig.107) is on the left.

In *Tryonicus parvus* (fig.94, 95, 323g) L1, pne, and dca are even further rotated, and the membranous part of the pne-wall is ventral. Compared with *Polyphaga* or *Cryptocercus*, L1 and pne are rotated 120°; compared with e.g. *Sphodromantis*, where L1 and pne are rotated in the opposite direction, the angle of rotation is 300°. Therefore, in comparing *T. parvus* with the other species, L1 should be viewed from ventrally (fig.323g, left picture). The anterior part of L1 (L1a in fig.323g) is a flat ribbon in the dorsal wall (rotation! former ventral wall) of the distinct but narrow anterior part of the pouch pne (fig.95). The position of the phallomere-gland opening is, having the L1-rotation in mind, exactly the same as in *Cryptocercus* or *T. angustus*. The sclerotisation of the two bulges dca posterior to L1a can be interpreted (fig.323g) as a complete sclerite-ring composed of the regions L1m, L1l, and L1r (like in *Ergaula* and *Cryptocercus*) and an additional expansion of L1 onto dca. The L1l-arm runs mesad because of the L1-rotation. L1m first extends far laterad, then it turns to the left, where it forms, like in *T. angustus*, a large plate and a broad hinge-like articulation A2 with L2.

Archiblatta, other Blattinae, and Eurycotis

In Archiblatta L1, pne, and dca (fig.53, 54, 323f) can be easily identified: They take a position in the central dorsal wall of the left complex. The anterior part of L1 (L1a-region in fig.323f) lies in a pouch pne. At its right margin L1 articulates with L2 (A2 in fig.54). The dca are membranous cushions at the left-posterior margin of L1 (fig.54); however, the dca are not very similar to those of e.g. *Cryptocercus* (fig.153). loa resembles loa of Mantodea (feature 12. in 6.1.1.). Some features can be regarded as derived: The pne-pouch is less deep and distinct than in all species discussed before (fig.53, 54). The L1a-region has become level as in *Tryonicus*. The phallomere-gland (P in fig.56) opens in the same position as in *Ergaula* – beneath the dca-processes (fig.54-56, 105, 106). (This situation has certainly been achieved independently in *Ergaula* and *Archiblatta*). There are no distinct arms L11 and L1m (and also no ring-formation or region L1r). The vestiges of L11 and L1m can be localised according to their characteristic relative positions (fig.323): L11 is left-anterior to the dca-cushions; L1m is right-anterior to the dca-cushions and bears articulation A2.

In other Blattinae (with *Deropeltis*, *Blatta*, and *Periplaneta* studied) L1 is similar to *Archiblatta*, but the dca-processes are rather variable, and the pne-pouch is less distinct

(as in *Eurycotis*, see below). The musculature of these species (not studied in *Archiblatta*) confirms the assumed homologies for L1 and pne: Like in e.g. *Mantoida* or *Cryptocercus*, there is a stout muscle from L1a to L4-sclerotisations (compare fig.53: L4C) in the left edge of the left complex (l2) and another one to L2 (compare fig.55) in the dorsal wall of the lve-pouch (l3). Muscle l1 is missing. A derived feature peculiar to Blattinae (and *Eurycotis*, fig.70) is the shift of the left insertion of muscle b4b to the anterior summit of the pne-pouch (discussion in 6.7.1.).

In *Eurycotis* (fig.65-67) the characteristics of L1a, L1l, L1m, and A2 (fig.323e) and the position of the phallomere-gland opening (P and edge 6 in fig.54, 55, 67, 68) are quite the same as in *Archiblatta*. The pouch-shape of **pne**, however, is by far less distinct. The processes posterior to L1 could be **dca** (as labelled in fig.66, 67) or **loa** (the right one?). The insertion of **l2** (fig.70) is still on the left part of the **pne**-pouch but has shifted from L1 to the adjacent membrane. (The position of the **l2**-insertion on L4 is the same as e.g. in *Mantoida*: discussion in 6.3.1.). Like in Blattinae, muscle **l1** is missing. Muscle **l3** from L1 to L2 is represented by three bundles (**l3a,b,c** in fig.71), which together occupy the same insertion area as the **l3** of Blattinae, and an apomorphic tripartition can be assumed. Muscle **b4b** inserts, like in the Blattinae, anteriorly on the **pne**-pouch (fig.70). The origin and homology of the sclerites L6A and L6B (fig.66, 322g) only found in *Eurycotis* is questionable: new sclerites or derivatives of L1? Homology with sclerite L8 of *Ergaula*, *Polyphaga* (fig.117, 3221,m), and *Lamproblatta* (fig.105) is also not very probable.

Lamproblatta

Like in the other species, L1 lies in the dorsal wall of the left complex, its anterior part L1a (fig.323k) is inside a deep pouch (**pne** in fig.177), and its right part articulates with L2 (A2 in fig.178). Furthermore, L1 and **pne** can be identified by the characteristic muscle connections with the area of L4 in the left edge of the left complex (l2 in fig.184; the L4-sclerotisations are highly modified, discussion in 6.3.4.) as well as with L2 (l3 in fig.187; the insertion on L2 is far posteriorly). Like in *Polyphaga, Ergaula*, and *Cryptocercus*, the l2-insertion on L1 has shifted far anteriad. Muscle l1 is missing.

As compared with other Blattaria and Mantodea, L1 and pne have shifted right-anteriad. Most of the anterior part of L1 (L1a in fig.323k) is level, but, in contrast to Blattinae, *Eurycotis*, and *Tryonicus*, there is a reminiscence of the hood-shape since the anteriormost part of L1 bends into the dorsal wall of pne (fig.177, 178). This dorsal part of L1a may even be regarded as a vestige of an anterior plateau which has been inclined posteriad. Sclerite arms (regions L11 and L1m) are not distinct. The part of L1 containing articulation A2 can be designated as the vestigial L1m-region (fig.323k; that A2 in fig.178 really is A2 is shown in 6.2.4.). The demarcation of L11 in fig.323k is tentative. For the process dca (fig.177) the homology with the dca (or loa, fig.54?) of the other species is questionable. Region L1r is missing (no sclerite ring). The phallomere-gland opens, like in *Archiblatta* or *Ergaula*, into the membrane extending ventrad from the posterior margin of L1 (P in fig.178); however, parts of L2 and L4 (with the processes paa and pda, fig.178) have shifted into the interspace between L1 and the opening (compare in 6.6.4.).

Anaplecta

In the previous species L1 and **pne** are in the central dorsal wall. In *Anaplecta* the corresponding area is membranous and just somewhat depressed (fig.209). This area is interpreted as the vestige of a **pne**-pouch, with L1 completely lost. This assumption is supported by the muscles l2 and l3 (fig.201, 221), which run to L4-sclerotisations in the left edge of the left complex (l2) and to L2 in the dorsal wall of the lve-pouch (l3). These are the same connections as in the species discussed before. l1 is, like in some other species, missing. (The homology of L4 and L2 is discussed in 6.2.4. and 6.3.4). Further support comes from McKittrick (1964): She identifies in another species of *Anaplecta* ("sp. C") a sclerite L1 (McKittrick's fig.112), which has the same position as the assumed **pne**-vestige in the species I studied.

Nahublattella

L1' and pne (fig.242, 243) show some characteristic features: They take a dorsal position. The level anterior part of L1' (L1a in fig.323n) occupies the ventral wall of a pouch pne. The posterior part of L1' completely sclerotises a bulge-like process (dca in fig.243, 244). This is interpreted as a sclerite-ring (regions L1l, L1m, and L1r in fig.323n) encircling the dca-process which has spread posteriad over the whole dca (similar to but more complete than in *Tryonicus parvus*, compare fig.323g and n). Further arguments for homology come from the muscles on pne: 11 (fig.249) runs leftward to the sclerite-ribbon L4d' (which is probably homologous with L4d of *Mantoida*, fig.44, and *Cryptocercus*, fig.150: discussion in 6.3.4.; compare 11 of *Mantoida*, fig.48, and *Cryptocercus*, fig.155). I3 (fig.250) runs to L2 in the dorsal wall of the lve-pouch (compare I3 of e.g. *Mantoida*, fig.50, *Polyphaga*, fig.128, *Cryptocercus*, fig.158, 159, and *Anaplecta*, fig.221); the homology relations of L2 and lve are discussed in 6.2.). I9a (fig.249) runs to the membrane to the right of L1' – as do I9 in *Anaplecta* (fig.221) and the posterior part of I9 in *Polyphaga* (fig.127, 129).

The right insertion of 12 (fig.249) has shifted away from the **pne**-pouch (discussion in 6.3.4.). Muscle 128, with both insertions on L1', is peculiar to *Nahublattella* (fig.250). Like in *Cryptocercus*, but in contrast to the other species, L1 and L2 are no longer in contact (articulation A2 lost).

Parcoblatta and Blaberus

There are no vestiges of L1 and pne. Muscles l1 and l3 have been lost. l2 has shifted in the same way as in *Nahublattella* (discussion in 6.3.4.).

6.2. Left complex II: Main sclerite L2 and associated elements

6.2.1. Comparison between Blattaria and Mantodea

In the Mantodean and in several Blattarian species the L2-sclerotisations, the lve-pouch, the vla-lobe, the processes **paa** and **pda**, and the genital opening show the same principal arrangement and similar positions relative to **pne** and L1. The proportions of these

elements, however, can be very different. To determine the homology relations between Blattaria and Mantodea and to reconstruct the ground-plan a comparison between *Mantoida, Polyphaga, Tryonicus, Archiblatta*, and *Eurycotis* is most useful. Some L4sclerotisations and the **pda**-process will be considered in this section, but the homology discussion of these elements will be completed in 6.3.

The right parts of L2 and of the lve-pouch are level in Mantodea (fig.11, 26, 34, 46) but curve dorsad and back to the left in most Blattaria (fig.54, 55, 94, 95, 118, 122). If this up- and recurved area is extensive, the walls of the lve-pouch cannot be designated as dorsal and ventral: The wall corresponding to the dorsal lve-wall of Mantodea (containing L2) is the inner lve-wall; the wall corresponding to the ventral lve-wall of Mantodea (mostly membranous) is the outer lve-wall (compare in 5.5.-5.7.). The respective walls of lve are homologous, the absence or presence of a curvature being the only difference. The vla-lobe usually shows the same curvature (in the figures vla is often pulled to the right), but the walls of vla will be designated throughout as dorsal and ventral.

Mantoida and Polyphaga have a lot of features in common:

- 1. The pouch **lve** (fig.46, 122) is flat and lies ventral to the pouch **pne**. However, in *Polyphaga* **lve** spans almost the whole width of the left complex and is on the whole very large, whereas in *Mantoida* **lve** is restricted to the right part and much smaller.
- Sclerite L2 (fig.46, 122) is arch-shaped and extends along the anterior and lateral edges of the lve-pouch (edges 7 in fig.46, 122). However, in *Mantoida* L2 is mainly restricted to the dorsal lve-wall (only its leftmost part bends into the ventral wall, compare fig.46, 47), whereas in *Polyphaga* L2 bends into the ventral wall along the edge 7 (compare 7 and L2 in fig.122 and 123).
- 3. The left part of L2 leaves the lve-pouch posteriorly, bends around the posterior edge of the left complex into the dorsal wall, and sclerotises a process (paa in fig.46, 117).
- 4. This L2-sclerotisation on **paa** is on its left connected with the sclerotisation of a closely adjacent process (**pda** in 44, 117). However, the shapes of both **paa** and **pda** are quite different in *Mantoida* and *Polyphaga*.
- 5. The right end of L2 articulates with the L1m-region (A2 in fig.45, 46, 118). However, the right parts of L2 are up- and recurved in *Polyphaga* but level in *Mantoida*.
- 6. A muscle (13 in fig.50, 128) runs from the dorsal wall of lve to pne (compare in 6.1.1.).
- 7. A muscle (14 in fig.50, 132) runs from the left edge of lve to sclerotisations in the left part of the left complex. However, the latter sclerotisations are very different in *Polyphaga* and *Mantoida*.
- 8. Ventral to the **lve**-pouch there is a broad ventral lobe (**vla** in fig.47, 123). The anterior part of the dorsal **vla**-wall is at the same time the ventral **lve**-wall.
- 9. The dorsal **vla**-wall is mostly membranous. The ejaculatory duct (**D** in fig.46, 47, 123, 124) opens into the right anterior part of this membrane.
- 10. The ventral vla-wall is part of the ventral wall of the left complex (fig.41, 115) and is largely sclerotised (by L4 or L4M, respectively).
- Two or three muscles inserting in the anterior ventral wall of the left complex run to the lve-pouch: The leftmost one inserts on the left anterior edge of lve (15 in fig.50, 133).

- 12. The anterior right muscle (or the anterior part of the right one in *Mantoida*) inserts on the right anterior edge of **Ive** (anterodorsal part of **I6** in fig.50; **I6a** in fig.133).
- 13. The posterior right muscle (or the posterior part of the right one in *Mantoida*) runs to the ejaculatory duct near its opening (posteroventral part of **16** in fig.52, **16b** in fig.132). Muscle **16** is undivided in *Mantoida*; in *Sphodromantis*, however, **16** is divided in the same way as in *Polyphaga* (compare **16a** and **16b** in fig.132, 133 and 16, 18).

All features listed for *Polyphaga* are also true of *Ergaula* (both species, muscles only investigated in *E. capucina*) – with the exception that the **pda**-process is missing.

Homology is assumed for all these similarities between *Mantoida* and *Polyphaga* and for all elements given the same name. 1.-13. are regarded as features of the common groundplan of Blattaria and Mantodea. Some of the mentioned differences between *Polyphaga* and *Mantoida* are bridged by various other Blattaria, which will be discussed subsequently; these species, however, are in some features rather different from *Mantoida* and/or *Polyphaga*.

The lve-pouch and L2 of *Tryonicus* (fig.95) are, like lve and L2 of *Polyphaga*, ventral to and to the right of the **pne**-pouch, and the right parts of L2 and lve curve dorsad and back to the left (compare fig.95 and 94). In contrast to *Polyphaga*, this up- and recurved part is by far more extensive, and it is directed anteriad and conceals the **pne**-pouch from dorsally. Some features correspond with both *Mantoida* and *Polyphaga*: The relative position of articulation A2 is the same; the dorsal lve-wall is largely sclerotised by L2; the left posterior part of L2 leaves the lve-pouch and provides the sclerotisation of an other process (**paa** in fig.94, 97), which is to the left connected with the sclerotisation of another process (**pda** in fig.94); the outer (= ventral) lve-wall is membranous; this membrane is at the same time the dorsal vla-wall (fig.87, 91) and contains the genital opening (D in fig.91, 92) anteriorly; the ventral vla-wall is part of the ventral wall of the left complex and is largely sclerotised (by L4G in fig.87).

In some features *Tryonicus* is more similar to *Mantoida* than *Polyphaga* is, and these similarities are regarded as further features of the common ground-plan of Blattaria and Mantodea:

- 14. The lve-pouch of *Tryonicus* does not extend as far to the left as in *Polyphaga* but is restricted to the right part of the left complex as in *Mantoida* (compare feature 1.; fig.46, 97, 122).
- 15. L2 does not, in contrast to *Polyphaga*, occupy the margins of the outer = ventral lvewall but is restricted to the inner = dorsal lve-wall (fig.92, 94-98); this situation approximates that in *Mantoida* (compare feature 2.).
- 16. The shapes of both **paa** and **pda** are quite similar in *Mantoida* and *Tryonicus* (fig.44, 96; compare feature 4.): both are short and bulge-like, and **paa** is somewhat upcurved.

As a consequence, some features of *Polyphaga* (and *Ergaula*) are assumed to be derived: (1) the extension of the **lve**-pouch almost to the left edge of the left complex; (2) the **L2**-sclerotisation in the marginal ventral **lve**-wall; (3) the special shapes of **paa** and **pda**.

In some other features *Tryonicus* is certainly derived: (1) L2 is much broader than in *Mantoida* and *Polyphaga* and has lost the arch-shape of the ground-plan since its dorsal part is directed anteriad (fig.94-97; compare feature 2). (2) Another feature concerns the

left parts of the **vla**-lobe and the **lve**-pouch. In all three species the invagination of the **lve**-pouch (edge **7** in e.g. fig.96-98) and the free left edge of the **vla**-lobe (edge **61** in e.g. fig.87, 98) start at the same point in the ventral wall of the left complex. Anterior to this point the ventral **vla**-wall is confluent with the remaining ventral wall of the left complex (e.g. fig.87). This point takes a different position in the three species. In *Tryonicus* it is far anteriorly (fig.87): The left edge **61** of **vla** extends far anteriad, and the invagination of the **lve**-pouch begins far anteriorly. In *Mantoida* this point is by far more posteriorly (**7** and **61** in fig.41, 47): The left edge **61** of **vla** does not extend so far anteriad, the ventral **vla**-wall is except for its posteriormost part confluent with the remaining ventral wall of the left complex, and the invagination of the **lve**-pouch begins far at the posterior edge of the left complex (fig.122, 123): The **vla**-lobe does not have a free left edge **61** at all, the ventral **vla**-wall is completely confluent with the remaining ventral wall of the left complex (fig.115), and the invagination of the **lve**-pouch begins most posteriorly. The ground-plan state of this feature is assumed to be somewhere in between the situation of *Mantoida* and that of *Polyphaga*:

17. The ventral **vla**-wall is for most or all of its length confluent with the remaining ventral wall of the left complex, the left edge **61** of the **vla**-lobe ends far posteriorly or is missing, and the invagination of the left(-ventral) part of the **lve**-pouch begins far posteriorly.

Archiblatta likewise has a second pouch (**Ive** in fig.54, 55) in the right part of the left complex. Like in *Polyphaga* and *Mantoida*, the sclerotisation **L2** (fig.55) runs like an arch along the edges of the pouch (**7** in fig.55). To the same extent as in *Tryonicus*, the right parts of **L2** and **Ive** curve dorsad and back to the left, and in this area **L2** is very broad. Similar to *Mantoida* (fig.46, 47), **L2** is mostly restricted to the inner = dorsal **Ive**-wall but bends into the outer = ventral **Ive**-wall in the posterior left part of **Ive** (at the posterior end of edge **7** in fig.55, 56). In some features *Archiblatta* corresponds with all previous species: **L2** articulates with **L1** (**A2** in fig.54, compare fig.94, 118, 45, 46). The left posterior part of **L2** leaves the **Ive**-pouch and runs onto a process (**paa** in fig.55, 56), which thus corresponds to the **paa** of the other species in this aspect of its relative position. The outer = ventral **Ive**-wall is the dorsal **vla**-wall (fig.53; **vla** is pulled to the right), is membranous, and receives the ejaculatory duct (**D** in fig.53). The ventral **vla**-wall is a right part of the ventral wall of the left complex and is largely sclerotised (by **L4G** in fig.54). Like in *Polyphaga* only, the **vla**-lobe does not have a left edge **61** but its ventral wall is entirely confluent with the remaining ventral wall of the left complex.

Derived features of *Archiblatta* are (1) that **paa** is mostly membranous, (2) that there is no connection between the sclerotisations of **paa** (L2) and **pda** (L4), and (3) that **paa** is quite far removed from **pda** (compare fig.44 and 53). The two latter features are probably correlated with a derived feature of the **lve**-pouch: (4) The posteroventral part of **lve** has strongly receded to the right (compare edges 7 in fig.122, 46, and 55) and is restricted to a narrow right part of the left complex. The anteroventral part of **lve** is still deeply invaginated to the left and has become tongue-like.

Eurycotis is similar to *Archiblatta* but in some features more derived (compare fig.67, 68 and 54, 55): (1) L2 is not an arch but a plate (probably the arms of the arch have fused).

(2) The posteroventral part of the **lve**-pouch is even more reduced than in *Archiblatta* (compare fig.55 and 68). (3) A deep notch (9 in fig.63, 69) separates the right main part of the **vla**-lobe from the remaining parts of the left complex. This notch lies within the **vla**-lobe and does not correspond to the left edge 61 of **vla** in *Tryonicus* (compare fig.63 and 87). Like in the other species, the posterior part of L2 extends onto a process (**paa** in fig.67, 68), which might thus be regarded as **paa** (compare fig.55). That **paa** is completely sclerotised is primitve compared with *Archiblatta*, but, like in *Archiblatta*, **paa** and its L2-sclerotisation have been far removed from **pda** and its L4-sclerotisation (fig.65).

The muscles of *Eurycotis*, compared with *Polyphaga* and *Mantoida*, confirm the assumed homologies: *Eurycotis* also has a muscular connection **13** from **L2** to the posterior part of **L1** (**13a,b,c** in fig.71, **13** in fig.50, 128; compare in 6.1.) and a muscle **14** from **L2** to the left edge of the left complex (fig.71, 50, 132). The **14** of *Eurycotis* and *Mantoida* and their insertion areas are extremely similar and reveal an additional feature of the common ground-plan of Blattaria and Mantodea:

18. 14 (fig.50, 71) runs from L2 in the lve-pouch to the swe-apodeme on L4-sclerotisations in the left edge of the left complex (sclerites L4 or L4H), where it inserts immediately ventral to muscle 12 (fig.49, 70) coming from the pne-pouch.

Several muscles of *Eurycotis* run from the ventral wall of the left complex to the **lve**pouch: **l6b** (fig.70, 71) runs to the ejaculatory duct near its opening, like **l6b** in *Polyphaga* (fig.132) and the ventral part of **l6** in *Mantoida* (fig.52). Another muscle (**l6a** in fig.73) runs to the anterior ventral wall of **lve**, and homology with either **l5** or **l6a** of *Polyphaga* (fig.133) and *Mantoida* (**l6a** = dorsal part of **l6** in fig.50) seems possible. Two other muscles (**l5a,b** in fig.72) insert on the ventral left edge of the **lve**-pouch, somewhat like **l5** in *Polyphaga* (fig.133); however, the ventral insertions of **l5a** and **l5b** are far posteriorly. Thus, for **l5a,b** and **l6a** of *Eurycotis* the homologies are not completely clear, but I suppose that the relations expressed by the designations are the most probable.

Concerning the common ground-plan of Blattaria and Mantodea, one question remains open: It is not decidable whether the right parts of L2 and lve are level (like in Mantodea) or up- and recurved (like in Blattaria).

The definition of the regions of main sclerite **L2** (fig.324) is based on the primitive archshape of **L2**, which is present in its typical form in *Mantoida*, *Archiblatta*, and *Polyphaga*. From the left to the right four **L2**-regions are distinguished:

- L2d (dorsal): The sclerotisation of the process paa.
- L2p (posterior): The part of the L2-arch in the left posterior part of the lve-pouch.
- L2a (anterior): The part of the L2-arch in the anterior part of the lve-pouch.
- L2m (median): The part of the L2-arch in the right part of the lve-pouch. L2m has an articulation A2 with L1.
- L2v (ventral): This is not defined as a separate region of L2. This term is used (mainly in fig.324) if large parts of L2 have invaded the ventral wall of the lve-pouch; these parts of L2 are not necessarily homologous in the species concerned.

The up- and recurved right parts of L2 of Blattaria belong to the regions L2m and L2a.

6.2.2. The elements in the common ground-plan of Blattaria and Mantodea

The features 1.-18. in 6.2.1. permit a reconstruction of the ground-plan morphology of L2, lve, paa, and vla, and of the genital opening (fig.321e,g): The lve-pouch lies ventral to the pne-pouch; it is quite broad but does not reach the left edge of the left complex (and does not have a recess to the right in its posterior part). L2 is largely restricted to the dorsal lve-wall and runs like an arch along the lateral and anterior edges of the lve-pouch. The left part (L2p-region) of the arch leaves the pouch posteriorly, and the posteriormost part (L2d-region) sclerotises the short, somewhat upcurved process paa. L2d is connected with the sclerotisation of the process pda, and paa and pda are close to each other. The right end of L2, or the dorsal left end in the case of L2-upcurving (L2m-region), articulates with L1m (A2). The ventral lve-wall is mostly membranous and is at the same time an anterior part of the dorsal vla-wall. The ejaculatory duct D opens into the right anterior part of this membrane. The ventral vla-wall is largely sclerotised and almost completely confluent with the ventral wall of the remaining left complex (i.e. the left edge 61 of vla is missing or does not extend far anteriad). Muscles l3, l4, l5, l6a, and l6b are present.

6.2.3. Homology relations and character states of the elements in Mantodea

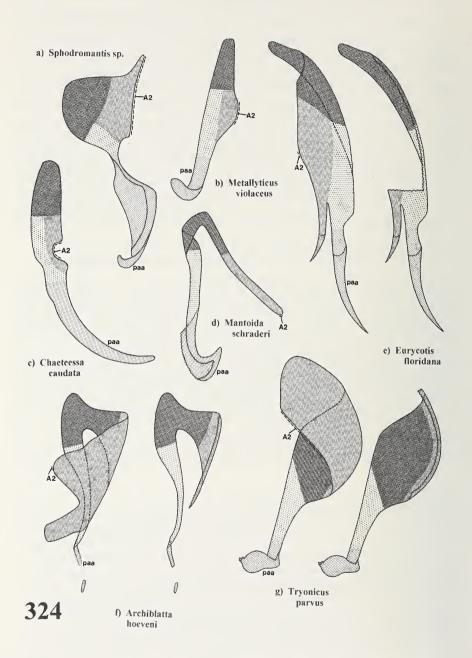
In *Chaeteessa, Metallyticus*, and *Sphodromantis*, the main features of L2, lve, and paa are like in *Mantoida* (fig.11, 26, 34, 45, 46): lve lies ventral to the pne-pouch. L2 is restricted to the dorsal lve-wall. The right parts of L2 and lve are level. Posteriorly L2 leaves the lve-pouch to provide the sclerotisation of a process which is therefore regarded as paa. The right margin of L2 articulates with L1 (A2; compare in 6.1.3.).

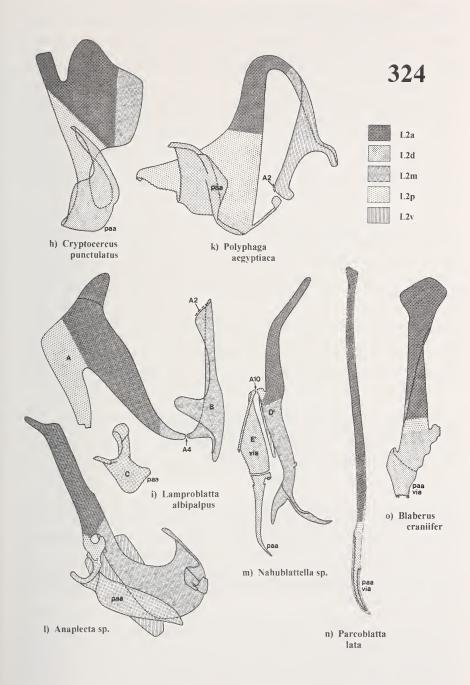
In contrast to *Mantoida, Metallyticus* and *Sphodromantis* have separated the sclerotisations of **paa** (L2) and **pda** (L4), and the processes are more distinct from each other and more prominent (fig.10-12, 23-26, 44-46). In *Chaeteessa* (fig.31, 32, 34) **paa** is very prominent, too, but a process **pda** is missing. Nevertheless, in this species, too, L2 and L4 are completely separated in the area concerned.

L2 of *Chaeteessa, Metallyticus*, and *Sphodromantis* has become plate- or ribbon-like and has spread over most of the dorsal **lve**-wall. Like in *Eurycotis*, the arms of the arch (regions

Fig.324: Left complex, homologous regions of main sclerite L2 (on pages 194, 195). – Only L2-sclerotisations are shown. Dorsal views. L2 is divided into the four regions L2a, L2d, L2m, and L2p (definition in 6.2.1.); the extensive parts of L2 in the ventral wall of pouch lve in *Polyphaga* are labelled L2v (not defined as a region of L2). If L2 is divided into several sclerites, these are labelled with the capital letters used in the text and in fig.1-319 (e.g. B = L2B). Undulate lines are cutting lines through sclerotisations (if L2 is fused with parts of other main sclerites, e.g. L4). In fig.324e,f,g the left drawing shows the complete L2, the right drawing shows L2 after removal of its dorsal parts. The part of the sclerite margin which forms articulation A2 with sclerite L1 is indicated by dashes. A4 and A10 are articulations between L2-sclerites. paa and via are processes occupied at least partly by L2-sclerotisations. The sclerite of *Parcoblatta* (fig.324n) and *Blaberus* (fig.324o) which is termed L2 in the text has to be designated correctly L2D+(L2E+L4N), compare in 6.2.4.; the L4N-part has been removed from the sclerite.







L2p and L2m) have probably fused (fig.324a-c, compare fig.324d). In *Chaeteessa* and *Metallyticus*, as compared with *Mantoida*, the **lve**-pouch has strongly narrowed (compare fig.26, 34 and 46), and L2 is also narrow. In *Sphodromantis* the anterior part of the **lve**-pouch is deeply invaginated to the left, and L2 forms a broad transverse tongue (fig.11). This resembles the situation in *Archiblatta* (compare the course of edge 7 in fig.11 and 55) – certainly a case of parallel evolution.

As in *Mantoida*, in *Chaeteessa*, *Metallyticus*, and *Sphodromantis* the membranous ventral **lve**-wall is a (left) anterior part of the dorsal **vla**-wall (fig.12, 27, 35), the ejaculatory duct (**D** in fig.11, 12, 26, 32) opens far anteriorly into the dorsal **vla**-wall, and the ventral **vla**-wall is part of the ventral wall of the left complex and is largely sclerotised (by **L4** or **L4A** in fig.6, 20, 28). The genital opening lies in *Sphodromantis* within the **lve**-pouch (like in *Mantoida* and *Polyphaga*); in *Metallyticus* and *Chaeteessa* it is a bit, or far, respectively, to the right of the pouch and outside of it.

The homology of **lve** and **L2** of *Mantoida* and *Sphodromantis* is confirmed by the musculature: **15**, **16a**, and **16b** of *Sphodromantis* (fig.15, 16, 18) show the same arrangement as **15** and **16** in *Mantoida* (fig.50, 52). Both species have a stout muscle **13** from **L2** to **L1** (fig.16, 50). Muscle **14** inserts on the left edge of **lve** (fig.15, 50) and has its opposite insertion (on **L4**-sclerotisations) immediately to the left of the dorsal **12**-insertion. However, the latter insertions of both **12** and **14** are in *Sphodromantis* by far more to the right than in *Mantoida*. This topic will be taken up again in 6.3.3. Muscle **18** of *Sphodromantis* (fig.16) consists of very few fibers at most (completely missing in some specimens). Snodgrass (1937) finds this muscle well-developed in *Tenodera sinensis* (Mantidae; muscle **13** in Snodgrass' fig.10D). Hence, **18** of *Sphodromantis* represents either a vestige or an early evolutionary stage of this muscle. Since **18** is missing in *Mantoida* and all Blattaria it is not assumed to be a muscle of the Mantodean ground-plan.

6.2.4. Homology relations and character states of the elements in Blattaria

Polyphaga, Ergaula, Tryonicus, Archiblatta, and *Eurycotis* These species have been sufficiently discussed in 6.2.1.

Lamproblatta

The pouch and the ventral lobe labelled **lve** and **vla** in fig.180 resemble in many respects the **lve** and **vla** of the previous species: The **lve**-pouch lies ventral to the **pne**-pouch. Its dorsal wall is largely sclerotised (**L2A**, **L2B**). Its ventral wall is mostly membranous, is at the same time the dorsal **vla**-wall (fig.180, 181), and contains the genital opening (**D** in fig.180, 181) in its right part. The ventral wall of **vla** is part of the ventral wall of the left complex and is partly sclerotised (**L4R** in fig.174, 181). However, in *Lamproblatta* there are two problems:

- Instead of only one sclerite L2 as in the former species there are two within the lve-pouch (L2A and L2B), and around articulation A4 the lve-pouch has a deep recess (fig.180). The questions arise if either only the part to the left of the recess (with L2A) corresponds to the lve-pouch of the other species, or if the part to the right of the recess (with L2B) is also a true part of lve, and whether L2B really is a part of L2 (or of L1:

a split off part of the L1m-region like L1B of *Sphodromantis*, fig.10?). This equals the question which of the articulations A2 (fig.178) and A4 (fig.180) is the true A2.

- In *Mantoida, Polyphaga*, and *Tryonicus* the left posterior part of L2 leaves the pouch and continues into the sclerotisation of the processes **paa** and **pda** (fig.46, 96, 118). In *Lamproblatta*, the left posterior part of L2A also has an extension that leaves the pouch (L4S in fig.178, 180), and right-anterior to this area there is a sclerotisation with two processes (**paa** and **pda** in fig.178, 179), which, however, is completely separated from L2A and L4S). The question arises if **paa** and **pda** of *Lamproblatta* are homologous with those of the other species.

These problems can be solved by a comparison of the musculature of *Lamproblatta* and the other species – especially *Polyphaga*.

- Lamproblatta also has a stout muscle from L1 to lve (l3 in fig.187; compare *Polyphaga*, l3 in fig.128). Its insertion area on lve is within the recess and also includes parts of L2B.
- Lamproblatta and Polyphaga have 4 muscles (15, 16a, s3, s12) inserting close to each other in the anteriormost ventral wall of the left complex (fig.133, 188); homology is assumed for all of them (s3 and s12 are discussed in 6.9.). In Polyphaga the two posterior muscles (15 and 16a) run to L2 at the left or right, respectively, anterior edge of the lve-pouch. In Lamproblatta 15 runs to L2A on the left of the recess, 16a runs to L2B on the right of the recess.
- The two l6b-bundles (fig.189) of Lamproblatta are assumed to be homologous with the l6b of the other species (Polyphaga: fig.132): The ventral insertion is posterior to that of l6a. The insertion in the dorsal vla-wall is somewhat different in Polyphaga and Lamproblatta (next to the genital opening or far posterior to it), but the insertions of the two bundles of Lamproblatta are similarly situated as the insertion of the one bundle of l6b of Sphodromantis (fig.18) or Cryptocercus (fig.155, 157, see below).
- Muscle 112 of Lamproblatta and Polyphaga inserts in the right part of the ventral = outer lve-wall very close to the dorsal insertion of 16a (fig.128, 129, 186, 188) and runs to a small sclerite in the right dorsal wall of the left complex (L8, homology discussion in 6.5.). Homology is assumed for these 112. In Lamproblatta 112 inserts to the right of the recess, on L2B. (L8 and 112 are derived features of Polyphaga, Ergaula, and Lamproblatta.)
- Muscle 110 of *Polyphaga* (fig.129) runs from L2 in the left dorsal wall of lve to the sclerotisation in between the processes paa and pda. *Lamproblatta* has a muscle (110 in fig.186) from the same area of lve to the sclerotisation between paa and pda. Such a muscle is missing in all species discussed before.

The course of **110** suggests homology for **paa** and **pda** of *Lamproblatta* and *Polyphaga*. Since the **pda**-sclerotisation is part of **L4**, the discussion of this topic will be continued in 6.3.4.

The insertions of 13, 16a, and 112 clearly demonstrate that L2B is a true part of L2 (regioning in fig.324i) and that the respective part of the pouch is a true part of lve. That the right part of L2B curves dorsad and leftward like the right part of L2 in other Blattaria and that the genital opening is to the right of the recess (fig.122, 180) supports this



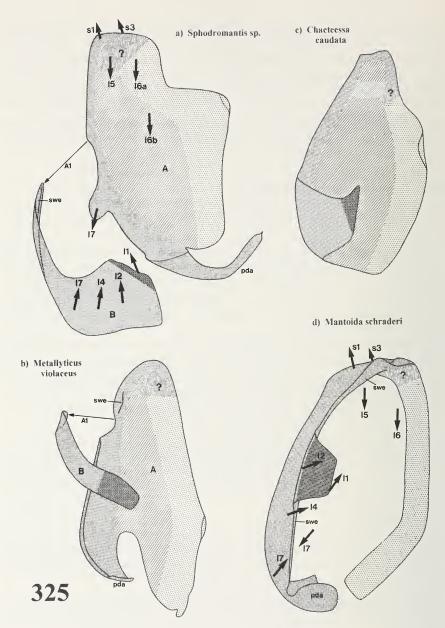
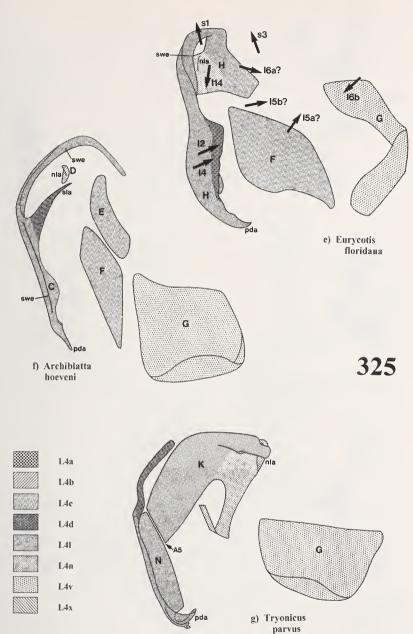
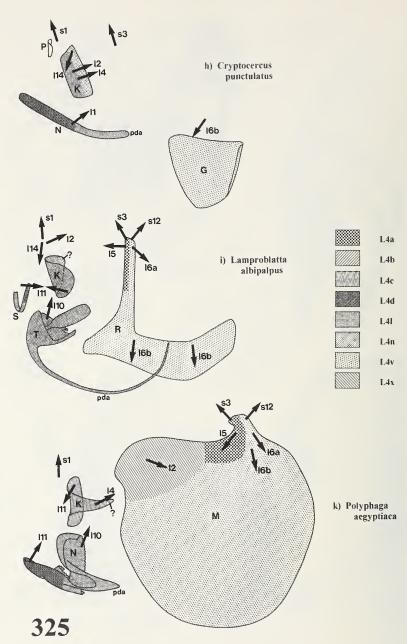


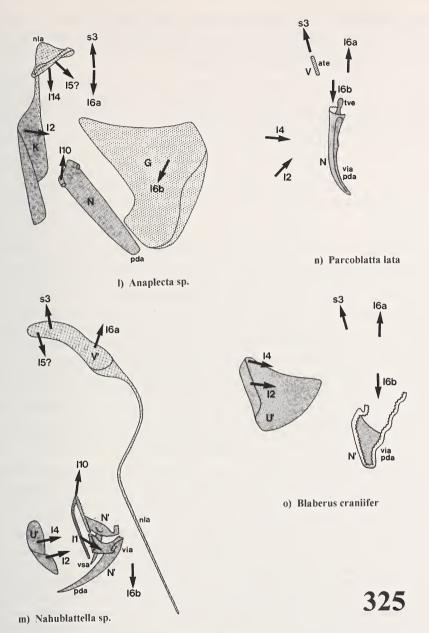
Fig.325: Left complex, homologous regions of main sclerite L4. – Only L4-sclerotisations are shown. Dorsal views. L4 is divided into the regions L4a, L4b, L4c, L4d, L4l, L4n, L4v, and L4x (definition in 6.3.1.). If L4 is divided into several sclerites, these are labelled with the capital letters used in the text and in fig.1-319 (e.g. B = L4B). The sclerites are mostly shown as they are arranged in the left



(contd.): complex; only in *Sphodromantis* (fig.325a) and *Metallyticus* (fig.325b) the dorsal sclerites **L4B** are shifted to the left (thin arrows). Undulate lines are cutting lines through sclerotisations (if **L4** is fused with parts of other main sclerites, e.g. **L2**). A1 and A5 are articulations between L4-sclerites. **pda**, via, nla, sla, vsa, tve, and swe are formative elements occupied at least partly by L4-



(contd.): sclerotisations. Broad arrows represent muscles inserted on or near L4-sclerotisations; 15 is certainly homologous in most species; if homology with these 15 is questionable, the muscle is labelled 15? (in *Anaplecta* and *Nahublattella* 15? are homologous). In Mantodea the presence of region L4c is questionable (? in fig.325a-d). The sclerite of *Parcoblatta* (fig.325n) and *Blaberus* (fig.325o)



(contd.): which is termed L2 in the text has to be designated correctly L2D+(L2E+L4N), compare in 6.2.4.; only the L4N-part is shown in the figures.

interpretation. Thus, the articulation between L2B and L1 (A2 in fig.178) is homologous with A2 of the other species. In correlation with the right-anteriad shift of L1 (compare in 6.1.4.), A2 of *Lamproblatta* is far anteriorly. The articulation A4 and the recess within the **lve**-pouch are derived features of *Lamproblatta*.

Another derived feature of *Lamproblatta* is the lack of muscle **14**. Another derived feature common to *Lamproblatta* and *Polyphaga* (and *Ergaula*) – in addition to **L8** and **112** – is that the **lve**-pouch and the **vla**-lobe extend nearly to the left edge of the left complex (compare edges 7 in fig.122 and 180).

Cryptocercus

The elements L2, lve, paa, and vla (fig.151, 152) can be clearly identified by many features corresponding with the other species: L2 lies ventral to the pne-pouch and is connected with it by a stout muscle (13 in fig.158, 159). The posterior part of L2 bends into the dorsal wall of the left complex, and the area of bending forms a bulge-like process (paa and L2d in fig.151, 152, 324h; compare Tryonicus, fig.95, 97, 324g). This dorsal part of L2 extends anteriad as far as to the opening of the phallomere-gland (P in fig.151, 152) – like in *Polyphaga* (fig.118, 120). The right anterior part of L2 occupies the dorsal wall of a pouch-like invagination (lve in fig.150-152), which, however, is restricted to the anteriormost part of the left complex. The ejaculatory duct (D in fig.150, 151; compare Mantoida, fig.46) opens into this lve-pouch. The ventral lve-wall is at the same time part of the dorsal vla-wall (fig.150-152), and the ventral vla-wall is partly sclerotised (L4G in fig.148, 152). Muscle 110 (fig.155) runs from L2 to the membrane to the left of paa. This membranous area is in between the processes **paa** and **pda** (**pda**, fig. 150, is discussed in 6.3.4.), and thus the left insertion of this 110 is like that of the 110 of *Polyphaga* (fig. 129) and Lamproblatta (fig.186). However, the right insertion is by far more posteriorly, and the homology of these 110 is not certain. Muscle 14 (fig.155, 158) runs from the anterior part of L2 to L4K in the left part of the left complex like l4 of Polyphaga (fig.132; l4 of Cryptocercus is strongly reduced; L4K is discussed in 6.3.4.).

Cryptocercus has some features which are, compared with the ground-plan, clearly derived: (1) L2 is, like in most Mantodea and *Eurycotis*, more plate-like, though the primitive arch is still recognisable (compare L2-regions in fig.324h and d,f,k). (2) The right part of L2 is, like in Mantodea, not upcurved (fig.151, 152). However, since the contact between L2 and L1 (articulation A2) has been lost, this is not interpreted as a primitive situation – as suggested by the outgroup comparison with Mantodea – but as a reduction of the right part of L2. Consequently, the right-dorsal part of the lve-pouch, which contains the upcurved part of L2 in other Blattaria, has been strongly reduced. (3) The vla-lobe is separated from the remaining ventral wall of the left complex as far as to the anterior margin of the left complex (edge 61 in fig.148). Accordingly, the invagination of the left-ventral part of lve has been strongly reduced. This is an extreme modification of the situation in *Tryonicus* (edge 61 in fig.87 and edge 7 in fig.97). (4) Of the muscles from the ventral wall of the left complex to the lve-pouch and the ejaculatory duct only one is present (l6b in fig.155). According to its insertions (anterior margin of L4G,

membrane posterior to genital opening), it is likely to be the homologue of **16b** of *Eurycotis* (fig.71), *Sphodromantis* (fig.18), and the other species. **15** and **16a** have been lost; this might be a consequence of the extreme reduction of the **lve**-pouch.

Anaplecta

The homology of the elements designated L2, lve, vla, paa, and pda in Anaplecta with the respective elements of the other species is suggested by the following features: The whole area labelled lve (fig.210-213), whose anterior part narrows to form the lveapodeme, is a large invagination to the anterior, which lies beneath the pne-"pouch" (fig.209). lve and pne are connected by a stout muscle (13 in fig.201, 222, 50, 128). L2 is mainly restricted to the dorsal wall of lve. Anteriorly, however, L2 also occupies the margins of the ventral lve-wall (fig.211, 225) - similar to Polyphaga (fig.123) and Lamproblatta (fig.181). That a phallomero-sternal muscle inserts on the lve-pouch resembles Eurycotis (s7 in fig.58, 200). That L2 forks at the base of the lve-apodeme is regarded as a vestige of the primitive arch-shape (compare fig.324l and d,k); this is confirmed by the morphology of the two branches of the fork: The right branch is upcurved at its right margin (fig.211-213), like the right part of L2 in other Blattaria. The cuticular area containing this part of L2 can therefore be regarded as the right dorsal (= inner) wall of the lve-pouch. Anterior to this right L2-part there opens, like in the other species (e.g. Mantoida, fig.46), the ejaculatory duct (D in fig.211). The left branch of L2 continues into a sclerotisation bearing two processes (paa in fig.211, pda in fig.214). This is the same situation as at the left end of the L2-arch of Mantoida, Polyphaga, and Tryonicus. Muscle 110 runs, like 110 of Polyphaga and Lamproblatta (fig.222, 129, 186), from the sclerotisation of paa and pda to L2 in the left-dorsal lve-wall. The relative positions of the vla-lobe (fig.205, 218-220) and its sclerite L4G (fig.205) in the ventral wall of the left complex are especially similar to those of vla and L4G of Tryonicus (fig.87, 205), with vla having a left edge (61 in fig.205) reaching far anteriad (farther than in Tryonicus, fig.87, but not as far as in Cryptocercus, fig.148).

In contrast to all other species, the edge of the **lve**-pouch is – except in the area of the **lve**-apodeme – not continuous throughout (compare edges 7 in fig.55, 122, 180, 211, 212) but interrupted by some apomorphic membranous foldings (fig.212-219): e.g. outfolding **vfa**, infolding **vpe** (fig.214, 215, 217). For that reason it is difficult to determine the homologies of the muscles of this area with the **l5**- and **l6**-muscles of the other species. **l6b** (fig.224) is probably homologous with **l6b** of e.g. *Sphodromantis, Eurycotis, Cryptocercus*, and *Lamproblatta* (fig.18, 71, 155, 189): All these **l6b** run from the sclerite plates in the ventral **vla**-wall, or from their vicinity, to the dorsal **vla**-wall. In *Anaplecta*, however, the dorsal insertion is not immediately behind the genital opening but is separated from it by the outfolding **vfa** (compare fig.223 and 224). **vfa** is therefore assumed to be evaginated from the anteriormost dorsal wall of **vla** and the ventral wall of the ejaculatory duct. This assumption is supported by two other muscles: **s10** inserts on the ejaculatory duct in *Nahublattella* (fig.229) and *Parcoblatta* (fig.132), *Cryptocercus* (fig.155), and *Eurycotis* (**l13h** in fig.72) runs from the ejaculatory duct to the dorsal **vla**-wall posterior to it. **l13**

of *Anaplecta* (fig.222) also inserts on the ejaculatory duct but bridges the **vfa**-outfolding on its way to its insertion on the dorsal base of **vla** (discussion of **I13** in 6.5.). Muscle **I6a** of *Anaplecta* (fig.222) resembles **I6a** of *Polyphaga* and *Lamproblatta* (fig.133, 188) in inserting ventrally behind **s3** and dorsally at the right anterior edge of the **lve**-pouch. Whether muscle **I5** is homologous with **I5** of the other species (fig.133, 188, 223) is questionable: The insertion on **L2** is similar in *Anaplecta* and e.g. *Polyphaga*; the anterior insertion (on **L4**), however, is situated quite differently in these two species. Homology is also unclear for the muscles **I25** and **I26** (fig.224).

As compared with the previous species, *Anaplecta* has some important derived features: (1) The anterior part of the **lve**-pouch is a tube-like **lve**-apodeme. (2) Edge 7 is interrupted by **vfa** and **vpe**. (3) The common sclerotisation of **paa** and **pda** is stout and ring-shaped at its base. (4) Muscle **l4** from **L2** to left parts of **L4** has been lost (like in *Lamproblatta*).

Nahublattella

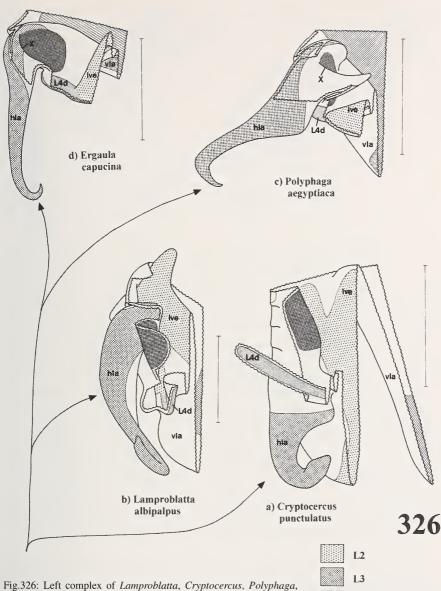
The part of the left complex comprising the large pouch **lve** (fig.242), the opening of the ejaculatory duct (**D** in fig.242), the processes, **via**, **vsa**, **paa**, and **pda** (fig.244, 245), and the sclerotisations **L2D'**, **L2E'**, and **L4N'** show a lot of similarities with the elements of *Anaplecta* discussed before:

All these elements lie in the center of the left complex and (antero-)ventral to the **pne**pouch. The anterior part of the **lve**-pouch (see edges **7** in fig.242) is a tube-like **lve**apodeme, whose dorsal wall is completely sclerotised, and whose ventral wall contains a membranous stripe (**44** in fig.206, 212, 239a, 245). Muscle **s7** runs from the **lve**-apodeme to the subgenital plate (fig.200, 234).

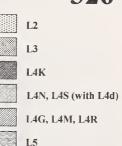
At the base of the apodeme, L2D' is somewhat forked (fig.243, 324m), like L2 in *Anaplecta* (fig.212, 324l): Extension 36 is the left branch, the posterior main part of L2D' is the right one. At the left branch there adjoins a ring-shaped sclerotisation lying at the base of some processes (paa and pda in *Anaplecta*, fig.211-214; via with vsa, paa, and pda in *Nahublattella*, fig.244, 245). A stout muscle 110 (fig.222, 250) runs from the lve-apodeme to the left part of this sclerite-ring. However, in *Nahublattella* the sclerite ring has become separated from the rest of L2 (L2D') by an articulation (A10 in fig.244). At the base of the left branch the L2-sclerotisation bends into the ventral lve-wall and forms a posterior extension (28 in fig.215, 245). However, in *Anaplecta* the cuticle around extension 28 forms a process (gta in fig.215), which is missing in *Nahublattella*.

The right branch of L2 or L2D', respectively, extends rightward in *Anaplecta* but more posteriad in *Nahublattella*. The relation between this sclerotisation and the dorsal wall of the ejaculatory duct (D in fig.245, 246) is the same in the two species (fig.211, 245), but only in *Nahublattella* the right-anterior margin of this L2-part folds narrowly back to the left (towards edge **38** in fig.245).

Two further muscles inserting on L2D' of *Nahublattella* correspond with muscles of the other species: 13 running to the **pne**-pouch (fig.250, compare e.g. fig.50, 71, 128, 221), and 14 running to L4-sclerotisations in the left part of the left complex (fig.249, compare e.g. fig.50, 71, 129; missing in *Anaplecta*; homology discussion of L4 in 6.3.4.).



Pig.320: Left complex of *Lamproblatta*, *Cryptocercus*, *Polyphaga*, and *Ergaula*, homology relations of the sclerotisations in the left part. – Only cuticular elements of the left part of the left complex are shown. Dorsal views. Scale 1mm. The sclerites are patterned differently according to their homology relations. **Ive, hla**, and **vla** are formative elements. The position of region **L4d** is given. Undulate lines are cutting lines. The branching black lines represent the assumed phylogeny.



The membranous lobe **vla** (fig.239a, 245-247) has similar features as the **vla**-lobe of e.g. *Sphodromantis* (fig.6,12), *Lamproblatta* (fig.174, 180), *Cryptocercus* (fig.148, 151), and *Eurycotis* (fig.63, 66): Its ventral wall is part of the ventral wall of the left complex. Its dorsal and its ventral walls are connected by a stout muscle **l6b** (fig.251, 252, 18, 71, 155, 188, 189). (These two features are also true of *Anaplecta*, fig.205, 218, 224). Its dorsal wall is at the same time the ventral wall of the **lve**-pouch. (This is not true of *Anaplecta* because of the membrane foldings between **lve** and **vla**, especially **vfa**).

Like the other species (with the exception of *Cryptocercus*), *Nahublattella* has muscles from the anterior ventral wall of the left complex to the L2-sclerotisations: 15 is certainly homologous with 15 of *Anaplecta* (similar posterior insertion on the left branch of L2; the homology of the anterior insertion is discussed in 6.3.4.), but, as in *Anaplecta*, homology with the 15 of the other species is questionable. Muscle 16a (fig.250) could be homologous with 16a of *Anaplecta* (and the other species); however, the insertion on the lve-apodeme is by far more anteriorly (fig.222, 250), and the insertion in the ventral wall is on sclerotisation. Alternatively, homology with 126 (fig.224) of *Anaplecta* seems possible.

Nahublattella shows some important derived features as compared with Anaplecta and, at least in the case of (2)-(6), all other previously discussed species: (1) The right branch of L2 (posterior part of L2D') is by far narrower (compare fig.213 and 245). Moreover, the whole right posterior dorsal part of the left complex - that part with the right L2-branch in its ventral wall (Anaplecta: fig.211-213) - is strongly reduced to form just the bifid psa-process (fig.245; compare fig.328a and b). (2) L2 has divided into L2D' and L2E' by articulation A10. (3) The sclerotisation at the common base of the processes paa and pda, which is ring-shaped in Anaplecta (fig.211-213), has lengthened to form a cylinder (fig.244). Hence, the processes paa and pda (and vsa) are now only the distal branches of a larger evagination, which has been defined as a "new" process via (paa, pda, and vsa are subordinate parts of via). The homologies of the single processes of Anaplecta and *Nahublattella* are hardly determinable, but in my view the relations expressed by the designations are the most probable. In accordance with Anaplecta, the sclerotisation of via is assumed to comprise a L2-part (L2E', roughly the L2d'-region; dorsally on via and near articulation A10; fig.324l,m) and L4N (ventrally on via and near the insertion of 110; fig.3251,m; discussion of L4N in 6.3.4.). (4) This sclerotisation of via is divided into a basal and a distal sclerite (39 in fig.241, 244). (5) There is no sclerotisation in the ventral wall of the vla-lobe (compare fig.205 and 239a). (6) The main muscle of the hlahook (114 in fig.249) has its anterior insertion on the lve-apodeme (discussion in 6.4.).

Parcoblatta, Blaberus, and other Blattellidae and Blaberidae

The L2-sclerotisations, the pouch lve, the processes via, paa, pda, and psa, the lobe vla, and the muscles s7, l4, and l10 have been studied not only in *Parcoblatta* and *Blaberus* but also in *Supella, Euphyllodromia, Loboptera, Ectobius* and *Nyctibora* (Blattellidae), *Nauphoeta* and *Blaptica* (Blaberidae) (muscles not studied in *Ectobius*). Morphology and homology of these elements are shown in fig.328. The morphology of all these species is derived from a situation similar to *Nahublattella*.

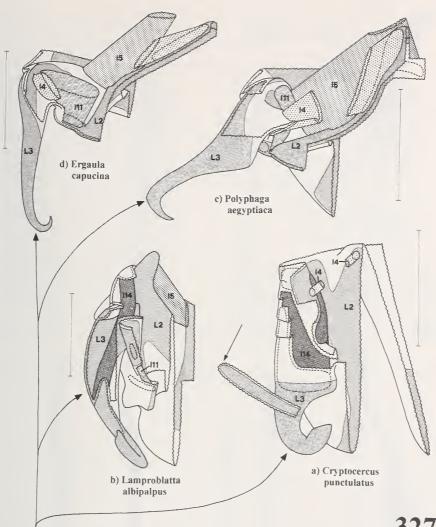
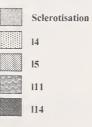
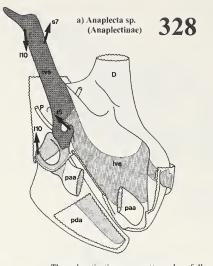


Fig.327: Left complex of *Lamproblatta*, *Cryptocercus*, *Polyphaga*, and *Ergaula*, homology relations of the muscles in the left part. – Cuticular elements are shown as in fig.326, but some parts are removed. Dorsal views. Scale 1mm. All sclerotisations are patterned in the same manner. The muscles **14**, **15**, **111**, and **114**, if present, are shown and patterned differently according to their homology relations. **14** is always cut through. The ventral insertion of **15** is shown only in *Lamproblatta*. Undulate lines are cutting lines. The branching black lines represent the assumed phylogeny.







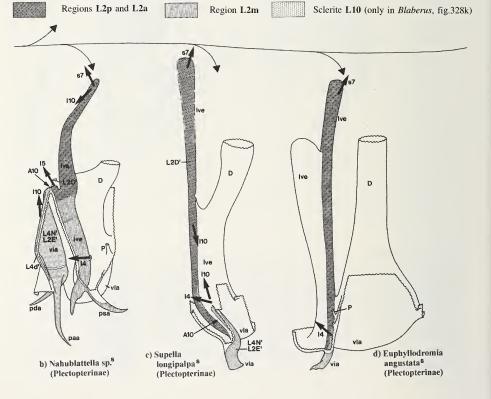
The sclerotisations are patterned as follows: Sclerotisation of processes pda (= posterior part of region L4l = main part of sclerite L4N) and paa (= region L2d = left posterior part of sclerite L2 or L2E), or of process via

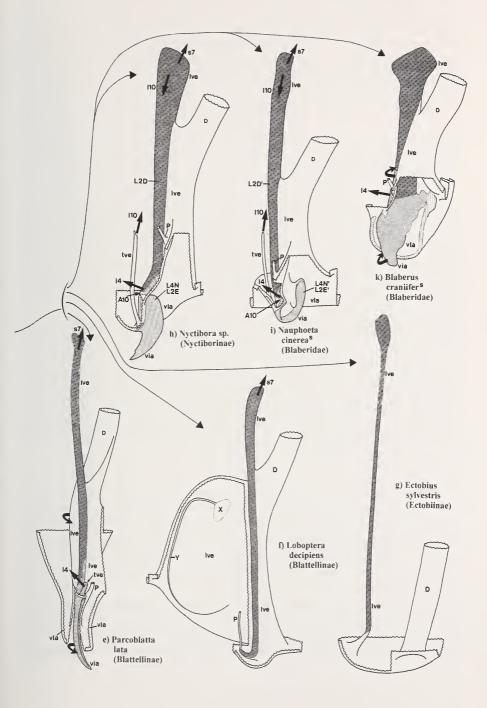
Blattellidae and Blaberidae. - The central part of the left complex is shown, with pouch lve, terminal parts of ejaculatory duct (D) and phallomeregland (P), processes paa, pda, via, and psa, ventral lobe vla, and tendon tve. Dorsal views. Patterned areas are sclerotised. Undulate lines are cutting lines through the cuticle. L2D and L2E are separate L2-sclerites, A10 is the articulation between them. Broad arrows represent the muscles 14, 15, 110, and s7 (not investigated in Ectobius, fig.328g). Curved arrows in Parcoblatta (fig.328e) and Blaberus (fig.328k) show the direction of rotations. X and Y are special elements of Loboptera. (Detailed information in 6.2.4.). Species with "S" behind their names have sidereversed phallomeres, and a mirrorimage of the original preparation is shown. The branching black lines represent the assumed phylogeny.

Fig.328: Left complex, evolution of

main sclerite L2 and sclerite L4N in







Pouch lve and its L2-sclerotisation The elements designated lve and L2 have a lot of features in common with lve and L2 of *Nahublattella* and/or *Anaplecta*: lve is a deep anteriad-directed invagination in the center of the left complex which is partly sclerotised (by L2; fig.210, 242, 268, 299, 328). The anterior part of this lve-pouch is a tube-like lve-apodeme with a membranous stripe in its ventral or right wall (44 in fig.206, 239a, 245, 266, 297a). A muscle s7 runs from the lve-apodeme to the left half of the subgenital plate (fig.221, 249, 276, 328); in Blaberidae, s7 is missing in Blaberus but present in Nauphoeta. A muscle 110 runs from the lve-apodeme to the base of the lvepouch; however, the positions of the **110**-insertions are not exactly the same in the various species, and in some species 110 is missing (fig.328d,e,f,k; discussion below). The ejaculatory duct joins the lve-pouch from the right side at the base of the lve-apodeme (like in Nahublattella; D in fig.242, 268, 299, 328). The genital opening is to the right of the apodeme – only in *Blaberus* its position is more dorsal (fig.328k), and only in Parcoblatta its position is more ventral (fig.328e). Only in Ectobius the ejaculatory duct opens far to the right of the apodeme (fig.328g). Where the ejaculatory duct joins the lvepouch, lve strongly widens, like in Nahublattella, and the right posterior part of lve is membranous (fig.328b-k; both is not true of Ectobius, fig.328g). The phallomere-gland opens into the posteriormost dorsal lve-wall (like in Nahublattella, P in fig.328; the phallomere-gland has been lost in Supella, fig.328c, and Ectobius, fig.328g). A muscle 14 runs from the L2-sclerotisation (fig.249, 276, 303, 328) to the left wall of the left complex, where it is attached to L4-sclerotisations if present (discussion of L4 in 6.3.4.). 14 was not found in Loboptera.

The remaining muscles of this area have been investigated only in *Parcoblatta* and *Blaberus*. Two further muscles having homologues in *Nahublattella* insert anteriorly on the **lve**-apodeme: **l14** or **l14a,b** (fig.249, 276, 303) run to the **hla**-hook (discussion of **l14** in 6.4.3.). **l6a** (fig.250, 277, 304) runs posteroventrad. In *Blaberus* and *Nahublattella* the posterior **l6a**-insertion is still in the anteriormost ventral wall of the left complex, but in *Parcoblatta* it has shifted far posteriad to the ventral wall of the genital pouch (fig.267). In *Blaberus* and *Parcoblatta* **l6a** has strongly enlarged. **l15** is restricted to *Nahublattella* (fig.249); **l42** is restricted to *Blaberus* (fig.304). *Parcoblatta* and *Blaberus* have lost muscle **l3** (from **lve** to **pne**, compare *Nahublattella*, fig.250).

Process via and its L2- and L4-sclerotisations All species except *Loboptera* (fig.328f) and *Ectobius* (fig.328g) have a sclerotised process behind the **lve**-pouch (via in fig.328), whose shape and size varies, and whose sclerotisation can be connected with (fig.328d,e,k) or separated from (fig.328c,h,i) the L2-sclerotisation in the **lve**-pouch. The question arises whether these processes are homologous with the via-process (fig.244) or with the **psa**-process (fig.245) of *Nahublattella*, and whether this homology relation is the same in all species.

Some similarities strongly suggest that via of *Nyctibora* is homologous with via of *Nahublattella*: The sclerotisation at the right base of via articulates (A10 in fig.328b,h) with the left posterior end of the L2-sclerite occupying the lve-apodeme. The basal sclerotisation of via forms a complete cylinder. A stout muscle runs from the lve-apodeme

to the left base of **via** (**110** in fig.250, 328b,h). Thus, the bipartition of **L2** (by **A10: L2D** and **L2E**) is assumed to be homologous in *Nyctibora* and *Nahublattella*, and for **via** of *Nyctibora* the same composition of **L2E** and **L4N** is assumed as for **via** of *Nahublattella* (further details on **L4N** in 6.3.4.). In some features *Nyctibora* is more derived than *Nahublattella*: (1) The right posterior branch of **L2D** (fig.244, 245; compare fig.328b and h) is reduced to a vestige, and the process **psa** is completely missing. (In *Nyctibora* the vestige can be identified by the insertion of **14**, which is much closer to articulation **A10** than in *Nahublattella*: fig.328b,h). (2) The posterior insertion of **110** is upon a long cuticular tendon (**tve** in fig.328h). (3) The **via**-process is no longer forked, and **paa** and **pda** (and **vsa**?) must be fused or partly reduced.

Nauphoeta (fig.328i) strongly resembles *Nyctibora*: The **via**-process, the articulation **A10**, the insertions of **l4** and **l10**, the **tve**-tendon, and the phallomere-gland are arranged in the same way (compare fig.328h and i). However, the basal sclerotisation of **via** is no longer a complete cylinder and does not reach the base of the **tve**-tendon.

Supella (fig.328c) is similar to *Nauphoeta*, but some features are different: The phallomeregland and the **tve**-tendon are missing. The sclerotisation of **via** has expanded anteriad along the right margin of **L2D'**, and articulation **A10** is therefore long and hinge-like. The right insertion of **l4** is in the usual position but has shifted from **L2D'** to the adjacent membrane. The posterior insertion of **l10** has shifted to the right; its position can be explained by a clockwise (as seen from behind) rotation of the **via**-process along its longitudinal axis (similar to *Parcoblatta*, see below). The anterior **l10**-insertion on **L2D'** is by far more posteriorly than in the other Blattellidae and Blaberidae having a **l10**; however, in *Anaplecta*, fig.222, and *Nahublattella*, fig.250, the **l10**-insertion also extends far posteriad.

Alternatively, one could assume that in *Supella* the process is not **via** but **psa** (compare fig.328b and c) and that **via** is missing. However, no muscle in any of the Dictyopteran species studied here would then have the same course as **l10** of *Supella* (from the anterior part of **L2** to its right part), and articulation **A10** of *Supella* would likewise have no homologue at least in Blattellidae and Blaberidae. Therefore, and since the respective area is quite similar in *Supella* and *Nauphoeta*, the process is more likely to be **via**.

Euphyllodromia (fig.328d), *Parcoblatta* (fig.328e), and *Blaberus* (fig.328k) have, in contrast to *Nahublattella*, *Nyctibora*, *Nauphoeta*, and *Supella*, the sclerotisation of **via** firmly connected with the **L2**-sclerotisation of the **lve**-pouch (like in *Anaplecta* and in the ground-plan), and muscle **110** is missing (however, **l42** of *Blaberus*, fig.304, might possibly be a **110** with its posterior insertion shifted far to the left). The basal sclerotisation of **via** is a complete cylinder (fig.328d, 273, 274, 300, 302). Muscle **14** is present (fig.328d,e,k). In *Parcoblatta* the **via**-process and the surrounding area have undergone a rotation (clockwise as seen from behind; lower curved arrow in fig.328e). This can be recognised by the following features: (1) The contact between the lumina of the **via**-process and of the rest of the left complex (fig.328e, 273, 274) is dorsal to the connection of the sclerotisations of **via** and **lve**, not to the left of this connection as e.g. in *Nyctibora* (fig.328h) and *Nahublattella* (fig.328b). (2) The right part of the **lve**-pouch, including the distal part of the ejaculatory duct, has partly wrapped around the **L2**-sclerite (again,

clockwise as seen from behind; upper curved arrow in fig.328e). (3) Posteriorly the sclerotisation on the left edge of the **lve**-pouch bends more and more into the dorsal **lve**-wall (compare fig.271 and 272). (4) The genital opening has been rotated in the same way and is now in the right ventral wall of the left complex (in between the lobes **47**, **48**, **49** in fig.266, 271).

In contrast, the area of **via** has been rotated counterclockwise in *Blaberus* (as seen from behind; curved arrows in fig.328k): The contact between the lumina of the **via**-process and of the rest of the left complex is situated ventral to the connection of the sclerotisations of **via** and **lve** (fig.328k, 300, 302). Posteriorly the sclerotisation on the left edge of the **lve**-pouch bends more and more into the ventral **lve**-wall (compare fig.299 and 300). In contrast to the other species (fig.328k,c,d,h,i), the genital opening is not exactly on the right side of the **lve**-pouch but more in its dorsal wall.

The **tve-**tendon is missing in *Euphyllodromia* and *Blaberus*. In *Parcoblatta* the invagination anteriorly on the **vge**-groove (**vge**, **tve** in fig.273) has exactly the same position as the **tve**-tendon in *Nyctibora* (fig.328e,h): At the base of **via**, opposite to where the sclerotisations of **via** and **lve** are connected. In *Parcoblatta* the right insertion of **14** has shifted to **tve**, and this might be the reason for the retention of **tve** despite the loss of muscle **110**.

The via of *Nyctibora* and *Nauphoeta* are clearly homologous with via of *Nahublattella*; the via-morphology of *Supella* and the remaining species can be derived from that of *Nyctibora* and *Nauphoeta*. Therefore, for all species shown in fig.328 it is assumed that the processes designated via are homologous. The presence of two sclerites L2D (in lve) and L2E+L4N (on via) is probably plesiomorphic. (Exact argumentation in 7.5.; the interpretation results from the situation in *Nahublattella*). In the species having these two sclerites fused, the resulting sclerite would have to be named correctly L2D+(L2E+L4N). I will simply designate it L2.

In most of the species shown in fig.328b-k, L2 or L2D occupy the entire left edge of the **lve**-pouch and the adjacent margins of the dorsal and ventral **lve**-walls (cross-section like in fig.270 or 301). This groove shape of L2 or L2D extends posteriad as far as to the base of **via** (articulation A10, if present). This is the case in *Nahublattella*, where, however, A10 is far anteriorly (fig.328b), and close to A10 there is a kink to the left (edge 7 at 36 in fig.242). Between A10 and the kink, L2D' bears the extension 28 (fig.245) into the ventral **lve**-wall, which has a homologue in *Anaplecta* (28 in fig.216). In *Supella, Parcoblatta, Nyctibora, Nauphoeta*, and *Blaberus* (fig.328c,e,h,i,k) L2 is also groove-shaped, but A10 or the **via**-base are by far more posteriorly, and there is no kink (except for a hint of one in *Nyctibora*) and no extension 28.

In *Euphyllodromia* (fig.328d) and *Loboptera* (fig.328f) the sclerotisation of the **lve**-pouch is – except for the anteriormost part – restricted to the dorsal wall (and not groove-shaped), and the membranous left edge of the **lve**-pouch is extensively invaginated. For a correct interpretation of these invaginations (origin, homology in *Euphyllodromia* and *Loboptera*?) further investigations are necessary. Only *Loboptera* has a bulge (**X** in fig.328f; thickened cuticle?) in the ventral wall of this invagination, which bears a sclerotised whip-like process (**Y** in fig.328f). (Since there is no **via**-process at the posterior end of **L2**, these

X- and **Y**-structures could possibly be **via**, which then would be in a rather primitive position; compare *Nahublattella*, fig.328b).

Ventral lobe vla *Parcoblatta* and *Blaberus* have retained a distinct vla-lobe. Like in *Nahublattella*, however, the L4-plate in the ventral vla-wall has been lost (compare e.g. L4G in *Anaplecta*, fig.205).

The vla-lobe of *Blaberus* (fig.297a, 299: beneath the via-process) can be identified as the true vla by some of its features in common with *Nahublattella* and other species: The dorsal vla-wall continues anteriad into the ventral lve-wall (fig.12, 181, 246, 300). The ventral vla-wall is part of the ventral wall of the left complex (fig.6, 174, 239a, 297a). The dorsal and ventral vla-walls are connected by a very stout muscle (l6b in fig.18, 188, 189, 251, 252, 305, 306). In contrast to all other species, however, *Blaberus* has the dorsal insertion of l6b on the L2-sclerotisation. This is a consequence of the rotation of via and of the posterior parts of L2 described above, by which extensive parts of L2 must have shifted into the insertion area of l6b.

Parcoblatta has a similar vla-lobe (fig.268-270), which, however, lies to the left and dorsal to the via-process. This location corresponds to the rotation of via and of the genital opening described above, in which vla has been involved, too. Muscle l6b (fig.278, 279) is in the same position as in *Blaberus*; its dorsal insertion, however, is on the membranous parts of the lve-pouch wrapped around the L2-sclerite; this situation is, again, an effect of the rotation.

In the other species (fig.328c,d,f,g,h,i; **l6b** not investigated) there is no distinct lobe **vla**. By comparing the relative positions of the phallomere elements adjacent to the **vla**-lobe in *Blaberus* and *Parcoblatta*, however, a membranous area that is assumed to be the last vestige of **vla** (fig.328c,d,h,i) can be determined. Only in *Loboptera* (fig.328f) and *Ectobius* (fig.328g) this is not possible because of extensive reductions in this part of the left complex.

A sclerotisation L10 on vla is only present in *Blaberus* (fig.299) and some other Blaberidae (e.g. *Blaptica*, fig.291). L10 is discussed in 6.3.4..

The muscles of this area have been investigated only in *Parcoblatta* and *Blaberus*. Muscle **15** of *Anaplecta* (fig.223) and *Nahublattella* (fig.251) is probably missing in *Parcoblatta* and *Blaberus* or might possibly be incorporated into muscle **16b**. *Nahublattella* (fig.251, 252), *Parcoblatta* (fig.277-279), and *Blaberus* (fig.305, 307) have some muscles in the ventral wall of the left complex (**130**, **131**, **132**, **137**, **138**, **140**, **144**, **145**), most of which are rather diffuse. These can be homologised only in part, and the most probable homologies are expressed by the designations.

6.3. Left complex III: Main sclerites L4 and L10 and associated elements

6.3.1. Comparison between Blattaria and Mantodea

The homology relations between Blattaria and Mantodea and the common ground-plan can be best deduced from a comparison between *Mantoida*, *Archiblatta*, *Eurycotis*, and

Tryonicus. Features of *Periplaneta* will also be discussed (no figures). In *Mantoida* L4 is one large sclerite, in the Blattarian species L4 is a group of sclerites.

L4 of *Archiblatta* (fig.53-57) and L4 of *Periplaneta* are very similar: 5 sclerites L4C, L4D, L4E, L4F, and L4G in the same arrangement. *Eurycotis* (fig.65-69) has three sclerites L4H, L4F, and L4G.

That both L4F and L4G are homologous in the three species is evident from the identical positions and similar outlines of the sclerites and from the fact that in *Eurycotis* as well as in *Periplaneta* the muscles 15 (fig.72) and 16b (fig.70, 71) insert on them. A special feature of *Eurycotis* is the **mla**-lobe (fig.63, 68, 69).

L4H of *Eurycotis* (fig.65-68, 325e) is composed of three parts which are homologous with L4C, L4D, and L4E of *Archiblatta* (fig.53, 57, 325f) and *Periplaneta*: (1) The left, crescent-shaped part of L4H corresponds to L4C. It lies in the left edge and in the anteriormost ventral wall, an apodeme swe runs along it, and its posteriormost part occupies a process pda. In *Eurycotis* and *Periplaneta* the muscles l2 and l4 (fig.70, 71) insert on the posterior part of swe and run to the pouches pne and lve. (2) The left part of the L4H-plate in the anterior ventral wall is homologous with L4D. It takes a position left-posterior to the right-anterior end of swe and bears a node-like process nla. *Eurycotis* and *Periplaneta* have a stout muscle from this sclerotisation to the hla-hook (l14c in fig.72). (3) The right part of the L4H-plate in the anterior ventral wall is homologous with L4E. It takes a position in between the right-anterior end of swe and L4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F. *Eurycotis* and *Periplaneta* have a muscle from this sclerotisation to the anterior part of the l4F.

The definition of the regions of main sclerite L4 is mainly based on the condition of L4 – as several isolated sclerites – in *Archiblatta* (compare fig.53-57 and fig.325f). The choice of *Archiblatta* as the type of reference is made for practical reasons and has nothing to do with an assumption of a primitive state. The positions of muscle insertions on the various L4-regions (not studied in *Archiblatta*) are taken from *Periplaneta* and *Eurycotis* – in accordance with the homology relations to *Archiblatta* discussed above. For *Archiblatta* and *Eurycotis* the regioning is shown in fig.325e,f.

- L4l (lateral): The sclerotisation homologous with sclerite L4C of Archiblatta, minus its dorsal extension to the right (L4d, see below). L4l bears the swe-apodeme and sclerotises the pda-process posteriorly. On L4l there are the left insertions of the muscles l2 and l4, which run to the pouches pne and lve.
- L4d (dorsal): The sclerotisation homologous with the dorsal, rightward directed extension of sclerite L4C of *Archiblatta*.
- L4n (node): The sclerotisation homologous with sclerite L4D of Archiblatta. L4n bears the evagination nla. On L4n there is the anterior insertion of muscle l14, which runs to the hla-hook.
- L4c (central): The sclerotisation homologous with the sclerites L4E and L4F of *Archiblatta*.
- L4v (ventral): The sclerotisation homologous with sclerite L4G in the ventral wall of the vla-lobe of *Archiblatta*. On L4v there is the ventral insertion of muscle l6b, which runs to the lve-pouch.

Eurycotis has no distinct **L4d**-region; the demarcation of **L4d** in fig.325e is tentative. In *Archiblatta* (fig.325f) and *Periplaneta* **L4d** is very distinct.

Three other L4-regions are not present in *Archiblatta* but are apomorphic sclerotisations of certain subgroups:

- L4a (anterior), L4x: Sclerotisations of Lamproblatta and Polyphaga (and Ergaula) which have developed by an expansion of the sclerites in the ventral wall of the vla-lobe (definition in 6.3.4.).
- L4b (between): A new sclerotisation in the ventral wall of the left complex of *Chaeteessa*, *Sphodromantis*, and *Metallyticus* (definition in 6.3.3.).

The left part of L4 of *Mantoida* (fig.44, 45) and the muscles inserting on it are very similar to the regions L4l and L4d of *Archiblatta*, *Periplaneta*, and *Eurycotis*; homology is assumed for the following similarities and elements (fig.325d,e,f):

- 1. Both the left part of L4 (*Mantoida*) and L4C (*Archiblatta*) occupy the whole left edge of the left complex and the anteriormost ventral wall (fig.44, 45, 53, 54).
- 2. An apodeme swe extends along most of this sclerotisation (fig.44, 45, 53, 54); anteriorly swe is massive and beam-like, posteriorly it is groove-like.
- 3. The posteriormost part of both L4 and L4C occupies a process (pda in fig.44, 53). However, only in *Mantoida* the sclerotisation of pda is connected with the L2-sclerotisation of the paa-process (compare in 6.2.1.).
- 4. Both L4 and L4C have a distinct dorsal extension to the right (L4d in fig.44, 53).
- Muscle 12 inserts on the swe-apodeme in the posterior half of L4 (*Mantoida*), or L4C (*Periplaneta*), or L4H (*Eurycotis*) (fig.49, 70). 12 runs to the pne-pouch and inserts on L1 (*Mantoida* and *Periplaneta*) or in the membrane to the left of L1 (*Eurycotis*).
- 6. Muscle 14 inserts on swe ventral to 12 (fig.50, 71) and runs to the lve-pouch.
- 7. Muscle **s1** (fig.48, 70), which comes from the left apophysis of the subgenital plate (fig.37, 59), inserts on that part of **L4**, **L4C**, or **L4H** in the anteriormost ventral wall.

The right part of L4 of *Mantoida* has some features in common with the regions L4v and L4c of *Archiblatta*, *Periplaneta*, and *Eurycotis* (fig.325d,e,f):

- 8. The posterior right part of L4 (fig.41, 47) has the same position in the ventral wall of the vla-lobe as the L4G-sclerites (= L4v-region) have in *Eurycotis* (fig.63, 66) and *Archiblatta* (fig.54).
- 9. The anterior right part of L4 is, like the anterior L4c-region in *Archiblatta* (L4E in fig.57) and *Eurycotis*, situated between the right anterior end of the L4l-region and the anterior end of the L4v-region.
- 10. A muscle running to the ventral wall of the ejaculatory duct inserts on or near the right part of L4 or on L4G, respectively: the posteroventral part of l6 in *Mantoida* (fig.52), l6b in *Eurycotis* (fig.70, 71) (compare in 6.2.1.).

These features 1.-10. suggest the homology relations shown in fig.325d,e,f. In *Mantoida*, the regions L4l, L4d, and L4v can be unambiguously identified, and the similarities 1.-8. and 10. can be regarded as features of the common ground-plan of Blattaria and Mantodea. L4c might be contained in the anterior right part of L4 (feature 9.), but this is not certain since in *Mantoida* the extension of the L4v-region to the anterior cannot be determined and the L4c-region is only indicated by its relative position in between L4v

and L41 (? in fig.325d). Hence, 9. is an uncertain ground-plan feature. In *Mantoida* there is no indication for the presence of a L4n-region, and a nla-process is missing.

Evidence from *Chaeteessa* (complete discussion in 6.3.3.) suggests that the **L4n**-region is also an element of the common ground-plan of Blattaria and Mantodea and that the lack of **L4n** in *Mantoida* is derived:

11. The heavier sclerotised transverse bridge in the anterior ventral wall of the left complex of *Chaeteessa* might, according to its very similar position, well be homologous with the **L4n**-region of *Eurycotis* (compare fig.31 and 65, 69; fig.325c,e).

Tryonicus has some features in common with *Archiblatta* and *Mantoida* which suggest the homology relations shown in fig.325d,f,g. The two sclerites L4K and L4N (fig.85, 97) together form a broad ribbon in the left wall of the left complex, which takes, like L4C in *Archiblatta* (fig.54), a position left-dorsal to the base of the hla-hook. Most of L4K and L4N is therefore assumed to represent the L4I-region. The nla-process (fig.97) on L4K corresponds to the nla of *Archiblatta* (fig.56) and *Eurycotis* (fig.68) in its shape, in its location in the anterior ventral wall, and in its position relative to the other L4-sclerotisations and to the hla-base. Thus, the part of L4K on nla is regarded as the L4n-region. That part of L4K which anterior to nla extends to the right (fig.53, 55) and is hence assumed to belong to the L4I-region. The ribbon-like extension of L4N (L4d in fig.88-95) corresponds in its shape and relative position with the L4d-region of *Mantoida* (fig.45) and especially *Archiblatta* (fig.53). The posteriormost part of L4N (L4N in fig.84-95) (pda in fig.91) like in *Mantoida* and *Archiblatta* (pda in fig.44, 53) and can be regarded as part of the L4I-region. Like in *Mantoida* but in contrast to

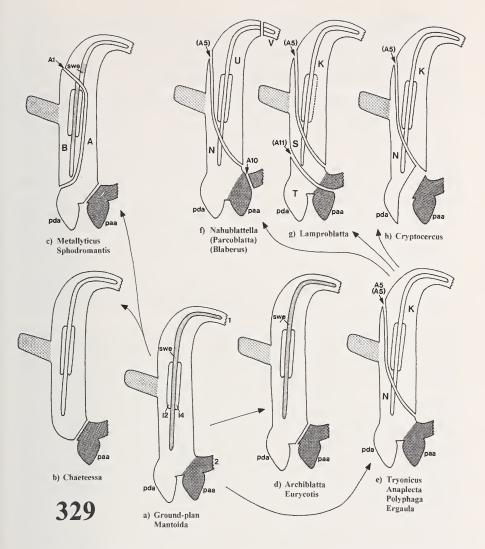
The following structures are shown:

Fig.329: Left complex, subdivisions of regions L4I and L4d into individual sclerites in Blattaria and Mantodea. – Region L4I (discussion in 6.3.) is in its primitive condition undivided and connected with region L2d posteriorly (between processes paa and pda). In many species L4I is connected with region L4n anteriorly (ground-plan condition unclear). The various subdivisions of L4I in the subgroups of Blattaria and Mantodea and the hypothetic directions of evolutionary transformation are shown. The various types – or further derivations of them – are present in the species listed. It is intended to emphasise principal similarities and differences in the subdivision of L4I. For comparability, all elements are left in their most primitive condition (like in fig.329a) – except for the subdivisions of L4I and of the included parts of L2. Inner views from the right side; dorsal \leftarrow , ventral \rightarrow , anterior \uparrow , posterior \downarrow .

The sclerite regions L4l (white) and L4d (light pattern) and parts of L2 (dark pattern; mainly region L2d on process paa). The right-anterior undulate line (1 in fig.329a) represents the removal of region L4n. The right-posterior undulate line (2 in fig.329a) represents the removal of the remainder of L2. L4d is always shown in its most primitive position, orientation, and shape, even if these have changed or if L4d has been lost.

⁻ The dividing lines which cause a division of these sclerotisations into individual sclerites. Dividing lines along which the respective sclerites are still in close contact are labelled A+Number (articulation, e.g. A5; like in the text); if the respective sclerites are farther away from each other the name of the respective articulation is put in brackets (e.g. (A5)).





- The individual L4-sclerites produced by the division of L4l and L4d. These are labelled with the capital letters used in the text and in fig.1-319 (e.g. **B** = L4B).
- The apodeme swe. swe is always shown in its most primitive condition and position (like in fig.329a), but only the parts of swe retained in the respective species are patterned and labelled.
- The insertion areas of the muscles **12** and **14**, which are also always shown in their most primitive condition and position (like in fig.329a).
- The processes pda (with its L4I-sclerotisation) and paa (with its L2d-sclerotisation).

The various types can be derived from each other in the way indicated by the arrows. *Mantoida* (fig.329a) conforms completely with the ground-plan. *Parcoblatta* and *Blaberus* can be derived from *Nahublattella* but differ in some respects (e.g. secondary fusion at A10).

Archiblatta, **paa** is bulge-shaped and its sclerotisation is connected with that of the **paa**process (fig.96). Sclerite **L4G** (fig.92, 325g) resembles **L4G** of *Archiblatta* (fig.325f) in its outline and its position in the ventral wall of the **vla**-lobe and probably represents the **L4v**-region; however, it cannot be excluded that, additionally, sclerotisations of the **L4c**region are contained in this **L4G** (in its left and anterior parts).

As compared with *Archiblatta*, *Eurycotis*, and *Mantoida*, some features of *Tryonicus* can be regarded as derived: The L4I-sclerotisation of *Tryonicus* is (1) broader and (2) divided into two sclerites (L4K, L4N; fig.325d,f,g) by the articulation A5 (fig.88, 97; compare fig.329a,d,e). (3) The swe-apodeme is completely missing. (4) L4d is directed more anteriad.

On the other hand, some features of *Tryonicus* can contribute to the common ground-plan of Blattaria and Mantodea:

- 12. As already stated in 6.2.1., feature 4., the connection of the sclerotisations of pda (L4I) and paa (L2d) present in *Tryonicus* and *Mantoida* is a ground-plan state. The separation of these sclerotisations in *Archiblatta* and *Eurycotis* is derived (compare feature 3.).
- 13. In *Tryonicus* and in *Mantoida* no sclerotisations can be unambiguously assigned to the L4c-region, and a sclerite corresponding to L4F of *Archiblatta* and *Eurycotis* is definitely missing. Thus, L4F, and possibly the whole L4c-region, can be regarded as a derived element of *Archiblatta* and *Eurycotis*.

Concerning the common ground-plan of Blattaria and Mantodea, four questions remain open: (1) It cannot be decided if there is a L4c-region. (No sclerotisations undoubtedly homologous with the sclerotisations defined as L4c in *Archiblatta, Eurycotis*, and *Periplaneta* have been identified in any other species). (2) It cannot be determined whether the L4l-region is connected with or separated from the L4v-region (or L4c-region, if present) in the anterior ventral wall of the left complex (? in fig.321e), since there is always a connection in Mantodea but never in Blattaria. (3) It cannot be decided if a nlaprocess is present (but compare in 7.5. (M), (N)). (4) In *Tryonicus, Eurycotis*, and, if the assumption in 11. is true, in *Chaeteessa* the regions L4l and L4n are firmly connected. Hence, the separation of L4l and L4n in *Archiblatta* (sclerites L4C and L4D) might be regarded as apomorphic. However, the position of the connection between L4n and L4l is rather different in *Tryonicus* (to the left of nla, fig.96, 97) and *Eurycotis* (to the right of nla, fig.67, 68), and these connections might be non-homologous. Thus, it seems better to regard the respective ground-plan state of L4n (connected with L4l or not) as unresolved.

6.3.2. The elements in the common ground-plan of Blattaria and Mantodea

The features 1.-13. in 6.3.1. permit the reconstruction of many ground-plan features of **L4**, **pda**, **vla**, and some adjacent elements: (fig.321e,g,i): **L4** is composed of sclerotisations in the left edge and in the anterior and right ventral wall of the left complex. The **L4**l-region is located in the left edge and in the anteriormost ventral wall. The **swe**-apodeme runs along most of **L4**l. **swe** is massive and beam-like anteriorly and groove-like posteriorly. There is a distinct dorsal extension **L4d** directed to the right and possibly

slightly anteriad. The posteriormost part of L4l completely sclerotises a bulge-like process pda. The sclerotisation of pda is connected with the sclerotisation of paa (region L2d). The right posterior part of L4 (region L4v) lies in the ventral vla-wall. The presence of the L4c-region is questionable, but a sclerite L4F is certainly missing. The L4n-region is present. The presence of the nla-process is unclear. The muscles l2, l4, and l6b are present. l2 and l4 have their L4-insertions close together on the swe-apodeme in the left edge of the left complex.

6.3.3. Homology relations and character states of the elements in Mantodea

In *Chaeteessa* (fig.28), *Metallyticus* (fig.20), and *Sphodromantis* (fig.6), the ventral wall of the left complex is completely sclerotised, not only along its margins as in *Mantoida* (fig.41). In *Chaeteessa*, however, the marginal ventral sclerotisation is distinctly heavier and is assumed to correspond to L4 of *Mantoida* (fig.325a-d) – composed of the ground-plan regions L4l and L4v (and possibly L4c). The anterior transverse bridge of heavier sclerotisation present in *Chaeteessa* is probably also a ground-plan element (L4n-region). The weaker sclerotisation of the remaining ventral wall is new and is defined as a further region of L4 (fig.325a-c):

L4b (between): The sclerotisation of the ventral wall of the left complex between the ground-plan regions L4l, L4n, and L4v (and possibly L4c).

In *Metallyticus* and *Sphodromantis* this **L4b**-sclerotisation is further derived in being as heavy as the ground-plan regions of **L4.** The presence of **L4n** is not assessable for this uniformity of the ventral sclerotisation (in fig.325a,b the interpretation is done in accordance with *Chaeteessa*). The presence of a region **L4c** is in *Chaeteessa*, *Metallyticus*, and *Sphodromantis* as uncertain as in *Mantoida* (? in fig.325a-d).

As compared with *Mantoida* (fig.44) or *Archiblatta* (fig.53), in *Chaeteessa* (fig.31), *Metallyticus* (fig.21), and *Sphodromantis* (fig.9) the L4-sclerotisation in the dorsal wall of the left complex has expanded to the right: In *Chaeteessa* and *Sphodromantis* L4 occupies most of the dorsal wall, in *Metallyticus* it is restricted to the anterior part. By this expansion L4 now covers the external opening of the **pne**-pouch from dorsally. Possibly in correlation with this expansion the **pne**-pouch has rotated to the right (clockwise as seen from behind; compare in 6.1.3.). These shifts are very obvious in *Chaeteessa* and *Sphodromantis* but less distinct in *Metallyticus*.

The muscle insertions on the L4I-region of *Mantoida* (fig.48-52, 325d) and on the dorsal part of L4 (L4B) of *Sphodromantis* (fig.15-17, 325a) also demonstrate these shifts: The muscles 11 (to L1 anteriorly in the pne-pouch), 12 (to L1 more posteriorly in the pne-pouch), 14 (to L2), and 17 (to the left posterior ventral wall of the left complex) are certainly homologous in the two species, but in *Sphodromantis* all insertions on L4 have shifted far to the right. These insertions also show that the extensive dorsal L4-sclerotisations of *Sphodromantis* (L4B) have not been produced by an expansion of the L4d-region (*Mantoida*: fig.44) but of the L4I-region (fig.325a): 12, 14, and 17 of *Mantoida* are not inserted on L4d but on L4l. At the most a small right-anterior part of L4B of *Sphodromantis* (posterior to the I1-insertion, fig.17) might be regarded as representing L4d (fig.325a). Whether this distribution of L4l and L4d in the dorsal wall is also true of

Chaeteessa and *Metallyticus* is unclear (no data for the musculature); in fig.325b,c L4l and L4d are demarcated in accordance with the situation in *Sphodromantis*. In any case, in *Chaeteessa, Metallyticus*, and *Sphodromantis* the L4d-region is no longer distinct from the L4l-region.

The **pda**-process of *Metallyticus* (fig.20, 23-26) is in its shape and in its position relative to the **paa**-process similar to **pda** of *Mantoida* (fig.44-46) and is likewise sclerotised by **L4**. Homology is assumed for the **pda** of the two species. However, in *Metallyticus* the sclerotisations of **pda** and **paa** are separated, and the processes themselves are more distinct from each other and by far longer. These two features also apply to *Sphodromantis* and *Mantis*: In *Mantis* (no figure) **pda** is shovel-shaped and far on the left side as in *Metallyticus*. In *Sphodromantis* (fig.9-12), certainly a close relative of *Mantis*, **pda** is long and slender and has shifted to the right. Thus, despite the different morphology of **pda** in *Mantoida* and *Sphodromantis*, these evolutionary stages suggest homology. In *Chaeteessa* the **pda**-process has been completely lost. (The one posterior process of *Chaeteessa*, fig.28, has proved to be **paa**; compare in 6.2.3.).

Only in *Sphodromantis* and *Metallyticus* (and *Mantis*, which will not be further considered) the dorsal and ventral parts of L4 have become separated by an articulation (A1 in fig.6, 10, 20, 24; sclerites L4A, L4B). The dividing line runs within the L4I-region (fig.325a,b). This is evident from the positions of the involved sclerotisations (compare fig.325a,b and c,d) and from the muscle insertions: In *Sphodromantis* (fig.325a) s1 inserts on L4A, but L2, L4, and L7 insert on L4B, and all these insertions belong to L4I (compare *Mantoida*, fig.325d). This division of L4I reminds one of the L4I-division in *Tryonicus* (by A5 in fig.88, 97, 325g). However, the courses of the dividing lines are different: The pdasclerotisation, for example, is part of the posterodorsal plate (L4N) in *Tryonicus* but part of the ventral plate (L4A) in *Metallyticus* and *Sphodromantis* (compare fig.329c and e). Thus, the articulations A1 and A5 are certainly not homologous, and the division of L4I is a case of parallel evolution.

The swe-apodeme is well-developed in *Mantoida* and *Archiblatta* (fig.45, 53). In *Chaeteessa* swe has been completely lost. *Metallyticus* has retained a vestige of swe on the left margin of the ventral L4A (fig.24). *Sphodromantis* has a vestige on the left margin of the dorsal L4B (fig.10,11). This suggests that swe has been cut through by the division into L4A and L4B and confirms that the L4I-region participates in both L4A and L4B.

6.3.4. Homology relations and character states of the elements in Blattaria

Archiblatta, Eurycotis, and Tryonicus

These species have been sufficiently discussed in 6.3.1.

Cryptocercus, Lamproblatta, Polyphaga, Ergaula, and Anaplecta

In *Tryonicus* (fig.325g) the L4l-region is divided by articulation A5: The anterior parts of L4l form, together with L4n, the L4K-sclerite; the posterior parts of L4l (with the pdasclerotisation) form, together with L4d, the L4N-sclerite. The connection of the sclerotisations of pda (L4l) and paa (L2d, fig.324g) is retained. The swe-apodeme has been lost. The **nla**-process is well-developed. The L4v-region is a plate in the ventral wall of the **vla**-lobe, the L4G-sclerite.

From this situation the morphology of the five species in the heading and the remaining Blattellidae and Blaberidae can be derived. In all these species, however, L4K and L4N are no longer articulated but far away from each other. In some species L4K or L4N undergo further divisions. In *Cryptocercus, Polyphaga, Ergaula*, and *Lamproblatta* the anterior sclerite L4K has been strongly reduced (fig.150, 124, 177). The sclerotisations of **paa** and **pda** remain in most species connected, and this connection often becomes very close. The L4v-region may retain its shape and position, or it becomes enlarged (*Lamproblatta, Polyphaga, Ergaula*) or lost (*Nahublattella, Parcoblatta, Blaberus*).

As a first point, the evolution of the L4N-sclerite and the processes pda and paa of the species in the heading will be discussed; then the L4K-sclerite with the nla-process and, at last, the sclerite in the ventral vla-wall will be considered. paa, pda, and vla have in part already been discussed in 6.2.4, but a discussion of these elements is only complete by considering them in context with the other L4-sclerotisations. This will be done in this section.

L4N-sclerite, processes pda and paa The homology of L4N of *Tryonicus* (fig.94-97), *Polyphaga* (fig.117, 118), *Ergaula, Cryptocercus* (fig.150, 151), and *Anaplecta* (fig.210-215) – and of its derivatives L4S and L4T in *Lamproblatta* (fig.177-180) – can best be deduced from a comparison of the prominent substructures. In *Tryonicus* these are: (1) The pda-sclerotisation (posteriormost L4I-region), including its close vicinity to and connection with the paa-sclerotisation (L2d-region). (2) The dorsal extension to the anterior (L4d-region).

Further evidence comes from the musculature, which, however, has not been studied in *Tryonicus*. The homology relations and the resulting regioning of these sclerotisations into **L4l** and **L4d** are shown in fig.325g,h,i,k,l.

The **pda** and **paa** of *Tryonicus, Lamproblatta, Polyphaga, Ergaula*, and *Anaplecta* have some features in common, none of which, however, is realised in all these species. But the whole of the similarities is sufficient to regard the **pda** and **paa** of all species as homologous.

- pda and paa are two processes with their sclerotisations firmly connected. (The sclerotisation of pda is designated as L4N or L4T, that of paa as L2 or L2C). *Ergaula*, however, has lost the right process paa.
- The position of pda and paa on the left complex is dorsal, far posterior, and far to the left near the left end of the lve-pouch. In *Lamproblatta*, however, their position relative to lve is more to the right (fig.179, 180).
- The common sclerotisation of paa and pda is, to the right (*Anaplecta*, fig.211) or to the ventral side (*Polyphaga*, fig.118, 122; *Ergaula*; *Tryonicus*, fig.97), firmly connected with the left end of the L2-sclerotisation in the lve-pouch. *Lamproblatta*, however, has lost this connection (fig.178, 179).
- On the common sclerotisation of **paa** and **pda** there inserts a muscle coming from the left part of the **lve**-pouch (**l10** in fig.129, 186, 222; compare in 6.2.4.; not analysed in *Tryonicus*).

- In *Polyphaga* and *Lamproblatta* the left process pda is long and pointed, the right process paa is somewhat saucer- or cup-shaped.
- In Polyphaga and Ergaula the pda-processes are nearly identical.

Polyphaga, Ergaula, and *Lamproblatta* (but not *Anaplecta*) have sclerotisations probably homologous with the **L4d**-region of *Tryonicus*:

- In *Tryonicus* the one end of L4N (to the right of pda) is connected with L2 occupying paa and the lve-pouch, and its other end has the extension L4d (fig.96), which is directed anteriad.
- In *Polyphaga* and *Ergaula* L4N is also connected with L2, and its opposite end has an extension, which, however, is directed to the left (L4d in fig.118).
- In Lamproblatta the sclerotisation L4S (fig.178, 180) is connected with L2 at the left end of the lve-pouch. The distal part of L4S resembles L4d of Polyphaga with the difference that it is not connected with the sclerotisation of pda and paa (compare fig.325i and k). I assume that in Lamproblatta a dividing line has formed which has separated the following sclerotisations from each other (fig.329e,g): Posterior to the line is the common sclerotisation of pda and paa (composed of L4T, a part of the former L4N-sclerite, and L2C, a part of the former L2-sclerite). Anterior to the line are (1) the part of the former L4N-sclerite (L4S) which maintains the connection with L2 at its one end and has the extension L4d at its other end (fig.178).
- The homology of the extensions L4d in *Lamproblatta*, *Polyphaga*, and *Ergaula* is confirmed by muscle 111, which inserts on or near L4d and runs to sclerite L4K (fig.128, 188, 327).
- Nahublattella (complete discussion below) also has an extension similar to L4d of Tryonicus or Polyphaga (L4d' in fig.242); it extends, like L4d of Polyphaga, from the common sclerotisation of paa and pda to the left. In Anaplecta L4d has been lost.

L4d is directed to the right in *Mantoida* (fig.44), right-anteriad in *Archiblatta* (fig.53), and anteriad in *Tryonicus* (fig.94). In *Polyphaga* and *Ergaula* (and *Nahublattella*) **L4d** has even further rotated (counterclockwise as seen from above) and is directed to the left. From such a position, **L4d** of *Lamproblatta* has additionally rotated 90° (clockwise as seen from behind) and shows a dorsoventral orientation (fig.178).

L4N of *Cryptocercus* (fig.150) is assumed to be homologous with the L4N of the other species and to have the same orientation as in *Polyphaga* and *Ergaula* (compare fig.117 and 150, 325h and k): Its left part is L4d, its right part is the pda-sclerotisation (fig.325h). This is suggested by the following features:

- L4N lies, like in the previous species, in the left dorsal wall of the left complex (fig. 150).
- The right part of L4N lies, like the right part of L4N of *Polyphaga* and the right-posterior part of L4N of *Tryonicus*, on a process (pda in fig.150, 118, 96).
- pda is, like in *Tryonicus*, *Mantoida*, and *Polyphaga*, situated to the left of (and somewhat dorsal to) the paa-process (fig.150, 96, 44, 117).
- Within the angle formed by the sclerites L4N and L2 (fig.150) there is the base of the hla-hook. *Tryonicus* shows the same relations (fig.97).
- In Cryptocercus and Mantoida, the muscles 11 (fig.48, 155) and 12 (fig.49, 156) run

from the **pne**-pouch to the left and dorsad (homology discussion in 6.1.1.). The insertion of **11** is in *Cryptocercus* on **L4N** (including **L4d**), in *Mantoida* on and to the right of **L4d**.

- In Cryptocercus and Mantoida, muscle 19 (fig.49, 155) runs transversely within the dorsal wall of the left complex (homology discussion in 6.5.). Its left insertion is in Cryptocercus anterior to the left end of the assumed L4d-part of L4N, in Mantoida near the right end of L4d; regarding the orientation of L4N assumed for Cryptocercus, these ends of the sclerites would be homologous. In Cryptocercus, however, 19 does not extend as far to the right as in Mantoida.

Some features of *Cryptocercus* are derived: (1) The sclerotisation of **pda** is largely reduced (fig.150, 329h), and, possibly as a consequence of this, (2) the sclerotisations of **pda** (L4) and **paa** (L2) have separated (like in *Chaeteessa*, *Metallyticus*, *Sphodromantis*, *Archiblatta*, and *Eurycotis*).

L4K-sclerite, process nla L4K of *Anaplecta* (fig.209, 212) is probably homologous with L4K of *Tryonicus* (fig.98); similarities are:

- The position in the left and left-ventral walls of the left complex.
- The anterior part is on a bulge-like evagination (**nla** in fig.97, 212). Like in *Tryonicus*, this sclerotisation is regarded as the **L4n**-region (fig.325g,l).
- The posterior part is plate-like, with a broadly truncate posterior margin, and this part partially encloses the retracted **hla**-hook from the left side (fig.85, 202). Like in *Tryonicus*, this sclerotisation is regarded as an anterior part of the **L4I**-region (fig.325g,l).

A comparison of the muscles of *Anaplecta* and *Eurycotis* confirms these assignments to L4n and L4l:

- That in both species the anterior insertion of the main muscle of the hla-hook (l14c or l14; fig.73, 222) is on or near the sclerotisation of the nla-bulge shows the homology of these nla. The nla-sclerotisation of *Eurycotis* is, by definition, the L4n-region (fig.325e,l).
- In *Eurycotis* and *Mantoida* the muscles 12 (fig.49, 70) and 14 (fig.50, 71) insert close to each other on the L4I-region and run to the pouches pne and lve, respectively. *Anaplecta* also has a muscle 12 from the posterior part of L4K to the pne-"pouch" (fig.221, compare in 6.1.4.); muscle 14 is missing. *Nahublattella* (complete discussion below), however, has both 12 and 14 (to the pouches pne and lve), and their left insertions are on a sclerotisation homologous with the posterior part of L4K of *Anaplecta* (L4U' in fig.249).

The narrow sclerotisation which extends in *Tryonicus* from the anteriormost part of L4K to the right (anterior to nla, fig.96, 97, 325g) is missing in *Anaplecta* (fig.325l). Since this sclerotisation probably corresponds to the anteriormost L4l-region of *Archiblatta* and *Mantoida* (bearing the anterior part of the swe-apodeme; fig.44, 53, 325d,f), this feature of *Anaplecta* is regarded as derived.

The course of the dividing line through the L4I-region of *Anaplecta* (separating L4K and L4N) can be deduced from the positions of the l2- and l4-insertions in *Eurycotis*,

Mantoida, *Anaplecta*, and *Nahublattella* and from the distribution of the other substructures: Anterior to the line (on L4K) there are the insertion of l2 (and l4 in *Nahublattella*) – and hence parts of the L4I-region – the L4n-region, the nla-process, and the l14-insertion. Posterior to the line (on L4N) are the pda-process – and hence posterior parts of the L4I-region – and the l10-insertion (and the L4d'-extension in *Nahublattella*). The course of the line is shown in fig.329e. The distribution of all cuticular elements present is the same as in L4K and L4N of *Tryonicus*, and the dividing lines of *Anaplecta* and *Tryonicus* (and *Nahublattella*) are strongly suggested to be homologous.

L4K of *Cryptocercus* (fig.150, 151) is homologous with L4K of *Tryonicus* and *Anaplecta* and is likewise composed of the anterior L4I-region and of the L4n-region. Both L4I and L4n, however, are strongly reduced. These relations are, firstly, suggested by similarities in the cuticular elements of *Tryonicus* and *Cryptocercus*:

- L4K of Cryptocercus has the same position like the left-dorsal half of L4K of Tryonicus: left-dorsal to the base of the hla-hook (compare fig.85, 97 and 145, 151). In Tryonicus this sclerotisation has been regarded as an anterior part of the L4I-region (fig.325g,h). Sclerite L4P of Cryptocercus (fig.151) probably corresponds to that part of L4K of Tryonicus immediately anterior to the hla-base. The right-ventral half of L4K of Tryonicus (fig.325g) with the anteriormost L4I-region (the anterior extension to the right) and the L4n-region (nla-sclerotisation) has been, like the nla-process itself, lost in Cryptocercus.

Secondly, the same relations result from a comparison of the muscle insertions of *Cryptocercus* and other species:

- The 12 and 14 of Cryptocercus (fig.155, 156), Mantoida (fig.49, 50), and Eurycotis (fig.70, 71) run from the pouches pne and lve to the leftmost part of the left complex, where their insertions are close to each other. Homology can be assumed. The left insertions are on the L41-region in Eurycotis and Mantoida (fig.325d,e), and on L4K in Cryptocercus. The contribution of the L41-region to L4K of Cryptocercus is thus confirmed (fig.325h). This can be only an anterior part of L41 since the posterior part (with the pda-sclerotisation) is included in sclerite L4N. In Anaplecta L4K also bears the left 12-insertion (fig.221), and in Nahublattella the homologue of the posterior part of L4K (L4U') bears the left 12- and 14-insertions (fig.249).
- The **l14** of *Cryptocercus* (fig.157), *Eurycotis* (fig.72), and *Anaplecta* (fig.222) run from the anterior left wall of the left complex to a large hook (**hla**) and are certainly homologous (discussion in 6.4.). *Cryptocercus* (fig.157) and *Eurycotis* (fig.70) have phallomero-sternal muscles s1+3 or s1 inserting immediately anterior to **l14**; the left part of s1+3 (= s1) is probably homologous with s1 of *Eurycotis* (s1 is missing in *Anaplecta*; discussion in 6.9.).
- In *Eurycotis* and *Anaplecta* 114 inserts on the L4n-region (on the nla-process, fig.72, 73, 222), and in *Eurycotis* s1 inserts at the border between L4n and the anterior L4l (fig.73, 325e). In *Cryptocercus* part of 114 inserts on L4K; this suggests that the L4n-region also contributes to L4K. The larger part of the 114-insertion and the entire s1+3-insertion, however, are on membrane (ventral and anterior to L4K); this suggests that the L4n-region as well as the anteroventral part of the L4l-region (corresponding to the

anterior extension to the right on L4K of *Tryonicus*) are strongly reduced. Probably as a consequence, the **nla**-process is missing.

Thus, L4K of *Cryptocercus* is composed of anterior parts of L4l (with the insertions of l2 and l4) and a highly reduced L4n (with part of the insertion area of l14). L4N of *Cryptocercus* is made of the posterior part of the L4l-region (with the reduced pda-sclerotisation) and of L4d. The distribution of all elements present is the same as in L4K and L4N of *Anaplecta* and *Tryonicus*, and homology can be assumed for L4K, for L4N, and for the dividing line between them (through L4l). The reduction of the L4n-region and the loss of the nla-process are derived features of *Cryptocercus*. The anteroventral part of L4l has also been lost in *Anaplecta*.

L4K of Lamproblatta (fig.177, 178) resembles L4K of Cryptocercus (fig.150, 151):

- The sclerites take the same position dorsal to the base of the hla-hook.
- A process nla on or near L4K is missing.

Anterior to L4K in *Lamproblatta* or on and anterior to L4K in *Cryptocercus* there insert some muscles having the same course, and the insertions on or near L4K show the same positions relative to each other:

- A muscle to the subgenital plate (s1 in fig.185; left part of s1+3 in fig.157).
- A muscle to sclerite L3 on the hla-hook (l14 in fig.184, 157). The l14-insertion is partly on L4K in *Cryptocercus* but completely on membrane in *Lamproblatta*.
- A muscle to the pne-pouch (12 in fig.184, 156). The 12-insertion is on L4K in Cryptocercus but on the membrane anterior to L4K in Lamproblatta.

Therefore, L4K of *Lamproblatta* and *Cryptocercus* are assumed to be homologous and to have the same composition: anterior part of L4l, vestiges of L4n. However, since in *Lamproblatta* the insertions of l2, l14, and s1 are exclusively on membrane and muscle l4 has been lost (compare fig.155), the muscles do not yield any direct evidence for the presence of the regions L4l and L4n and for the distribution of L4l and L4n within sclerite L4K. The distribution can only be deduced from a comparison with *Cryptocercus*, as it is done in fig.325h,i. That the l14-insertion is completely on membrane could be interpreted as a further reduction of the L4n-region as compared with *Cryptocercus*. That the l2-insertion is anterior to L4K (not on L4K as in *Cryptocercus*) is interpreted as a shift of this insertion to the anterior, not as a reduction of the respective L4l-sclerotisation (comparison with *Polyphaga*, see below).

L4K of *Polyphaga* is situated not in the dorsal but in the posteroventral part of the hlabase (fig.122-124; compare fig.151, 178). It is assumed to be homologous with L4K of *Lamproblatta* and *Cryptocercus* and to have shifted and rotated (clockwise as seen from the left) ventrad around the posterior part of the hla-base. This is suggested by the following features:

- In Polyphaga and Lamproblatta L4K is broadly horseshoe-shaped and curves into the base of the hla-hook. (According to the assumed shift and rotation in Polyphaga the latter is almost 180° the orientation of the sclerite is in Lamproblatta and Polyphaga opposite). In Cryptocercus this curvature of L4K is missing.
- In *Polyphaga* and *Cryptocercus* L4K bears the insertion of a muscle coming from the left-anterior part of L2 (l4 in fig.132, 155). l4 is missing in *Lamproblatta*.

- In Polyphaga and Lamproblatta L4K bears the insertion of a muscle coming from the sclerotisation L4d (or from the adjacent membrane; 111 in fig.128, 184). Taking the assumed rotation of L4K in Polyphaga into account, the insertion on L4K is in exactly the same position. 111 is missing in Cryptocercus and all other species and is a derived feature of Polyphaga, Ergaula, and Lamproblatta.

The insertion of muscle s1 (fig.127) has retained the same position as in *Lamproblatta* (fig.185) and *Cryptocercus* (left part of s1+3 in fig.158, 159): on the basal line anterior to the hla-base. The hla-muscle 114, present in all other Blattaria studied (discussion in 6.4.), is missing in *Polyphaga*, and the hla-hook and its sclerite L3 are bare of muscles. The function of 114 has probably been taken over by the very stout 14, which does not insert on L3 but on the dorsal part of L4K situated within the hla-base.

The muscles 12 are certainly homologous in Polyphaga, Lamproblatta, Cryptocercus (fig.128, 184, 156), Mantoida, Eurycotis, and Anaplecta (fig.49, 70, 221; discussion in 6.1.). The ground-plan positions of the **12**-insertions are shown by the three latter species: right insertion in the left wall of the **pne**-pouch; left insertion roughly in the middle of the left edge of the left complex. In Cryptocercus, Lamproblatta, and Polyphaga, as a first point, the right 12-insertion has shifted anteriad to the top of the pne-pouch (compare in 6.1.). As a second point, the left insertion also shows a gradual shift to the anterior and takes a position (1) more anteriorly than in the ground-plan but still on the L4Isclerotisation (L4K) in Cryptocercus, (2) even more anteriorly and anterior to the L4Isclerotisation (L4K) in Lamproblatta, and (3) still more anteriorly, and ventrally, but again on sclerotisation (L4M) in *Polyphaga*. The various stages of this l2-shift are regarded as synapomorphies of the species concerned. The l2-insertion is assumed to have shifted away from the L4I-region (Lamproblatta, Polyphaga) and to have later reached a position on another sclerotisation formed by an enlargement of the ventral sclerotisation of the vlalobe (*Polyphaga*; this aspect is discussed below). Hence, contrary to the definition of L4I in 6.3.1., the sclerotisation bearing the 12-insertion in *Polyphaga* is not assigned to L41 since the fact that the shifted 12 inserts on sclerotisation is not the result of a concomitant shift or expansion of L4l.

In *Polyphaga* the contribution of the L4I-region to L4K can be directly deduced from the l4-insertion on L4K. For the presence of L4n, however, there is, like in *Lamproblatta*, no direct evidence (the nla-process and muscle l14 are missing). L4K is hence assumed to be mainly made of anterior parts of L4I, with little (like in *Cryptocercus*) or no contribution from L4n.

The situation in *Ergaula capucina* (fig.326d, 327d) can be derived from that in *Polyphaga* (fig.326c, 327c): **L4K** is likewise ventral to the **hla**-base but has shifted even further anteriad. The dorsal part of **L4K**, which bends into the **hla**-base, is distinctly shorter (compare edges **X** in fig.326c and d) and fused to the ventral anterior margin of sclerite **L3** (along edge **X** and more anteriorly). A muscle coming from the same part of **L2** as **l4** in *Polyphaga*, which is certainly homologous with this **l4**, inserts on this compound sclerite (mainly along edge **X**: **l4** in fig.327d). Muscle **l11** has the same insertions as in *Polyphaga* and *Lamproblatta* (fig.327b,c,d) but is much stouter. The muscles **l2** and **s1** insert like in *Polyphaga*.

The morphology of L4K and l4 of *Ergaula* could easily be mistaken as corresponding with the situation in Blattellidae (*Anaplecta* excluded) and Blaberidae: In the latter groups the main muscle of the hla-hook (l14 in fig.249, 276, 303) runs from the anteriormost part of L2 to sclerite L3. The situation in *Anaplecta* (fig.222) suggests that this hla-muscle is a true l14 whose anterior insertion has been translocated from the L4n-region (with nla) to L2 (discussion in 6.4.3.). Looking at *Ergaula* only, the "hla-muscle" (l4) with its course from L2 anteriorly in the lve-pouch to the "base of L3" (= L4K) could easily be misinterpreted as the "l14", with the "translocation" of its anterior insertion to L2 being a synapomorphy of *Ergaula* and the respective Blattellidae and Blaberidae. However, the situations in *Polyphaga* and *Lamproblatta* clearly show that in *Ergaula* the muscle is l4 (not 114), the sclerite is L4K (not the basal part of L3), and the similarity with Blattellidae is a case of convergence.

Ventral sclerite plate Sclerite L4G in the ventral wall of the vla-lobe of *Eurycotis* and *Archiblatta* is, by definition, the region L4v (fig.325e,f). L4G of *Tryonicus* (fig.325g) probably also corresponds exactly to the L4v-region, but it cannot be excluded that parts of the L4c-region are contained in the sclerite (compare in 6.3.1.). In the latter case, L4G of *Tryonicus* and L4G of *Eurycotis* and *Archiblatta* would be only partly homologous. The ventral plate of *Anaplecta* (L4G in fig.205) lies similarly in the ventral wall of the vla-lobe like L4G of *Tryonicus* (fig.87), and in both species L4G is, apart from the ventral parts of sclerite L4K, the only sclerotisation in the ventral wall of the left complex. This indicates that L4G of *Tryonicus* and *Anaplecta* are strictly homologous. The ventral plates of *Cryptocercus* (L4G in fig.148), *Lamproblatta* (L4R in fig.174), and *Polyphaga* (L4M in fig.115) also lie in the ventral vall and can be assumed to be at least in part homologous with each other and with the L4G-plates of the other species. These plates, however, are rather different in their relative sizes, and the homology relations should be analysed in detail.

Some evidence for the exact homology relations comes from the muscles 12, 15, 16a, 16b (homology discussion in 6.1. and 6.2.), and s3 (homology discussion in 6.9.). In *Sphodromantis, Mantoida, Polyphaga*, and *Lamproblatta* s3, 12, 15, 16a, and 16b can be homologised one by one (with the exception that in *Mantoida* 16a and 16b have fused). In *Cryptocercus* homology is clear for s3 (right part of s1+3), 12, and 16b; 15 and 16a have been lost. In *Eurycotis* and *Anaplecta* homology is also clear for s3, 12, and 16b; as regards 15 and 16a, homology with the 15 and 16a of the other species is questionable. The relations between the ventral plates and the insertions of 16b, s3, and 12 are different in the various species:

- In Eurycotis, Cryptocercus, and Anaplecta only 16b (fig.70, 157, 224) inserts, at least in part, on the ventral plate (L4G in fig.63, 148, 205). s3 (fig.70, 157, 222) inserts in the membrane left-anterior to L4G. The positions of these insertions may suggest (but do not prove) that the L4G of Anaplecta and Cryptocercus are strictly homologous with L4G of Eurycotis (i.e. only L4v but no parts of L4c or of other sclerotisations are included; fig.325e,h,l). The same might be assumed for Tryonicus since its L4G is similar to L4G of Anaplecta (fig.325g).

- Lamproblatta and Polyphaga (and Ergaula) differ from the previous species (fig.132, 133, 188, 189): (1) Not only l6b but also s3 inserts on the ventral plate (L4R in fig.174; L4M in fig.115). This is certainly a derived feature. (2) l5 and l6a also insert on the ventral plate, but since the homology relations with the respective muscles of the previous species are uncertain, this feature is not interpretable (no L4c-region is included in fig.325i,k, but its absence is questionable). (3) A special muscle s12 from the right half of the subgenital plate runs to the ventral plate and inserts immediately to the right of s3. The presence of s12 is also a derived feature.
- Polyphaga (and Ergaula) shows an additional derived feature already mentioned above: Muscle 12 inserts on the ventral plate L4M (fig.128).

The derived condition that, in *Lamproblatta* and *Polyphaga* (and *Ergaula*), the insertions of some muscles are now on the sclerotisation of the ventral plate (at least s3 in *Lamproblatta* and s3 and l2 in *Polyphaga*) is interpreted as an expansion of this plate, and the sclerotisations bearing these insertions are defined as new regions of L4:

- L4a (anterior): The sclerotisation of the insertion area of s3. (The s3-insertion has not changed its position.)
- L4x : The sclerotisation of the insertion area of l2. (The l2-insertion has shifted ventrad.)

According to this interpretation, L4M and L4R are not strictly homologous with each other and with the L4G of the other species: L4R evolved from L4G by expansion (new region L4a), and L4M evolved from L4R by an additional expansion (new region L4x). In *Ergaula* a small anterior part of L4M (with the insertions of s3 and s12) has split off to form a sclerite of its own (compare fig.3221 and m).

Nahublattella

The homologue of sclerite L4K of *Anaplecta* (fig.209) has divided into two sclerites L4U' and L4V' (fig.242). L4U' resembles the posterior part of L4K: It has the same position on the left edge of the left complex, the same position relative to the hla-hook, and a similar shape (curved plate). L4V' resembles the anterior part of L4K: It lies in the anteriormost ventral wall of the left complex and forms a process (nla in fig.242, 248). The homology of the nla-processes of the two species is, regarding their different shape, debatable.

These relations are supported by the muscles: L4U' bears the insertions of l2 and l4 (fig.249). l2 runs to the basalmost part of the hla-hook (membrane 30). l2 of Anaplecta (fig.221) runs to the pne-pouch next to the hla-base (30 in fig.210, 211). I assume homology for the l2 of the two species and a slight shift of the right insertion in *Nahublattella*. l4 of *Nahublattella* inserts immediately ventral to l2 and runs to the lve-pouch, exactly like l4 of e.g. *Eurycotis* (fig.70, 71) and *Cryptocercus* (fig.155, 156). As mentioned above, l4 has been lost in *Anaplecta*. L4V' bears the insertions of l5, l6a, and s3 (fig.250, 251). l5 has its posterior insertion like l5 of *Anaplecta* (fig.223) at the left base of the lve-apodeme, and homology is highly probable for these l5; that the anterior insertion is on the anterior part of L4K in *Anaplecta* confirms the homology between this part of L4K and L4V' (and, maybe, the homology of the nla-processes, too). The insertions of s3 and l6a in *Anaplecta*, however, are on the ate-tendon to the right of the

sclerotisation (fig.222). (Homology is quite certain for s3 but not for l6a; discussion in 6.9. and 6.2.4.). That the dorsal insertion of s3 is on a sclerotisation is not comparable with the situation in Lamproblatta and Polyphaga: In the latter species the sclerotisation concerned is an expansion of the ventral plate (L4a-region); in Nahublattella the sclerotisation with the s3-insertion is an expansion of the former L4K-sclerite. (The respective area of the sclerite could be defined as a new sclerite region, but this is omitted). Thus, L4U' is assumed to consist of the same parts of the L4l-region as the posterior part of L4K in Anaplecta. L4V' roughly corresponds to the L4n-region (fig.325m, compare fig.3251); however, the line dividing the two sclerites does certainly not exactly correspond to the border between L4l and L4n; this is only the case – by definition – in Archiblatta. The homology relations of the processes **paa** and **pda** of Anaplecta and **via**, **paa**, **pda**, and vsa of Nahublattella (fig.241, 244) and of their sclerotisations have been discussed in 6.2.4.. L4N' of Nahublattella is probably the left-ventral sclerotisation of the viaprocess (including **pda** and **vsa**; fig.325m). The ribbon-like extension **L4d'** at the left base of via (fig.244, 250) closely resembles L4d of Polyphaga and Tryonicus (fig.94, 97, 118, 129) in its position relative to **paa** and **pda** and their sclerotisation and to the **l10**-insertion. Like in Polyphaga and Cryptocercus, L4d' is directed to the left. Like in Mantoida and Cryptocercus, L4d' has a muscle running to the pne-pouch (11 in fig.48, 155, 249). Homology is assumed for the L4d and l1 of all species. In Nahublattella the whole area containing via, pda, paa, and L4d' is sunken anteriad into the left complex and has become the left part of an expanded lve-pouch, and L4d' lies in the left edge of this enlarged lve-pouch and runs posteriad (L4d' is, so to speak, invaginated). This is in contrast to all other species; only Anaplecta shows a slight anteriad invagination of the paa+pda-sclerotisation (but L4d has been lost). The separation of the paa+pdasclerotisation from the L2-sclerotisations in the lve-pouch reminds of Lamproblatta (fig.177-179), but the division of the sclerotisations is different and non-homologous: L4d is connected with the paa+pda-sclerotisation in Nahublattella but with the sclerotisation of the lve-pouch in Lamproblatta (compare fig.329f and g).

The identification of the vla-lobe (fig.245-247) was done in in 6.2.4.. That there is no sclerite plate in its ventral wall (L4v-region; compare L4G of *Anaplecta*, fig.224) is a derived feature.

Parcoblatta, Blaberus, and other Blattelidae and Blaberidae

Sclerite L4U' of *Blaberus* has the same shape, relative position, and muscle insertions as L4U' of *Nahublattella* (fig.242, 249, 299, 303). In both species l4 runs to sclerite L2, and l2 runs to the hla-base (30 in fig.249, 303). In *Parcoblatta*, the morphology of the left part of the left complex (compare fig.268-270 and 299-301) and the arrangement of l2 and l4 (compare fig.276 and 303) are nearly the same as in *Blaberus*; however, sclerite L4U has been lost. In *Nyctibora* L4U is present and very similar to L4U' of *Blaberus*.

Blaberus and *Parcoblatta* both have a tendon-like invagination (ate in fig.268, 271, 302) near the ventral basal line of the left complex. ate is also present, and in the same position, in other Blattellidae and Blaberidae (investigated species: those listed in 5.15.; *Blaptica*: fig.291). The homology of these **ate**-tendons is confirmed by the insertion of a phallomero-

sternal muscle (s3b, studied in *Parcoblatta*, *Blaberus*, and *Blaptica*; fig.277, 304; homology discussion of s3b in 6.9.) and by the presence of a sclerite in the dorsal wall of the tendon (L4V or L4V', which, however, is present only in *Parcoblatta*, *Nyctibora*, and *Blaptica*, fig.289, 291).

The evolutionary origin of tendon **ate** and sclerite **L4V** of these species is unclear. The terms used express the possible homologies with structures being in similar positions in *Anaplecta* (**ate** in fig.212) and *Nahublattella* (**L4V'** in fig.244):

- ate and, if present, L4V resemble both ate of Anaplecta and L4V' of Nahublattella in bearing the insertion of at least part of (1) muscle s3 (s3b of Parcoblatta and Blaberus, fig.277, 304) and (2) muscle l6a (only Blaberus, fig.304). However, it is impossible that both homologies of ate and L4V are true in a strict sense since in Anaplecta the ate-tendon and the sclerotisation homologous with L4V' of Nahublattella (anterior L4K) are located side by side.
- As a combined hypothesis accepting a partial homology of the ate-tendons and a strict homology of the L4V-sclerites, it might be assumed that in the more derived Blattellidae and Blaberidae, as compared with *Anaplecta*, the cuticular area forming the ate-tendon has expanded basad and that by this process L4V has become integrated into the tendon. *Nahublattella* could be an intermediate, with the anteriormost ventral part of the left complex being a very broad ate-"tendon", and with L4V' integrated into this "tendon". In the other Blattellidae and Blaberidae this anterior part with L4V' must then be assumed to have become very narrow, and L4V' has become smaller. If this is true, the ate-tendon of *Anaplecta* would be homologous with the distalmost part of the ate-tendon (anterior to L4V, if present) of e.g. *Supella, Euphyllodromia, Parcoblatta, Nyctibora, Blaptica*, and *Blaberus*.
- However, the lack of a sclerotisation within the ate of *Supella*, *Euphyllodromia*, and other species might suggest that L4V of *Parcoblatta*, *Nyctibora*, and *Blaptica* is a new element not homologous with L4V' of *Nahublattella*. If this is true, ate of *Anaplecta* could be strictly homologous with ate of the more derived Blattellidae and Blaberidae. These questions concerning ate and L4V cannot be settled here.

The sclerotisation of the **via**-process has been assumed, in accordance with the situation in *Nahublattella*, to be composed of L4N and L2E (posterior L4I-region and L2d-region: fig.325m,n,o and 324m,n,o; discussion in 6.2.4.). Since the primary processes **paa** and **pda** are no longer distinguishable in these **via**-processes (fig.328c-k), the exact arrangement of L4N and L2E is less clear than in *Nahublattella*. In determining the position of the L4N- and L2E-sclerotisations on **via** of *Parcoblatta* and *Blaberus* one must consider the rotation of the **via**-process. An extension corresponding to L4d' of *Nahublattella* is missing in all species (compare fig.328b and c-k), and the L4d-region is assumed to have been lost like in *Anaplecta* (fig.325l,m,n,o and 324l,m,n,o).

At least *Parcoblatta, Nyctibora, Blaptica, Nauphoeta*, and *Blaberus* (the other species not investigated) lack, like *Nahublattella*, a sclerite plate in the ventral wall of the **vla**-lobe (fig.266, 268, 297): The **L4v**-region has been lost. Sclerite **L10'** of *Blaberus* (fig.299) and the small sclerites **L10'** of *Blaptica* (fig.291) are not assumed to be descendants of **L4v** but new sclerotisations having evolved within Blaberidae. In the blaberid *Nauphoeta* **L10'** is missing.

6.4. Left complex IV: Main sclerite L3 and associated elements

6.4.1. Comparison between Blattaria and Mantodea

In *Archiblatta* (fig.53-55) the **hla**-hook is an evagination of the anterior left ventral wall of the left complex, and its base is immediately beneath the arched anterior part of the **L4C**-sclerite = **L4l**-region. *Mantoida* has no process in the corresponding part of the ventral wall (fig.45, 46), and the neighboring processes **paa** and **pda** have proved to be homologous with **paa** and **pda** of *Archiblatta* and other Blattaria. The elements of the left complexes of *Chaeteessa, Metallyticus*, and *Sphodromantis* – including the processes **paa** and **pda** – have all been homologue of the **hla**-hook; **hla**, and also its sclerite **L3** and its main muscle **l14**, are restricted to Blattaria.

6.4.2. The elements in the common ground-plan of Blattaria and Mantodea

Since hla is present in all Blattaria (discussion in 6.4.3.) but absent in all Mantodea, its presence in the common ground-plan cannot be reliably decided. However, a comparison of the copulation habits of Blattaria and Mantodea might indicate that the lack of hla and L3 in Mantodea is a derived feature.

In Blattaria copulation has several successive phases (data from Scudder 1971, who refers to Gupta 1947): In *Periplaneta*, in phase (1), the male places itself in front of the female, with its rear end facing the female. Then the female climbs upon the back of the male, both animals facing the same direction. In this phase the **hla**-hook of the male makes the first contact of the genital regions: It seizes the terminal lobes of the female subgenital plate (Scudder: "initial seizing"). (2) This connection being established, the male rotates ca. 180° in the horizontal plane (clockwise as seen from above). (3) After this rotation the animals are again in a line, with their rear ends still in contact. Now other phallomere elements establish a firmer contact – mainly the seizing apparatus formed by the posterior part of the male's right phallomere (Scudder: "final holding"). Scudder describes a several-phase process with similar positions for some subgroups of Ensifera. But, of course, the connection of male and female genitalia is established by completely different structures. Nevertheless, it seems plausible that a copulation procedure with a sequence of these positions might be plesiomorphic for a higher taxon including at least Orthoptera and Dictyoptera.

Mantodea have a different copulation procedure, which Scudder regards as apomorphic: The male mounts the female (often by jumping) and then clings to the female thorax with its grasping legs. Holding this position, the male curves its terminal abdomen to the right and pushes it into the female genital pouch from laterally (e.g. Kumar 1973). Together with the modified fore legs, the very special feeding habits of Mantodea (lurking predators) are certainly derived. It might be plausible that changes in behaviour correlated with these new feeding habits might have caused changes in the copulation procedure, (So to speak, it is no longer advisable for the male to place itself in front of the female in the way Blattaria do).

Thus, the outgroup comparison with Ensifera as well as biological properties of Mantodea suggest that the copulation procedure of Blattaria is plesiomorphic and that of Mantodea apomorphic: Phase (1), in which Blattaria make use of their **hla**-hook, can be regarded as secondarily lost in Mantodea. Additionally, since the phallomeres of Mantodea and e.g. *Periplaneta* are rather similar in their morphology (and completely different from those of Ensifera), it might be assumed that the way the Mantodean phallomeres functioned before the copulation procedure has changed was similar to that of the Blattarian phallomeres, and that a **hla**-hook was present for initial seizing. Though these ideas are highly speculative, it is at least plausible that **hla** and the associated elements **L3** and **l14** were present in the common ground-plan of Blattaria and Mantodea and have been lost in Mantodea. The same might also be true of the **nla**-process, which is present in many Blattaria (fig.69, 98, 212) but never in Mantodea. **nla** and **l14** are lost an additional loss of **nla** could be expected.

6.4.3. Homology relations and character states of the elements in Blattaria

The **hla**-hook is present in all Blattaria. The homology of all these **hla** is suggested by their position in the leftmost part of the left complex, by their similar shape, and by the presence of a special sclerite **L3** occupying the distal part of **hla** (**L3**, however, can be very different in its extension). Apart from these superficial features, additional similarities between certain species confirm this homology assumption. The most important question in this context is whether the main muscles of the **hla**-hooks (called **l14** in most species) are homologous.

Archiblatta, Periplaneta, and Eurycotis

The homology of L3, hla, and l14 of these species is quite evident. (1) The hla-base takes the same relative position: right-ventral to the L4l-region, left-posterior to the L4n-region with the nla-process, and left-anterior to sclerite L4F (fig.54, 56, 66, 67). (2) L3 occupies the entire hla except for the basalmost part (30 in fig.65-67). (3) The tip of hla is two-pointed (fig.53, 65). (4) In *Periplaneta* and *Eurycotis* the main muscle of hla (l14c in fig.72) comes from the L4n-region on and near the nla-process and inserts immediately behind s1 (fig.70). However, only *Eurycotis* has one accessory hla-muscle l14d (fig.73) – possibly a subdivision of l14c.

Cryptocercus and Lamproblatta

The **hla**-base has a similar position relative to the insertions of **l2** and **s1** (fig.156, 157, 184, 185) as in *Eurycotis* (fig.70), and the anterior insertion of the main muscle of **hla** (**l14** in fig.157, 184, 185) is likewise immediately behind the **s1**-insertion (fig.157, 158, 184, 185). Thus, homology can be assumed for the **hla**, **L3**, and **l14** of these three species (homology discussion of **s1** in 6.9.). *Cryptocercus* has one accessory **hla**-muscle **l19** (fig.156); *Lamproblatta* has two, **l22** and **l23** (fig.184-186). These accessory muscles and **l14d** of *Eurycotis* all have different insertions, and homology relations are not assumed. In *Cryptocercus* – as compared with the previous species, *Polyphaga*, and *Ergaula* (see below) – the base of **hla** is more posteriorly, and **hla** is shorter (fig.151).

Polyphaga and Ergaula

The homology of **hla** with **hla** of *Lamproblatta* and *Cryptocercus* is suggested mainly by the similar position of the **hla**-base posterior to the **s1**-insertion (fig.127, 157, 185) and by the similar relations between the **hla**-base and sclerite **L4K** (discussion in 6.3.4.). A muscle inserting directly on **hla** or **L3** (**l14**) is missing; the very stout **l4** has probably taken over the function of **l14** (discussion in 6.3.4.).

Tryonicus

The hla-base has the same relative position as in *Eurycotis*: right-ventral to the L4I-region and left-posterior to the nla-process (sclerites L4K and L4N in fig.97). hla and L3 of the two species are certainly homologous. *Tryonicus*, however, shows three special features as compared with the species discussed so far: (1) The hla-base is distinctly more posteriorly (compare fig.87, 97 and 63, 67). However, this is also true of *Cryptocercus*. (2) The introversible membranous basal part of hla (30 in fig.97) is by far more extensive, and, consequently, hla can be retracted more deeply into the left complex. (3) The basal margin of L3 is connected with L4 (L4K) by the sclerite ribbon L3a (fig.89,98). This last feature is restricted to *Tryonicus*.

Anaplecta, Nahublattella, Parcoblatta, and Blaberus

The two first-mentioned peculiarities of *Tryonicus* are more pronounced: The hla-base is at the posterior edge of the left complex, and the membranous basal part of hla (30 in fig.210, 242, 269, 300) is so extensive that hla can be retracted into the left complex except for its distalmost part only (fig.210, 242, 269) or even completely (fig.295a). (These two features have also been investigated and found in all other Blattellidae and Blaberidae listed in 5.15.). Another feature common to these 4 species is the membranous infolding **fpe** separating the left part of the left complex (with the hla-base) from the right part (fig.210, 243, 268, 299). These similarities clearly suggest the homology of hla and L3 in the 4 species. With *Tryonicus* as an intermediate, homology can also be assumed with hla and L3 of the previous species.

Additionally, the homology of hla and L3 in *Anaplecta* and *Eurycotis* is more directly suggested by the anterior insertion of the hla-muscle l14 or l14c,d, which is, in both species, on and near the nla-process (fig.72, 73, 222). In *Nahublattella, Parcoblatta*, and *Blaberus*, however, the anterior insertion of the main muscle of hla (l14 or l14a,b in fig.249, 276, 303) is on L2D' or L2, on top of the lve-apodeme (L2a-region). In *Anaplecta*, interestingly enough, the top of the lve-apodeme and the nla-process are firmly connected with each other (fig.222). This might suggest that all Blattellidae and Blaberidae have gone through an evolutionary stage showing this connection, and that, at that time, muscle l14 has shifted from L4n to L2a. Homology is assumed for all hla-muscles l14. (The shift of l14 will be disscussed in a functional context in 7.5.).

Of these 4 species only *Parcoblatta* and *Blaberus* show the following features: (1) **114** is divided into two bundles (**114a** and **114b** in fig.276, 303; the division in *Eurycotis* mentioned above is clearly not homologous with this division). (2) There is a muscle within the membranous basal part **30** of **hla** (**136** in fig.276, 303). (3) The distal part of

hla has a groove hge with a notch 45 in its ventral wall (fig.266, 297a). In the species studied only in part (listed in 5.15.), the hge-groove and the notch 45 are distinctly present in *Supella, Euphyllodromia, Loboptera, Byrsotria*, and *Blaptica*; *Nyctibora* has only hge but no notch 45; in *Ectobius* and *Nauphoeta* the hge-groove is quite indistinct, and the notch 45 is missing. (114 and 136 have not been investigated in these species). Muscle 146 is peculiar to *Blaberus* (fig.304, left part).

6.5. Left complex V: Further main sclerites and muscles

Some Blattaria and Mantodea have small sclerites in the dorsal wall of the vla-lobe, which I have designated L5. L5 of Metallyticus (fig.26, 27) and Cryptocercus (fig.151, 155) is posterior to the genital opening. L5 of Periplaneta (no figure) lies more anteriorly, within the terminal part of the ejaculatory duct. L5 of Polyphaga (fig.123, 124) is far to the left of the genital opening. L5 of Ergaula is situated like in Polyphaga but is tranversely orientated and approaches the genital opening more closely (fig.322m). In Anaplecta and Nahublattella, the extension 28 of the L2- or L2D'-sclerite (fig.214, 215, 245) takes a very similar position relative to the other parts of L2 and to the genital opening as L5 of Polyphaga (fig.123) and might be homologous with it. The sclerites L10' of Blaberus and Blaptica (fig.291, 300) lie either in the dorsal vla-wall (Blaptica) or along the posterior edge of the vla-lobe (*Blaberus*); whether they show any kind of homology relation with the L5 of the other species is unclear, and improbable in my view. Sclerites in the dorsal vla-wall are missing in Mantoida, Chaeteessa, Sphodromantis, Archiblatta, Eurycotis, Tryonicus, Lamproblatta, and Parcoblatta. It cannot be decided whether L5 is a ground-plan element of Blattaria and Mantodea and has been lost several times, or whether such sclerites have developed several times independently.

Sclerite L7 is present only in *Polyphaga, Ergaula*, and *Lamproblatta*. These L7 (fig.115, 174) take the same relative position between the sclerite plate of the vla-lobe (L4M, L4R) and the right phallomere and are therefore assumed to be homologous. L7 is regarded as an element of the left complex since in a specimen of *Polyphaga* with its external genitalia consisting of two right phallomeres only there was no trace of L7 (compare in 3.1.). Only in *Polyphaga* and *Ergaula* the area containing L7 is elaborated as a special lobe-like evagination (lba in fig.115; in *Ergaula* the morphology is the same, but L7 and lba are larger). The lba-lobe is assumedly homologous with the rightmost part of the vla-lobe of the other species. (If this is true, not the vla-lobe of *Polyphaga* and *Ergaula* alone but the vla- and lba-lobes together are the strict homologue of the vla-lobe of the other species. That lba is not alone the homologue of the vla of the other species and that L7 is not the homologue of the L4G-plates is clearly shown by the muscles l5, l6a, and l6b, compare in 6.2.1.. L7 and lba are bare of muscles).

Sclerite L8 is likewise restricted to *Polyphaga, Ergaula*, and *Lamproblatta* (L8 = neoformation N of Grandcolas & Deleporte 1992). These L8 take the same position in the right dorsal wall of the left complex, but they differ somewhat in their position relative to the **pne**-pouch (fig.117, in *Ergaula* similar; fig.177). However, it must be considered that in *Lamproblatta*, as compared with *Polyphaga*, the **pne**-pouch has shifted right-

anteriad (compare in 6.1.4.). The homology of the L8-sclerites is also strongly supported by the insertions of three muscles (**112**, **b2**, **19**) in their immediate vicinity: **112** (fig.128, 129, 186, 188; discussion in 6.2.4.) runs to the right ventral (or outer) wall of the **Ive**pouch, with its insertion close to that of **16a** (fig.133, 188). **b2** (fig.127, 184; discussion in 6.8.) runs to the ventral part of the right phallomere, where the insertion, however, has a slightly different position in *Polyphaga* and *Ergaula* on the one hand (**R3**, fig.141) and *Lamproblatta* on the other (membrane next to **R2**, fig.198). **19** (fig.127, 184, 170; discussion below) runs to the left dorsal wall of the left complex. **L8** and the three muscles are assumed to be homologous, and **L8** and **112** are regarded as derived features of these species.

Ergaula and *Eurycotis* have sclerites in the dorsal wall of the **pne**-pouch (**L9** in fig.322m; **L6A** and **L6B** in fig.66), but **L9** and **L6** are probably not homologous. Sclerite **L11** (fig.91) is peculiar to *Tryonicus*.

Many species have transverse muscles within the dorsal wall of the left complex, which have been termed 19: Mantoida (fig.49), Eurycotis (fig.70), Polyphaga (fig.127, 129), Cryptocercus (fig.155), Lamproblatta (fig.170, 185), Anaplecta (fig.221), Nahublattella (19a and 19b in fig.249), Sphodromantis (fig.17; "b4, 19?" might be the homologue of either 19 or b4a and b4b of Mantoida: compare in 6.7.3.). However, the exact position and the extension of these 19 can be rather different. The homology of 19 of Lamproblatta and Polyphaga (and Ergaula) is highly probable since the right insertion is on or near sclerite L8 and close to the insertions of 112 and b2. In *Polyphaga* (and *Ergaula*) as well as in Anaplecta and Nahublattella 19 has its left insertion, at least in part, on the right wall of the **pne**-pouch. This relation between **pne** and **19** is assumed to have been lost in Lamproblatta by the right-anteriad shift of pne and L1 (compare in 6.1.4.). 19 of Eurycotis has a similar position like the dorsal part of 19 of Polyphaga. In Cryptocercus 19 is far on the left; that its left insertion is next to the L4d-region (left part of sclerite L4N in fig.155) and close to the l1-insertion resembles the situation in Mantoida (fig.48, 49; compare in 6.3.4.), but this close relation between 19 and L4d is in contrast to Polyphaga (fig.127, 128). On the other hand, 19 of *Mantoida* is farther to the right than 19 of *Cryptocercus*, and its overall position is similar to that of 19 of Eurycotis and the dorsal part of 19 of Polyphaga. In my view, these similarities are sufficient to assume homology for all these 19-muscles and to regard 19 as an element of the common ground-plan of Blattaria and Mantodea. In the evolution of 19, some shifts might have occurred, or different parts of 19 might have been reduced or enlarged in the various species.

Some Blattaria have muscles from the ejaculatory duct **D** to that part of the dorsal wall of the **vla**-lobe posterior to the genital opening; these have been termed **l13**: *Polyphaga* (fig.132), *Cryptocercus* (fig.155), *Lamproblatta* (fig.188), *Anaplecta* (fig.222), *Eurycotis* (**l13h** in fig.72). Homology is tentatively assumed for them though their positions are somewhat different. In *Anaplecta* **l13** bridges the base of the **vfa**-outfolding (an outfolding from the anteriormost dorsal wall of **vla**, compare in 6.2.4.). In *Eurycotis* some other diffuse muscles within the **vla**-lobe have been assigned to **l13** (**l13a,b,c,d,e,f,g,i** in fig.71-73); these could be new muscles, or some of them might be split off parts of the true **l13** (*Archiblatta, Blatta, Periplaneta*, and *Deropeltis* not investigated). In Mantodea no **l13**-

muscles have been found; however, muscle **b3** of *Sphodromantis* (fig.15) has its right insertion not far from the dorsal **vla**-wall and might be a shifted **l13**. Hence, it is not clear if **l13** is present in the common ground-plan of Blattaria and Mantodea.

Mantoida and *Cryptocercus* have a longitudinal muscle in the posterior left ventral wall of the left complex (**17** in fig.52, 158). Since the position is very similar these **17** could well be homologous. However, the respective part of the left complex is very different in the two species (presence or absence of the **hla**-hook, highly modified **L4**-sclerotisations in *Cryptocercus*), and it is not possible to compare the relative position of **17** in the two species. Therefore, the homology of these muscles must be regarded as highly questionable. **17** of *Sphodromantis* (fig.15) is certainly homologous with **17** of *Mantoida* but has undergone a shift (compare in 6.3.3.). *Nahublattella, Parcoblatta*, and *Blaberus* also have longitudinal muscles in the ventral wall of the left complex (**130** in fig.251, 307; **130a,b** in fig.278, 279); these **130** are assumed to be homologous, but since they take a rather different position homology with **17** of *Cryptocercus* is not assumed.

6.6. Left complex VI: The position of the phallomere-gland opening

The opening of the phallomere-gland **P** certainly has its primitive position within the membranous part of the **pne**-wall (discussion in 6.1.1.). It opens far anteriorly into this membrane in *Mantoida* (fig.45), *Chaeteessa* (fig.32), and *Sphodromantis* (fig.10), and far posteriorly and on the left side in *Cryptocercus* (fig.153, 154), *Polyphaga* (fig.120, 121), *Tryonicus angustus* (fig.107, 108), and – considering the rotation of the **pne**-pouch – *Tryonicus parvus* (fig.95, 96).

In *Ergaula capensis* the opening has, as compared with *Polyphaga*, shifted only a short distance; by this shift, however, it has reached a position left-ventral to the **dca**-processes and outside the **pne**-wall (compare fig.106 and 121). In *Eurycotis* (fig.67, 68) and *Archiblatta* (fig.54-56) the opening is likewise ventral to the **dca**-processes and is assumed to have undergone a similar shift. In *Lamproblatta* the opening has the same position relative to the posterior margin of **L1** (fig.177, 178) as in the previous three species but is farther away from **L1**, and the processes **paa** and **pda** have, as compared with e.g. *Polyphaga* (fig.118), shifted to the right (relative to the left posterior end of the **lve**-pouch; compare in 6.3.4.) and are assumed to have intruded into the interspace between **L1** and the phallomere-gland_opening.

In *Nahublattella* the opening has a similar position relative to sclerite L1 and the dcaprocess as in e.g. *Ergaula* (fig.243, 244, 328b) but has shifted far anteriad within the membrane ventral to dca and lies in the posterior right dorsal wall of the lve-pouch – posterior to the dorsal wall of the ejaculatory duct D. The muscles l27 and l29 (fig.249) are derived features of *Nahublattella*. In *Parcoblatta* (fig.270, 328e), *Blaberus* (fig.300, 328k), *Euphyllodromia* (fig.328d), *Nyctibora* (fig.328h), and *Nauphoeta* (fig.328i) the opening has a similar position as in *Nahublattella* but is slightly more to the left and close to sclerite L2 or L2D. The situation in Anaplecta is difficult to interpret. This concerns the presence of two outlet ducts with their openings close to each other (P in fig.216), the position of these two openings, and the presence of a muscle 125 (fig.224) inserting between them. Taking a situation like in Nahublattella as a starting point, the position of the openings could be explained by the assumption of a further shift to the left within the dorsal wall of the ejaculatory duct, and then ventrad to beneath the lve-apodeme. However, the preceding shift assumed for Nahublattella would have hardly been possible in Anaplecta since between the membrane posteroventral to the pne-pouch (fig.209) and the posterior dorsal wall of the ejaculatory duct there are still extensive right parts of L2 (fig.211) "blocking" this shift. The openings of Anaplecta are in one respect similarly situated as in Lamproblatta: more or less ventral to the processes paa and pda (compare fig.210 and 178). The position relative to the lve-pouch, however, is completely different: dorsal to lve in Lamproblatta, ventral to lve in Anaplecta. Possibly, the outlet ducts of Anaplecta are new organs. In this case, for the remaining Blattellidae and for Blaberidae the possibility has to be considered that their glands and outlet ducts are homologous with those of Anaplecta (or one of them) and not with those of the other species. As a point possibly interesting in this context, the spermathecae of the female genitalia have also been replaced by completely new organs in Blattellidae and Blaberidae (McKittrick 1964).

6.7. The elements of the right phallomere

6.7.1. Comparison between Blattaria and Mantodea

The homology relations and the ground-plan of the elements of the right phallomere can best be deduced from a comparison of *Eurycotis, Chaeteessa*, and *Mantoida*.

The cuticular elements of the right phallomeres of *Eurycotis* and *Chaeteessa* show the following similarities:

- 1. A sclerite **R3** occupies the anteriormost ventral wall of the right phallomere (fig.28, 77).
- 2. At least the right and the right anterior margins of **R3** form a groove-like apodeme **age** (fig.28, 77).
- 3. The right posterior end of R3 articulates (A3 in fig.28, 77) with more posterior sclerites (*Eurycotis*: R1F in fig.77; *Chaeteessa*: R1B in fig.28).
- 4. The anterior part of both **R1F** and **R1B** extends to the left and reaches an edge (16 in fig.28, 77) along which it bends sharply dorsad.
- 5. Then this sclerotisation arches dorsad and then to the left. The arching in a dorsal direction is extensive in *Eurycotis*; in *Chaeteessa* it is less pronounced and the sclerotisation extends mainly to the left.
- 6. The posterior margin of this sclerotisation forms a posteriad-directed ridge (**pva** in fig.28, 78).
- 7. To the left of (*Chaeteessa*) or left-ventral to (*Eurycotis*) this **pva**-ridge the right phallomere has a large central invagination (**cbe** in fig.29a, 31, 77, 78; the whole of **cbe** is a part of the ventral wall of the right phallomere).

- 8. The posterior part of the right phallomere is composed of a dorsal lobe (**fda** in fig.31, 74) and a ventral tooth or ridge (**pia** in fig.28, 29a, 77, 78). **fda** and **pia** are connected along the right edge of the right phallomere, and they diverge to the left like the two halves of an opened book. In *Eurycotis* **pia** is as large as **fda** (and two-pointed); in *Chaeteessa* **pia** is much smaller than **fda**.
- 9. The dorsal wall of the **fda**-lobe is sclerotised (*Eurycotis*: **R1H** in fig.74; *Chaeteessa*: **R1A** in fig.31).
- 10. The posteroventral part of both R1F and R1B (fig.28, 77; posterior to edge 16) extends onto the pia-tooth. However, the sclerotisations of the anterior and of the posterior parts of pia are connected in *Chaeteessa* (R1B) but separated in *Eurycotis* (R1F anteriorly and R1G posteriorly, which articulate in A9).
- 11. A large membranous area (17 in fig.28, 77) is present at the posterior right edge of the right phallomere, between the right margins of the sclerotisations of fda and pia.

Homology is assumed for all these similarities, for all elements given the same name, and for the compared sclerotisations taking the same relative positions. 1.-11. are assumed to be features of the common ground-plan of Blattaria and Mantodea.

A further similarity between *Chaeteessa* and *Eurycotis* is that the sclerotisation adjoining articulation A3 posteriorly (R1B in fig.28; R1F in fig.74, 77) is separated from the dorsal sclerotisation of the fda-lobe (R1A in fig.28, 31, 32, R1H in fig.74, 77) by membrane (4 in fig.28, 32, A8 in fig.74). The dividing lines 4 and A8, however, are probably non-homologous (discussion below).

There are also some essential differences between *Eurycotis* and *Chaeteessa*: (1) The separation or connection of the anterior and posterior sclerotisations of **pia** (compare feature 10.). (2) Only *Eurycotis* has a sclerite **R2** (fig.77), which articulates with **R3** (**A7** in fig.75, 77) and **R1F** (**A6** in fig.75). (3) In *Eurycotis* the **cbe**-invagination has a summit in the center of the right phallomere and becomes shallower to the left of this summit (where **R2** adjoins; fig.75, 78); in *Chaeteessa* the **cbe**-invagination becomes continuously deeper to the left (fig.29a, 31). (4) Only *Eurycotis* has a **tre**-tendon in the anteriormost dorsal wall of the **fda**-lobe (fig.74). (5) Only *Eurycotis* has the sclerotisations of **pia** (**R1G**) and **fda** (**R1H**) connected with each other posterior to the membranous area **17** (by a narrow sclerite bridge; fig.77, 78).

As regards the right phallomeres of *Mantoida* and *Chaeteessa*, homology is quite evident for most elements: Sclerite **R3** has the same shape and position and a similar **age**-apodeme (compare fig.28, 29a and 41, 43). The right posterior end of **R3** articulates (**A3** in fig.28, 41) with the sclerotisation adjoining posteriorly (**R1E** or **R1B**). However, only in *Mantoida* the groove-shape of the sclerotisation extends from **R3** (**age**) far beyond **A3** onto the posterior sclerite **R1E** (fig.41, 43). The posterior part of the right phallomere is, like in *Chaeteessa*, composed of a large dorsal lobe (**fda** in fig.44) with a sclerotisations (**R1E** in fig.41, 43). However, in *Mantoida* the sclerotisations in the dorsal **fda**-wall and those on **pia** are interconnected anteriorly by a broad sclerite bridge (**R1E** in fig.41, 44; no membranous stripe **4** as in *Chaeteessa*, fig.28). Behind this bridge there is, like in *Chaeteessa*, a large membranous area (**17** in fig.41). In the ventral wall of the right

phallomere, to the left of A3 and anterior to **pia**, *Mantoida* has likewise a tooth-like evagination (**pva** in fig.41, 28). Its sclerotisation, however, is isolated (**R1D** in fig.41); this is in contrast to both *Chaeteessa* and *Eurycotis* (fig.28, 77, 78) and is assumed to be a derived feature. The edge 16 of *Chaeteessa* and *Eurycotis* (fig.28, 77) has also been lost. The large central invagination **cbe** to the left of the **pva**-tooth resembles **cbe** of *Chaeteessa* (fig.43, 29a).

Taking the homology hypotheses assumed so far as a basis, the muscles of the right phallomere are rather similar in *Mantoida* and *Eurycotis*, and the assumed homologies of the cuticular elements (1.-11.) are confirmed:

- 12. Some phallomero-sternal muscles insert along the anterior margin of R3 (s2 and s4 in fig.42, 82; homology discussion in 6.9.).
- 13. The s2-insertion on R3 extends to the right as far as to a keel-apodeme on the ageapodeme (3 in fig.41, 42, 44 and 74, 77, 82). (Keel 3 is missing in *Chaeteessa*.)
- Muscle r1 (fig.48, 79) inserts on the right part of R3, immediately to the right of the s2-insertion and the keel 3, and runs to the dorsal wall of the fda-lobe.
- Muscle r2 (fig.49, 80) runs from R3 to the cbe-invagination (compare fig.44, 74 and 49, 80). The right part of the posterior r2-insertion is on the R1-sclerotisation that forms the pva-tooth more posteriorly (fig.50, 80).
- 16. Muscle r3 (fig.49, 50, 80) runs from the right wall of the right phallomere to the left where it inserts mainly in the dorsal wall of the pia-tooth. The rest of the left r3-insertion is on the ventral fda-wall in *Mantoida* (compare fig.49 and 50) but in the ventral pia-wall in *Eurycotis* (compare fig.80 and 82).

These muscles and the keel **3** are assumed to be homologous and to be features of the common ground-plan of Blattaria and Mantodea. Muscle r4 is only present in *Mantoida* (fig.49), the muscles r5 (fig.80) and r6 (fig.79) only in *Eurycotis*.

Furthermore, *Mantoida* and *Eurycotis* have in common that (1) the **age**-apodeme extends as far as to articulation **A3** (fig.41, 44, 74) and that (2) even the sclerotisation posterior to **A3** is groove-shaped (fig.41; **rge** in fig.74, 77). Both is not the case in *Chaeteessa* (fig.28). (1) is assumed to be a feature of the ground-plan of Blattaria and Mantodea: 17. The **age**-apodeme reaches articulation **A3**.

As regards (2), however, the grooves posterior to A3 take different positions relative to the right **r3**-insertion (ventral to **r3** in *Mantoida*, dorsal to **r3** in *Eurycotis*) and are regarded as non-homologous.

Main sclerite **R1** is differently divided in the species discussed so far; the questions arise (1) which of these divisions are homologous and (2) when have these divisions evolved. The separation of the **pva**-sclerotisation (sclerite **R1D**, fig.41) in *Mantoida* is certainly apomorphic (compare above).

Chaeteessa and *Eurycotis* have the dividing lines **4** (fig.28, 32) and **A8** (fig.74) in a similar position. *Sphodromantis* has a dividing line (**4** in fig.6, 14) in the same position as **4** of *Chaeteessa*, which is not membranous but only weaker sclerotised than the sclerites **R1A** and **R1B**. This weak stripe **4** of *Sphodromantis* and the membranous stripe **4** of *Chaeteessa* are assumed to be homologous. Muscle **r3** of *Sphodromantis* (fig.16, 19) has the same

course as r3 of *Mantoida* (fig.49, 50). The right insertion of r3 is posterodorsal to stripe 4 (compare fig.14 and 16, 19). In *Eurycotis*, however, the right insertion of r3 is anteroventral to articulation A8 (compare fig.74 and 80). Thus, homology is highly improbable for the dividing lines 4 and A8. The dividing line 4 is thus missing not only in *Mantoida* but also in *Eurycotis*, and it is not a feature of the common ground-plan of Blattaria and Mantodea but a derived feature of a Mantodean subgroup containing at least *Chaeteessa* and *Sphodromantis*.

The question remains whether the articulations A8 and A9, both missing in Mantodea, could be elements of the common ground-plan of Blattaria and Mantodea. In Eurycotis, A8, A9, and the sclerite bridge between R1G and R1H (behind membrane 17 in fig.77, 78) are assumed to be functionally correlated: The posterior part of the right phallomere - composed of fda and pia - can perform a swinging or flapping movement, with A8 and A9 defining the axis. During this movement the membrane 17 is folded and stretched again, and the sclerite bridge may have the function to stabilise R1G and R1H against each other. Muscle r3 (fig.80) moves the flap posterolaterad; r1 and r6 (and possibly s8 on the tre-tendon; fig.79) pull it anteromediad. In Mantoida and Chaeteessa nothing suggests that such a flap-mechanism has ever been present. Thus, from this functional point of view, A8, A9, and the posterior bridge are probably derived features of Eurycotis (and other Blattaria, see in 6.7.6.). However, this view is debatable: In the copulation of Periplaneta the hla-hook has its function in the "initial seizing" and the flap-mechanism in the "final holding". Since the copulation habits of Mantodea are derived, the flapmechanism could be in the same way completely obliterated as the hla-hook and some correlated elements possibly are on the left side (compare in 6.4.2.). On the other hand, the right phallomeres of *Mantoida* and *Chaeteessa* (fig.28, 41) also seem to have the ability to grasp (mainly by the **pia**- and **pva**-teeth), and the final holding could well have been performed by other structures different from the flap-mechanism in the common groundplan of Blattaria and Mantodea. Thus, it is improbable but cannot be completely excluded that A8, A9, and the posterior bridge are elements of the common ground-plan of Blattaria and Mantodea.

As a result, the articulations A8 and A9 separating the sclerites R1F, R1G, and R1H are probably derived elements of Blattaria. The dividing line between R1E and R1D (*Mantoida*) as well as the dividing line 4 between R1A and R1B (*Chaeteessa* and *Sphodromantis*) are certainly derived features of Mantodean subgroups. If these hypotheses are true,

18. R1 is an undivided sclerite in the common ground-plan of Blattaria and Mantodea.

However, if A8 and A9 should prove to be elements of this ground-plan, R1 would have to be regarded as tripartite – composed of R1F, R1G, and R1H like in *Eurycotis*. (When R1 will subsequently be assumed to be undivided in this ground-plan, this must be seen with these reservations in terms of A8 and A9).

A further difference between *Eurycotis* (and some other Blattaria) and all Mantodea studied is the presence or absence of the **tre**-tendon and muscle **s8** and the different condition of the muscles **b4**.

Muscles connecting dorsal parts of the left complex and of the right phallomere have been termed b4. Mantoida, Eurycotis, and Polyphaga have two such muscles, b4a and b4b (fig.36, 48, 58, 109); Cryptocercus has three, b4a, b4b, and b4c (fig.143a). The Blattarian species have the right insertions of all b4-muscles on the tre-tendon, deeply immersed in the body, and the homology of the b4-group as a whole is rather certain. Mantoida has the right insertions of both b4a and b4b on the left dorsal anterior margin of the fda-lobe (fig.48; the **b4b**-insertion is not shown; it is immediately posterior to the **b4a**-insertion). Since the external origin of tre is at the dorsal anterior margin of fda, the right insertions of the b4-muscles take the same relative position in Blattaria and Mantoida. The left insertions, however, take rather different positions, and some shifts must have taken place: The left b4a-insertion is in Eurycotis and Polyphaga on the utmost right part of the lvepouch (fig.70, 129, 130), in Mantoida on an infolding to the right of the lve-pouch (fig.48, compare fig.46). These positions are quite similar. The left b4b-insertion is in Polyphaga (fig.127) in the anterior right dorsal wall of the left complex, far right-dorsal to the pnepouch; in Eurycotis (fig.70) it is on the top of the pne-pouch; in Mantoida the position is intermediate - dorsal to the pne-pouch, but next to its base (fig.48).

On the basis of these relations, it is in my view acceptable to regard the **b4a** and **b4b** of *Mantoida*, *Eurycotis*, and *Polyphaga* as strictly homologous and to assume homology between these **b4**-muscles as a whole and the **b4**-group of *Cryptocercus*. The immersion of the right insertions (by **tre**) can possibly be regarded as the derived condition and as an autapomorphy of Blattaria or of a Blattarian subgroup. The same might be assumed for the cooperating **s8**-muscle (homology discussion in 6.9.).

- 19. Muscles b4a and b4b are present.
- 20. Muscle s8 and the tre-tendon are probably absent.

For the simple sclerites **R2** and **R3** there is no necessity for defining regions. The complicated main sclerite **R1** will be divided into regions, which mainly (and arbitrarily) correspond to the division into individual sclerites in *Eurycotis*, and which are defined as follows (fig.331e, 332e):

- R1d (dorsal): The sclerotisation homologous with sclerite R1H of *Eurycotis* (fig.74) on the fda-lobe. On R1d there are the posterior insertions of the muscles r1 and r6 (fig.80).
- R1v (ventral): The sclerotisation homologous with sclerite R1G of *Eurycotis* (fig.77, 78) on the posterior part of the pia-tooth. On R1v there is the left insertion of muscle r3 (fig.80, 82).
- R1t (tooth): The sclerotisation homologous with that part of sclerite R1F of *Eurycotis* which extends dorsad from the edge 16 (fig.77, 78) and lies in the right-dorsal wall of the cbe-invagination. Along its posterior margin R1t forms the ridge or tooth pva. On R1t there is the insertion of the right dorsal part of muscle r2 (fig.80, 81).

R1t there is the insertion of the right-dorsal part of muscle r2 (fig.80, 81).

- R1c (central): The sclerotisation homologous with the remainder of sclerite R1F of *Eurycotis*, which adjoins sclerite R3 posteriorly and articulates with it (A3), which extends onto the anterior part of the pia-tooth, and which forms the groove rge along its dorsal margin. R1c is situated centrally between the other R1-regions: The border to R1d is articulation A8; the border to R1v is articulation A9; the border to R1t is edge 16. On R1c there is the right insertion of r3 (fig.80, 82), the anterior insertion of r6 (fig.79; on rge), and the posterior insertion of r5 (fig.80; on rge).

In fig.331c and 332c this regioning of **R1** is transferred to *Chaeteessa* – according to the homology relations assumed above (features 1.-11.). In *Chaeteessa* (and *Sphodromantis*) the **R1c**-region extends far dorsad into the **R1A**-sclerite (fig.331c); this results from the position of the right **r3**-insertion in *Sphodromantis* (compare fig.16 and 331a,c). The regioning of **R1** of *Mantoida* is shown in fig.331d, 332d.

6.7.2. The elements in the common ground-plan of Blattaria and Mantodea

The features 1.-20. in 6.7.1. permit the reconstruction of many ground-plan features of the right phallomere (fig.321f,h): **R3** is a curved plate in the anteriormost ventral wall. The right posterior end of R3 articulates with the R1c-region (A3). At least the right and right anterior margins of R3 form a groove-like age-apodeme, which reaches articulation A3 (but the groove does not exceed A3). R1 is (probably) an undivided sclerite, with all its regions firmly connected. Along edge 16 the regions R1c and R1t are sharply angled to each other. **R1t** forms a posteriad-directed tooth or ridge **pva** at its posterior margin. A large central invagination cbe is situated to the left of or left-ventral to pva. The dorsal lobe **fda** and the ventral tooth **pia** are distinct; they are connected along the right edge of the right phallomere and diverge to the left. Their walls are largely sclerotised by R1 (regions **R1d** and **R1v**). In the posterior right edge of the right phallomere there is a membranous area 17. The parts of R1 in the dorsal wall of fda and those on pia are interconnected anterior to membrane 17. Posterior to membrane 17 there is no dorsoventral connection (like in Mantodea) or, at most, a very narrow one (like in Eurycotis). The muscles r1, r2, r3, s2, and s4 are present. The insertions of s2 and r1 are separated by the keel-apodeme 3. The tre-tendon and the articulations A8 and A9 are probably missing. It is unclear if **R2**, the articulations A6 and A7, and the muscles r4, r5, and r6 are present or not.

6.7.3. Homology relations and character states of the elements in Mantodea

The **R3**-sclerites of *Chaeteessa* (fig.28), *Mantoida* (fig.41), *Metallyticus* (fig.20), and *Sphodromantis* (fig.6) are very similar. The **age**-apodeme is always deeper in its left part, where it is more or less plate-like (this is least distinct in *Chaeteessa*). In *Metallyticus* and *Sphodromantis* this deepening of **age** is very abrupt. Only in *Sphodromantis* this left part of **age** has developed a curvature to the posterior and back to the right (fig.6, 8). Only in *Chaeteessa* the left marginal part of **R3** bends dorsad into the **cbe**-wall (fig.29a, 32). Two other derived features of *Chaeteessa* are that the utmost right-posterior part of **age** and the keel-apodeme **3** have been lost (fig.28). In the other species the groove-like shape of the sclerotisation even exceeds **A3** (distinct in *Mantoida*, fig.41, 43, and *Sphodromantis*, fig.6, 8; less distinct in *Metallyticus*, fig.20, 21); the keel **3** has been retained (fig.6, 13, 20, 23, 41, 44). At least in *Sphodromantis* and *Mantoida* keel **3** separates the insertion areas of **s2** and **r1** (fig.15, 48). The apodeme **are** (fig.6, 8) is a derived feature of *Sphodromantis*.

The **cbe**-invagination becomes in all four species continuously deeper to the left (fig.6, 8, 20, 21, 28, 29a, 41, 43), and a sclerite **R2** is always missing (compare *Eurycotis*, fig.77, 78; but see below: *Metallyticus*).

The posterior part of the right phallomere is in all species composed of a large dorsal lobe **fda** (fig.13, 23, 31, 44) and a smaller, leftward projecting ventral tooth **pia** (fig.6, 20, 28, 41). In *Sphodromantis* **pia** has become very small by a reduction of its posterior part. Left-anterior to **pia** there is always another tooth-like process **pva** (fig.6, 20, 28, 41).

The R1-sclerotisations are very similar in Chaeteessa and Sphodromantis (fig.6, 28): The sclerotisation posterior to articulation A3 is connected with the pva-sclerotisation and with the pia-sclerotisation (sclerites R1B) but more or less separated (by 4 in fig.6, 28) from the dorsal fda-sclerotisation (sclerites R1A). The dividing line 4 is also present in Metallyticus (fig.20), but R1A has expanded far into the right ventral wall and occupies the ventral wall of pia (fig.20, 21). Only Mantoida shows the plesiomorphic state with the dividing line 4 missing. In both Metallyticus and Mantoida the pva-sclerotisation has been separated from the R1-sclerotisation posterior to A3 (sclerites R1D in fig.20, 41). That in Mantoida these elements are really pva and parts of R1 (and not R2) is shown by the posterior insertion of r2, whose right part inserts on that sclerite (fig.49, 50; compare fig.16, 19). (R2 of Eurycotis bears the left part of the r2-insertion; compare feature 15. in 6.7.1.). In Metallyticus, however, R1D adjoins the left posterior end of R3 in a similar way as **R2** in *Eurycotis* (fig.20, 77) and could really be the homologue of **R2**. But regarding the situations in the other Mantodea, it is certainly more probable that the tooth is the true pva and R1D the respective part of R1. A definite decision might come from an investigation of muscle r2.

The membranous area **17** has retained its primitive condition only in *Chaeteessa* and *Mantoida* (fig.28, 41). In *Metallyticus* it has been largely reduced by the expansion of **R1A** onto **pia** (fig.20). In *Sphodromantis* it has lost its boundary to the membranous ventral **fda**-wall by the reduction of the posterior part of the **pia**-tooth and its sclerotisation (fig.6). The regioning of **R1** of *Chaeteessa* (6.7.1.) can be transferred to *Mantoida*, *Metallyticus*, and *Sphodromantis* (fig.331a-d and 332a-d). One minor problem is the exact course of the boundary between the regions **R1c** and **R1t**, since the edge **16** is distinct only in *Chaeteessa* (fig.28). In *Mantoida* and *Metallyticus* sclerite **R1D** is tentatively equated with the **R1t**-region, but it probably does not exactly correspond to **R1t** as defined in *Eurycotis*. The muscles of *Sphodromantis* (fig.15, 16, 19) are very similar to those of *Mantoida* (fig.48, 49, 50), but **r4** is much stouter. Of the muscles connecting the right phallomere and the left complex dorsally (**b4a** and **b4b** in *Mantoida* is questionable since both its insertions are on the left complex. This muscle of *Sphodromantis* could also be homologous with **19** of *Mantoida* (fig.49; compare in 6.5.).

6.7.4. Homology relations and character states of the elements in Blattaria I: The anteroventral elements

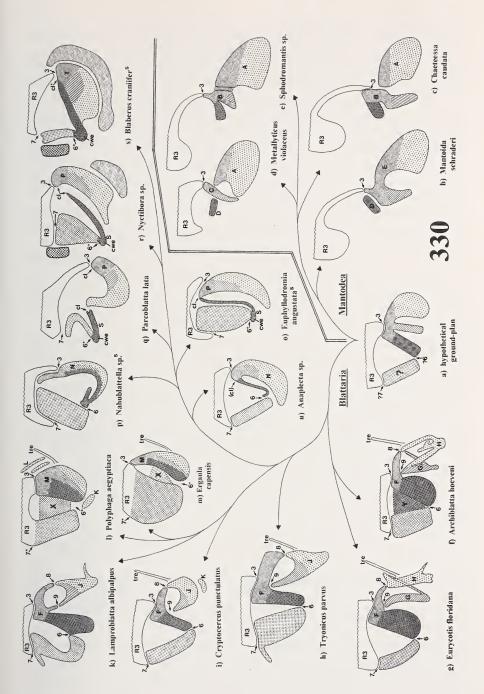
In the following discussions in 6.7.4., 6.7.5., and 6.7.6., data of some species are included whose right phallomeres have been studied only in part: *Archiblatta, Ergaula, Euphyllodromia, Nyctibora, Byrsotria* (fig.330f,m,o,r, 318, 319), and *Supella. Archiblatta* resembles *Eurycotis* (fig.330g); *Ergaula* resembles *Polyphaga* (fig.3301); *Nyctibora* and *Byrsotria* resemble *Blaberus* (fig.330s). For a discussion of the right phallomeres it is

useful to consider first the anteroventral elements (**R2**, **R3**, **cbe**-invagination; 6.7.4.) and the **tre**-tendon (6.7.5.), whose homology relations are quite evident. Then the posterodorsal parts will be discussed (**R1**, **pva**-tooth, **fda**-lobe, **pia**-tooth; 6.7.6.).

The sclerites **R2** and **R3** and the **cbe**-invagination can, as compared with *Eurycotis*, easily be identified in all Blattaria: **R3** (fig.77, 102, 137, 163, 193, 229, 257, 284, 312a) is always a plate in the anteriormost (right-)ventral wall of the right phallomere, and parts of its anterior and lateral margins nearly always form a groove-like age-apodeme. The right and left posterior margins of R3 articulate with more posterior sclerites: A3 (between R3 and R1) is always distinct. A7 (between R3 and R2) is in most cases also a true articulation; sometimes, however, **R2** and **R3** are fused in this place (*Polyphaga, Ergaula*; A7*), or **R2** and **R3** are more distant from each other and no longer articulated (*Parcoblatta*, *Blaberus*; the term A7 is still used to designate the homology of the respective areas). Posterior to the central part of R3 (between A3 and A7 or A7*), the ventral wall of the right phallomere always bends dorsad and (more or less strongly) anteriad to form a central invagination (cbe in fig.78, 104, 138, 164, 195, 230, 258, 285, 313). cbe is variable in its extension and distinctness and is sclerotised to a varied extent. cbe always has its summit in the center of the right phallomere and a descent in the left-ventral direction (missing in Mantodea), at whose base that sclerite adjoins which also approaches R3 in A7 or A7* (R2 in fig.75, 100, 135, 161, 191, 227, 255, 282, 310). R2 has either as a whole the shape of a ridge, often projected into teeth or bulges (fig.75, 76, 135, 136, 161, 162, 191, 192, 227, 228, 281, 285, 287, 310, 311), or **R2** is more plate-like but likewise beset with toothlike evaginations (fig.99, 100, 102, 254-256). All species with the muscles studied have a stout r2 from R3 to the cbe-invagination (fig.80, 81, 140, 141, 166, 167, 197, 198, 231, 232, 259, 260, 286, 287, 314, 316), and some phallomero-sternal muscles insert at the anterior margin of **R3** (fig.82, 142, 168, 199, 233, 261, 288, 317; homology discussion in 6.9.). According to these corresponding relative positions and similarities in shape, homology is assumed for the sclerites **R2** and **R3**, the articulations **A3** and **A7** (or **A7***), the cbe-invaginations, the age-apodemes, and the r2-muscles of all species.

Fig.330: Right phallomere, evolution of main sclerites. – The sclerotisations of the right phallomere are shown. The view is roughly cranial but the phallomeres are unfolded. For correct orientation compare fig.1-319.

Of sclerite R3 (white) only the posterior part is shown (anterior part cut off along undulate line). The other sclerotisations are shown completely and patterned differently. The individual sclerites of R1 are labelled with the capital letters used in the text and in fig.1-319 (e.g. F = R1F). X (part of sclerite R2) and Y (part of region R1t) are sclerotisations occupying the **cbe**-invagination (compare in 6.7.4.). Articulations between sclerites are labelled with the numbers used in the text and in fig.1-319 (e.g. 6 = A6). If an articulation has been lost by fusion of the respective sclerites, the point of fusion is labelled by adding * to the name of the lost articulation (e.g. $6^* = A6^*$). tre and cwe are formative elements. cl represents a certain dividing line between R1-sclerotisations (compare in 6.7.6.). Species with "S" behind their names have side-reversed phallomeres, and a mirror-image of the original preparation is shown. The branching black lines represent the assumed phylogeny. The ground-plan is in some respects unclear (? in fig.330a; discussion in 6.7.1.): Presence of sclerite R2 and of articulations A6 and A7.



The **age**-apodeme is rather variable in its extension: In *Polyphaga* (fig.137), *Ergaula*, *Anaplecta* (fig.229), *Archiblatta*, and *Eurycotis* (fig.77), it is restricted to the right and the right anterior margins of **R3**. In *Cryptocercus* (fig.163) and *Nahublattella* (fig.257) it extends along the whole anterior and lateral margins of **R3**. In *Parcoblatta* (fig.284), *Nyctibora*, *Byrsotria*, and *Blaberus* (fig.312a) it is restricted to the anterior marginal areas of **R3**. Only in *Tryonicus* (fig.102) and *Lamproblatta* (fig.193) **age** has been lost.

R2 is an isolated sclerite in *Eurycotis* (fig.75, 76), *Tryonicus* (fig.100, 101), *Cryptocercus* (fig.161, 162), *Lamproblatta* (fig.190, 191), *Anaplecta* (fig.227, 228), *Nahublattella* (fig.254, 255), and *Supella*. In all these species, the right-ventral end of **R2** articulates distinctly with **R3** (A7), the left-dorsal end of **R2** articulates distinctly with **R1**-sclerotisations in the dorsal wall of the **cbe**-invagination (A6), and **R2** is restricted to the ventral base or, at least, to the ventral wall of the **cbe**-invagination.

In *Euphyllodromia* (fig.3300), *Parcoblatta* (fig.282, 283, 330q), *Nyctibora* (fig.330r), *Blaberus* (fig.310, 311, 330s), and *Byrsotria* **R2** is fused to **R1**-sclerotisations (**R1S** or **R1T**) in the area corresponding to the **A6**-articulation of the other species. The point of fusion is **A6***, with the **cwe**-thickening (fig.282, 310) in its immediate vicinity. This topic will be discussed in 6.7.6. In *Parcoblatta* and *Blaberus* **R2** is in close vicinity to **R3** in the area corresponding to the **A7**-articulation of the other species (**A7** in fig.284, 312a) but is not articulated with **R3**. In *Euphyllodromia* and *Byrsotria* (fig.318) **A7** is a distinct articulation. In *Nyctibora* **A7** is distinct and hinge-like (fig.319).

In Polyphaga (fig.135-137, 330l) and Ergaula (fig.330m) R2 and R3 are clearly identifiable by their shapes (R3 is a broad curved plate, R2 forms a dental ridge) and by their positions relative to each other, to **cbe**, and to the **r2**-insertions (fig.141; compare Cryptocercus, fig.161, 167, and Eurycotis, fig.75, 81; r2 not investigated in Ergaula). However, in both species R2 shows two peculiarities: (1) R2 is fused to R3. (2) R2 has spread over the **cbe**-invagination (sclerotisation \mathbf{X} in fig.330l,m) and is broadly fused to **R1**-sclerotisations in the dorsal **cbe**-wall; hence, the whole of **cbe** is sclerotised (fig.134, 135). (1): In *Polyphaga* the stripe of weaker sclerotisation A7* (fig.135, 137) takes the same position as articulation A7 in other Blattaria and is assumed to be the line of fusion between R2 and R3 (and a vestige of A7). In Ergaula R2 and R3 are fused without any vestige of A7 (no weak line), and the border is not exactly determinable. Moreover, R2 of Ergaula has become so broad that **R3** is for most of its breadth confluent with **R2** (compare fig.330l and m). (2): In Polyphaga the sclerotisation of cbe has a weak line (13 in fig.134, 138) and an adjacent notch within the sclerite margin; these structures are assumed to mark the border between R2 and R1, and the sclerotisation in the ventral wall and on the summit of cbe is assumedly part of R2 (X in fig.3301). R1 is restricted to the posterior dorsal wall of **cbe**. In *Ergaula* **R1** and **R2** are firmly connected (no weak line); the interpretation of the cbe-sclerotisation is done in accordance with Polyphaga (X in fig.330m is part of R2).

Archiblatta also has the whole **cbe**-invagination sclerotised (compare fig.74 and 75 of *Eurycotis*; the sclerotisation concerned is **Y** in fig.330f, compare fig.330g), but, in contrast to *Polyphaga*, the **cbe**-sclerotisation is slightly weaker near the ventral base of the **cbe**-invagination; in the corresponding area of *Eurycotis* **R2** has its dorsal margin and the

membranous ventral wall of **cbe** adjoins (fig.75, 76). Therefore, in *Archiblatta* the sclerotisation of **cbe** is assumed to have developed by an expansion of **R1** (**R1t**-region, fig.330f). The situation in *Eurycotis*, with **R1** occupying a large part of **cbe** (fig.75; compare e.g. fig.99, 160), can be regarded as a primitive stage of such a development. As a result, homology is assumed for the **cbe**-sclerotisations of *Polyphaga* and *Ergaula* (mainly part of **R2**, **X** in fig.330f), but the **cbe**-sclerotisation of *Archiblatta* (mainly part of region **R1t**, **Y** in fig.330f) is not homologous with these. In *Archiblatta* **R2** and the heavier sclerotised dorsal parts of **R1** are still distinctly articulated (**A6**) in the left-dorsal wall of the **cbe**-invagination (like in *Eurycotis*, **A6** in fig.75). In *Polyphaga* and *Ergaula* this articulation is missing (**A6*** in fig.135, 137).

The shape of **R2** is rather variable. Details are shown in the figures. Some peculiar features are: In *Parcoblatta* (fig.285) **R2** is strongly curved. In *Nahublattella* **R2** bears the conspicuous elements **42** and **43** (fig.254). Only in *Tryonicus* and *Lamproblatta* **R2** has an extension posterior to articulation **A7** (**R2m** in fig.102, 91 and 174, 193), which lies in the rightmost part of the **vla**-lobe (left complex), and which has in *Lamproblatta* a close contact with sclerite **L7**.

Only *Blaberus* (fig.311, 312a), *Byrsotria*, and *Nyctibora* (fig.319) have a peculiar sclerite **R5** ventral to **R2** and **A7**. The **R5** of the three species take exactly the same relative position and are certainly homologous.

As regards the muscles, **r9** is specific to *Polyphaga* (fig.141; *Ergaula* not studied), and **r8** is specific to *Cryptocercus* (fig.167). Since the posterior insertions take completely different positions, a homology of **r8** and **r9** is most unlikely.

6.7.5. Homology relations and character states of the elements in Blattaria II: The tre-tendon

A tre-tendon is present in Archiblatta (fig.330f), Eurycotis (fig.74, 330g), Tryonicus (fig.99, 330h), Ergaula (fig.330m), Polyphaga (fig.134, 330l), and Cryptocercus (fig.160, 330i). Homology is ascertained by the similar position of the tre-base in the anterior dorsal wall of the right phallomere, by a muscle from the right half of the sugenital plate (s8), and by two or three muscles from the dorsal part of the left complex (b4-group; fig.79, 139, 165; muscles not studied in Tryonicus). A muscle from tre to R3 is specific to Cryptocercus (r7 in fig.165). In Lamproblatta as well as in Anaplecta, Nahublattella, Parcoblatta, Blaberus, and all other Blattellidae and Blaberidae studied (fig.330n-s) tre, s8, and b4 are missing.

6.7.6. Homology relations and character states of the elements in Blattaria III: The posterodorsal elements

The elements discussed here are those dorsal and posterior to the summit of the **cbe**invagination and posterior to articulation **A3** (compare fig.321f,h). In *Eurycotis* (fig.74-77), for example, this part of the right phallomere is composed of the dorsal wall of **cbe**, the ridge **pva**, the dorsal lobe **fda**, and the ventral tooth **pia**, and it contains the sclerotisations comprised in **R1**: three sclerites **R1F**, **R1G**, and **R1H**. This part of the right phallomere has undergone very complicated evolutionary changes.

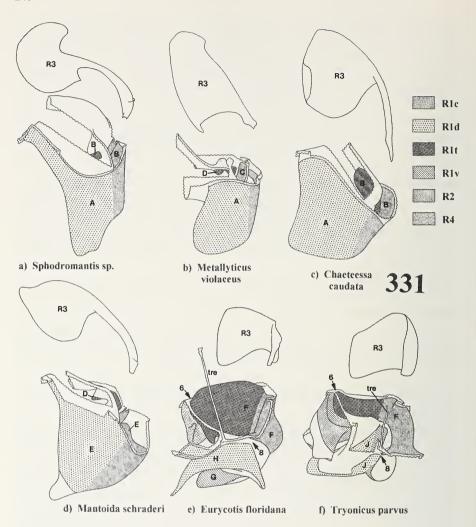
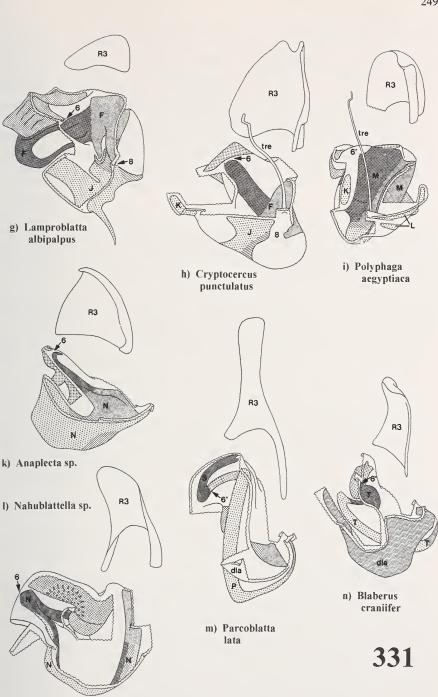


Fig.331: Right phallomere, homology of main sclerites and homologous regions of main sclerite **R1** (dorsal views). – The cuticular elements of the right phallomere are shown, but some membranous parts are removed. Patterned areas are sclerotised, white areas are (except for sclerite **R3**) membranous. Undulate lines are cutting lines. **R3** is separated from the remainder of the right phallomere and shifted anteriad. **R5** of *Blaberus* is not shown. The individual sclerites of **R1** are labelled with the capital letters used in the text and in fig.1-319 (e.g. **F** = **R1F**). Articulations between sclerites are labelled with the numbers used in the text and in fig.1-319 (e.g. **6** = **A6**). Articulations **A3** and **A7** are not labelled (see fig.332). If an articulation has been lost by fusion of the respective sclerites, the point of fusion is labelled by adding * to the name of the lost articulation (e.g. **6*** = **A6***). tre and cwe are formative elements.



Eurycotis and Archiblatta

For *Eurycotis* this area has been fully discussed in 6.7.1.. In *Archiblatta*, like in *Eurycotis* and in all Mantodea, the **fda**-lobe and the **pia**-tooth are both very distinct. Sclerite **R1F** (fig.330f, regions **R1c** and **R1t**) closely resembles **R1F** of *Eurycotis*. The sclerotisations of the **R1d**- and **R1v**-regions are more complicated than in *Eurycotis* (compare fig.330f and g) but similarly structured in a dorsal (**R1H** = **R1d**) and a ventral (**R1G** = **R1v**) sclerite. The sclerite bridge connecting **R1H** and **R1G** in *Eurycotis* (behind membrane **17** in fig.77; fig.330g) has a short gap in *Archiblatta*; instead, there is a ribbon-like connection between **R1H** and **R1G** across the ventral wall of the **fda**-lobe (compare fig.330f and g).

Tryonicus, Cryptocercus, and Lamproblatta

In these species (fig.99-104, 160-164, 190-195) the posterodorsal part of the right phallomere has only two sclerites **R1F** and **R1J**. **R1F** corresponds to **R1F** of *Eurycotis* (fig.74-78). The area posterior to **R1F** contains **R1J**, which is a fusion product of **R1H** and **R1G** of *Eurycotis*, and is an undivided lobe (**fda**, no ventral tooth **pia** present). The regioning of **R1** is shown in fig.331e,f,g,h and 332e,f,g,h.

R1F (fig.102, 163, 193) is in these species, like in *Eurycotis*, somewhat horseshoe-shaped (open to the left, with a dorsal and a ventral arm), and along **R1F** there are the following structures in common, which are all regarded as homologous:

- The ventromedian end of **R1F** articulates with **R2** (A6 in fig.75, 100, 160, 164, 190).
- The ventral arm lies in the dorsal wall of the **cbe**-invagination (fig.74, 99, 160, 190). It bears a ridge (**pva** in fig.80, 99, 164, 190, 196), which is formed by cuticular evagination in *Eurycotis*, *Tryonicus*, and *Lamproblatta*, and by cuticular thickening in *Cryptocercus* (cross-section in fig.164).
- At the base of this ventral arm the posterior margin of R1F articulates with sclerite R1J (A9 in fig.102, 103, 190, 193), or the sclerites are at least in close vicinity (A9 in fig.163, 166). This corresponds to the position of articulation A9 of *Eurycotis* (compare fig.77 and 78). Special features of *Lamproblatta* are the extension 20 of that part of R1F bearing A9 and the immersion of the whole articulation.
- From A9 R1F extends to articulation A3; then it curves into the dorsal wall of the right phallomere.
- At its dorsomedian end R1F has another articulation with R1J (A8 in fig.99, 190) or, at least, closely approaches R1J (A8 in fig.160). This corresponds to the position of articulation A8 of *Eurycotis* (fig.74).
- This dorsal arm of R1F has, like in *Eurycotis*, a sclerotised groove at its dorsal margin, between the articulations A3 and A8 (rge in fig.74, 77, 99, 102, 160, 163, 190, 193).

R1J (fig.99, 102-104, 160-164, 166, 190, 192-195, 197) bears both the articulations **A8** (like **R1H** or region **R1d** in *Eurycotis*) and **A9** (like **R1G** or region **R1v** in *Eurycotis*) and is therefore regarded as a compound sclerite **R1d+R1v** (fig.331f,g,h, 332f,g,h). Thus, in contrast to the situation in Mantodea, *Eurycotis*, and *Archiblatta*, there is now a very broad connection between the **R1d-** and **R1v**-regions posterior to membrane **17**, and this is clearly a derived feature. For *Cryptocercus, Lamproblatta*, and *Eurycotis* these relations

are confirmed by a comparison of the muscles, since **R1J** bears insertions which are in *Eurycotis* either on **R1H** or on **R1G**:

- Muscle r3 of Cryptocercus and Lamproblatta (fig.166, 197) runs from that part of R1F posterior to articulation A3 to the right margin of R1J. It is assumed to be homologous with r3 of Eurycotis (fig.80), which inserts on R1G.
- Muscle r6 of Lamproblatta (fig.196) runs from the rge-groove to the dorsal wall of the right phallomere, like r6 of Eurycotis (fig.79). The left insertion is partly on R1J in Lamproblatta and on R1H in Eurycotis. Such a muscle is missing in Cryptocercus.
- Muscle **r1** of *Cryptocercus* (fig.165) runs from the **age**-apodeme on **R3** to the dorsal wall of the right phallomere, like **r1** of *Eurycotis* (fig.79). The posterior insertion is partly on **R1J** in *Cryptocercus* and on **R1H** in *Eurycotis*. Such a muscle is missing in *Lamproblatta*.

The **fda**-lobe of *Tryonicus* (fig.99, 102-104), *Cryptocercus* (fig.160-164, 166), and *Lamproblatta* (fig.190-195, 197) largely corresponds to **fda** of *Eurycotis*. However, parts of its ventral wall assumedly correspond to the **pia**-walls of *Eurycotis* (after having been leveled). Thus, the **fda**-lobes of these species are not strictly homologous with **fda** of *Eurycotis*. The levelling of **pia** is also a derived feature.

At least *Lamproblatta* has a similar flap-mechanism as *Eurycotis* (with **fda** being the flap and the stout **A8** and **A9** defining the axis of movement). To what extent this is also practised in *Tryonicus* and *Cryptocercus* is questionable since the articulations **A8** and **A9** are by far less distinct.

Polyphaga and Ergaula

In *Polyphaga* the posterodorsal part of the right phallomere contains the large sclerite **R1M** and the smaller sclerites **R1K** and **R1L** (fig.134). The regions **R1c**, **R1t**, **R1d**, and **R1v** can be identified and demarcated by their characteristic features (fig.331i, 332i), but some points remain unclear.

- R1M articulates with R3 (A3 in fig.137) and forms a rge-groove on its dorsal margin (from A3 to the posterior: fig.134, 137, 140), and rge bears the insertion of a stout muscle (r6 in fig.140). These features resemble the R1c-region of *Eurycotis* (fig.331e,i, 332e,i) and the other species. In contrast to the other species, the rge-groove extends much farther posteriad (compare fig.74, 99, 160, 190).
- To the left of A3 R1M bends around an edge (16 in fig.137) to occupy the dorsal wall of the cbe-invagination. The right part of muscle r2, coming from R3, inserts at the anterior margin of this part of R1M (fig.140). More to the left this part of R1M forms a ridge (pva in fig.139, 137, 138). These features resemble the R1t-region of *Eurycotis* (fig.331e,i, 332e,i) and the other species. In contrast to the other species, the pva-ridge is not transversely but longitudinally orientated (compare pva in fig.139 and 80, 99, 197). However, the shape of pva of *Polyphaga* is not so different from pva of *Lamproblatta* (compare fig.139 and 197), if a lengthening of pva along the longitudinal axis and a shortening along the transverse axis is assumed for *Polyphaga*.
- Corresponding to the probable lengthening of pva and rge to the posterior, it is assumed



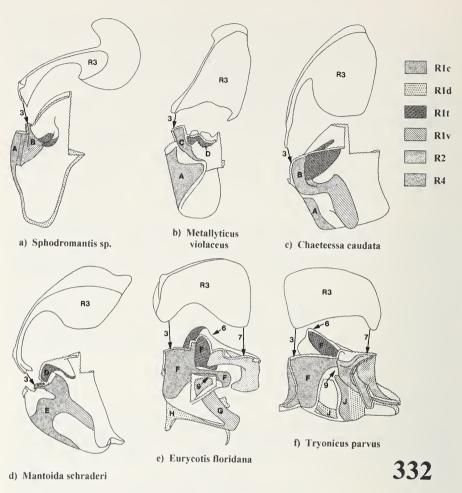
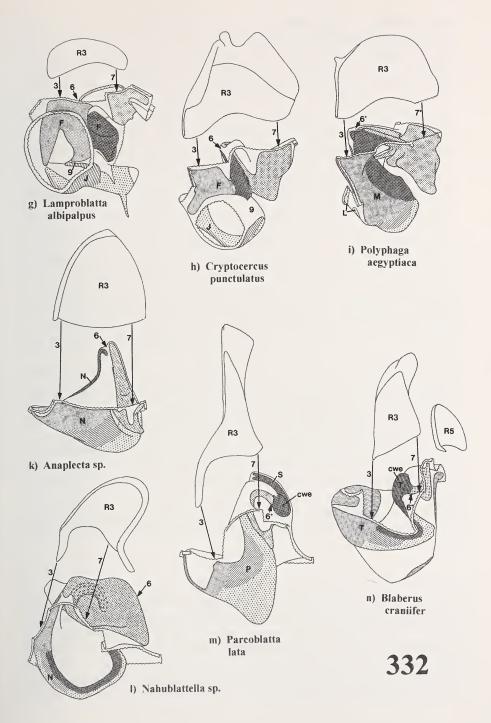


Fig.332: Right phallomere, homology of main sclerites and homologous regions of main sclerite **R1** (ventral views). – The cuticular elements of the right phallomere are shown, but some membranous parts are removed. Patterned areas are sclerotised, white areas are (except for sclerites **R3** and **R5**) membranous. Undulate lines are cutting lines. **R3** and, in *Blaberus*, **R5** are separated from the remainder of the right phallomere and shifted anteriad. The individual sclerites of **R1** are labelled with the capital letters used in the text and in fig.1-319 (e.g. **F** = **R1F**). Articulations between sclerites are labelled with the numbers used in the text and in fig.1-319 (e.g. **6** = **A6**). If an articulation has been lost by fusion of the respective sclerites, the point of fusion is labelled by adding * to the name of the lost articulation (e.g. **6*** = **A6***). The articulation points of **A3** and **A7** are connected by arrows. **cwe** is a formative element.



that in *Polyphaga* the regions **R1t** and **R1c** have considerably expanded posteriad and make up most of **R1M** (fig.331i, 332i).

- Muscle r1 (fig.139) has its anterior insertion on the right margin of R3, like r1 and r5 of *Eurycotis*. The dorsal part of r1 has its posterior insertion in the dorsal wall of the right phallomere (on the R1L-sclerites), like r1 of *Eurycotis* (fig.79). This suggests that the two R1L-sclerites are part of the R1d-region (fig.331i). The ventral part of r1 has its posterior insertion on the rge-groove (fig.134, 139), similar to r5 of *Eurycotis* (fig.80) but more posteriorly. Thus, r1 of *Polyphaga* is certainly homologous with r1 of *Eurycotis* but possibly also includes the homologue of r5. r1 of *Cryptocercus* closely resembles r1 of *Polyphaga*, but a contribution of a r5-part is less probable since no fibers insert on rge or R1F (fig.165).
- Eurycotis (fig.79) and Lamproblatta (fig.196) have the left insertion of r6 in the dorsal wall of the fda-lobe and, by definition, in the R1d-region. r6 of Polyphaga has a very similar course; its left insertion is on sclerite R1K, which is therefore assumed to belong to the R1d-region (fig.331i). Thus, the R1d-sclerotisations of Polyphaga have become rather fragmented (3 sclerites) and far removed from each other (as the insertions of r1 and r6 are). However, R1K and R1L could also be new elements not homologous with sclerotisations of other Blattaria. R1K of Cryptocercus (fig.160) could well be homologous with R1K of Polyphaga. However, since r6 is missing in Cryptocercus, the somewhat similar position of the sclerites is the only indication for homology.
- Further parts of the regions R1d and R1v might be included in the posterior part of R1M. Compared with *Cryptocercus* (fig.160, 163) or *Lamproblatta* (fig.190, 193), this would correspond to a fusion of R1F and R1J across the membrane 17 and the articulations A8 and A9. This is possibly indicated by the complete loss of muscle r3, which in *Cryptocercus* (fig.166) and *Lamproblatta* (fig.196, 197) moves R1F and R1J upon each other: The loss of r3 could be the consequence of such a fusion. In fig.3301, 331i, and 332i R1d and R1v are shown according to this assumption.

The posterior part of the right phallomere is assumed to be composed of the **fda**-lobe and of the **pva**-ridge (fig.136-138). Like in *Lamproblatta, Cryptocercus*, and *Tryonicus*, the ventral tooth **pia** has been lost, and its leveled vestiges are assumed to be contained in the ventral wall of **fda**.

R1M of *Ergaula* is very similar to that of *Polyphaga* but narrower (compare fig.330l and m). **R1L** and **R1K**, however, are missing. The regioning of **R1M** is assumed to be the same as in *Polyphaga*.

Anaplecta

The posterodorsal part of the right phallomere is, like in *Tryonicus, Cryptocercus*, and *Lamproblatta*, an undivided lobe (**fda** in fig.226-230; no **pia**-tooth present). In contrast to these species (with **R1F** and **R1J**), however, there is only one sclerite present (**R1N** in fig.226-230), which somewhat resembles **R1M** of *Polyphaga* (fig.134, 137). The regioning of **R1N** is assumed to be as follows (fig.331k, 332k):

 The part of R1N immediately posterior to articulation A3 (fig.229) is the R1c-region. However, the rge-groove is missing.

- The extension 34 of R1N, which to the left of A3 bends into the cbe-invagination (compare fig.229 and 230; fig.226), exactly corresponds with the R1t-region of the other species by its relative position, by its articulation with R2 (A6 in fig.226, 227, 230), and by bearing the insertion of the right part of muscle r2 (fig.231) (compare fig.331k and 331f,h). R1t of *Anaplecta* is somewhat thickened to the outside (pva) but does not form a true ridge.
- The posterior main part of R1N takes the same position as the R1J-sclerite in *Tryonicus*, *Cryptocercus*, and *Lamproblatta* and is probably composed of the regions R1d and R1v.

R1N is assumed to have developed by a fusion of the former **R1F** and **R1J** across the membrane **17** and the articulations **A8** and **A9** (compare fig.160, 163 and 226, 229), as it has also been assumed for **R1M** of *Polyphaga* (compare fig.331i and k, 332i and k). Moreover, like in *Polyphaga*, muscle **r3** has been lost. In contrast to *Polyphaga*, however, the **R1t**-region retains the same degree of independence and the same transverse orientation as it has in e.g. *Eurycotis* and *Tryonicus*, and there are no free sclerites **R1L** and **R1K**. Thus, it is not clear if **R1N** of *Anaplecta* and **R1M** of *Polyphaga* are strictly homologous and if the fusion of the former **R1F** and **R1J** and the loss of **r3** are homologous in the two species. (Therefore the sclerites are given different names).

The muscle connecting **R3** and **R1N** (**r1** in fig.231) could be homologous with **r1** or **r5** or both muscles of *Eurycotis* (fig.79, 80) and with the **r1** of *Polyphaga* (fig.139) and the other species. Like in *Polyphaga*, the muscle will be named **r1** in *Anaplecta* (and in the other Blattellidae and Blaberidae discussed below).

Nahublattella

The posterodorsal part of the right phallomere is, like in *Anaplecta*, an undivided lobe (fda in fig.253, 256: no ventral tooth pia) with one sclerite (R1N'), but the R1t'-sclerotisation seems to be missing (compare 34 in fig.226). However, similar to the left end of 34 in *Anaplecta*, the left end of R1N' (34 in fig.253) articulates with R2' (A6 in fig.254, 255, 226) and curves back to the right like a hook. Therefore, the R1t'-region is assumed to have fused to the main part of R1N' lying posterior to it (fig.3311). Apart from this difference, R1N' of *Nahublattella* is regioned in the same way as R1N of *Anaplecta* (compare fig.331k and l, 332k and l). A peculiar feature of *Nahublattella* is the hinge-like shape of articulation A3 (fig.253, 257). Muscle r1 is certainly homologous with r1 of *Anaplecta* (fig.231, 259). Muscle r10 (fig.259) is specific to *Nahublattella*.

Supella

The posterodorsal part of the right phallomere is again an undivided lobe **fda** with one large sclerite **R1N'**, but **R1N'** has expanded over the whole **fda**-lobe and over the whole dorsal wall of the **cbe**-invagination. The **R1t'**-region must have been firmly integrated into this sclerotisation. Articulation **A6**, indicating the left end of the **R1t'**-region, is distinct and, like in *Nahublattella*, on the summit of **cbe** (compare fig.253, 254). A hook-like or curved sclerotisation near **A6**, however, is not present. *Supella* resembles *Nahublattella* in probably having **R1t'** completely integrated into **R1N'**, but because of the large expansion of **R1N'** in *Supella* the situations in the two species are hardly comparable.

Parcoblatta, Blaberus, and other Blattellidae and Blaberidae

The morphology of the posterodorsal part of the right phallomere of *Parcoblatta* and *Blaberus* is in some repects very different from *Anaplecta* and *Nahublattella*. Concerned are two areas, which will be discussed separately: (1) the **R1t**-region and (2) the dorsal lobe **fda**. The essence of the changes having taken place can be understood by considering the morphology of some more blattellid and blaberid species included in this investigation.

The **R1t**-region Within Blattellidae and Blaberidae the **R1t**-region (with **pva**) undergoes some changes which also involve **R2**. These developments are shown in fig.330n-s.

Anaplecta has **R1t** (fig.330n, **34** in fig.226) in the same relative position as e.g. *Tryonicus* (fig 330h): situated in the dorsal wall of **cbe**, connected with **R1c** to the right, articulated with **R2** to the left (**A6**). In contrast to *Tryonicus*, the left end of **R1t** shows the hook-like curvature, which is a derived feature.

Euphyllodromia has a similar ribbon-like sclerotisation in the dorsal wall of **cbe** (fig.3300), which by its position can be identified as the **R1t'**-region. The right end of this **R1t'** approaches, like in *Anaplecta*, the **R1c'**-sclerotisation immediately behind articulation **A3** but is narrowly separated from **R1c'** by membrane (at **cl** in fig.3300). The left end of **R1t'** shows, like in *Anaplecta*, a hook-like curvature, but this curved part is swollen to the interior of the phallomere by extensive thickening of the cuticle (**cwe** in fig.3300). Moreover, the left end of **R1t'** is not articulated with **R2'** but fused to it (at **6*** in fig.3300). Thus, the former sclerite **R1N'** has divided (at **cl**) into two new sclerites: **R1S'** (**R1t'**-region, now firmly connected with **R2'**) and **R1P'** (rest of the former **R1N'**). The separation of **R1t'** from **R1c'**, its fusion to **R2'**, and the **cwe**-thickening are derived features.

Nyctibora shows the same situation (fig.330r), but **R1t** (sclerite **R1S**) and the rest of **R1** (sclerite **R1P**) are slightly farther removed from each other. (i.e. the two points of division, called **cl** again, are farther away from each other). The **cwe**-thickening and its curvature are very distinct (fig.319).

In *Parcoblatta, Blaberus*, and *Byrsotria* the fusion of **R1t** and **R2** and the **cwe**-thickening are very similar to *Nyctibora* (fig.282, 283, 285 and 309, 310, 313), and **cwe** marks the border between **R2** and **R1t** (with **cwe** belonging to **R1t**). However, the condition of the right end of **R1t** varies: In *Parcoblatta* (fig.330q, 281, 282) this end of the **R1t**-region (sclerite **R1S**) is still free. It has been far removed from its previous point of contact with **R1c** (sclerite **R1P**) (or, in other words, **R1t** has been shortened; compare the **cl**-points in fig.330q and r). Instead, it has approached the opposite end of sclerite **R1P**.

In *Blaberus* (fig.330s, 309) and *Byrsotria* (fig.318) the **R1t'**-region is firmly connected with the rest of **R1'**. From the phylogenetic context, discussed later in 7.3., it follows that this is due to a secondary fusion of the sclerites **R1S** and **R1P** and does not correspond to the primary connection of these sclerotisations within the **R1N**-sclerite of *Anaplecta, Nahublattella*, and *Supella* (fig.226, 253, 330n,o,p). Therefore, the resulting sclerite, though having the same composition as **R1N**, is named differently: **R1T'**. (The sclerotisations contained within **R1N** and **R1T'** are homologous throughout but the

sclerites themselves are not). Whether the fusion of **R1S** and **R1P** to form **R1T'** had as its starting point a similar situation as in *Nyctibora*, or if it was preceded by a shortening of **R1S** like in *Parcoblatta*, is unclear. (In the regioning of **R1T'** in fig.330s the former situation has been assumed, compare fig.330r).

The **R1t**-morphology of all these species also shows that the complete incorporation of **R1t'** into sclerite **R1N'** in *Nahublattella* and, in a different way, in *Supella* is in both cases a special derivation, and that the situations in *Euphyllodromia, Parcoblatta, Nyctibora*, and *Blaberus* are derived from a situation similar to *Anaplecta* (fig.330n), with **R1t** connected with **R1c** only at its right end.

The **pva**-ridge on **R1t** is very low in *Euphyllodromia, Parcoblatta* (fig.282), and *Nyctibora* (fig.319) and has been completely lost in *Blaberus* and *Byrsotria*.

The situation in *Nahublattella* could be interpreted in another way: That part of **R1N'** which near **A6** curves back to the right (right part of **34** in fig.253) could alone be the **R1t'**-region, which is shortened like in *Parcoblatta* and, by this, far away from **R1c'** with its right end. According to this (improbable) interpretation, the **R1'**-morphology of *Nahublattella* would be likewise much more primitive than in *Euphyllodromia, Parcoblatta*, *Nyctibora*, and *Blaberus*: There would be no fusion between **R1t'** and **R2'**, and **cwe** would be missing. Instead, some features would have to be regarded as derived peculiarities of *Nahublattella*: a fusion between the left end of **R1t'** and the left end of the posterior **R1N'**-sclerotisation (next to articulation **A6**); a reduction of the hook-curvature at the left end of **R1t'** (in the same area); an extreme shortening of **R1t'** (which in any case would be a parallelism as compared with *Parcoblatta*). In my view, the interpretation of **R1N'** of *Nahublattella* made above is by far more probable.

The dorsal lobe **fda** In *Parcoblatta* (fig.280, 281), *Nyctibora* (fig.319), *Byrsotria* (fig.318), and *Blaberus* (fig.308, 309) the posterodorsal part of the right phallomere is not an undivided lobe as in *Anaplecta* (**fda** in fig.226), *Nahublattella* (**fda** in fig.253), *Supella*, and *Euphyllodromia*, but it is, from posteriorly, divided into two lobes lying one above the other: **dla** (dorsally) and **fda** (ventrally).

Sclerite **R1P** of *Parcoblatta* resembles **R1N** of *Anaplecta*: Both sclerites articulate with **R3** (A3 in fig.226, 229, 281, 284), have a similar shape, and largely occupy the walls of a posterior lobe (fda in fig.226, 281). Homology is assumed for **R1P** and **R1N** – minus the **R1t**-region of **R1N** (compare fig.330n and q). Consequently, the ventral lobe fda of *Parcoblatta* is assumedly the homologue of fda of *Anaplecta*. Apart from r2 (fig.286), the right phallomere of both *Parcoblatta* and *Anaplecta* has only one further muscle (r1 in fig.231, 286), which has the same course and is assumed to be homologous. The posterior insertion of r1 is in the anteriormost dorsal wall of fda in *Anaplecta* but in the anteriormost dorsal wall of dla in *Parcoblatta*. Thus, it can be assumed that the dla-lobe is a new outfolding originating from the anterior dorsal wall of the formerly undivided fda. Hence, fda of *Parcoblatta* is not strictly homologous with the fda of *Anaplecta* and the other species. (Moreover, like in e.g. *Anaplecta*, the ventral fda-wall of *Parcoblatta* probably still contains the leveled vestiges of the **pia**-walls. Thus, the homology between

the **fda** of *Parcoblatta* and the **fda** of e.g. *Eurycotis*, which has a well-developed **pia**tooth, is not strict in even two respects).

In *Nyctibora* (fig.319) sclerite **R1P** and the **fda**- and **dla**-lobes take the same relative positions as in *Parcoblatta* (fig.280, 281). Additionally, however, there is a sclerite in the dorsal wall of **dla** (**R4** in fig.319, 330r). In *Byrsotria* (fig.318) the situation is essentially the same as in *Nyctibora*, but sclerite **R4'** is in two respects more derived: (1) It is expanded to the right and in contact with sclerite **R1T'** (**59** in fig.318). (2) The left end of **R4'** (**60** in fig.318) bends around the left edge of **dla** into its left ventral wall. In *Blaberus* (fig.308) the situation is like in *Byrsotria*, but **R4'** is even further expanded to the right and curves into the ventral wall of the right phallomere (**59** in fig.308, 309) where it closely approaches articulation **A3**. Moreover, the **dla**-lobe is extremely enlarged as compared with the **fda**-lobe (compare fig.308 and 318). In *Nyctibora, Byrsotria*, and *Blaberus* (fig.314) the posterior insertion of **r1** is, like in *Parcoblatta*, in the anteriormost dorsal **dla**-wall, but it is also on sclerite **R4**. (According to the insertion of **r1**, **R4** would have to be classified, by definition, as a **R1d**-sclerotisation. **R4** is preferred).

The right phallomere of *Blaberus* has, in contrast to *Parcoblatta* and *Anaplecta*, not only the plesiomorphic muscles **r1** and **r2**, but also some further, certainly new muscles. Two of them run from **R4'** to the right ventral wall of **fda** (**r11a** and **r11b** in fig.314). One muscle having the same course is also present in *Nyctibora*. The other muscles (**r12-r18** in fig.314-317) have been found only in *Blaberus* (*Byrsotria* not studied); however, no specimen of *Blaberus* had all these muscles.

6.8. The muscles connecting the left complex and the right phallomere

The **b4**-muscles have been discussed in 6.7.1., 6.7.3., and 6.7.5., muscle **b3** of *Sphodromantis* in 6.5.

b2-muscles are present in *Sphodromantis* (fig.15), *Mantoida* (fig.49), *Polyphaga* (fig.110, 127, 141), *Lamproblatta* (fig.184, 198), and *Anaplecta* (fig.224, 232). The position of the right insertion is quite similar in all species: on the left part of **R3** in *Sphodromantis* and *Mantoida*, on the left margin of **R3** in *Polyphaga*, next to the left margin of **R3** and **R2** in *Anaplecta*, and next to the left margin of **R2** in *Lamproblatta*. That the **b2** of *Lamproblatta* and *Polyphaga* are homologous despite the somewhat different position of the right insertion is strongly suggested by the very similar position of the left insertion: in the right dorsal wall of the left complex, next to **L8** and to the insertions of **I9** and **I12**. In *Anaplecta*, and *Anaplecta*, and in the left anterior ventral wall of **vla** in *Sphodromantis*. The **b2** are certainly homologous in *Polyphaga* and *Lamproblatta* on the one hand and in *Mantoida* and *Sphodromantis* on the other. Whether the **b2** of these two groupings and of *Anaplecta* are homologous and whether **b2** is a muscle of the common ground-plan of Blattaria and Mantodea or a derived element of these groupings is questionable.

b1-muscles are only present in Mantodea: *Sphodromantis* (fig.15, 16), *Mantoida* (fig.43). Muscle **b5** of *Nahublattella* (fig.235, 259, 261) and the muscles **b6** (fig.294, 316) and **b7** (fig.317) of *Blaberus* are certainly apomorphic muscles of these species.

6.9. The phallomero-sternal muscles

Phallomero-sternal muscles inserted on the right half of the subgenital plate have even numbers, those inserted on the left half have odd numbers. The insertions on the subgenital plate are designated as ventral, those on the phallomere complex or on the walls of the genital pouch as dorsal. The anterior margin of the subgenital plate is the whole margin between the right and left contacts between the subgenital plate and the paratergites of segment 9 (compare fig.333a). Fig.333 gives an overview and a homology hypothesis for the phallomero-sternal musculature of the species studied.

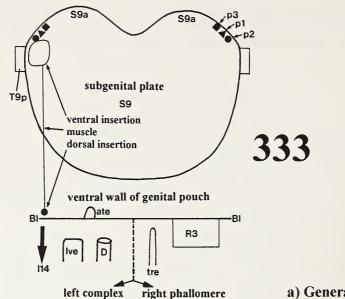
Mantoida has 6 phallomero-sternal muscles **s1-s6**, whose ventral and dorsal insertions are arranged almost symmetrically (fig.40, 333b). In my view, this is the most primitive situation within the species studied, and it is taken as the basis for the terminology. **s1** and **s2**, **s3** and **s4**, as well as **s5** and **s6** are assumed to be pairs of primitively symmetrical muscles. Thus, three pairs of **primary muscles** will be distinguished, which in the primitive situation have the following **basic arrangement**:

- s1 and s2 insert on the median anterior margin of the subgenital plate and run to the ventral basal line of the left complex or right phallomere, where they insert more laterally.
- s3 and s4 insert on the median anterior part of the subgenital plate, closely behind s1 and s2, and run to the ventral basal line of the left complex or right phallomere where they insert more medially than s1 and s2.
- s5 and s6 insert on the lateral anterior margin of the subgenital plate and run to the lateral or lateroventral walls of the genital pouch.

Within this basic arrangement, the dorsal insertion of s^2 and s^4 is on the anterior margin of sclerite **R3**, that of s^1 is on anteriormost parts of **L4**-sclerotisations (anterior **L4**-region), and that of s^3 is on a membranous part of the basal line.

The evolution of the phallomero-sternal musculature comprises two kinds of development: (1) The morphology of the 6 **primary muscles** changes by subdivision, fusion, shift, or loss. The exact homology relations are often difficult to determine. (2) New **secondary muscles** develop, each with a characteristic course by which the homology relations can in most cases easily be determined. These secondary muscles are:

- s7 inserts on the median anterior part of the left side of the subgenital plate and runs to the anterior part of the lve-pouch.
- s8 inserts on the median anterior part of the right side of the subgenital plate and runs to the top of the tre-tendon.
- s10 inserts on the median anterior part of the right side of the subgenital plate and runs to the ejaculatory duct or to an area near its opening.
- s12 inserts on the median (or more lateral) anterior part of the right side of the subgenital



a) General scheme

Fig.333: Phallomero-sternal muscles, homology relations. – **333a** gives an overview of the mode of representation and the symbols and abbreviations used. **333b-1** (following pages) show the morphology of the various species.

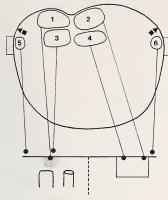
The anterior half of each figure shows the subgenital plate, with

- ventral insertions of muscles p1, p2, and p3 (\blacktriangle , \bigcirc , and \blacksquare).
- ventral insertions of all phallomero-sternal muscles.
- paratergites of abdominal segment 9 T9p.

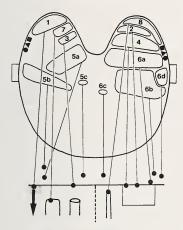
The posterior half of each figure shows the phallomere complex, with

- ventral basal line BI = border between ventral + lateral walls of phallomere complex (below BI) and ventral + lateral walls of genital pouch (above BI).
- border between left complex and right phallomere = median vertical broken line.
- muscle **l14** (to hook **hla**; base of arrow = anterior insertion area).
- pouch lve (with L2-sclerotisations).
- ejaculatory duct D.
- tendon ate.
- tendon tre.
- sclerite R3.
- dorsal insertions of all phallomero-sternal muscles (\bullet) .
- if the dorsal insertion area of a muscle on or near the basal line of the left complex is sclerotised this is shown by a stippled field around the insertion.

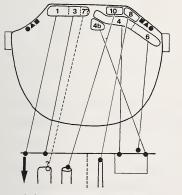
The shape of the subgenital plate is generalised, only the shape of the anterior margin (with apophyses **S9a**) corresponds with the special condition in the respective species. The numbers in the ventral insertion areas of the phallomero-sternal muscles are the numbers used in the text and in fig.1-319 to designate the muscles (e.g. 2 = s2, 5a = s5a). Species with "S" behind their names have the phallomeres and the subgenital plate side-reversed, and a mirror-image of the original preparation is shown. Dorsal views. Further information in 6.9..



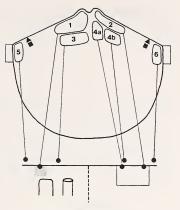
b) Mantoida schraderi



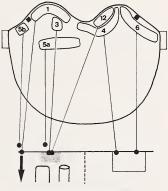
d) Eurycotis floridana



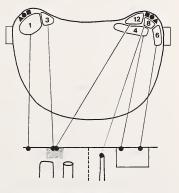
f) Cryptocercus punctulatus



c) Sphodromantis sp.



e) Lamproblatta albipalpus



g) Polyphaga aegyptiaca

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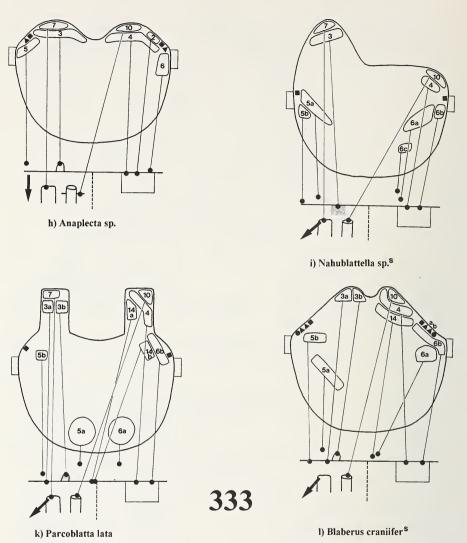


plate and runs to the ventral basal line of the left complex where it inserts immediately to the right of s3.

- s14 inserts on the median anterior part of the right side of the subgenital plate and runs to the ventral wall of the genital pouch beneath the rightmost part of the left complex.

Mantoida and Sphodromantis

Mantoida (fig.40, 42, 333b) conforms with the basic arrangement, except that the dorsal insertion of s3 is posterior (not median) to that of s1. *Sphodromantis* (fig.5, 7, 333c) is also close to the basic arrangement, but the right side has four muscles instead of three.

The arrangement of the insertion areas on the subgenital plate (compare s4 in fig.40 and s4a and s4b in fig.5) and on the phallomere complex (s4a and s4b insert medially like s4 in *Mantoida*, compare fig.7 and 42) suggests that this is due to a division of s4. Both species lack secondary muscles.

Eurycotis

s1, s2, s3, and s4 conform with the basic arrangement (fig.62, 64, 333d), but s2 is rather weak. s5 and s6 have divided into three (s5a,b,c) or four (s6a,b,c,d) bundles, whose insertions occupy a large area in the anterior half of the subgenital plate. However, most of these bundles are very diffuse and indistinctly bordered to each other. Moreover, it is not clear if really all these muscles are derivatives of s5 and s6 or if some of them are new. Of the secondary muscles s7 and s8 (fig.58) are present.

Anaplecta

The left side has three muscles (fig.204, 207, 333h): The secondary s7 (fig.200) runs, like in *Eurycotis*, to the **lve**-pouch. s5 can be identified by the lateral position of its ventral insertion and by its dorsal insertion on the lateroventral wall of the genital-pouch. The third muscle (named s3) might be s1 or s3. It has its ventral insertion posterior to s7 like s3 of *Eurycotis* (s1 of *Eurycotis* inserts anterior to s7, compare fig.333d and h) and has its dorsal insertion immediately anterior to l6a (fig.222) like s3 in *Eurycotis* (fig.70, 73), *Polyphaga*, and *Lamproblatta* (discussion below; fig.133, 188). This muscle of *Anaplecta* is therefore regarded as s3; s1 is missing.

On the right side all three primary muscles are present (s2, s4, s6 in fig.204, 207, 333h). Their ventral insertions resemble the basic arrangement (s2 and s4 on the median anterior margin; s2 anterior to s4; s6 far laterally), but the s4-insertion is very broad, and s2 is strongly reduced. The insertions on R3 are side by side: s4 on the left, s2 in a small central area, s6 on the right. Thus, in contrast to the basic arrangement, the dorsal insertion of s6 has expanded to the anterior right margin of R3 (but still occupies parts of the membranous ventral wall of the genital-pouch as well). The same is true of s6a of *Eurycotis* (fig.64, 333d), whose assignment to s6 is thus confirmed by the morphology of s6 of *Anaplecta*. Noticeably, just s1 and s2 have been reduced – two muscles which probably are a pair.

Apart from s7 there is another secondary muscle: s10, which, however, is not directly inserted on the ejaculatory duct but more ventrally on the infolding between the lobes vla and vfa (fig.221, 222; in fig.333h this infolding is symbolised by a transverse line beneath the ejaculatory duct; compare discussion in 6.2.4.).

Lamproblatta and Polyphaga

On the left side, s1 and s3 of *Lamproblatta* (fig.173, 175, 333e) conform with the basic arrangement, but the dorsal insertion of s1 is rather far on the left (compare *Eurycotis*, fig.70). In *Polyphaga* (fig.113, 116, 333g) both insertions of s3 and the dorsal insertion of s1 exactly correspond with *Lamproblatta* (s1: membrane anterior to the hla-hook, fig.127, 185), but the ventral insertion of s1 has shifted leftward, too, and is not anterior to but to the left of s3. Homology can certainly be assumed for s1 as well as for s3. The

leftward shift of the dorsal **s1**-insertion, and that of the ventral **s1**-insertion in *Polyphaga*, might be correlated with a parallel shift of the **hla**-base from the left ventral wall (fig.65) to the left edge of the left complex (fig.117, 177). *Lamproblatta* has a lateral muscle (**s5b**) showing the **s5**-insertions of the basic arrangement. Whether another muscle having a much more median position (**s5a**) is a true part of **s5** or a new muscle is unclear. In *Polyphaga* **s5** is completely missing – possibly in consequence of the leftward shift of **s1**.

On the right side, the s4 of *Polyphaga* and *Lamproblatta* conform with the basic arrangement and are easily identified. To the right of s4 both species have another muscle (named s6) running to the anterior margin of R3, which might be s2 or s6. Its ventral insertion is lateral (not posterior) to s4 (fig.113, 173) and extends far laterad. This suggests it to be s6. Its dorsal insertion on the right anterior margin of R3 only, not on the genital pouch, however, might suggest it to be s2. But since the dorsal insertion of s6 of *Anaplecta* and *Eurycotis* has expanded to the anterior margin of R3, and since s2 is small in *Eurycotis* and very small in *Anaplecta*, these muscles of *Lamproblatta* and *Polyphaga* are interpreted as s6; s2 is assumed to have been lost. The restriction of the dorsal s6-insertion to the anterior margin of R3 (no longer in the wall of the genital pouch) would be a derived feature of *Polyphaga* and *Lamproblatta*. (If the muscle should be s2, the laterad expansion of its ventral insertion would be a derived feature).

Of the secondary muscles **s12** is present (fig.133, 188): The dorsal and ventral insertions are exactly the same in both species, and, in addition, the dorsal insertion is surrounded in the same way by **s3**, **l5**, and **l6a**. **s12** is peculiar to *Lamproblatta* and *Polyphaga* (and *Ergaula*) and is assumedly a synapomorphy of these species. **s8** is present in *Polyphaga* (and *Ergaula*) but missing in *Lamproblatta*.

Cryptocercus

The left side (fig.147, 149, 333f) has only one muscle **s1+3**, with its ventral insertion along the median (not the lateral) anterior margin of the subgenital plate and its dorsal insertion only on and near the basal line (not in the more posterior walls of the genital pouch, fig.158). By this extension of the insertion areas **s5** is unlikely to contribute to this muscle and is, like in *Polyphaga*, assumed to have been lost. **s1+3** seems to be composed of the previous **s1** and **s3** since its dorsal insertion includes the areas occupied by **s1** and **s3** in *Polyphaga* and *Lamproblatta* (**s1**: anterior to the **hla**-base, compare fig.157 and 127, 185; **s3**: farther on the right, compare fig.158 and 133, 188). The dorsal insertion of **s1+3** reaches with some fibers sclerite **L2** in the **lve**-pouch; this insertion is typical for **s7**, which may therefore also contribute to **s1+3** (fig.333f).

The right side has one large primary muscle, s2+4+6, and, in some specimens only, a very small one, s4b. The ventral insertion area of the large muscle includes, in contrast to s1+3 on the left side, the lateral anterior margin of the subgenital plate (fig.147, 333f). Thus, this muscle assumedly contains not only the median s2 and s4 but also the lateral s6. This is also suggested by the fact that the dorsal insertion (fig.168) extends far into the ventral wall of the genital pouch (compare s6a of *Eurycotis*, fig.64). In this latter feature *Cryptocercus* would be more primitive than *Polyphaga* and *Lamproblatta*, whose s6-insertion is restricted to the anterior margin of R3. To what extent each s2 and s4 contribute

to the large muscle is unclear; s2 might also be missing like in *Lamproblatta* and *Polyphaga* (as shown in fig.333f). The small s4b can, according to its ventral insertion posterior to the large muscle (fig.147), only be a split off part of s4 or a new muscle but certainly not s2.

It is difficult to interpret this highly peculiar condition of the primary muscles. At least, the outgroup comparison between the other Blattaria and *Mantoida* strongly suggests that it is not primitive for Dictyoptera. Possibly, extensive fusions of muscles have taken place. However, there is still another possible explanation: It could be due to a retention of a nymph-like situation in adult morphology (a neotenic trait), with the differentiation of the single primary muscles not yet completed. This question could possibly be settled by an investigation of the ontogeny of the phallomero-sternal muscles in other Blattaria.

Two secondary muscles are present: s8 to the tre-tendon and s10 to the ejaculatory duct near its opening.

Nahublattella

Nahublattella (fig.237, 240, 333i) closely resembles *Anaplecta*, but the ventral insertions of the median (on the apophyses **S9a**) and of the lateral (more posteriorly on the subgenital plate) muscles are extremely far away from each other.

The secondary muscles **s7** and **s10** are easily identified by their insertions (fig.249). **s3** can, like in *Anaplecta*, be identified by its ventral insertion posterior to **s7** (compare fig.333h and i). **s1** is missing. The muscle from the right apophysis **S9a** to the anterior margin of **R3** is probably **s4**, not **s2**. (Since in *Anaplecta* **s2** is reduced, a loss of **s2** seems for *Nahublattella* more probable than a loss of **s4**).

The lateral primary muscles **s5** and **s6** have undergone a division (or new muscles have been added). The dorsal as well as the ventral insertion areas of **s5a** and **s5b** together have the same extension as those of **s5** of *Anaplecta* (compare fig.204 and 237, fig.207 and 240), and a division can readily be assumed. The same correspondence is found for the ventral insertion areas of **s6a** and **s6b** or **s6**, respectively (compare fig.204 and 237). Dorsally **s6b** of *Nahublattella* inserts on the anterior margin of **R3** like **s6** of *Anaplecta*; the dorsal insertion of **s6a**, however, is completely different (compare fig.207 and 240). Thus, for **s6a** the derivation from **s6** is not certain. The question is the same for the very delicate muscles **s6c** (fig.237, 240).

Parcoblatta and Blaberus

The phallomero-sternal musculature (fig.265, 267, 333k and 296, 298, 333l) can be derived from *Nahublattella* but also shows some differences.

In *Parcoblatta*, the secondary muscles s7 and s10 as well as the primary muscle s4 conform with *Nahublattella*. s3 also inserts like in *Nahublattella* but is divided into two bundles s3a and s3b (compare fig.237, 240 and 265, 267). Dorsally s3b inserts on the ate-tendon, s3a to the left of ate (fig.267). s1 and – with the same reservations as in *Nahublattella* – s2 are missing. Concerning these muscles, the situation in *Blaberus* is the same except that s7 is missing. (However, s7 is present in its typical position in *Nauphoeta*, another member of Blaberidae; fig.328i).

The lateral primary muscles of the left side (s5) are in *Blaberus* divided into three bundles (s5a – 2 bundles – and s5b in fig.296, 298). s5a and s5b resemble s5a and s5b of *Nahublattella* in their dorsal insertions: s5b near the anterior margin of L4U' (fig.250, 304); s5a in the left ventral wall of the genital pouch (fig.240, 298). The ventral insertions are similar, too, but in *Blaberus* s5a has shifted posteriad (fig.237, 296). The division of s5 into s5a and s5b is assumedly homologous in *Nahublattella* and *Blaberus*. The smaller median muscle of *Blaberus*, also named s5a, is regarded as a median subdivision of s5a. In *Parcoblatta* s5 shows a similar and certainly homologous division: The ventral and dorsal insertions of s5b are similar to *Blaberus* (fig.265, 267, 296, 298), but the dorsal one is somewhat more posteriorly. The ventral and dorsal insertions of s5a are, as compared with *Blaberus*, by far more posteriorly.

The lateral primary muscles of the right side (s6) are in *Blaberus* present as two bundles (s6a and s6b in fig.296, 298), which are certainly homologous with s6a and s6b of *Nahublattella*: The dorsal insertions of s6a and s6b take the same positions as in *Nahublattella* (fig.240, 298), but s6b has considerably expanded posteriad and now occupies the whole right margin of **R3'**. As regards the ventral insertions, s6a has, as compared with s6b, shifted far posteriad (like s5a on the left side!). s6 of *Parcoblatta* shows the same division: The dorsal and ventral insertions of s6b are situated like in *Blaberus* (fig.265, 296), and the dorsal insertion likewise occupies the whole right margin of **R3**. The dorsal and ventral insertions of s6a are, as compared with *Blaberus*, shifted even farther posteriad (like those of s5a on the left side!).

Only *Parcoblatta* and *Blaberus* have the secondary muscle **s14**, which is divided into two bundles in *Parcoblatta* (**s14a,b**). The ventral insertion is closely behind **s4** (fig.265, 296, 333k,l). The dorsal insertion is immediately anterior to **s6a** in *Blaberus*, but, corresponding to the posteriad shift of **s6a**, far anterior to **s6a** in *Parcoblatta*.

6.10. The subgenital plate and associated structures

The subgenital plate is poor in complex structures, and hardly any character is valuable for the phylogenetic analysis in the frame of this study.

In most species the subgenital plate is asymmetrical, but the degree of asymmetry varies, and various parts are concerned: mainly the styli **S9s** and the dorsal sclerotisation **S9d** in *Anaplecta* (fig.204); the posterior edge of the plate in *Polyphaga* (fig.113, 114); mainly **S9d** in *Sphodromantis* (fig.5), *Lamproblatta* (fig.173), *Parcoblatta* (fig.265), and *Blaberus* (fig.296); **S9d** and the apophyses **S9a** in *Nahublattella* (fig.237, 238); the apophyses **S9a** and the lateral and posterior edges in *Mantoida* (fig.40); the whole subgenital plate in *Metallyticus* (fig.22). Only in *Chaeteessa* (fig.30), *Eurycotis* (fig.62), *Tryonicus* (fig.86), and *Cryptocercus* (fig.147) the subgenital plate is symmetrical or nearly so.

The apophyses **S9a** can be very different in their length and distinctness (compare *Chaeteessa*, fig.30, and *Parcoblatta*, fig.265). *Cryptocercus* has no apophyses at all (fig.147). As explained in 3.1., the areas designated as apophyses **S9d** are not in all species homologous in a strict sense, but short apophyses may be homologous with only the anterior parts of long apophyses. Such relations are obvious if *Parcoblatta* (fig.265) is

compared with *Blaberus* (fig.296): In *Parcoblatta* the recess between the apophyses extends far beyond the insertions of **s14a**,**b** and nearly reaches the level of the **p3**-insertions. In *Blaberus* the recess does not even reach **s14**, much less **p3**. Thus, either the recess has deepened in *Parcoblatta*, or the posterior parts of the apophyses have fused with each other in *Blaberus*. To compare more distantly related species in this regard, however, is hardly possible.

The area designated as the dorsal sclerotisation **S9d** is rather variable and is certainly not strictly homologous in all species. In *Metallyticus* (fig.22), *Eurycotis* (fig.62), *Tryonicus* (fig.86), *Polyphaga* (fig.114), *Lamproblatta* (fig.173), *Anaplecta* (fig.204), *Parcoblatta* (fig.265), and *Blaberus* (fig.296) **S9d** is restricted to the more posterior part or even to the margins of the dorsal wall of the subgenital plate (= posterior ventral wall of genital pouch), and it is firmly connected with the ventral sclerotisation around the lateral and posterior edges of the plate. In *Sphodromantis* (fig.5) and *Cryptocercus* (fig.147) **S9d** is also continuous with the ventral sclerotisation but extends by far more to the anterior. In *Mantoida* (fig.40), *Chaeteessa* (fig.30), and *Nahublattella* (fig.237, 238) **S9d** also extends far anteriad but is isolated from the ventral sclerotisation. In *Nahublattella* **S9d** is highly elaborated (division, bristles, muscles **133** and **134**; fig.238, 240).

In several species, some patterns in the sclerotisation of the subgenital plate remind one of its presumable composition (true sternite, two coxites; compare in 3.1.) and might therefore be regarded as primitive. In *Eurycotis* (fig.62), *Polyphaga* (fig.113), *Cryptocercus* (fig.147), and *Nahublattella* (fig.237) an anterior (sternite?) and a posterior (transversely fused coxites?) sclerotisation are separated by a membranous field – except for a lengthwise connection of the lateralmost parts. In *Parcoblatta* (fig.265), *Blaberus* (fig.296), and *Lamproblatta* (fig.173; the anterior sclerotisation is very narrow) the field is no longer membranous but still distinctly weaker sclerotised than the other parts. In *Tryonicus* (fig.86) the sclerotisation is weaker in the anterior third, but there is no heavier sclerotisation along the anterior margin. In *Mantoida* such a zoning of the plate is only slightly indicated (not shown in fig.40). In the other Mantodea (fig.5, 22, 30) and in *Anaplecta* (fig.204) the sclerotisation of the subgenital plate is uniform. An interesting feature of *Cryptocercus* (fig.147) is that in the posterior part of the plate the lateral areas are distinctly heavier sclerotised than the median area; the transverse fusion of the coxites is probably not complete.

6.11. The peripheral muscles

The muscles **p1**, **p2**, and **p3** are, if present, always inserted close to each other on the anterior margin of the subgenital plate, between the median and the lateral primary phallomero-sternal muscles (fig.333b-1; fig.5, 40, 62, 113, 147, 173, 204, 237, 265, 296). The **p1**-muscles are ventral muscles of segment 9. Their posterior insertions are on or close to the **Pv**-sclerites or, if separate **Pv**-sclerites are absent, on the anterior margin of the paraprocts **Pp**. The **p1** are rather weak (*Sphodromantis*, fig.1; *Mantoida*, fig.36; *Polyphaga*, fig.109) or even consist of very few fibers only (*Eurycotis*, fig.58; *Anaplecta*,

fig.200; *Blaberus*, fig.293), or they are completely missing (*Lamproblatta, Nahublattella, Parcoblatta*). In *Polyphaga, Anaplecta*, and some specimens of *Eurycotis* the **p1** are divided into two bundles on one or on both sides.

In *Blaberus* the **p1** are also divided into two bundles on each side; the one bundle shows the usual insertion on the anterior margin of the paraproct, but the other bundle inserts on its posterior margin (fig.293), and this is certainly a derived feature. According to McKittrick (1964), the posterior insertions of the ventral muscles of segment 9 take the same positions in the females of most Blaberidae, too.

In *Cryptocercus* the **p1** (fig.143a) are extremely broad. This is unlikely to be a primitive state since the **p1** are by far narrower in all other Blattaria as well as in *Mantoida* and *Sphodromantis*. In last-instar nymphs of *Blaberus* and *Eurycotis* the **p1** are by far broader than in the respective adults (though not as broad as in *Cryptocercus*). Hence, the broadness of **p1** of *Cryptocercus* might be a neotenic trait.

The **p2**-muscles are dorsoventral muscles of segment 9. They are either very delicate (*Eurycotis*, fig.58; *Polyphaga*, fig.109; *Cryptocercus*, fig.143a; *Blaberus*, fig.293) or completely missing (*Mantoida, Sphodromantis, Lamproblatta, Anaplecta, Nahublattella, Parcoblatta*).

A special feature of *Eurycotis* is that the **p2** as well as the serially homologous muscles of abdominal segment 8 pass through two pairs of eyelets in the vasa deferentia (as shown in fig.58: **Vd**, **p2**, and **p2(8)**). Snodgrass (1937) finds such eyelets, in the same arrangement, also in *Blatta orientalis* (Blattinae); Pipa (1988), fig.7, describes eyelets for *Periplaneta americana*. I have additionally investigated the vasa deferentia of *Deropeltis*, *Periplaneta, Parcoblatta*, and *Blaberus*: There are no traces of eyelets in *Blaberus*. Eyelets or vestiges of them have been found in *Periplaneta, Deropeltis*, and *Parcoblatta*, but either the passage is more or less narrowed, or there is only a thickening of the vas deferens without any passage. The degree of eyelet reduction can be rather different in the four places (often asymmetrical; this was also the case in some specimens of *Eurycotis*) and in different specimens of a species. If passages were present in these species, these were never passed through by muscles (though very thin **p2** were often present). Pipa (1988), however, finds the **p2** passing through the eyelets in *Periplaneta* (**S-9** in his fig.7).

In last-instar nymphs of *Eurycotis* and *Blaberus* p2 and p2(8) are by far stronger than in the adults of the same species, and they all run through eyelets in the vasa deferentia. The eyelets and their penetration by p2 and p2(8) are assumed to be nymphal features, which in the adults can be retained to rather various extents (even within a single species). The same seems to be true of the muscles themselves. A far-reaching retention of these structures in the adult is thus regarded as a neotenic trait.

The **p3**-muscles (rectal muscles) are present in all species and have a similar fan-shape throughout. In *Cryptocercus* they are divided into two fans on each side.

The **p4**-muscles have their anterior insertions always far laterally on the anterior margin of tergite 9 **T9**. In many species they additionally extend onto the paratergites **T9p** (*Mantoida*, fig.36, 37; *Cryptocercus*, fig.143a; *Lamproblatta*, fig.169, 170; *Anaplecta*, fig.200, 201; *Parcoblatta*, fig.262, 263; *Blaberus*, fig.293, 294). In the latter case, except

in *Cryptocercus*, the **p4** are divided into several bundles: throughout their length in *Mantoida* and *Parcoblatta*; only anteriorly in *Lamproblatta*, *Anaplecta*, and *Blaberus*.

The posterior insertions take rather different positions: on the lateral anterior margin of tergite 10 **T10** (*Cryptocercus*, fig.143a,b), or on the anterior margin of the paratergites 10 **T10p** (*Sphodromantis*, fig.1; *Mantoida*, fig.36, 37; *Parcoblatta*, fig.262, 263), or in the membrane median to **T10p** (*Anaplecta*, fig.200); in this latter case they can be far anteriorly (*Lamproblatta*, fig.169, 170; *Polyphaga*, fig.109; *Blaberus*, fig.294) or extremely far medially (left **p4** of *Eurycotis*, fig.58; *Nahublattella*, fig.255).

In *Cryptocercus* the insertion on tergite 9 and that on tergite 10 take the same relative position (compare fig.143a and b), and **p4** is clearly a dorsal muscle of segment 9. The **p4** of the other species are assumed to be the same dorsal muscles, and the posterior insertion is assumed to have undergone a ventromediad and anteriad shift which is variously pronounced in the different species. The homology of these **p4** is suggested by the constant position of the anterior insertion and by the following fact: In last-instar nymphs of *Eurycotis* and *Blaberus* the posterior **p4**-insertion is by far more laterally than in the adults; that means, it shifts mediad during late ontogeny. In the various species, the final position of the posterior **p4**-insertion in the adult might depend on the extent to which the adult character state prevails against the nymphal state. A dorsolateral position (like in *Cryptocercus*, fig.143b) is probably a neotenic trait. However, in some species this could also be a primitive feature.

In *Periplaneta americana*, whose posterior **p4**-insertions have a similar ventromedian position as in *Eurycotis* (compare fig.58), the innervation of **p4** is known (Pipa 1988): It is accomplished by a nerve-branch (the common base of **4A1c** and **4A1d** in Pipa) which innervates, apart from **p4** (**359**, **360** in Pipa), the various groups of dorsal muscles (**M** and **MDM9** in Pipa). This is consistent with the assumption that even those **p4** having their posterior insertions far medially are true, though modified, dorsal muscles.

The **p5**-muscles are dorsoventral muscles of segment 10. Dorsally they always insert on the lateral anterior margin of tergite 10 **T10**. Their ventral insertions are on or near the **Pv**-sclerites (*Eurycotis*, fig.58; *Lamproblatta*, fig.169; *Anaplecta*, fig.200; *Nahublattella*, fig.235) or, if separate **Pv**-sclerites are missing, on the anterior margin of the paraprocts **Pp** (*Sphodromantis*, fig.1; *Polyphaga*, fig.109; *Cryptocercus*, fig.143a; *Parcoblatta*, fig.262; *Blaberus*, fig.293). Hence, the position of the **p5**-insertions (**Pv**-sclerites or anterior margin of paraprocts **Pp**) differs in the same way as in the **p1**-muscles. The insertions of **p1** and **p5** suggest that in those species without separate **Pv**-sclerites the **Pv**sclerotisations have become incorporated into the anterior part of the paraprocts. (This part of the paraprocts is then labelled **Pv** in the figures). Moreover, the insertions of **p1** and **p5** (ventral muscles of segment 9, dorsoventral muscles of segment 10) suggest that the **Pv**-sclerites (or the **Pv**-parts of the paraprocts) are sternal sclerotisations of abdominal segment 10. However, this question cannot be finally settled here.

The **p6**-muscles are dorsoventral muscles of segment 9. The dorsal insertion is always far laterally on tergite 9 **T9**. The ventral insertion is either close to the line of contact between the lateral margin of the subgenital plate and the paratergite of segment 9 **T9p** (*Eurycotis*, fig.69; *Cryptocercus*, fig.146; *Lamproblatta*, fig.172; *Anaplecta*, fig.203; left muscle of

Polyphaga, fig.112a) or slightly posterior to this area (*Mantoida*, fig.39; *Sphodromantis*, fig.4). In *Polyphaga* the ventral insertion has expanded into the lateral wall of the genital pouch (left muscle, fig.112a) or has completely shifted to this area (right muscle, fig.112b). In *Nahublattella* (fig.237), *Parcoblatta* (fig.265), and *Blaberus* (fig.296) the **p6** insert distinctly more medially on the subgenital plate.

The **p7**-muscles have their anterior insertions far medially in the membrane anterior to paraprocts and **Pv**-sclerotisations; their posterior insertions are far laterally where the paratergites 10 **T10p** meet the paraprocts **Pp** (articulations **A99**; lateral to the posterior **p1**-insertions). **p7** is well-developed in *Mantoida* (fig.37), *Sphodromantis* (fig.2), *Lamproblatta* (fig.170), and *Cryptocercus* (fig.144). In *Lamproblatta* the posterior (or lateral) insertion of the left **p7** has distinctly shifted anteriad. In *Eurycotis* **p7** is represented by only very few fibers (fig.59). In the other species no **p7** have been found.

The muscles **p8** and **p9** will not be discussed: Their homology relations are uncertain since they are not clearly distinguishable from other muscles of the anal region. The muscles **p10** of *Cryptocercus* (fig.144) are probably subdivisions of the **p5**-muscles.

The muscles **p1-p7** are certainly present in the common ground-plan of Blattaria and Mantodea. **p3** and **p5** are very uniform in the species studied. The differences in the morphology of **p1**, **p2**, and **p4** are assumed to be of limited value for a phylogenetic analysis, because these differences probably depend on the extent to which nymphal features are retained in the adult. As regards **p6**, the mediad shift of the ventral insertions could be a synapomorphy of the species concerned.

6.12. The terminal part of the abdomen

The homology relations of most elements of this area are quite evident and need no discussion. The homologies concerning the supraanal lobe **spl**, the epiproct **Ep**, and the tergite 10 **T10** are discussed in 3.1. The homology between the **Pv**-sclerites and the anterior part of the paraprocts **Pp** (in species without separate **Pv**-sclerites) is discussed in 6.11.. There are hardly any features valuable for a phylogenetic analysis, but the following features are worth mentioning and might gain some more value in future investigations including more species.

The area where the paraproct **Pp**, the **Pv**-sclerite, and the paratergite 10 **T10p** meet each other shows in several species some peculiarities. However, the ground-plan condition of this area is in most respects uncertain since Mantodea seem to have this area highly modified – similar to but certainly independently of certain Blattaria – and since the outgroup comparison with other Ectognatha suffers from the uncertainty of homology relations. Hence, the plesiomorphic or apomorphic nature of these peculiarities is debatable. A rather primitive condition might be assumed to be represented in e.g. *Eurycotis* (fig.59): The lateral tip of **Pp** articulates (**A99**) with the ventromedian tip of **T10p** laterally and is in close vicinity to the lateral end of a completely free **Pv** anteriorly. If the **Pv**-sclerites really represent the medially divided sternite 10 (compare in 6.11.), their complete isolation could be plesiomorphic. *Tryonicus* (fig.83), whose **Pv**-sclerites are

fused to the paraprocts laterally, would then have a more derived state of this character. In the species having no separate **Pv**-sclerites by fusion to **Pp** (e.g. *Mantoida*, fig.37, *Polyphaga*, fig.110) this character would be even more derived. The fusion of the paraprocts **Pp** and the paratergites **T10p** in *Lamproblatta* (partial; fig.170), *Anaplecta, Mantoida*, and *Sphodromantis* (complete; fig.201, 37, 2), corresponding to a partial (*Lamproblatta*) or complete loss of **A99**, is certainly a derived feature. The presence of two articulations per side is peculiar to *Lamproblatta* (**A97** and **A99** in fig.169, 170). The interpretation in this species is done in accordance with *Eurycotis* (fig.59) and *Tryonicus* (fig.83): The lateral articulation is the true **A99**; **A97** is assumed to be a new articulation within the paraproct **Pp**.

In several species tergite 10 **T10** has undergone a complete longitudinal division by a median stripe of membrane (*Polyphaga*, fig.109; *Nahublattella*, fig.234; *Blaberus*, fig.293) – certainly a case of threefold parallel evolution. The membranous area **21** of *Lamproblatta* (fig.169) might represent an early stage of such a division.

The articulation **A98** between the cercal base and tergite 10 has been lost only in *Polyphaga* and *Blaberus* – certainly another case of parallel evolution.

The ventral sclerotisation of tergite 10 **T10v** is only in *Anaplecta* separated from the dorsal main part of **T10** (fig.200). However, the **T10v**-sclerites of *Anaplecta* could also be homologous with the **Cc**-sclerites of the other species (compare fig.200 and e.g. 58).

The various paired sclerites median to the cercal base (Ca, Cb, Cc) are certainly homologous in the way expressed by the designations. All three pairs are present only in (some) Blattaria but not in *Mantoida* and *Sphodromantis* (*Chaeteessa* and *Metallyticus* not investigated). Sclerites median to the cercal base are also present in e.g. Caelifera (Snodgrass 1935, fig.7), but whether there is any kind of homology with the Blattarian sclerites is unknown. It is therefore also unclear if some or all of these sclerites are elements of the Dictyopteran ground-plan or derived features of Blattarian subgroups or of Blattaria as a whole. **Ca**-sclerites are present in *Eurycotis* (fig.58, 59), *Tryonicus* (fig.83, 84), *Lamproblatta* (fig.169, 170), *Anaplecta* (fig.200), *Nahublattella* (fig.234, 235), and *Parcoblatta* (fig.262, 263), and they are crescent-shaped in most species. Except in *Tryonicus* and *Lamproblatta* (fig.58, 59), *Tryonicus* (fig.83, 84), *Lamproblatta* (fig.169, 170), and possibly *Anaplecta* (fig.200: **T10v**?). **Cb**-sclerites are peculiar to *Lamproblatta* (fig.169, 170), and *Blaberus* all three pairs are missing, but in *Polyphaga* and *Blaberus* at least the **Ca**-bulges are distinct.

A distinct supraanal lobe **spl** has been found in Mantodea (fig.1, 36) and in *Eurycotis, Tryonicus, Cryptocercus, Lamproblatta, Parcoblatta*, and *Blaberus* (fig.58, 83, 143a, 169, 262, 293). An epiproct **Ep** is present in Mantodea (fig.1, 36) but never in Blattaria.

6.13. The asymmetry of the phallomere complex

The right phallomeres of the Mantodean species, especially *Chaeteessa*, and of *Eurycotis* are very similar in the arrangement of the sclerotisations (**R1**, **R2**), the formative elements (invagination cbe, lobe fda, tooth pia, ridge pva, apodeme age), the main muscles (r1,

r2, **r3**, **s2**, **s4**), and some morphological details (keel **3**, edge **16**). The right phallomeres of all other Blattaria studied can be derived from that of *Eurycotis* without any problems; especially the area comprising sclerites **R2** and **R3**, invagination **cbe**, and muscle **r2** is very similar in all species. Therefore, homology is assumed for all these right phallomeres. This assumption also includes those species with the right phallomere situated on the left side (*Nahublattella, Supella, Euphyllodromia, Byrsotria*, and *Blaberus* investigated in this paper): The right phallomeres of these species can be integrated into the homology hypothesis without any problems, and the right phallomeres of *Blaberus* and *Byrsotria* (situated on the left side) and the right phallomere of *Nyctibora* (situated on the right side) are nearly identical. (The only principal difference is the fusion of **R1P** and **R1S** to form **R1T** in the two blaberid species).

The left complexes of *Mantoida* and of *Archiblatta* and *Eurycotis* are guite similar in the principal arrangement of the sclerotisations (L1, L2, L4), the formative elements (e.g. pouches lve and pne, ventral lobe vla, apodeme swe, processes paa and pda), the main muscles (12, 13, 14, 16, 19, s1, s3), the genital opening, and some morphological details (L4d-region). Most of the morphological gaps between these species are bridged by other Blattaria, e.g. Tryonicus (shape of **paa** and **pda** and relation between them), Polyphaga (shape of sclerites L1 and L2, position of phallomere-gland opening), or Cryptocercus (muscle 11). The left complexes of the other Blattaria (e.g. Parcoblatta) can be extremely different from those of Archiblatta and Eurycotis, but the morphology of each species can, if several other species are included in the comparison, be traced back to the basic pattern. Therefore, homology is assumed for all these left complexes. This assumption likewise includes those species with the left complex situated on the right side (Nahublattella, Supella, Euphyllodromia, Blaptica, Nauphoeta, and Blaberus investigated in this paper): The left complexes of these species can be integrated into the homology hypothesis without any problems. The left complex of Blaberus (situated on the right side) and the left complex of *Parcoblatta* (situated on the left side) are very similar; concerned are the principal arrangement and shape of most cuticular elements, the course of most muscles, as well as many details (ate-tendon, hge-groove, notch 45). Differences between Blaberus and Parcoblatta are in most cases bridged by other species of Blattellidae and Blaberidae: Loboptera and Nyctibora (orientation as in Parcoblatta) have, like Blaberus, a sclerite L4U, which is missing in *Parcoblatta*. Nauphoeta (orientation as in Blaberus) has, like Parcoblatta, a muscle s7, which is missing in Blaberus. The lve-apodeme and the viaprocess of Nyctibora (orientation as in Parcoblatta) and Nauphoeta (orientation as in Blaberus) are very similar and do not show the strong differences as present between Parcoblatta and Blaberus (which are due to the differently directed rotation of this area).

From the homology of the right phallomeres and from that of the left complexes it follows that the asymmetry of the whole phallomere complex is homologous in all species studied. Thus, the asymmetry of the phallomere complex is a feature of the common ground-plan of Blattaria and Mantodea (and maybe Isoptera). Moreover, from a comparison of the ground-plan morphologies of the left complex (fig.321e,g) and of the right phallomere (fig.321f,h) it follows that the asymmetry of the phallomere complex was in the common ground-plan of Blattaria and Mantodea already as extreme and of the same very special kind as in the extant species. For the investigated members of Plectopterinae

(*Nahublattella, Euphyllodromia, Supella*) and Blaberidae (*Nauphoeta, Byrsotria, Blaptica, Blaberus*) it is thus evident that the phallomere complex has undergone a change of its left-right-asymmetry (like a mirror-image): This hypothesis first proposed by Bohn (1987) is strongly supported, and many new arguments are now available. According to e.g. Bohn (1987), the phallomere complex of most species of *Ectobius* (Ectobiinae) is normally orientated, but some species show the same orientation as Blaberidae. The latter species, as compared with the former, clearly show that a reversal of the left-right-asymmetry must be regarded as a possible evolutionary pathway.

Vestiges of a bilateral symmetry or side-homologies within the phallomere complex are revealed in only very few respects: (1) The primary phallomero-sternal muscles s1 and s2 are assumed to be side-homologous (e.g. fig.37), and side-homology might also be assumed for the areas of their dorsal insertions. In the primitive case (compare in 6.9.) these are the anterior L4I-region on the left side and the anterior margin of R3 on the right side. These two areas have additionally in common that the margin of the sclerotisation is more or less groove- or beam-like (apodemes swe or age). (2) The transverse phallomere muscles (b-muscles) might be assumed to have primitively a symmetrical course, and their left and right insertion areas might be side-homologous. The situation in Mantoida might be interpreted in this way: The insertions of muscle b1 (fig.43) are next to those of the side-homologous s3 (left side) and s4 (right side), and the resulting side-homology would again concern the (median) anterior margins of R3 and L4. (3) The dorsal transverse muscles b4a and b4b (fig.48, 58, 109) have their right insertions close to each other, but the left insertions are quite distant from each other. From their course it can be at most deduced that there is some kind of side-homology between the dorsomedian parts of the left complex and of the right phallomere. As regards the remaining parts of the right phallomere and of the left complex, there are in no species any similarities in the positions, in the special shapes, or in the muscular connections of elements which show similar spatial interrelationships on both sides. Hence, no further side-homologies can be reliably assumed.

The primary phallic lobes of nymphal Blattaria and Mantodea are certainly homologous with those of the other Ectognatha (in a more or less strict sense). In most other Ectognatha (also in the most primitive: Archaeognatha, Zygentoma) the external genitalia developing from these phallic lobes are bilaterally symmetrical. Hence, the male external genitalia have certainly been bilaterally symmetrical in some early members of the common stemgroup of Blattaria and Mantodea. However, from this it cannot be concluded that there must be extensive vestiges of this bilateral symmetry in the sclerotisations, in the formative elements, or in the musculature of the phallomere complex of the extant species (or of the later members of the common stem-group): The sclerotisations, muscles, and formative elements present in the common ground-plan of Blattaria and Mantodea (or more or less extensive parts of them) might have evolved later – at a time when the extreme asymmetry had already established. Nothing is known about homology relations between these Dictyopteran phallomere elements and the elements of the male genitalia in other insect groups, and hence there is no information about which elements have already been present when the Dictyoptera branched off from their (unknown; Kristensen 1995) sister-group.

7. THE GROUND-PLAN AND THE EVOLUTION OF THE PHALLOMERE COMPLEX AND THE PHYLOGENY OF BLATTARIA AND MANTODEA

In the sections of chapter 6 many features of the common ground-plan of Blattaria and Mantodea have been reconstructed. This ground-plan will be given completely in 7.1. In 7.2. and 7.3. the evolution of the phallomere complex will be described for Mantodea and for Blattaria. This will be done in accordance with a phylogenetic hypothesis which results as the most parsimonious solution from the distribution of the phallomere character states analysed in chapter 6. In this description, all derived character states present in the various subgroups will be listed, and these derivations are regarded as autapomorphies of the respective subgroups. For each autapomorphy the section of chapter 6. in which the

the respective subgroups. For each autaponorphy the section of chapter 6. In which the respective feature has been discussed will be given. Most of the phallomere characters are consistent with each other in the distribution of their states over the subgroups defined in 7.2. and 7.3., and this phylogenetic hypothesis is thus highly supported. Some derived character states which appear as autapomorphies of single species in the frame of the sample of species included in this investigation and which are uninformative in the present analysis will also be mentioned, since in later investigations they might be detected in other species, too, and might then serve as synapomorphies and help in integrating further species into this phylogenetic hypothesis.

A survey of all assumed aut/synapomorphies is given in 7.4. – together with a phylogenetic tree (diagram 1) showing the most parsimonious solution.

For some characters the polarity of the states does not become unambiguously clear from the discussions and informations given in chapter 6, and the respective interpretations given in 7.1.-7.4. are not yet sufficiently substantiated. The evolution of these characters and the polarity of their states will be discussed in 7.5. The single topics will be designated with letters and referred to in 7.2. and 7.3.

For some characters there will, despite the previous discussions, remain some doubt in terms of polarity. In some other characters whose polarity is rather clear the distribution of the character states over the species is in some way inconsistent with the phylogenetic hypothesis in 7.2. and 7.3. The respective (possibly or clearly) derived character states and the groupings they would suggest will be listed in 7.6. All these inconsistencies will be also mentioned in 7.2. and 7.3.

In the following discussions, assemblages of species regarded as holophyletic are called "subgroups" and numbered according to their hierarchy. Assemblages not regarded as holophyletic are called "groupings" and are designated with capital letters for cross reference. The character states which are assumed to be autapomorphies of subgroups, and also the character states whose role as possible autapomorphies is discussed but regarded as improbable, are, for easy reference, numbered (bold printed and put in brackets, e.g. **(45)**).

7.1. The common ground-plan of the phallomere complex of Blattaria and Mantodea

Fig.321a-p shows the cuticular elements and the muscles of the phallomere complex and of the other parts of the male postabdomen as they have assumedly been in this ground-plan. For some characters, however, the ground-plan state could not be resolved, since

- the respective elements are (probably) present in the ground-plan of Blattaria but completely absent in Mantodea: presence or absence of hla-hook, nla-process, dcaprocesses, tre-tendon, sclerites L3 and R2, articulations A6, A7, A8, and A9, muscles l14, r6, s7, and s8.
- the respective elements are present in the ground-plan of Mantodea but completely absent in Blattaria: presence or absence of muscles **r4** and **b1**.
- the condition of the respective elements is different in the ground-plan of Mantodea and in that of Blattaria: connection or separation of the L4-sclerotisations L4v/L4c, L4l, and L4n in the anterior ventral wall of the left complex; presence or absence of the curvature (dorsad and back to the left) of the right parts of L2 and lve; connection or separation of the R1-regions R1d and R1v posterior to the membranous area 17.
- the homology relations between Blattaria and Mantodea or within Blattaria are questionable: presence or absence of loa-process, L5-sclerite, L4c-region, muscles 17, 113/b3, and b2.

In fig.321 the elements or properties concerned are omitted or supplied with questionmarks. As regards all the data in 7.1., compare in 6.1.2., 6.2.2., 6.3.2., 6.4.2., 6.5., 6.7.2., 6.8., 6.9., and 7.5.

Cuticular elements

Left complex

L1 is an undivided sclerite in the central dorsal wall of the left complex. A large anterior part of L1 (L1a-region) is situated in a deep and distinct **pne**-pouch and is hood-shaped (but not plateau-like anteriorly). The right posterior part of L1 is a distinct arm-like extension (L1m-region). There is possibly another arm-like extension formed by the left posterior part of L1 (L11-region). L1l and L1m do not join each other ventrally to form a sclerite-ring (no region L1r). The membranous part of the **pne**-wall has a roughly dorsal position and receives the opening of the phallomere-gland **P**. A completely sclerotised **loa**-process is probably present. Whether there are **dca**-processes is not decidable (omitted in fig.321).

The pouch **lve** lies ventral to the **pne**-pouch. **L2** is an undivided arch-shaped sclerite which extends along the edges **7** of the **lve**-pouch and is (almost) completely restricted to its dorsal wall. Whether the right parts of **lve** and **L2** are level or curved dorsad and back to the left is not decidable (curvature omitted in fig.321). The right end of **L2**, or its dorsal left end if the right parts of **L2** are up- and recurved, (**L2m**-region) shows a narrow (not hinge-like) articulation **A2** with **L1m**. The left end of **L2** (**L2p**-region) leaves the **lve**-pouch posteriorly, bends into the dorsal wall, and forms the sclerotisation of the **paa**-process (**L2d**-region). **paa** is completely sclerotised, short, and somewhat upcurved. The

ventral wall of the **lve**-pouch is mostly membranous and is at the same time the left anterior part of the dorsal wall of the **vla**-lobe. The ejaculatory duct **D** opens into the right anterior part of the dorsal **vla**-wall. Both the presence of a small sclerite **L5** in the dorsal **vla**-wall and, if present, its exact position are questionable.

The ventral wall of **vla** is part of the ventral wall of the left complex and is sclerotised by the right posterior part of **L4** (**L4v**-region). The ventral wall of **vla** is for most or all of its length confluent with the remaining ventral wall of the left complex (i.e. only most posteriorly **vla** has a free left edge **61**, or edge **61** is missing). The **lve**-pouch and the **vla**lobe are rather broad but do not reach the left edge of the left complex.

The L4-sclerotisations as a whole form an arch (open posteriorly) in the ventral wall and at the left edge of the left complex. The L4-regions L4l, L4d, L4n, and L4v (and possibly L4c) are present. The ventral wall between these regions is membranous. The morphology of the L4I- and L4d-regions is like in Mantoida and Archiblatta: L4I is undivided and extends over the left anterior margin and the whole left edge of the left complex. In the dorsal wall L4I is restricted to the left margin. The posteriormost part of L4I sclerotises a short bulge-like process **pda**, which takes a position immediately to the left of the **paa**process and whose sclerotisation is connected with the L2d-sclerotisation of paa. The sweapodeme extends over most of the length of L4l. In its anterior part swe is beam-shaped by cuticular thickening, in its posterior part it is groove-like. L4d is distinctly prominent from the outline of L4I and directed to the right (and possibly slightly anteriad). The L4nregion is present; whether it is connected with or separated from the L4I-region cannot be decided. The **nla**-process on **L4n** is possibly also present (according to its functional correlation with 114 and hla; discussion in 7.5. (M), (N)). Whether the L4c-region is present is not decidable. (If it should be present, it is certainly firmly connected with the L4v-region right-posterior to it. At least, there is no separate sclerite L4F). Whether the L4v-region (or the L4c-region, if present) is connected with or separated from the anterior end of the L4I-region is unclear. The hla-hook and its L3-sclerite are probably present. If this is true, the ground-plan condition of hla and L3 can be assumed to correspond with the ground-plan situation in Blattaria (compare in 7.3.).

Right phallomere

Sclerite **R3** lies more or less transversely in the anteriormost ventral wall. At least its right margin and the right part of its anterior margin form a groove- (or somewhat beam-) like apodeme **age**, which reaches the **A3**-articulation. The **age**-groove bears a keel **3**. The right posterior end of **R3** has an articulation **A3** with the **R1c**-region. Posterior to **R3** the ventral wall of the right phallomere curves dorsad and somewhat anteriad to form a large invagination **cbe**.

The posterior part of the right phallomere is composed of a dorsal lobe **fda** and of a ventral tooth **pia**. **fda** and **pia** are confluent along the right edge of the right phallomere and diverge to the left. **R1** is probably an undivided sclerite (or, with less probability, it is divided into three sclerites **R1F**, **R1G**, and **R1H**, corresponding to the regions **R1c+R1t**, **R1v**, and **R1d**, by the articulations **A8** and **A9**). **R1** occupies the area behind the **A3**-articulation (**R1c**-region), part of the right-dorsal wall of the **cbe**-invagination (**R1t**-

region), the dorsal (and possibly part of the ventral) wall of **fda** (**R1d**-region), and the dorsal and ventral walls of **pia** (**R1v**-region). The regions **R1c** and **R1t** form a distinct angle along the edge **16**. Along its posterior margin **R1t** has a posteriad-directed ridge **pva**. At and near the posterior right edge of the right phallomere there is a membranous area **17**. The dorsal and ventral parts of **R1** are (probably) connected anterior to this area but separated (or only narrowly connected) posterior to it.

The presence of sclerite R2 and of its articulations A6 and A7 is questionable. The membranous stripe 4 and probably the articulations A8 and A9 and the tre-tendon are missing.

Musculature

Muscles certainly present in the ground-plan (all shown in fig.3211-p)

11: from the dorsal wall of the pne-pouch to the L4d-region (6.1.1., 6.3.4.).

12: from sclerite L1 in the left wall of the **pne**-pouch to the swe-apodeme on the L4l-region (6.1.1., 6.3.1.).

13: from sclerite L1 in the posterior ventral wall of the **pne**-pouch to the L2a-region in the anterior dorsal wall of the lve-pouch (6.1.1., 6.2.1.).

14: from the L2a-region anteriorly in the lve-pouch to the swe-apodeme on the L4l-region (insertion on swe ventral to 12) (6.2.1., 6.3.1.).

15: from the anterior ventral wall of the left complex to the left(-anterior) edge of the **lve**-pouch (6.2.1.).

l6a: from the anterior ventral wall of the left complex to the right(-anterior) edge of the **lve**-pouch (6.2.1.).

l6b: from the ventral wall of the left complex to the membranous ventral wall of the **lve**pouch at or near the genital opening (ventral insertion posterior to l6a) (6.2.1.).

19: transversely within the dorsal wall of the left complex (6.5.).

b4a, **b4b**: connect the dorsal parts of the left complex and of the right phallomere; left insertions in the right marginal area of the **lve**-pouch (**b4a**) or in the dorsal wall dorsal to the **pne**-pouch (**b4b**); right insertions on the dorsal anterior margin of the **fda**-lobe (on the **tre**-tendon, if it is present in the ground-plan) (6.7.1.).

r1: from the right margin of sclerite R3 (to the right of keel 3) to the R1d-region in the dorsal wall of the fda-lobe (6.7.1).

r2: from sclerite **R3** to the **cbe**-invagination and the **R1t**-region (and to sclerite **R2**, if it is present in the ground-plan) (6.7.1.).

r3: from the R1c-region to the R1v-region in the dorsal wall of the pia-tooth (6.7.1.).

s1: from the left median anterior margin of the subgenital plate to the lateral ventral basal line of the left complex (on the L4l-region) (6.9.).

s2: from the right median anterior margin of the subgenital plate to the lateral ventral basal line of the right phallomere (on sclerite R3, to the left of keel 3) (6.9.).

s3: from the left median anterior part of the subgenital plate (posterior to s1) to the median ventral basal line of the left complex (on membrane) (6.9.).

s4: from the right median anterior part of the subgenital plate (posterior to s2) to the median ventral basal line of the right phallomere (on sclerite R3) (6.9.).

s5: from the left anterior margin of the subgenital plate to the left(-ventral) wall of the genital-pouch (6.9.).

s6: from the right anterior margin of the subgenital plate to the right(-ventral) wall of the genital-pouch (6.9.).

Muscles possibly present in the ground-plan (except for 114 omitted from fig.3211-p)

17: longitudinally within the posterior left ventral wall of the left complex (6.5.).

113, **b3**: from the ejaculatory duct **D** to the dorsal wall of the **vla**-lobe posterior to or to the right of the genital opening (6.5.).

114: from the L4n-region on or near the nla-process to sclerite L3 and the hla-hook (6.4.2.).

r4: from the R1d-region in the left dorsal wall of the fda-lobe to the left ventral wall of the fda-lobe (6.7.1., 6.7.3.).

r6: from the R1c-region to the R1d-region (6.7.1., 6.7.6.).

b1: from the left margin of sclerite **R3** to the anteriormost ventral wall of the left complex (6.8.).

b2: from the left margin of sclerite **R3** to an area next to the right end of the **lve**-pouch (6.8.).

s7: from the left anterior margin of the subgenital plate to the lve-pouch (6.2.4., 6.9.).

s8: from the right anterior margin of the subgenital plate to the tre-tendon (6.7.1., 6.9.).

Asymmetry

From the features of the ground-plans of the left complex and the right phallomere it follows that the very special kind of extreme asymmetry – corresponding to the extreme differences between these two ground-plans – is also a feature of the ground-plan.

7.2. The evolution of the phallomere complex and the phylogeny in Mantodea (= subgroup 1.)

The ground-plan of the phallomere complex of Mantodea

The features listed subsequently can be ascribed to the ground-plan of Mantodea since they are true either of all investigated species (cuticular elements) or at least of *Sphodromantis* and *Mantoida* (muscles), which are representatives of the two basal sistergroups (subgroups 1.1. and 1.2., see below). For all these features it is not clear whether they belong to the common ground-plan of Blattaria and Mantodea or whether they are autapomorphic for Mantodea.

The following elements are absent: sclerotisations L3, R2; formative elements dcaprocesses, hla-hook, nla-process, tre-tendon; articulations A6, A7, A8, A9; muscles 114, r6, s7, s8. The L4-sclerotisations L4v/L4c, L4l, and L4n are firmly connected with each other in the anterior ventral wall. The right parts of L2 and lve are level. The loa-process is present. The membranous part of the **pne**-wall is right-dorsal. The regions **R1d** and **R1v** are not connected with each other posterior to the membranous area 17. The muscles r4 and b1 are present.

Subgroup 1.1.: Mantoida

The phallomere complex of *Mantoida* is rather close to the Mantodean ground-plan but also has some derived features: On the left complex, the L4n-region has been lost (6.3.1., 6.3.3.). On the right phallomere, the **R1t**-region (with the **pva**-tooth) has been separated from the **R1c**-region (6.7.1., 6.7.3.; like in *Metallyticus*: compare (**G**) in 7.5. and grouping B (123) in 7.6.).

Subgroup 1.2.: Chaeteessa + (Metallyticus + Sphodromantis)

There are some conspicuous autapomorphies on the left complex: The ventral wall has developed the L4b-region which occupies all the interspaces between the primary L4regions L4v/L4c, L4n, and L4l and makes the ventral wall completely, or nearly so, sclerotised (6.3.3.; in the primitive case L4b is distinctly weaker than the primary L4regions). The swe-apodeme has been lost or reduced to vestiges (6.3.3). In the dorsal wall the L4I-region has strongly expanded to the right (6.3.3.). The L4d-region, if present at all, is no longer prominent from the outline of the L4I-region (6.3.3.). Possibly in correlation with this expansion of L4I, the pne-pouch has rotated (clockwise as seen from behind), and the membranous part of its wall is somewhat more on its right side (6.1.3). The distal part of the right posterior extension L1m of sclerite L1 curves into the dorsal wall of the lve-pouch, and L1m and L2 are therefore in the same plane in the area of articulation A2 (6.1.3.). The lve-pouch has become distinctly narrower (6.2.3.). Sclerite L2 in its dorsal wall has lost its primitive arch-shape (probably by a fusion of the arms of the arch, L2p and L2m) and is now ribbon- or plate-like (6.2.3.; compare (B) in 7.5.). At least the **paa**-process has distinctly lengthened (6.2.3.). Whether the **pda**-process has lengthened is not assessable since **pda** has been lost in *Chaeteessa*. The sclerotisations of pda and paa (or, to apply this character state also to *Chaeteessa*, the L4- and L2sclerotisations of the corresponding area) have been separated from each other (6.2.3.; compare (A) in 7.5.).

On the right phallomere, the **R1c**-region has been divided by the membranous stripe 4 separating the sclerites **R1A** and **R1B** (**R1A** and **R1C** in *Metallyticus*) (6.7.1., 6.7.3.).

Subgroup 1.2.1.: Chaeteessa

The left complex has a membranous pouch **pbe** between the **pne**- and **lve**-pouches (fig.34). The **pda**-process has been lost (6.2.3., 6.3.3.). The **loa**-process has also been lost (assuming that at least the **loa** of *Mantoida* and *Metallyticus* + *Sphodromantis* are homologous; 6.1.3.). The **vla**-lobe has considerably broadened as compared with the narrow **lve**-pouch (fig.32), and the genital opening has come into a position far to the right of the **lve**-pouch (6.2.3.). On the right phallomere, sclerite **R3** bends to the right along its left margin (6.7.3., fig.32). The keel-apodeme **3** has been lost (6.7.3.). The utmost right-posterior part of the **age**-groove (near articulation **A3**) has been reduced (6.7.3.).

Subgroup 1.2.2.: Metallyticus + Sphodromantis

Most of the autapomorphies are on the left complex: The dorsal and ventral parts of L4 have been separated by articulation A1 at the left edge of the left complex, which divides

the L4I-region (sclerites L4A and L4B; 6.3.3.). The ventral wall is uniformly sclerotised since the L4b-region has become as heavy as the primary L4-regions (6.3.3.). The L1-extension L1m, which curves into the dorsal lve-wall, as well as articulation A2 have become much broader (6.1.3.). The curving part of L1m now also sclerotises the afaprocess on the anterior part of the edge 1 between the pne- and lve-pouches (6.1.3.; this autapomorphy is uncertain since the homology of afa with the elements called afa in *Mantoida* and *Chaeteessa* is not certain, and since afa of *Metallyticus* is nothing but a shallow bulge). Sclerite L1 is, at least in its posterior part, divided by a stripe of membrane 2 within the L1m-region (6.1.3.). On the right phallomere, the deepening of the left part of the age-groove is very abrupt – certainly also a derived condition (6.7.3.).

Subgroup 1.2.2.1.: Metallyticus

The restriction of L4 (L4B-sclerite) to the anterior part of the dorsal wall seems to be an autapomorphy since in both *Chaeteessa* and *Sphodromantis* L4 or L4B occupies the whole dorsal wall (6.3.3.). The phallomere-gland P has probably been lost (at least, it was not found; 6.1.3.). The R1t-region (with the pva-tooth) has separated from the R1c-region (6.7.1., 6.7.3.; like in *Mantoida*: compare (G) in 7.5. and grouping B (123) in 7.6.). The R1d-region has expanded into the ventral wall of the pia-tooth and has largely ousted the membranous area 17 (6.7.3.).

Subgroup 1.2.2.2.: Sphodromantis

There are several autapomorphies: The L1m-extension has become extremely broad (fig.323a; 6.1.3.). Sclerite L1 is now completely divided by the membranous stripe 2 (sclerites L1A and L1B; 6.1.3.). The afa-process is highly elaborated (fig.10; 6.1.3.). The membranous part of the **pne**-wall with the phallomere-gland opening has undergone a further rotation to the ventral side of the **pne**-pouch (6.1.3.). The sclerotisation of the **loa**-process has been strongly reduced (6.1.3.). The tongue-like deepening of the anterior part of the **lve**-pouch and of L2 to the left is certainly also a derived feature since **lve** and L2 are narrow in both *Metallyticus* and *Chaeteessa* (6.2.3.). On the right phallomere, the crescent-like curvature and the extreme deepening of the left part of the **age**-apodeme and the resulting pouch **5** in the ventral wall of the genital pouch are derived features (fig.6; 6.7.3.). The posterior part of the **pia**-tooth and its **R1v**-sclerotisation have been reduced (6.7.3.).

LaGreca (1954) investigated the phallomeres of species of Amorphoscelididae, Eremiaphilidae, Hymenopodidae, Mantidae, and Empusidae. Not many of the characters dealt with in this chapter are recognisable in the figures of LaGreca or discussed in his text, but at least three features are present in all these species: (1) L4 always occupies the whole ventral wall of the left complex: the L4b-sclerotisation is present. (2) The sclerotisations of **paa** and **pda** are always separated from each other. (3) L4 is always divided into a dorsal (L4B) and a ventral (L4A) sclerite: the articulation A1 is present. (1) and (2) suggest that these families all belong to subgroup 1.2.; (3) additionally suggests that they all belong to subgroup 1.2.2.

7.3. The evolution of the phallomere complex and the phylogeny in Blattaria (= subgroup 2.)

The ground-plan of the phallomere complex of Blattaria

The features listed subsequently can be ascribed to the ground-plan of Blattaria since they are true either of all investigated species or at least of representatives of the two basal sister-groups (subgroups 2.1. and 2.2., see below). For all these features, except for the last-mentioned (reduction of muscle s2), it is not clear whether they belong to the common ground-plan of Blattaria and Mantodea or whether they are autapomorphic for Blattaria.

The following elements or properties are assumed to be present in the ground-plan since they are found in all Blattaria: sclerotisations L3, R2; formative element hla-hook. The L4v-region (in close connection with the L4c-region, if L4c is present) is separated from the regions L4l and L4n. The regions R1d and R1v are connected with each other (narrowly in the primitive case) posterior to the membranous area 17.

The following elements or properties occur only in part of the investigated species but are assumed to be present in the ground-plan since they are found in representatives of the two basal sister-groups: formative elements dca-processes, nla-process, tre-tendon, rge-groove; articulations A6, A7, A8, A9; muscles 114, r6, s7, and s8. The right parts of L2 and of the lve-pouch curve dorsad and back to the left. The base of the hla-hook is in the left anterior ventral wall of the left complex, and the introversible membranous basal part 30 of hla is narrow. The dca are two membranous cushions posterior to L1.

Another ground-plan feature of Blattaria might be that muscle s2 is distinctly thinner than its left counterpart s1 (6.9.); this situation is distinct in *Eurycotis* (fig.62: s2 weak), *Polyphaga*, and *Lamproblatta* (fig.113, 173: s2 lost). In *Cryptocercus* the condition of s2is not assessable; in *Anaplecta, Nahublattella, Parcoblatta*, and *Blaberus* s2 is also weak or absent, but, in addition, s1 has been completely lost. The situation in the Mantodean ground-plan, with s1 and s2 of similar stoutness (*Mantoida*, fig.37, 40), is regarded as more primitive than in all Blattaria since both muscles as well as their symmetry are preserved. An asymmetry in the stoutness of s1 and s2 is assumed to be an autapomorphy of Blattaria.

Subgroup 2.1.: Archiblatta + Eurycotis (and Periplaneta, Blatta, Deropeltis)

Periplaneta, Deropeltis, and *Blatta* have been studied only in part, but at least all the derived features listed subsequently are also present in these species. The muscles have not been studied in *Archiblatta*.

Most of the autapomorphies are on the left complex: The anteroventral part of the **lve**pouch is, like a tongue, deeply invaginated to the left (6.2.1.). The posteroventral part of the **lve**-pouch has strongly receded to the right (or is even more reduced: *Eurycotis*; 6.2.1.). Correlated with the latter feature is that the **paa**-process has shifted far to the right and is far away from the **pda**-process (6.2.1.). The sclerotisations of **pda** (**L4I**-region) and **paa** (**L2d**-region) have, possibly again in correlation with the previous feature, been separated from each other (like in some other subgroups: compare (A) in 7.5.; 6.2.1.). The **L4c**region is highly elaborated (or **L4c** is as a whole an autapomorphy of this subgroup), and

there is a distinct L4F-sclerite (6.2.1., 6.3.1.). The ventral insertions of the I5-muscles (I5a and I5b in *Eurycotis*) have shifted posteriad and take a position on L4F (6.3.1.; I5a,b could also be new muscles, but this would be a derived condition, too). The left insertion of muscle b4b has shifted to the top of the pne-pouch (6.1.4., 6.7.1.). The anterior part of sclerite L1 (L1a-region) has been leveled, and muscle I1 has been lost (6.1.4.; these two derived features, however, are also present in other subgroups of Blattaria: compare grouping M (24) and grouping R (25) in 7.6.). On the right phallomere, the R1t-region has enlarged and occupies most of the cbe-invagination including its anterior wall (6.7.4.).

Subgroup 2.1.1.: Archiblatta

On the left complex, the **paa**-process has lost most of its sclerotisation (**L2d**-region; 6.2.1.). On the right phallomere, the **R1t**-region has expanded to sclerotise the entire **cbe**-invagination and has developed a broad connection with sclerite **R2** (6.7.4.). Whether the condition that the anterior **L4c**-region forms an isolated sclerite **L4E** is an apomorphy of *Archiblatta* (and *Periplaneta, Blatta, Deropeltis*) or the plesiomorphic state within subgroup 2.1. is not decidable (6.3.1.).

Subgroup 2.1.2.: Eurycotis

As compared with *Archiblatta*, the cuticular elements of the left complex show many derived features: The L4d-region, if present at all, is no longer prominent from the outline of the L4l-region (6.3.1.). The posteroventral part of the lve-pouch is extremely reduced (6.2.1.). Sclerite L2 has lost the arch-shape and is plate-like: the arms of the arch, L2m and L2p, are assumed to have fused (6.2.1.; like in the Mantodean subgroup 1.2.; compare (B) in 7.5.). Within the vla-lobe there is a deep incision 9 (6.2.1.). The mla-lobe covering L4F has evaginated from the ventral wall (6.3.1., fig.63, 69). The pne-pouch has become less distinct (6.1.4.). The left-dorsal wall of pne contains two probably new sclerites L6A,B (6.1.4., 6.5.).

As compared with *Periplaneta, Blatta*, and *Deropeltis*, the muscles also show some derived features: The right insertion of **12** has shifted from sclerite **L1** to the adjacent membrane (6.1.4.). Muscle **13** has divided into three bundles **13a,b,c** (6.1.4., 6.2.1.).

The following derived features have not been investigated in the other species of subgroup 2.1., and it is unclear whether they are autapomorphies of *Eurycotis* or of a larger holophyletic group within 2.1.: **115** (fig.70) and some muscles in the **mla**- and **vla**-lobes (**113**-group except for **113h**, 6.5.; **116**, **117**, **118**; fig.71-73) are new. The presence of a separate muscle **r5** might also be derived (fig.80; this could be a new muscle or a split off part of **r1**).

Subgroup 2.2.: *Tryonicus* + (*Cryptocercus* + (*Lamproblatta* + (*Ergaula* + *Polyphaga*))) + (*Anaplecta* + (*Nahublattella* + (*Supella* + (*Euphyllodromia* + (*Parcoblatta* + (*Nyctibora* + (*Blaberus* + *Nauphoeta* + *Blaptica* + *Byrsotria*))))))

The most important synapomorphy of all these species is that the L4l-region reveals the same division (at A5 or (A5) in fig.329e-h): As far as the various substructures are preserved, and as far as no further subdivisions of the L4l-region have occurred, the

anteroventral part of L4I bears the left insertions of I2 and I4 and forms a sclerite (L4K) together with the L4n-region or its vestiges. The posterodorsal part contains the pdasclerotisation and, at its connection with the paa-sclerotisation (L2d-region), the posterior 110-insertion and forms a sclerite (L4N) together with the L4d-region. The L4I-, L4d-, and L4n-morphology of all species comprised in this subgroup follows this description (muscles not known in *Tryonicus*) or can be derived from this situation (6.3.1., 6.3.4.). (This division is completely different from the division of the L4I-region in the Mantodean subgroup 1.2.2. where the I2- and I4-insertions are together with L4d on the dorsal L4B and the pda-sclerotisations together with the L4n-region on the ventral L4A; fig.329c).

Additionally, the **swe**-apodeme has been completely lost (6.3.1., 6.3.4.). (**swe** has also been reduced in the Mantodean subgroup 1.2. The loss might be correlated with the division of **L4l** in both groups, for which region there is now no longer any need to be stiffened. For *Chaeteessa*, however, this explanation does not fit).

In its ground-plan subgroup 2.2. probably possesses a muscle **110** from the **lve**-pouch to the common sclerotisation of **paa** and **pda**. (However, this feature is not investigated in *Tryonicus*, and homology is not certain for **110** of *Cryptocercus*. In some members of subgroup 2.2.3.2.2.2. **110** is missing, but this is certainly a secondary loss, compare (**R**) in 7.5). **110** might be a posterior part of the ground-plan muscle **14**, which might have divided together with the **L4I**-region (its left insertion area); in this case, the similar division of **14** and the shift of the posterior part of its left insertion to the **paa**- and **pda**-sclerotisation would be an autapomorphy of this subgroup.

The L4d-sclerotisation has rotated (counterclockwise as seen from above): In *Tryonicus* L4d is directed anteriad; in the other species L4d is directed to the left, or, after a further rotation, dorsad (*Lamproblatta*), or it has been lost (6.3.4.).

On the right phallomere, the **pia**-tooth has been lost (6.7.6.). The regions **R1d** and **R1v** have developed a broad connection at the posterior edge of the **fda**-lobe (i.e. the former sclerites **R1G** and **R1H** have broadly fused to form **R1J**; 6.7.6.). (In some more derived species **R1J** has additionally fused with **R1F**, the sclerites **R1M** or **R1N** being the results). A possible autapomorphy is the extreme reduction of muscle s2 (more than in the Blattarian ground-plan and in subgroup 2.1.); this feature, however, has not been investigated in

Tryonicus and is not assessable in *Cryptocercus* (6.9.). Another possible autapomorphy is the sclerite-ring formed by the posterior part of L1 (by the regions L1l, L1m, and L1r; 6.1.1., 6.1.4.; compare (P) in 7.5.).

For this subgroup 2.2. there are two possibilities for the next subordinate sister-group relation; both are supported by derived character states or possible autapomorphies. Hence, there is a trichotomy not resolvable with the present state of knowledge. Alternative B, followed in fig.322 and 330, might be more probable.

Alternative A: Holophyly of Subgroup 2.2.1. + Subgroup 2.2.3. is supported by two derived character states of the hla-hook: The introversible membranous basal part 30 of hla has become more extensive (hla can therefore be retracted more deeply into the left complex; 6.4.3.). The base of hla has shifted posteriad (6.4.3.). These two features are possibly intercorrelated (compare (M), (N) in 7.5.). A posteriad shift of the hla-base, however, is also present in *Cryptocercus* (fig.151; compare in 7.7.).

Alternative B: Holophyly of Subgroup 2.2.2. + Subgroup 2.2.3. is supported by three derived character states of the regions L4I and L4d: The anterior and posterior parts of the L4I-region (in the primitive case included in the L4K- and L4N-sclerites) are still hinged to each other in *Tryonicus* (articulation A5) but always far removed from each other in the species of these subgroups (6.3.4.). The utmost right-anterior part of the L4I-region, which in *Tryonicus* extends rightward anterior to the L4n-region, has been lost (6.3.4.). The L4d-ribbon has further rotated (counterclockwise as seen from above) and is now directed to the left. (In *Lamproblatta* L4d has additionally rotated into a dorsoventral orientation; in subgroup 2.2.3. L4d has been preserved only in *Nahublattella*; 6.3.4).

Subgroup 2.2.1.: Tryonicus (parvus and angustus)

Synapomorphies of the two species of *Tryonicus* investigated in this paper are the rotation of the **pne**-pouch (counterclockwise as seen from behind; extreme in *T. parvus*; 6.1.4.), the enlargement and plate-like condition of the **L1m**-region, and the consequently hinge-like condition of articulation **A2** (6.1.4.). The anterior part of **L1** has become level (6.1.4.; like in the subgroups 2.1. and 2.2.3.; compare grouping M (24) in 7.6.).

Derived features of *T. parvus* (characters not investigated in *T. angustus*) are the extension **R2m** of sclerite **R2** and the loss of the **age**-apodeme (6.7.4.; both features also in *Lamproblatta*: compare grouping G (**129**) and (**130**) in 7.6.). The sclerite bridge **L3a** which connects the **L4n**-region and **L3** and crosses the **hla**-base **30** is probably also derived (6.4.3.).

Subgroup 2.2.2.: Cryptocercus + (Lamproblatta + (Ergaula + Polyphaga))

On the left complex, sclerite L4K has been reduced in a specific way (6.3.4.): The parts of L4K which in the ground-plan of subgroup 2.2. (as in *Tryonicus*) take a position right-ventral to the hla-base have been lost. This concerns mainly the L4n-region, and the anterior insertion of 114 (muscle lost in *Polyphaga* and *Ergaula*) is at least mostly on membrane. (In *Polyphaga* and *Ergaula* this reduced L4K has shifted to the ventral side of the hla-base). Probably in consequence of this L4n-reduction, the nla-process has been lost (6.3.4.).

Muscle 12 shows a shift to the anterior: This concerns a gradual anteriad shift of the left insertion (6.3.4.; least distinct in *Cryptocercus*: insertion still on sclerite L4K, i.e. on region L4I) and a complete anteriad shift of the right insertion from the left wall of the **pne**-pouch to its top (6.1.4.). That the anterior face of the **pne**-pouch, i.e. of sclerite L1, has become plateau-like (6.1.4.; distinct in *Cryptocercus*, *Polyphaga*, and *Ergaula*; plateau vestigial in *Lamproblatta*) is possibly correlated with the shift of 12 and is also assumed to be an autapomorphy of this subgroup. That this feature of **pne** is obsolete in *Lamproblatta* is assumed to be a secondarily derived condition, possibly correlated with the apomorphic right-anteriad shift of **pne** by which the insertion angle of 12 on **pne** has become very acute.

Another possible autapomorphy might be the complete loss of muscle s2, which feature, however, is not assessable in *Cryptocercus* (6.9.). (The loss of s2 in subgroup 2.2.3.2. is certainly a case of parallel evolution since s2 is present in *Anaplecta*.)

Subgroup 2.2.2.1.: Cryptocercus

The left complex has several derived features: The right part of L2, which is upcurved in other Blattaria, has been reduced; in consequence, the respective right-dorsal part of the **lve**-pouch has been reduced, and the contact between L2 and L1 (articulation A2) has been lost (6.1.4., 6.2.4.). The left edge **61** of the **vla**-lobe has expanded almost to the anterior margin of the left complex; in consequence, the left-ventral part of the **lve**-pouch has also been extremely reduced (6.2.4.; a similar derived state is present in *Tryonicus* and in *Anaplecta*: compare grouping C (**124**) in 7.6.). The **pda**-sclerotisation (**L2d**-region) (6.2.4., 6.3.4.; the separation of **L4l** and **L2d** has also developed in the subgroups 1.2. and 2.1.: compare (A) in 7.5.). New muscles of *Cryptocercus* are **l19** (6.4.3.), **r7** (6.7.5.), **r8** (6.7.4.), and possibly **l7** (6.5., 7.1.).

There are some derived features in the phallomero-sternal and peripheral musculature: s1 and s3, and probably s7, are fused (6.9.). s4 and s6, and s2 if present, are fused (6.9.). The p1 are extremely broad. The posterior insertions of the p4 are far laterally (6.11.). All these seemingly primitive features are assumed to be neotenic traits; that they are not primitive but derived results from the outgroup comparison with Mantodea. s5 has been lost (6.9.; like in *Polyphaga* and *Ergaula*: compare grouping H (131) in 7.6.).

Subgroup 2.2.2.2.: Lamproblatta + (Ergaula + Polyphaga)

There are many autapomorphies on the left complex: L8 is a new sclerite in the right dorsal wall, with the insertions of 112, 19, and b2 upon it or in its immediate vicinity (6.5.). L7 is a new sclerite on the right part of the vla-lobe (or on the lba-lobe, which is a right part of vla; 6.5.). The muscles 111 (6.3.4.), 112 (6.2.4., 6.5.), and s12 (6.2.4., 6.3.4., 6.9.) are also peculiar to this subgroup. The L4-plate in the ventral vla-wall has expanded to include the dorsal insertion of muscle s3 (new region L4a, larger sclerites L4R and L4M; 6.3.4.). The left insertion of 12 has shifted further anteriad and away from sclerite L4K (or region L4l; 6.3.4.). The lve-pouch has expanded almost to the left edge of the left complex (6.2.1., 6.2.4.).

Other derived features could possibly also be autapomorphies of this subgroup: If s2-parts should be included in s2+4+6 of *Cryptocercus*, s2 would have been lost in subgroup 2.2.2.2. (6.9.). The dorsal insertion of muscle s6 has become restricted to the anterior margin of sclerite R3 (6.9.). (If the muscle named s6 should be s2, the loss of s6 and the laterad expansion of the ventral s2-insertion would be the autapomorphies). If s7 really is a ground-plan muscle of Blattaria (compare (L) in 7.5.), and if vestiges of s7 are included in s1+3 of *Cryptocercus*, the complete loss of s7 would also be an autapomorphy of subgroup 2.2.2.2. (6.9.). Muscle 11 has been lost. (However, 11 has certainly been lost several times: at least also in the subgroups 2.1., 2.2.3.1., and 2.2.3.2.2.; compare grouping R (25) in 7.6.)

Subgroup 2.2.2.1.: Lamproblatta

There are many derived features on the left complex. L2 has divided into three sclerites: L2A in the left part of the lve-pouch, L2B in the right part of the lve-pouch, and L2C

on the **paa**-process (6.2.4.). **L4N** has divided into two sclerites: **L4S** containing the **L4d**region, **L4T** on the **pda**-process (6.3.4.). (The sclerotisations of **paa** and **pda** remain connected). Around articulation **A4** (between **L2A** and **L2B**) the **lve**-pouch has developed a deep recess (6.2.4.). The **L4d**-region has rotated into a dorsoventral orientation (6.3.4.). The processes **paa** and **pda** have elaborated the cuticular invaginations **boe** and **sbe** (fig.182). The muscles **l20**, **l21**, **l22**, **l23**, and **l24** have evolved (fig.184-188). Muscle **l4** has been lost (like in *Anaplecta*: compare grouping K (**133**) in 7.6.).

On the right phallomere, the **tre**-tendon and its muscles **b4** and **s8** have been lost (6.7.5.; like in subgroup 2.2.3.: compare (I) in 7.5. and grouping E (73) in 7.6.). The **age**-groove has been lost (6.7.4.), and sclerite **R2** bears an extension **R2m** to the left (6.7.4.; both features also in *Tryonicus*: compare grouping G (129) and (130) in 7.6.). Muscle **r1** has been lost (6.7.6.).

Subgroup 2.2.2.2.: Ergaula + Polyphaga

There are many autapomorphies on the left complex: Sclerite L4K has shifted to the posteroventral part of the hla-base (6.3.4.). Muscle 114 has been lost and functionally replaced by 14 (6.3.4., 6.4.3.). The L4-plate in the ventral vla-wall has undergone a further expansion (new region L4x, larger sclerite L4M) and includes now the left insertion area of 12 (6.3.4.). Additionally, this 12-insertion has shifted further anteriad and also ventrad (6.3.4.). The cuticular area around sclerite L7 has been elaborated as a new lobe lba which represents the rightmost part of the vla-lobe (6.5.).

On the right phallomere, the sclerites **R2** and **R3** have fused (and articulation **A7** has been lost; 6.7.4.). The large sclerite **R1M** has developed, either by a posteriad expansion of the former **R1F** alone or, more probably, by an additional fusion of **R1F** and parts of the former **R1J** (6.7.6.; with the loss of the membranous area **17** and of the articulations **A8** and **A9** as a result; compare (**H**) in 7.5.). Probably in correlation with this feature (in its latter interpretation) muscle **r3** has been lost (6.7.6.). (The fusion of **R1F** and **R1J** and the loss of **r3** have also been achieved in subgroup 2.2.3.: compare grouping **F** (**128**) and (**64**) in 7.6.). The **rge**-groove on the **R1c**-region has distinctly expanded posteriad (6.7.6.). The **pva**-ridge on the **R1t**-region has achieved a longitudinal orientation and has likewise expanded posteriad (6.7.6.). (That means, within the **R1M**-sclerite, as compared with the **R1F**-sclerites, the regions **R1c** and **R1t** have expanded posteriad). Sclerite **R2** has expanded to occupy most of the **cbe**-invagination and is connected with **R1t** in the dorsal wall of **cbe** (6.7.4.). The articulation **A6** between **R2** and **R1t** has been lost (6.7.4.). (Since a fusion of **R1t** and **R2** does not necessarily result in a loss of **A6** – compare *Archiblatta* in 6.7.4. – each of the two latter features is regarded as an autapomorphy of its own.)

Subgroup 2.2.2.2.1.: Polyphaga

The only derived feature known as compared with *Ergaula* is the ventral gap in the sclerite ring formed by the posterior part of L1 (6.1.4.). **r9** is a new muscle (6.7.4.; *Ergaula* not investigated).

Subgroup 2.2.2.2.2.: Ergaula (capensis and capucina)

On the left complex, the anteriormost part of L4M has split off to form an isolated sclerite (with the insertions of s3 and s12; 6.3.4.). Sclerite L4K has shifted somewhat farther anteriad (6.3.4.). The dorsal part of L4K within the hla-base has shortened and fused to the ventral anterior margin of sclerite L3 (6.3.4.). Muscle l11 has distinctly enlarged (6.3.4.; investigated only in *E. capucina*). The paa-process has been lost (6.3.4.). On the right phallomere, R2 has broadened, and R3 is now for most of its breadth confluent with R2 (6.7.4.). The weak lines A7* and 13, representing the fusion lines between R2 and R3 or R2 and R1t, respectively, in *Polyphaga*, have been lost (6.7.4.).

Subgroup 2.2.3.: Anaplecta + (Nahublattella + (Supella + (Euphyllodromia + (Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria)))))

All these species belong to Blattellidae and Blaberidae sensu McKittrick (1964). In *Anaplecta, Nahublattella, Parcoblatta*, and *Blaberus* the whole phallomere complex has been investigated, including its muscles. In the other species only certain parts or elements have been studied, or their presence or absence has been checked (mainly the elements listed in 5.15.). It will be exactly specified which derived features are known to be present in which of these species. *Ectobius* and *Loboptera* will not be considered in the following analysis since too few features have been investigated to correctly assess and assign these species, which are probably highly modified in their phallomere morphology.

At least the following apomorphies are present in all species comprised in this subgroup: On the left complex, the introversible membranous basal part **30** of the **hla**-hook has become very extensive, and **hla** can be almost completely retracted (6.4.3.). The **hla**-base has shifted to the left posterior edge of the left complex (6.4.3.). (These two features are possibly intercorrelated; compare (**M**), (**N**) in 7.5). The left part of the left complex, which contains the **hla**-base, has been separated from the parts more to the right by the **fpe**infolding (6.4.3.). The anterior part of the **lve**-pouch has been elaborated as a tube-like **lve**-apodeme (6.2.4.). The common sclerotisation of the processes **paa** and **pda** has become stout and ring-shaped in its basal part (6.2.4.). (The resulting very close relation of **paa** and **pda** and their sclerotisations might be the basis for the formation of the **via**-process with an elongated common basal part of **paa** and **pda** in subgroup 2.2.3.2.).

On the right phallomere, the **tre**-tendon and its muscles **b4** and **s8** have been lost (like in *Lamproblatta*: compare (I) in 7.5. and grouping E (73) in 7.6.; 6.7.5.). Sclerite **R1N** has developed by a fusion of the former **R1F** and **R1J** (6.7.6.; the loss of the membranous area 17 and of the articulations A8 and A9 are concomitant derivations; all regions of **R1** are now included in one sclerite, like in the common ground-plan of Blattaria and Mantodea). The loss of muscle **r3** is probably correlated with this feature (6.7.6.). (The fusion of **R1F** and **R1J** and the loss of **r3** have also been achieved in subgroup 2.2.2.2.2.: compare grouping F (128) and (64) in 7.6. and (H) in 7.5.). The **rge**-groove on the **R1c**-region has been lost (6.7.6.). (This feature is absent in *Supella*; it is assumed to be rendered unrecognisable by the extreme expansion of sclerite **R1N'**. In subgroup 2.2.3.2.2.2. this curved area forms the **cwe**-thickening).

The following features have been investigated only in *Anaplecta, Nahublattella, Parcoblatta*, and *Blaberus*: Muscle **s1** has been lost (6.9.). The muscles termed **15** in *Anaplecta* and *Nahublattella* are possibly new muscles, or they are true **15** with the anterior insertion shifted to the **L4n**-region; in any case, one of these character states is probably apomorphic for subgroup 2.2.3. (6.2.4., 6.3.4.). (In *Parcoblatta* and *Blaberus* **15** has been lost or integrated into **16b**).

Subgroup 2.2.3.1.: Anaplecta

On the left complex, sclerite L1 and the dca-processes have been lost (like in all or many species of subgroup 2.2.3.2.2., compare grouping L (109) and (110) in 7.6.; according to McKittrick (1964), however, L1 is present in another species of *Anaplecta*; 6.1.4.). The **pne**-pouch is therefore completely membranous; it has been reduced to a shallow depression in the central dorsal wall. (**pne** has been lost completely in all or many species of subgroup 2.2.3.2.2., compare (111) in 7.4. and grouping L (134) in 7.6.; 6.1.4.). Muscle **14** has been lost (6.2.4., 6.3.4.; like in *Lamproblatta*: compare grouping K (133) in 7.6.). Some membranous foldings have developed in the area of the **lve**-pouch, e.g. **vfa** and **vpe** (6.2.4.). The **gta**-process (6.2.4., fig.215) and the **vte**-tendon (fig.208) have evolved. The L4d-region has been lost (like in subgroup 2.2.3.2.2., compare grouping L (95) in 7.6.). Muscle **11** has been lost (like in some other subgroups: compare groupings L and R (25) in 7.6.). **125** and **126** are probably muscles peculiar to *Anaplecta*. (However, **126** might be homologous with **16a** of *Nahublattella*, *Parcoblatta*, and *Blaberus*; 6.3.4.).

The firm connection between the **nla**-process and the top of the **lve**-apodeme might be autapomorphic for *Anaplecta*. According to its possible correlation with the translocation of the anterior **l14**-insertion from **nla** to the top of **lve**, however, this feature could also be an autapomorphy of the whole subgroup 2.2.3., lost again at the base of subgroup 2.2.3.2. (compare (**M**), (**N**) in 7.5.; 6.4.3.). That there are two phallomere gland openings in a rather peculiar position – possibly new organs – could also be either a derived feature of *Anaplecta* alone or an autapomorphy of subgroup 2.2.3.2, with a loss of one opening and a deplacement of the other at the base of subgroup 2.2.3.2. (6.6.).

Subgroup 2.2.3.2.: Nahublattella + (Supella + (Euphyllodromia + (Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria)))))

At least the following apomorphies are present in all species listed: The anterior insertion of muscle **114** has been translocated from the **L4n**-region on the **nla**-process to the **L2a**-region on top of the **lve**-apodeme (6.4.3.; or, if the homology of the **l14**-muscles should not be true, a new muscle from **L2a** to **L3** has developed). The size of the **vla**-lobe has distinctly decreased (6.2.4., 6.3.4.). The common base of the **paa**- and **pda**-processes has been elongated and forms, together with **paa** and **pda**, the **via**-process (6.2.4., 6.3.4.). The right posterior dorsal part of the left complex – the part dorsal to the right half of the **lve**-pouch – has been reduced (6.2.4.). The division of **L2** into **L2D** (within the **lve**-pouch) and **L2E** (together with **L4N** on the **via**-process) by articulation **A10** can also be regarded as an autapomorphy of this group since this situation is present in both the subordinate

sister groups 2.2.3.2.1. and 2.2.3.2.2. (The absence of this division and of A10 is regarded as a secondary loss having occurred several times within subgroup 2.2.3.2.2.; compare (\mathbf{Q}) in 7.5.).

The following derived features are also assumed to be autapomorphies of subgroup 2.2.3.2. but have been investigated in sufficient detail only in *Nahublattella, Parcoblatta*, and *Blaberus*: On the left complex, L4K has divided into the sclerites L4U and L4V – with the nla-process probably still present on L4V' of *Nahublattella*. (In *Blaberus* and possibly also in *Parcoblatta* L4V has been completely lost; 6.3.4.). Sclerite L4G (L4v-region) in the ventral vla-wall has been lost (6.3.4.). The right insertion of muscle l2 has shifted to the membranous basal part 30 of the hla-hook (6.3.4.). Muscle l30 has developed, having a longitudinal course in the ventral wall of the left complex (6.5.). Muscle s2 has been lost (6.9.; like in subgroup 2.2.2. or 2.2.2.2.: compare above). The muscles s5 and s6 have divided into s5a and s5b or s6a and s6b, respectively (or, new muscles s5a and s6a have developed; 6.9.). The p4-insertions on the subgenital plate have shifted mesad (6.11.).

Subgroup 2.2.3.2.1.: Nahublattella

On the left complex, the sclerotisation L2E'+L4N' of the via-process has been divided transversely by the membrane ring **39** (6.2.4.). The **nla**-process, if actually **nla**, has become whip-shaped (6.3.4.). The central dorsal wall contains a bristle area **35** (fig.242). The muscles **127**, **128**, **129**, **131**, **132**, and **135** have developed (fig.240, 249-252). On the right phallomere, sclerite **R2** has become plate-like, bearing the highly elaborated evaginations **42** and **43** (6.7.4.). The **R1t**-region has probably completely fused to the rest of sclerite **R1N'** (6.7.6.). There is a new muscle **r10** (6.7.6.). The dorsal sclerotisation **S9d** of the subgenital plate is highly elaborated, divided, and provided with the new muscles **133** and **134** (6.10.).

Subgroup 2.2.3.2.2.: Supella + (Euphyllodromia + (Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria))))

At least the following apomorphies are present in all species listed (with the exceptions mentioned): On the left complex, the **hla**-hook has evolved a **hge**-groove with a notch **45** in its ventral wall (6.4.3.). (In *Nyctibora* and *Nauphoeta* **hge** is not that distinct, and the notch **45** is missing; this is probably due to secondary reduction). The **ate**-tendon has become very long and narrow (6.3.4.). (That **ate** is rather short in *Blaberus, Blaptica*, and *Byrsotria* is interpreted as a secondary reduction since **ate** is very long and thin in *Nauphoeta*, and since these four species form the holophyletic subgroup 2.2.3.2.2.2.2.2.2.). Sclerite **L4V** (essentially the **L4n**-region) has been reduced to a small sclerite in the dorsal wall of the **ate**-tendon or has been completely lost (or, **L4V** has been generally lost, and the sclerite within **ate**, present only in *Parcoblatta, Nyctibora*, and *Blaptica*, is a new one; 6.3.4.). The **nla**-process has been lost (6.3.4.). The **right** posterior branch of **L2**, whose distal part sclerotises the **psa**-process in *Nahublattella*, has been completely lost (6.2.4.). The **L4d**-region has been lost (6.3.4.; like in *Anaplecta*: compare grouping L (**95**) in 7.6.). The

ventral extension **28** of **L2**, present in *Anaplecta* and *Nahublattella*, is lacking; this is also assumed to be a derived feature (6.2.4.; compare the possible homology of **28** and **L5**: 6.5.).

The following apomorphies have been investigated only in *Parcoblatta* and *Blaberus*; they might be autapomorphies of the whole subgroup 2.2.3.2.2. or of any subordinate subgroup containing at least Parcoblatta and Blaberus: The age-groove has been restricted to the anterior part of sclerite R3, i.e. the age-part along the posterior right margin of R3 has been lost (6.7.4.). Muscle 114 has divided into two bundles 114a and 114b (6.4.3.; this division is completely different from the division of **114** in *Eurycotis*). Muscle **13** has been lost (6.1.4., 6.2.4.). The muscles 136, 137, and 138 have developed (fig. 276-278, 303, 307; 6.4.3.). Muscle 15 has been lost or integrated into 16b (6.2.4.). Muscle 16a has distinctly enlarged (6.2.4.), Muscle s3 has divided into two bundles s3a and s3b (6.9.). The muscles s14 or s14a,b are new (6.9.). The dorsal insertion of muscle s6b has expanded posteriad to occupy the entire right margin of sclerite $\mathbf{R3}$ (6.9.). The ventral insertions of s5a and s6a have shifted posteriad (6.9.). Some further derived features of *Parcoblatta* and Blaberus are also present in Anaplecta but not in Nahublattella (compare grouping L (109), (110), (134), and (25) in 7.6.): Sclerite L1, the dca-process(es), and muscle 11 have been lost. The **pne**-pouch, which has become indistinct in *Anaplecta*, has been completely lost (6.1.4.).

Subgroup 2.2.3.2.2.1.: Supella

Sclerite **R1N'** has extremely expanded to occupy the entire dorsal wall of the **cbe**invagination (6.7.6.). The indistinctness of the **R1t'**-region and the lack of a hook-like curvature at the median end of **R1t'** (present in the ground-plan of subgroup 2.2.3.) are assumed to be results of this expansion.

Subgroup 2.2.3.2.2.: Euphyllodromia + (Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria)))

All species listed share some very important derived features on the right phallomere (6.7.6.): The median ends of the **R1t**-region and of sclerite **R2** have fused: loss of articulation **A6**. (**A6** is still an articulation in *Supella*). The hook-curvature at the median end of **R1t** has been elaborated as the **cwe**-thickening. At its lateral end, **R1t** has been separated from the **R1c**-region: resulting sclerites **R1P** and **R1S**. (In *Supella* **R1c** and **R1t** are still connected with each other. In *Blaberus, Nauphoeta, Blaptica*, and *Byrsotria* **R1t** and **R1c** are also connected: sclerite **R1T'**; this situation is interpreted as a secondary fusion of these regions and as a synapomorphy of these species, which view is suggested by the assumed autapomorphies of the subgroups 2.2.3.2.2.2.2. and 2.2.3.2.2.2.2.2.).

Subgroup 2.2.3.2.2.1.: Euphyllodromia

No derived features restricted to this species have so far been found in the phallomere complex. According to the phylogenetic hypothesis presented here, the following derived features appear as autapomorphies: The L2-sclerotisations of the via-process and of the lve-pouch have fused secondarily (i.e. articulation A10 has been lost). Muscle 110 has

been lost. (Both features also in some other species of subgroup 2.2.3.2.2.2.; compare (Q), (R) in 7.5.). The membranous left wall of the lve-pouch has deeply invaginated to the left (also in *Loboptera*; 6.2.4.).

Subgroup 2.2.3.2.2.2.: Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria))

Only two possible autapomorphies are present in the phallomere complex: First, the presence of a **dla**-lobe (6.7.6.). **dla** is clearly missing in *Supella* and *Euphyllodromia*. **dla** is also missing in *Nauphoeta*, but in this species the whole right phallomere has been strongly reduced. Second, the presence of a **tve**-tendon (6.2.4.). **tve** is absent in *Supella* and *Euphyllodromia* and present in all members of this subgroup except *Blaberus* (*Byrsotria* not investigated). The absence in *Blaberus* is regarded as a secondary loss. The holophyly of this subgroup is strongly supported by a clearly derived feature of the females: They perform a rotation of the ootheca within the vestibulum (into a horizontal orientation; McKittrick 1964; termed advanced rotation by Roth 1967). In *Supella* and *Euphyllodromia* the ootheca retains a vertical orientation till it is dropped (Roth 1967).

Subgroup 2.2.3.2.2.2.1.: Parcoblatta

Features of *Parcoblatta* which are derived as compared with all other investigated species are the **vge**-groove on the **via**-process (fig.273) and the rotation of the **via**-process and of some adjacent elements, which includes a ventrad shift of the genital opening (6.2.4.). Some other features of *Parcoblatta* are derived at least as compared with *Blaberus* (characters mostly not investigated in the other species of subgroup 2.2.3.2.2.): Sclerite **L4U** has been lost (6.3.4.). Sclerite **R2** has become distinctly curved (6.7.4.). The muscles **139** and **140** have developed (fig.277, 278). Muscle **s14** has divided into two bundles **s14a** and **s14b** (6.9.). The ventral insertion of muscle **I6a** and the dorsal and ventral insertions of muscles **s5a** and **s6a** have shifted very far posteriad (6.2.4., 6.9.). Further derived features are the fusion of the **L2**-sclerotisations of the **via**-process and of the **lve**-pouch and the loss of muscle **I10** (6.2.4.; both like in *Euphyllodromia* and *Blaberus*: compare (**Q**), (**R**) in 7.5.).

Subgroup 2.2.3.2.2.2.2.: Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria) This subgroup has some probably derived features on the right phallomere: There are two new sclerites, **R5** in the ventral part of the right phallomere (6.7.4.) and **R4** in the dorsal wall of the **dla**-lobe (6.7.6.). A new muscle **r11** runs from the dorsal **dla**-wall to the ventral **fda**-wall (6.7.6.).

Subgroup 2.2.3.2.2.2.2.1.: Nyctibora

No certain autapomorphies have so far been found in the phallomere complex.

Subgroup 2.2.3.2.2.2.2.2.: Blaberus + Nauphoeta + Blaptica + Byrsotria

In all listed members of this subgroup, which corresponds to the Blaberidae sensu McKittrick (1964), the **R1t'**-region (sclerite **R1S'**) and the rest of **R1'** (sclerite **R1P'**) have

fused secondarily to form sclerite **R1T'**. Only in *Blaberus, Blaptica*, and *Byrsotria* the **ate**-tendon has shortened and broadened (6.3.4.; **ate** has remained long and thin in *Nauphoeta*), the **L10'**-sclerotisation has evolved (one sclerite in *Blaberus*, fig.299; many small sclerites in *Blaptica*, fig.291, and *Byrsotria*), and the **via**-process and some adjacent elements have rotated (6.2.4.; this rotation, which includes a dorsad shift of the genital opening, is most advanced in *Blaberus*).

At least *Blaberus* has developed the muscles **l41** to **l46** and **r12** to **r18** (not investigated in *Nauphoeta, Blaptica*, and *Byrsotria*) and lost muscle **s7** (6.9.; **s7** is present at least in *Nauphoeta*; not investigated in *Byrsotria* and *Blaptica*).

7.4. Survey of phylogeny and aut/synapomorphies

In this section, the character states assumed to be autapomorphies of subgroups are listed, and a phylogenetic tree is given (diagram 1). The autapomorphies are termed by bold printed numbers put in brackets. Some symbols give additional information: 1: The same apomorphic character state has evolved in at least one other subgroup, too, and homology is not contradicted by morphological data or functional arguments but only by the distribution of the apomorphic states of other characters (i.e. by parsimony). The apomorphic state has the same number in all subgroups concerned. ?: The position of the autapomorphy in the tree is questionable, due either to lack of investigations or to not definitely interpretable morphology. (? element): The homology of the named element and hence that of the respective apomorphic state in the various species included in the subgroup is questionable. // separates different conceivable morphological interpretations of character states. In the tree bold print, the brackets, and the symbols except for ? are omitted.

The plesiomorphic character states are given in brackets, and for each state the taxon or subgroup is named within the range of which it is plesiomorphic: (1) In most cases one character state is listed which is plesiomorphic within Blattaria and Mantodea as a whole (i.e. which is present in the common ground-plan of Blattaria and Mantodea). Such a character state is preceded by "BM:" = Blattaria + Mantodea. (2) In some cases one character state is listed whose categorisation as plesiomorphic is related to the range of Blattaria, of Mantodea, or of a subgroup of Blattaria or Mantodea which is superordinate to the subgroup under consideration and includes it. Such a character state is preceded by the name of the respective superordinate subgroup or taxon, e.g. "SG2.2.:" = subgroup 2.2., "SG1.:" = Mantodea, "SG2.:" = Blattaria. This is practised if the character concerns a property of an element whose presence in the common ground-plan of Blattaria and Mantodea is uncertain, if the character is for any reason not assessable in the species outside the named subgroup or taxon, or if an exact description of the character is only possible within the named subgroup (e.g. if a sclerotisation has divided and changed its shape previously, and the character concerns a further derivation of such a sclerotisation). (3) If several states of a character form a transformation series, all states which are more plesiomorphic than the named apomorphic state are listed, and for each of them the respective subgroup is given as in (1) and (2).

Subgroup 1.2.: (1) Region L4b between L4-regions L4l, L4v, and L4n present (BM: L4b absent) (2) Apodeme swe vestigial or absent (BM: swe present along most of L4l) (3) Region L4l in dorsal wall extending far to the right (BM: L4l restricted to left margin of dorsal wall) (4) Region L4d not prominent from outline of region L4l // L4d absent (BM: L4d prominent from outline of L4l // L4d present) (5) Membranous part of pne-wall on right side (BM: Membranous part of pne-wall dorsal or right-dorsal) (6) Distal part of region L1m curving into dorsal wall of pouch lve (BM: Distal part of L1m not curving into dorsal wall of lve) (7) Pouch lve narrow (BM: lve moderately broad) (8) Sclerite L2 ribbon- or plate-like (BM: L2 arch-shaped) (9) Process paa long (BM: paa very short) (10) Sclerotisations of processes pda and paa = regions L4l and L2d separated (BM: L4l and L2d connected) (11) Region R1c divided by stripe 4: sclerites R1A, R1B, or R1A, R1C (BM: R1c undivided, stripe 4 absent).

Subgroup 1.2.2.: (12) Region L4l divided by articulation A1: sclerites L4A, L4B (BM: L4l undivided, A1 absent) (13) Region L4b as heavily sclerotised as L4-regions L4l, L4v, and L4n (SG1.2.: L4b weaker than L4-regions L4l, L4v, and L4n; BM: L4b absent) (14) Region L1m rather broad (BM: L1m very narrow) (15)(? afa) Region L1m occupying process afa (SG1.: afa membranous) (16) Region L1m divided in its posterior part by membranous stripe 2 (BM: L1m completely undivided, stripe 2 absent) (17) Left part of apodeme age abruptly deepened (BM: Left part of age not or gradually deepened).

Subgroup 2.1.: (18) Pouch lve: anteroventral part deeply invaginated to the left, posteroventral part receded to the right (BM: anterior or anteroventral part of lve not deeper invaginated to the left than posterior part, posterior or posteroventral part not receded to the right) (19) Process paa far on the right and far to the right of process pda (BM: paa on the left and immediately to the right of pda) (20) Sclerotisations of processes pda and paa = regions L4l and L2d separated (BM: L4l and L2d connected) (21) Sclerite L4F present (BM: L4F absent) (22) Ventral insertion of muscle 15 far posteriorly and on sclerite L4F // Muscle 15 present, not homologous with 15 of other subgroups (BM: Ventral insertion of 15 far anteriorly // This 15 absent) (23) Left insertion of muscle b4b on top of pouch pne (BM: L1a hood-shaped) (25)! Muscle 11 absent (BM: 11 present) (26) Region R1t rather large, occupying entire dorsal and anterior walls of invagination cbe (BM: R1t narrow, occupying only part of dorsal wall of cbe).

Subgroup 2.2.: (27) Region L4l divided by articulation A5 or by a farther separation homologous with A5: sclerites L4K, L4N (BM: L4l undivided, A5 absent) (28) Apodeme swe absent (BM: swe present along most of L4l) (29)? Muscle 110 present // Muscle 14 divided into anterior and posterior (= 110) bundle and posterior bundle shifted posteriad to common sclerotisation of pda and paa (BM: 110 absent // 14 undivided and inserted on L4l left-anterior to sclerotisation of paa and pda) (30) Region L4d directed anteriad (BM: L4d directed to the right or right-anteriad) (31) Tooth pia absent (BM: pia present) (32) Regions R1d and R1v broadly connected posterior to membrane 17: sclerite R1J (BM: R1d and R1v not or narrowly connected posterior to membrane 17) (33)? Muscle s2 extremely reduced (SG2.: s2 moderately reduced; BM: s2 not reduced and as strong as s1) (34) Posterior part of sclerite L1 forming a ring (BM: L1 not forming a ring).

? Subgroups 2.2.1. + 2.2.3.: (35) Membranous basal part 30 of hook hla extensive, hla rather deeply retractable (SG2.: 30 very narrow, hla hardly retractable) (36) Base of hla in middle part of left ventral wall of left complex (SG2.: Base of hla in anterior left ventral **?** Subgroups 2.2.2. + 2.2.3.: (37) Anterior and posterior parts of region L4l far separated (SG2.2.: Anterior and posterior parts of L4l separated but articulated in A5; BM: L4l undivided) (38) Utmost right-anterior part of region L4l absent (BM: Utmost right-anterior part of L4l present) (39) Region L4d directed to the left (SG2.2.: L4d directed anteriad; BM: L4d directed to the right or right-anteriad) (29)? Muscle 110 present // Muscle 14 divided into anterior and posterior (= 110) bundle and posterior bundle shifted posteriad to common sclerotisation of paa and paa (BM: 110 absent // 14 undivided and inserted on L4l left-anterior to sclerotisation of paa and pda) (33)? Muscle s2 extremely reduced (SG2.: s2 moderately reduced; BM: s2 not reduced and as strong as s1).

Subgroup 2.2.1.: (40) Membranous part of pne-wall on left side (BM: Membranous part of pne-wall dorsal or right-dorsal) (41) Region L1m plate-like, articulation A2 very broad (BM: L1m ribbon-like, A2 narrow) (24)! Region L1a level (BM: L1a hood-shaped).

Subgroup 2.2.2.: (42) Right-ventral part of sclerite L4K missing (SG2.2.: Right-ventral part of L4K present) (43) Process nla absent (SG2.: nla present) (44) Right insertion of muscle l2 on top of pouch pne (BM: Right insertion of l2 in left wall of pne) (45) Left insertion of muscle l2 on anterior left edge of left complex, on sclerite L4K and region L4l (BM: Left insertion of l2 in posterior two thirds of left edge of left complex, on region L4l) (46) Anterior face of pouch pne and sclerite L1 plateau-like (BM: Anterior face of pouch pne and sclerite L1 plateau-like (BM: Anterior face of pouch pne and sclerite L1 plateau-like (SG2.: s2 moderately reduced; BM: s2 not reduced and as strong as s1).

Subgroup 2.2.2.2: (48) Sclerite L8 present (BM: L8 absent) (49) Sclerite L7 present (BM: L7 absent) (50) Muscle 111 present (BM: 111 absent) (51) Muscle 112 present (BM: 112 absent) (52) Muscle s12 present (BM: s12 absent) (53) Region L4a bearing dorsal s3-insertion present (BM: L4a absent, dorsal s3-insertion on membrane) (54) Left insertion of muscle 12 on anterior left edge of left complex, anterior to sclerite L4K and region L4l (SG2.2.2.: Left insertion of 12 on anterior left edge of left complex, on L4K and L4l; BM: Left insertion of 12 in posterior two thirds of left edge of left complex, on L4l) (55) Pouch lve almost reaching left edge of left complex (BM: lve by far not reaching left edge of left complex) (56) Muscle s7 absent (SG2.2.: s2 extremely reduced; SG2.: s2 moderately reduced; BM: s2 not reduced and as strong as s1).

Subgroup 2.2.2.2.: (57) (Dorsal part of) Sclerite L4K in posteroventral part of hla-base (SG2.2.2.: L4K in dorsal part of hla-base; SG2.2.: Dorsal part of L4K left-dorsal to hla-base) (58) Muscle 114 absent (SG2.: 114 present) (59) Region L4x bearing left l2-insertion present (BM: L4x absent) (60) Left insertion of muscle 12 in left anterior ventral wall of left complex, on region L4x (SG2.2.2.: Left insertion of l2 on anterior left edge of left complex, anterior to L4K and L41; SG2.2.2.: Left insertion of l2 in posterior two thirds of left edge of left complex, on L4K and L41; BM: Left insertion of l2 in posterior two thirds of left edge of left complex, on L4K and L41; BM: Left insertion of l2 in posterior two thirds of left edge of left complex, on L4H (61) Lobe lba present (BM: lba absent) (62) Sclerites R2

and R3 fused, articulation A7 absent (SG2.: R2 and R3 separated, articulated in A7) (63) Regions R1t and R1c reaching far posteriad and probably fused with parts of the broadly interconnected regions R1d and R1v: articulations A8 and A9 and membranous area 17 absent; sclerite R1M (SG2.2.: R1t and R1c restricted to a more anterior area and separated from the broadly interconnected R1d and R1v: A8, A9, and 17 present; SG2.: R1t and R1c restricted to a more anterior area and separated from the narrowly interconnected R1d and R1v: A8, A9, and 17 present; BM: R1t restricted to a more anterior area and not connected with R1d or R1v; R1c restricted to a more anterior area ventrally but possibly extending more posteriad dorsally, connected with both R1d and R1v which are separated from each other: A8 and A9 absent but 17 present) (64)! Muscle r3 absent (BM: r3 present) (65) Ridge pva longitudinally orientated (BM: pva transversely or obliquely orientated) (66) Sclerite R2 occupying left-ventral and anterior walls of invagination cbe, broadly connected with R1t (SG2.: R2 restricted to left-ventral base of cbe, separated from R1t) (67) Articulation A6 absent (SG2.: A6 present).

Subgroup 2.2.3.: (68) Membranous basal part 30 of hook hla very extensive, hla very deeply retractable (SGs2.2.1.+2.2.3.: 30 extensive, hla rather deeply retractable; SG2.: 30 very narrow, hla hardly retractable) (69) Base of hook hla at left posterior edge of left complex (SGs2.2.1.+2.2.3.: Base of hla in middle part of left ventral wall; SG2.: Base of hla in anterior left ventral wall) (70) Infolding fpe present (BM: fpe absent) (71) lveapodeme present (BM: lve-apodeme absent) (72) Common sclerotisation of processes paa and pda with stout basal ring (BM: Common sclerotisation of paa and pda not with stout basal ring) (73)! Tendon tre and muscles s8 and b4a,b absent (SG2.: tre, s8, and b4a,b present; BM: presence of tre and s8 unclear, b4a,b present) (74) Region R1c fused with the broadly interconnected regions R1d and R1v: articulations A8 and A9 and membranous area 17 absent; sclerite R1N (SG2.2.: R1c separated from the broadly interconnected R1d and R1v: A8, A9, and 17 present; SG2.: R1c separated from the narrowly interconnected R1d and R1v: A8, A9, and 17 present; BM: R1c connected with both R1d and R1v which are separated from each other: A8 and A9 absent but 17 present) (64)! Muscle r3 absent (BM: r3 present) (75) Groove rge absent (SG2.: rge present) (76) Hook-curvature on median end of region R1t present (BM: Hook-curvature on R1t absent) (77) Muscle s1 absent (BM: s1 present) (78) Ventral insertion of muscle 15 on region L4n near process nla // Muscle 15 present, not homologous with 15 of other subgroups (SG2: Ventral insertion of 15 not on L4n // This 15 absent).

Subgroup 2.2.3.2.: (79) Anterior insertion of muscle 114 on region L2a (SG2.: Anterior insertion of 114 on L4n) (80) Lobe vla small (BM: vla large) (81) Process via present (BM: via absent) (82) Right posterior dorsal part of left complex small (SG2.: Right posterior dorsal part of left complex large) (83) Sclerite L2 divided by articulation A10: sclerites L2D, L2E (BM: L2 undivided, A10 absent) (84) Sclerite L4K divided: sclerites L4U, L4V (SG2.2.: L4K undivided) (85) Region L4v absent (BM: L4v present) (86) Right insertion of muscle 12 on basal part 30 of hook hla (BM: Right insertion of 12 in left wall of pouch pne on L1) (87) Muscle 130 present (BM: 130 absent) (47)! Muscle s2 absent (SG2.2.: s2 extremely reduced; SG2.: s2 moderately reduced; BM: s2 not reduced and as strong as s1) (88) Muscles s5 and s6 divided into s5a,b and s6a,b (BM: s5 and s6

undivided) (89) Insertions of muscles p4 median to lateral margin of subgenital plate (BM: Insertions of p4 on lateral margin of subgenital plate).

Subgroup 2.2.3.2.2.: (90) Groove hge and notch 45 on hook hla present (SG2.: hge and 45 absent) (91) Tendon ate very long and narrow, including region L4n or the membranous area corresponding to L4n (SG2.2.3.: tendon ate short and broad, not including L4n) (92) Region L4n = sclerite L4V very small or absent, not forming a process nla // Region L4n and process nla absent, new sclerite L4V present (SG2.: L4n large, forming a process nla // L4n and nla present, sclerite L4V absent) (93) Process via unbranched, subordinate processes paa and pda not distinct (SG2.2.3.2.: via branching into distinct paa and pda; BM: via absent, paa and pda distinct) (94) Right posterior branch of sclerite L2 or L2D = right arm of L2-arch absent (BM: Right posterior branch of L2 = right arm of L2-arch absent (BM: L4d present) (96) Extension 28 on sclerite L2 absent (SG2.2.3.: 28 present; BM: 28 or sclerite L5 possibly present).

Subgroup 2.2.3.2.2., or 2.2.3.2.2.2., or 2.2.3.2.2.2.: (97) That part of apodeme age along posterior right margin of sclerite R3 absent, age by far not reaching articulation A3 (BM: That part of age along posterior right margin of R3 present, age reaching A3) (98) Muscle 114 divided into two bundles 114a and b (SG2.: 114 undivided) (99) Muscle 13 absent (BM: 13 present) (100) Muscle 136 present (BM: 136 absent) (101) Muscle 137 present (BM: 137 absent) (102) Muscle 138 present (BM: 138 absent) (103) Muscle 15 absent // Muscle 15 integrated into muscle l6b (BM or SG2.2.3.: 15 present // 15 not integrated into l6b) (104) Muscle l6a very large (SG2.: l6a of moderate size) (105) Muscle s3 divided into two bundles s3a and b (BM: s3 undivided) (106) Muscle s14 present (BM: s14 absent) (107) Dorsal insertion of muscle s6b on entire right margin of sclerite R3 (SG2.2.3.2.: Dorsal insertion of s6b restricted to anterior right margin of R3; SG2.2.: Dorsal insertion of s6 restricted to anterior right margin of R3; BM: Dorsal insertion of s6 in ventral wall of genital pouch, possibly extending to anterior margin of R3) (108) Ventral insertions of muscles s5a and, less distinctly, s6a more posteriorly (SG2.2.3.2.: Ventral insertions of s5a and s6a more anteriorly; BM: Ventral insertions of s5 and s6 more anteriorly) (109)! Sclerite L1 absent (BM: L1 present) (110)! Process(es) dca absent (SG2.: dca present) (25)! Muscle 11 absent (BM: 11 present) (111) Pouch pne absent (BM: pne present).

Subgroup 2.2.3.2.2.: (112) Median ends of region R1t and sclerite R2 connected: articulation A6 absent (SG2.: Median ends of R1t and R2 separated and articulated in A6) (113) Thickening cwe present (BM: cwe absent) (114) Region R1t separated from region R1c: sclerites R1P, R1S (BM: R1t connected with R1c).

Subgroup 2.2.3.2.2.2.: (115) Lobe dla present (BM: dla absent) (116) Tendon tve present (BM: tve absent) (117) Females: Advanced rotation of ootheca present (BM: Advanced rotation absent).

Subgroup 2.2.3.2.2.2.2.: (118) Sclerite R5 present (BM: R5 absent) (119) Sclerite R4 present (BM: R4 absent) (120) Muscle r11 present (BM: r11 absent).

Subgroup 2.2.3.2.2.2.2.: (121) Region R1t connected with region R1c: sclerite R1T (SG2.2.3.2.2.2.: R1t separated from R1c; BM: R1t connected with R1c).

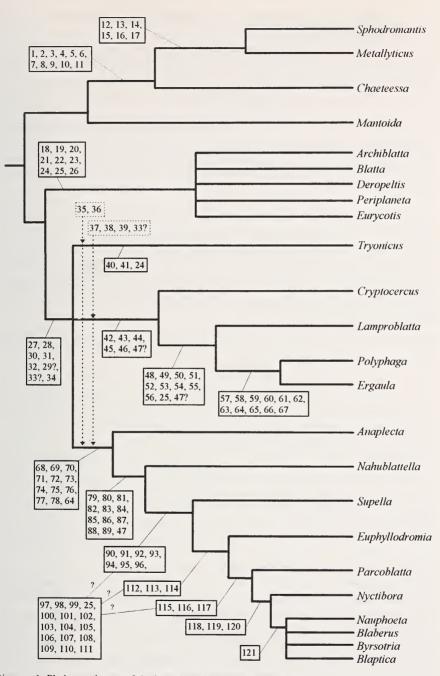


Diagram 1: Phylogenetic tree of the investigated representatives of Blattaria and Mantodea, with the assumed aut/synapomorphies

7.5. Remarks on the polarity and evolution of some characters

For some characters the polarity assumptions in 7.1.-7.4. are not yet sufficiently substantiated. In some cases the polarity question can be settled by a detailed discussion of morphology, homology, or functional intercorrelations. The respective discussions will be largely independent of the phylogenetic hypothesis in 7.4. In other cases a solution of the polarity question can only be approached by a reciprocal consideration of the various arguments or possibilities in terms of parsimony, i.e. a weighing of the various possible polarity assumptions against the assumed autapomorphies of the subgroups defined in 7.2. and 7.3. and against the outgroup comparison between Blattaria and Mantodea. The respective discussions will be done in interdependence with the phylogenetic hypothesis in 7.4.

The following discussions under (A)-(C) are concerned with the polarity of three characters for which the outgroup comparison between Blattaria and Mantodea is somewhat conflicting since the same two character states are present in Blattaria as well as in Mantodea. The question arises whether **that** character state represents the ground-plan condition which in 7.1. has been assumed to do this. (A similar conflict is also present in (G), which will be discussed below.) These discussions will be independent of the phylogenetic hypothesis in 7.4.

(A) The connection or separation of the sclerotisations L2d and L4l of the processes paa and pda and the length of paa and pda

The area bearing the **paa**- and **pda**-processes is very similar in *Mantoida* (fig.44) and *Tryonicus* (fig.91): The sclerotisations of **paa** (**L2d**-region) and **pda** (**L4l**-region) are connected; **paa** and **pda** are completely sclerotised, are bulge-like and short, **paa** being somewhat upcurved, and are close to each other. This has been regarded as the condition of the common ground-plan of Blattaria and Mantodea (6.2.1., 7.1.).

In other Blattaria (6.2.1., 6.2.4.) as well as in other Mantodea (6.2.3.) paa and pda can be longer, and their sclerotisations can be separated from each other. Alternatively, these two states could be regarded as the ground-plan states of the respective characters, but there are some arguments against this view: Within Blattaria the sclerotisations are separated in Eurycotis (fig.66), Archiblatta (fig.53), and Cryptocercus (fig.150). This separation is accompanied (1) by a reduction of the sclerotisation of one of the processes in Cryptocercus (pda) and in Archiblatta (paa), and (2) by a far separation of the processes themselves in *Eurycotis* and *Archiblatta*, which feature is correlated with the recess of the posterior part of the lve-pouch to the right (6.2.1.). These accompanying conditions in Eurycotis, Archiblatta, and Cryptocercus are clearly derived features, and the separation of the **paa**- and **pda**-sclerotisations can easily be explained as correlated with these and as being derived, too. In Chaeteessa (fig.34), Metallyticus (fig.25), and Sphodromantis (fig.11) the sclerotisations of **paa** and **pda** are separated from each other without being reduced (with the exception that in *Chaeteessa* the whole **pda** has been lost), and the close vicinity of **paa** and **pda** has been retained. According to these very different concomitant circumstances in Cryptocercus, in Archiblatta + Eurycotis, and in the respective Mantodea, the separation of the sclerotisations of **paa** and **pda** is clearly suggested to have been achieved three times independently. As regards the length of **paa** and **pda**, there are no peculiar similarities in the shape of **paa** and **pda** in the Blattaria and Mantodea with these processes being long. On the other hand, both **paa** and **pda** of *Tryonicus* and *Mantoida* are rather similar, and in my view it is the most parsimonious solution to regard this similarity as homologous and as representing the ground-plan condition.

(B) Sclerite L2 arch-shaped or plate-like

In *Mantoida, Archiblatta*, and *Polyphaga* L2 extends like an arch along the margins of the **lve**-pouch, and this has been regarded as the condition of the common ground-plan of Blattaria and Mantodea (6.2.1., 7.1.; fig.324d,f,k). A plate-like L2, with the arms of the arch (regions L2p and L2m) probably fused to each other, is present in *Chaeteessa, Metallyticus*, and *Sphodromantis* (6.2.3.), but also in *Eurycotis* (6.2.1.) and, more or less, in *Cryptocercus* (6.2.4.) (fig.324a,b,c,e,h). However, in *Cryptocercus* the indistinctness of the arch-shape of L2 is due to a reduction of the right part of the L2-arch (L2m-region, area of articulation A2) and to a broadening of the L2a-region; vestiges of the arch-shape are still recognisable in this L2. *Eurycotis* has the posterior left-ventral part of L2 extremely reduced. In the respective Mantodea L2 is only narrowed, with none of its parts reduced. Thus, the morphology of L2 and the concomitant circumstances of its plate-like condition are very different in *Eurycotis*, in *Cryptocercus*, and in the respective Mantodea, and this supports the assumption that the plate-like shape of L2 has arisen several times by parallel evolution. Additionally, the plate-like condition is in my opinion more liable to homoplasy than the more complicated arch-shape.

(C) Sclerite L5 present or absent and in various positions

As discussed in 6.5., the presence of L5 – somewhere in the dorsal wall of the vla-lobe – might be the plesiomorphic state for Blattaria or even Blattaria + Mantodea, but a definite decision is not possible. The extension 28 (a ventral part of sclerite L2) of *Anaplecta* and *Nahublattella* is possibly homologous with L5 (6.5.). The position of L5 or 28 is very similar in *Polyphaga, Anaplecta*, and *Nahublattella*, but since it is not clear which position of L5 or 28 has to be regarded as primitive, this similarity could also represent the plesiomorphic state of Blattaria or Blattaria + Mantodea. At the present state of knowledge, the characters of these sclerotisations are not yet suitable for a phylogenetic analysis since the polarity of their states remain uncertain.

The following discussions will be concerned with the polarity of some characters, termed (**D**)-(**K**) and (**L**), for which the reciprocal outgroup comparison between Blattaria and Mantodea suggests another polarity as it has been assumed in 7.1.-7.4. (or, in the case of (**G**), this outgroup comparison is conflicting in the same way as in the characters discussed under (**A**)-(**C**)). The question is whether some features present in some Blattaria but not in the other Blattaria and in Mantodea are either ground-plan features of Blattaria as stated in 7.3. or autapomorphies of Blattarian subgroups as it is suggested by the outgroup comparison. If the polarity assumptions are based on the latter alternative, then (1) the distribution of the states of the characters (**D**)-(**K**) would suggest a phylogenetic hypothesis

which is roughly the reverse of the hypothesis in 7.4., and (2) the distribution of the states of character (**L**) would suggest a grouping which is inconsistent with the phylogenetic hypothesis in 7.4. as well as with the hypothesis discussed in the context of the characters (**D**)-(**K**). These possibilities have to be tested.

(D) The presence or absence of the curvature of the right parts of sclerite L2 and of the lve-pouch

That the right parts of **L2** and of **lve** curve dorsad and back to the left (6.2.1., 6.2.4.) has been assumed to be a ground-plan feature of Blattaria. This curvature is present in *Archiblatta, Eurycotis, Tryonicus, Lamproblatta, Polyphaga, Ergaula*, and *Anaplecta* but absent in the remaining Blattaria (*Cryptocercus* and subgroup 2.2.3.2.) and in Mantodea.

(E) The presence or absence of the nla-process

The **nla**-process (6.3.1., 6.3.4.) has been assumed to be a ground-plan element of Blattaria. **nla** is present in *Eurycotis, Archiblatta, Tryonicus, Anaplecta*, and probably *Nahublattella* but absent in the remaining Blattaria (subgroups 2.2.2. and 2.2.3.2.2.) and in Mantodea.

(F) The presence or absence of the dca-processes

The **dca**-processes (6.1.1., 6.1.4.) have been assumed to be ground-plan elements of Blattaria. **dca** are present in *Eurycotis, Archiblatta, Tryonicus, Cryptocercus, Polyphaga, Ergaula, Nahublattella*, and possibly *Lamproblatta* but absent in the remaining Blattaria (*Anaplecta* and subgroup 2.2.3.2.2.) and in Mantodea.

(G) The connection or separation of the regions R1t and R1c

In most Blattaria and in the Mantodea *Chaeteessa* and *Sphodromantis* the regions **R1t** and **R1c** are firmly connected, and several Blattaria and *Chaeteessa* have a distinct edge **16**, which has been defined as the border between **R1t** and **R1c**. This has been regarded as the condition of the common ground-plan of Blattaria and Mantodea (6.7.1., 7.1.). In most Blattaria of subgroup 2.2.3.2.2.2. (6.7.6.) as well as in the Mantodea *Mantoida* and *Metallyticus* (6.7.3.) **R1t** and **R1c** are separated from each other.

(H) The connection or separation of the regions R1c, R1d, and R1v (= the absence or presence of the articulations A8 and A9)

That **R1d** as well as **R1v** are separated from **R1c** by the articulations **A8** and **A9** (6.7.1., 6.7.6.), respectively, has been regarded as a ground-plan feature of Blattaria (7.3.). This separation is present in *Eurycotis, Archiblatta, Tryonicus, Lamproblatta*, and *Cryptocercus*, but in the Blattarian subgroups 2.2.2.2.2. and 2.2.3. (6.7.6.) and in the ground-plan of Mantodea (6.7.1.) the regions **R1c**, **R1d**, and **R1v** are contained in one sclerite.

(I) The presence or absence of the **tre**-tendon and of muscle **s8**

The **tre**-tendon (6.7.1., 6.7.5.) and the **s8**-muscle (6.9.) have been assumed to be groundplan elements of Blattaria. **tre** and **s8** are present in *Eurycotis, Archiblatta, Tryonicus, Cryptocercus, Polyphaga*, and *Ergaula* (**s8** not studied in *Archiblatta* and *Tryonicus*) but absent in the remaining Blattaria (*Lamproblatta* and subgroup 2.2.3.) and in Mantodea.

(J) The presence or absence of the rge-groove

The **rge**-groove (6.7.1., 6.7.6.) has been assumed to be a ground-plan element of Blattaria. **rge** is present in *Archiblatta, Eurycotis, Tryonicus, Cryptocercus, Lamproblatta, Polyphaga*, and *Ergaula* but absent in the remaining Blattaria (subgroup 2.2.3.) and in Mantodea.

(K) The presence or absence of muscle r6

The **r6**-muscle (6.7.1., 6.7.6.) has been assumed to be a ground-plan element of Blattaria. **r6** is present in *Eurycotis, Lamproblatta, Polyphaga*, and *Ergaula* but absent in the remaining Blattaria (*Cryptocercus* and subgroup 2.2.3.) and in Mantodea. (*Archiblatta* and *Tryonicus* not investigated.)

A comparison between Blattaria and their outgroup Mantodea could hence lead to the view that in the ground-plan of Blattaria **nla**, **dca**, **rge**, **tre**, **s8**, **r6**, and the curvature of **L2** and **lve** are missing; **R1t** is separated from **R1c**, and **R1c** is connected with both **R1v** and **R1d** (no articulations **A8** and **A9**). The counterparts of these character states would then have developed within Blattaria and would have to be regarded as possible autapomorphies of Blattarian subgroups. From this view the following phylogenetic hypothesis could arise:

- 1. Part of subgroup 2.2.3.2.2. (*Euphyllodromia* + *Parcoblatta* + *Nyctibora*) is the sistergroup of the other Blattaria. All remaining Blattaria have connected **R1t** and **R1c**.
- 2. The next offshoots are Supella and Blaberus + Nauphoeta + Blaptica + Byrsotria. All remaining Blattaria have developed the nla-process (lost again in Lamproblatta, Ergaula, Polyphaga, and Cryptocercus) and the dca-processes (lost again in Anaplecta), and they have also developed, for example, sclerite L1 and region L4d (both lost again in Anaplecta), the posterior branching of sclerite L2 (the first hint of the arch-shape), and the division of the via-process into paa and pda.
- 3. The next offshoot is *Nahublattella*. All remaining Blattaria have developed the curvature of L2 and lve (lost again in *Cryptocercus*), and they have also translocated the anterior insertion of 114 from the lve-apodeme to nla (L4n-region).
- The next offshoot is *Anaplecta*. All the remaining Blattaria have developed the rgegroove, and they have also lost the lve-apodeme and developed the contact between L1 and L2 (articulation A2).
- 5. The next dichotomy would be questionable: *Polyphaga* + *Ergaula* could be the next offshoot, with the remaining species having separated **R1c+R1t** and **R1d+R1v** (new articulations **A8** and **A9**). Alternatively, *Lamproblatta* could be the next offshoot, with the remaining species having developed the **tre**-tendon and the **s8**-muscle. Alternatively, *Cryptocercus* could be the next offshoot, with the remaining species having developed the **tre**-tendon and the **s8**-muscle. Alternatively, *Cryptocercus* could be the next offshoot, with the remaining species having developed the **r6**-muscle.
- 6. *Eurycotis* + *Archiblatta* and *Tryonicus* would represent a holophyletic group which has, for example, developed a close contact between the anterior and posterior parts of the L4I-region and rotated the L4d-region anteriad (from a formerly leftward orientation).
- 7. In Eurycotis + Archiblatta the anterior and the posterior parts of the L4I-region have

fused, L4l has developed an swe-apodeme, and the right phallomere has evolved the **pia**-tooth.

This phylogenetic hypothesis as a whole is for several reasons extremely improbable: (1) If accepting it, one would have to suppose that many of the features assumed for the common ground-plan of Blattaria and Mantodea (listed in 7.1.) are not ground-plan features but similarities born by parallel evolution. Some of the most important have been mentioned in the steps 1.-7. Concerned are, for example: the detailed similarities of the L4I- and L4d-regions and of the swe-apodeme in Mantoida and Archiblatta; the archshape of L2 and articulation A2 in e.g. Mantoida, Archiblatta, and Polyphaga; the similar morphology of **paa** and **pda** in *Mantoida* and *Tryonicus*; the **pia**-teeth of Mantodea and e.g. Archiblatta. (2) In addition, some elements present in subgroup 2.2.3. or its subordinate subgroups but not in the other Blattaria and in Mantodea would have to be regarded as ground-plan elements of Blattaria, e.g. the hook-like curvature of the median end of the R1t-region, inclusive of its cwe-thickening, and the lve-apodeme. Features of the females could be added to this "ground-plan" list, e.g. the advanced rotation of the ootheca. Hence, as regards (1) and (2), this hypothesis would be extremely conflicting with the outgroup comparison of Blattaria with Mantodea – much more than the hypothesis proposed in 7.4. (3) This hypothesis is rather inconsistent in itself: In several cases secondary reductions (e.g. nla-process in 2.) or parallel evolution (situation in 5.) have to be assumed. If the developments contained in the steps 1.-5, are arranged in another way, some of these assumptions could be avoided, but they would only unavoidably be replaced by other assumptions of secondary loss or parallel evolution.

If only some or even only one of the polarity statements of this alternative hypothesis are accepted, this would still cause extensive inconsistencies either with the ground-plan hypothesis given in 7.1. or with the clusters of assumed autapomorphies given in 7.4. If it is, for example, supposed that rge(J) is an autapomorphy of a Blattarian subgroup comprising Archiblatta, Eurycotis (and the other species assigned to subgroup 2.1.), Tryonicus, Lamproblatta, Cryptocercus, Polyphaga, and Ergaula, and that the lack of rge in subgroup 2.2.3. is the plesiomorphic state, one would have to accept many cases of parallel evolution. Which features would have to be regarded as having evolved several times depends on which type of phallomere complex is regarded as plesiomorphic for this hypothetical grouping: (1) If the basic phallomere morphology is supposed to resemble Archiblatta, all the assumed autapomorphies of subgroup 2.2. must have developed two times independently. (2) If the basic phallomere morphology is supposed to resemble either Tryonicus, Lamproblatta, Cryptocercus, Polyphaga, or Ergaula, most of the similarities in the morphology of the L4I- and L4d-regions (including the similar insertions of I2 and 14; 6.3.1.) and in the posterior part of the right phallomere (with the fda-lobe and the piatooth; 6.7.1.), which have revealed in the comparison between Mantoida, Chaeteessa, Archiblatta, and Eurycotis, would have to be regarded as cases of parallel evolution. (3) If any combination or mixture of these types is supposed to represent the basic phallomere morphology, the extent of parallel evolution having to be accepted could not be decreased, but only the assumptions of parallel evolution necessary for (1) and (2) would mingle. If it is assumed that either tre and s8, or r6, or the L2-curvature, or nla, or dca, or the separation of R1d and R1v from R1c, or the connection of R1c and R1t is an

autapomorphy of the respective Blattarian grouping, one would likewise have to accept extensive parallel evolution – in an analogous way as described for **rge**. It is thus certainly by far most parsimonious to regard all these elements or properties as ground-plan features of Blattaria (like in 7.3.) and to assume secondary loss or change when any of these elements or properties is missing in any of the Blattarian species investigated in this paper. As regards (E) and (I), this view is additionally supported by arguments concerning the functional intercorrelation of phallomere elements. (E): compare discussion in 7.5. (M), (N). (I): The b4-muscles, which in Blattaria and Mantodea (6.7.1., 7.1.). When present all together, **tre**, **b4**, and **s8** are certainly functionally intercorrelated elements (and in this case the function of the b4-muscles is certainly different from that of the b4-muscles of *Mantoida*). If reduction occurs in such a situation, all three elements can be expected to be concerned. Hence, the lack of **b4** in *Lamproblatta* and subgroup 2.2.3. (there are no muscles in a similar position as **b4a** and **b4b** are in *Mantoida*) might indicate that **tre** and **s8** were present in former times.

(L) The presence or absence of muscle s7

s7 (6.9.) has been assumed to be a ground-plan element of Blattaria (7.3.). s7 is present in the subgroups 2.1. and 2.2.3., and vestiges are probably present in subgroup 2.2.2.1. (*Cryptocercus*). In subgroup 2.2.2.2. and in Mantodea s7 is absent. Subgroup 2.2.1. (*Tryonicus*) has not been investigated. Hence, s7 could be a synapomorphy of the subgroups 2.1. and 2.2.3. and possibly *Cryptocercus*. However, since this assumption would be inconsistent with the assumed autapomorphies of the subgroups 2.2. and 2.2.2., it is assumed that s7 has been lost secondarily in *Lamproblatta*, *Polyphaga*, and *Ergaula*. The lack of s7 in *Blaberus* (6.9.) is certainly a secondary loss since s7 is present in all other investigated species of subgroup 2.2.3. (inclusive of *Nauphoeta*).

The following discussions under $(\mathbf{M}) + (\mathbf{N})$ and (\mathbf{O}) will be concerned with the polarity of some characters of Blattaria for which an outgroup comparison with Mantodea is not possible since the respective elements (**hla**-hook or **dca**-processes) are present in all Blattaria (**hla**) or at least in the Blattarian ground-plan (**dca**, compare (**F**)) but not in Mantodea. A result can be achieved in interdependence with the phylogenetic hypothesis in 7.4., but, mainly in the case of (**M**) and (**N**), also independently of this hypothesis, if correlations with other elements for which an outgroup comparison with Mantodea is possible are considered.

(M) The position of the hla-base

(N) The extension of the membranous basal part 30 of hla

The **hla**-hook and the **L3**-sclerite are present in all Blattaria (6.4.3.). In 7.3. it has been stated that in the ground-plan of Blattaria the **hla**-base takes a position in the left anterior ventral wall of the left complex, and that the introversible membranous basal part **30** of **hla** is narrow (and **hla** is therefore – almost – non-retractable). These statements have to be substantiated.

In dependence on the phylogenetic hypothesis in 7.4. the following view results: A far anterior position of the **hla**-base and a narrow membrane **30** (and a non-retractable **hla**) are present in the subgroups 2.1. and 2.2.2.2. Thus, the more posterior position of the **hla**-base and the expansion of membrane **30** (and the retractility of **hla**) can be regarded as apomorphic states developed in the Blattarian subgroups 2.2.1. and 2.2.3. (These are possibly synapomorphies of these two subgroups: (**35**) and (**36**) in 7.4.) The more posterior position of the **hla**-base in subgroup 2.2.2.1. (*Cryptocercus*, fig.151), which is, however, not accompanied by an expansion of membrane **30** and by a retractility of **hla**, is also derived.

There are some functional intercorrelations between the various characters of **hla** and between these and some other elements of the left complex. Consequently, the species with a completely retractable **hla** (subgroup 2.2.3.) show some concomitant morphological and functional differences to the species with a non-retractable **hla**. Since for some of these correlated elements or properties an outgroup comparison with Mantodea and hence a polarisation of the respective character states is possible, these intercorrelations permit assessing the polarity of the characters of **hla** independently of the phylogenetic hypothesis in 7.4. The following intercorrelations and evolutionary changes are assumed:

- The extension of membrane 30, the retractility of hla, and the position of the hla-base are intercorrelated: If membrane 30 is more extensive, hla can be retracted more deeply, and then its base can be more posteriorly without having hla exceeding the subgenital plate or protruding from the genital pouch during its inactive state. From a functional point of view, the more posterior position of the hla-base and the retractility of hla might have the advantage that in its active state (during copulation) hla protrudes farther from the genital pouch and is more flexible. In *Cryptocercus* the more posterior position of the hla-base is not accompanied by an extension of membrane 30 but by a shortening of hla (fig.151).
- The retractility of hla, the length of 114, and the positions of the 114-insertions are intercorrelated: In the species with a non-retractable hla, muscle 114 has only the function to move hla; to accomplish this function 114 has to be contracted for a short distance only and can be rather short (fig.72). In the species retracting hla (subgroup 2.2.3.) the contraction distance of 114 has to be much longer, and thus 114 itself has to be longer. This requirement is in part fulfilled by the posteriad shift of the posterior 114-insertion together with the hla-base. Additionally, the anterior insertion of 114 has shifted anteriad. At a first stage, this latter shift has evidently been achieved by an anteriad shift of the L4n-region and of the nla-process, the primary anterior insertion area of 114; this stage is represented by Anaplecta (fig.222; compare Eurycotis, fig.68, 72, where nla and the 114-insertion are by far more posteriorly). At a later stage, in subgroup 2.2.3.2., the anterior 114-insertion has been translocated to the top of the lvepouch (lve-apodeme), and the 114-insertion is shifted further anteriad by a lengthening of this lve-apodeme (compare Anaplecta, fig.222, and Nahublattella and Parcoblatta, fig.249, 276).
- The position of the anterior l14-insertion and the condition of the swe- and lve-apodemes and of the nla-process are intercorrelated: A lot of force seems to be needed in operating

the hla-hook, and there are various supporting mechanisms present to give the contraction of 114 the proper effect, which is the movement or retraction of hla, and to prevent a distortion of the areas (1) between the anterior 114-insertion and the hla-base and (2) immediately around the anterior 114-insertion. (1) In Archiblatta and Eurycotis the swe-apodeme might fulfil the former function (fig.53, 65), and since the morphology of the L4l-region and swe is the same in Mantoida (6.3.1.), this mechanism is certainly the most primitive within Blattaria. In subgroup 2.2.3. swe is missing, and the anterior part of the lve-pouch has been elaborated as a long and stiff lve-apodeme. In Anaplecta (subgroup 2.2.3.1.) the anterior l14-insertion on nla is stabilised by nla being firmly rested upon the top of the lve-apodeme (fig.209, 210). The stabilisation of nla might be the primary function of the lve-apodeme. In the further evolution (in subgroup 2.2.3.2.) the anterior **114**-insertion has shifted even further anteriad, and it has apparently been preferred to make more directly use of the supporting function of the lve-apodeme and to translocate the l14-insertion to the top of lve. (2) The nla-process itself assumedly has the function to stiffen the immediate vicinity of the anterior 114-insertion – as long as this insertion is on the L4n-region - and is therefore bulge-like (Archiblatta, Eurycotis, Anaplecta). In species having this insertion translocated to the lve-apodeme, nla is either modified in its shape (Nahublattella: whip-like) or has been lost (Parcoblatta, Blaberus).

The position of the anterior 114-insertion and the condition of the phallomero-sternal muscles s1 and s7 are intercorrelated: Additional stabilisation preventing a distortion of the area around the anterior 114-insertion is probably achieved by muscles conducting much of the force which 114 exerts to this area to the left apophysis of the subgenital plate. These phallomero-sternal muscles insert immediately anterior to the anterior 114-insertion. In *Eurycotis* this function is accomplished by muscle s1, which inserts between nla and the anterior end of swe (fig.70). In the species of subgroup 2.2.3., where the supporting function has been transferred from swe to lve and where, except for *Anaplecta*, the muscle-insertion for which the support is needed (114) has been transferred from nla to lve, the function of the "conductor"-muscle has consequently been transferred from s1 to s7, which inserts anteriorly on the lve-apodeme. Muscle s1 has been lost in all species using a lve-apodeme for support (subgroup 2.2.3.; 6.9.).

The reciprocal outgroup comparison between Blattaria and Mantodea clearly suggests that **swe** (6.3.1.) and **s1** (6.9.) are ground-plan elements, that the primitive position of the **L4n**-region is like in *Eurycotis* (6.3.1.; compare fig.325c and e), and that there is no tube-shaped **lve**-apodeme. The lack of **swe** and **s1**, the extremely far anterior position of the **L4n**-region (or its lack), and the **lve**-apodeme – the features of subgroup 2.2.3. – are certainly derivations within Blattaria. Especially the assumedly primitive position of the **L4n**-region (like in *Eurycotis*) is clearly correlated with a short **114** inserted on **L4n**, with the non-retractility of **hla**, and with a narrow basal membrane **30**, and a non-retractable **hla** can be expected to have its base far anteriorly. Thus, the non-retractable **hla** with its base in the anterior ventral wall can be assumed to represent the plesiomorphic condition within Blattaria. A bulge-shaped **nla**-process can also be regarded as a ground-plan element of Blattaria, since it is present in all species showing the primitive condition that the anterior **114**-insertion is on a well-developed **L4n**-region.

In subgroup 2.2.2. the anterior **114**-insertion (**114** present in *Lamproblatta* and *Cryptocercus* only) is not stabilised by cuticular elements (no **swe**- or **Ive**-apodemes, **L4n**-region highly reduced, no **nla**-process), and the force acting on the anterior **114**-insertion seems to be completely conducted to the subgenital plate by the **s1**-muscles, which insert immediately anterior to **114** (fig.157, 158, 184, 185). Also in subgroup 2.2.1. (*Tryonicus*) neither an **swe**- nor an **Ive**-apodeme are present, and the stabilisation by cuticular elements does not seem to be very effective. The mechanism cannot be assessed here since the muscles are not known.

(O) The shape and sclerotisation of the dca-processes

dca-processes are restricted to Blattaria (6.1.1.) and are probably ground-plan elements of this taxon (7.5. (F)). In 7.3. it has been stated that in the Blattarian ground-plan the dca are two cushion-like processes posterior to L1; this statement has to be substantiated. Two membranous cushion-like dca are present in representatives of both of the basic Blattarian subgroups 2.1. (*Archiblatta*, fig.54) and 2.2. (*Tryonicus angustus*, fig.107, *Cryptocercus*, fig.153, *Polyphaga*, fig.120). More or less completely sclerotised (by L1) dca are also present in both the subgroups 2.1. (*Eurycotis*, fig.67) and 2.2. (*Tryonicus parvus*, fig.94, *Nahublattella*, fig.243, 244), but since the L1m-region is a ribbon-like extension in the common ground-plan of Blattaria and Mantodea, and since L1m is expanded posteriad in the species with sclerotised dca-processes (to contribute to this sclerotisation), this is assumed to be a derived state.

In those species of subgroup 2.2. having two membranous **dca**, these **dca** are very similar (fig.107, 120, 153; these are the members of the subgroups 2.2.1. and 2.2.2. except *Ergaula*, fig.105, and *Lamproblatta*, fig.177, but no member of subgroup 2.2.3. is concerned). This peculiar shape of the **dca** is assumed to represent the plesiomorphic state within subgroup 2.2. It is not regarded as a synapomorphy of the respective species since this assumption would be inconsistent with the many assumed autapomorphies of the subgroups 2.2.1. and 2.2.2.2. It also cannot be regarded as a synapomorphy of the subgroups 2.2.1. and 2.2.2. (assuming a secondary change in *Lamproblatta* and, less drastic, in *Ergaula*) since the shape of the **dca** in their primitive membranous condition is not assessable in subgroup 2.2.3.: Here the **dca** are either completely sclerotised (*Nahublattella*) or missing (remaining species), situations which are both derived.

A sclerotised peak **18** in between the **dca**-processes is present only in *Tryonicus angustus* and *Cryptocercus* (fig.107, 153). To regard this as a synapomorphy (assuming secondary loss in *Tryonicus parvus*) would be inconsistent with the autapomorphies of subgroup 2.2.2. only, but in my view the latter are much more conclusive. The peaks **18** are possibly non-homologous in the two species, or they are again an element of the ground-plan of subgroup 2.2.

The following discussions under (P) and (Q) + (R) will be concerned with the evolution of three characters for which the polarity is essentially clear, but for which the distribution of the character states within Blattaria is somewhat in conflict with the phylogenetic hypothesis in 7.4.; reversals of apomorphic character states seem to have taken place.

These discussions will be dependent on the phylogenetic hypothesis in 7.4., and the soundness of the results depends on the soundness of this hypothesis.

(P) The presence or absence of a sclerite-ring formed by the regions L1m, L1l, and L1r A distinct extension L1m and possibly also a less distinct extension L1l are elements of the common ground-plan of Blattaria and Mantodea (6.1.1.). However, only in species of the Blattarian subgroup 2.2. L1l and L1m curve ventrad (L1m does this in a different way as in the Mantodean subgroup 1.2.) and approach each other again (6.1.4.); this is certainly a derived feature. The sclerite ring is either complete (*Ergaula, Cryptocercus*), and in this case it sometimes additionally expands onto the dca-processes (*Tryonicus parvus, Nahublattella*), or the ring has a short gap (ventrally in *Polyphaga*, dorsally in *Tryonicus angustus*). In all other species of subgroup 2.2. the feature is not assessable since L1 has been completely lost. The only exception is *Lamproblatta*, which shows no trace of a ring though L1 is present; this is assumed to be a secondary loss because of the many assumed autapomorphies of the subgroups 2.2.2. and 2.2.2.2. A complete sclerite-ring is assumed to be a ground-plan feature of subgroup 2.2.

(Q) The connection or separation of the sclerotisations of the lve-pouch (L2 or L2D) and of the **paa-** and **pda-** (or **via-**) processes (L2+L4N or L2E+L4N)

(R) The absence or presence of muscle 110

In the common ground-plan of Blattaria and Mantodea the sclerotisation of the **lve**-pouch is connected with the common sclerotisation of the **paa**- and **pda**-processes (6.2.1., 7.1.), and the same is true of the ground- plan of the Blattarian subgroup 2.2. Muscle **110** connecting these two sclerotisations is certainly also a ground-plan element of subgroup 2.2. (or at least of 2.2.2. + 2.2.3. since *Tryonicus* = 2.2.1. has not been investigated; 7.3.). Within subgroup 2.2.3. (fig.328) the sclerotisations concerned can be connected with (*Anaplecta, Euphyllodromia, Parcoblatta, Blaberus*) or separated from each other (*Nahublattella, Supella, Nyctibora, Nauphoeta*), and muscle **110** is present in the species showing a connection (except for *Anaplecta*).

Anaplecta is the first offshoot within subgroup 2.2.3. The outgroup comparison with the subgroups 2.2.1. and 2.2.2. clearly suggests that the connection of the sclerotisations in *Anaplecta* corresponds to the plesiomorphic state. *Nahublattella* is the next offshoot, and *Supella* follows. Both species show an (probably homologous) apomorphic separation of these sclerotisations. In the remaining species, which together form subgroup 2.2.3.2.2.2., both conditions – connection and separation – are represented. The outgroup comparison with the subgroups 2.2.3.2.1. (*Nahublattella*) and 2.2.3.2.2.1. (*Supella*) clearly suggests that within subgroup 2.2.3.2.2.2. the connected condition is an apomorphic state. This view is supported by the additional lack of **110**, which is certainly a derived feature, and which is probably correlated with this secondary fusion of the sclerotisations.

The fusion of the sclerotisations and the concomitant loss of **110** have certainly happened several times within subgroup 2.2.3.2.2.2. since the distribution of these two derived features is completely inconsistent with some other well-founded groupings: (1) *Blaberus*

and *Nauphoeta* are (together with *Blaptica* and *Byrsotria*) members of the certainly holophyletic taxon Blaberidae (McKittrick 1964). (2) *Parcoblatta, Nyctibora, Blaberus*, and *Nauphoeta* are (together with *Blaptica* and *Byrsotria*) members of a probably holophyletic taxon characterised by the autapomorphies (115)-(117) in 7.4. Hence, the fusion and the loss of 110 can be assumed to have been achieved independently in *Euphyllodromia, Parcoblatta*, and *Blaberus*.

7.6. Conflicts in the distribution of character states

In some characters whose polarity is rather clear, the distribution of the apomorphic state(s) over the taxa is inconsistent with the phylogenetic hypothesis in 7.4.; these inconsistencies can only be resolved by the assumption of either parallel evolution in the one or reversal in the other species. In some other characters, mainly in those discussed in 7.5., the polarity might be supposed to be the reverse of that assumed in 7.4. or resulting from the discussions in 7.5. The discussions have shown that this is highly improbable, but some doubt may remain. If the reverse polarity is supposed, the distribution of the surmisedly apomorphic character states would also be inconsistent with the hypothesis in 7.4.

The (clearly or possibly) apomorphic character states concerned and the groupings they might support will be listed here, and the respective plesiomorphic states are given in brackets. This will be done in the same manner as in 7.4. The conclusiveness of many of these "apomorphic" states is in terms of their value as possible autapomorphies decreased by uncertain homology relations, uncertain or even improbable polarity assumptions, or other circumstances. Some of these states, however, could really be autapomorphies and could hazard the phylogenetic hypothesis in 7.4., but only in very few points.

In the species provided with "?" some or all of the respective characters have not been investigated. For muscles the two insertion areas are given.

Arguments for alternative groupings within Mantodea

Grouping A: *Metallyticus* + (*Mantoida* + *Sphodromantis*)

(122) Groove on region R1c behind articulation A3 and anterodorsal to right insertion of muscle r3 present (BM: Groove on R1c behind A3 and anterodorsal to left r3-insertion absent).

This groove on the **R1c**-region (fig.6, 20, 41), not homologous with the **rge**-groove of Blattaria (6.7.1.), is very distinct in *Mantoida* and *Sphodromantis*, hardly recognisable in *Metallyticus*, and missing in *Chaeteessa* (6.7.3.). To interpret the groove as an autapomorphy of this grouping would be in conflict with the many assumed autapomorphies of the subgroups 1.2. and, since the groove is more distinct in *Mantoida* and *Sphodromantis*, 1.2.2. It is assumed that this groove is a ground-plan element of Mantodea and has been reduced in *Metallyticus* and lost in *Chaeteessa*. The following fact supports this interpretation: The **age**-apodeme on **R3** probably reaches articulation **A3** in the common ground-plan of Blattaria and Mantodea (6.7.1., 7.1.), but in *Chaeteessa* **age** does not reach **A3**, and the right posterior part of **age** has evidently been reduced. A concomitant reduction of the groove posterior to **A3** would be plausible.

Grouping B: Mantoida + Metallyticus

(123) Region R1t separated from region R1c: sclerites R1D, R1E or R1D, R1C (BM: R1t connected with R1c).

To regard this certainly apomorphic state (6.7.1., 6.7.3.; compare (G) in 7.5.) as a synapomorphy of *Metallyticus* and *Mantoida* would be in conflict with the assumed autapomorphies of the Mantodean subgroups 1.2. and 1.2.2. Parallelism is thus highly suggested. Support for this assumption comes from the fact that some other taxa have also achieved this state independently: According to LaGreca (1955), at least *Amorphoscelis abyssinica* (Amorphoscelididae), *Tarachodes insidiator* (Mantidae), and *Polyspilota* sp. (Mantidae) (fig.13, 15, 16 in LaGreca) show the separation of **R1t** from **R1c**, and this is certainly not a synapomorphy of these distantly related species. The Blattarian subgroup 2.2.3.2.2.2. also shows the separation of **R1t** and **R1c**.

Arguments for alternative groupings within the Blattarian subgroup 2.2.

Grouping C: Tryonicus + Cryptocercus + (Anaplecta + (Nahublattella + (Supella + (Euphyllodromia + (Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria))))))

(124) Left edge 61 of lobe vla extending far anteriad (BM: 61 ending far posteriorly). This certainly apomorphic state (6.2.1., 6.2.4.) is distinct in *Tryonicus, Cryptocercus, Anaplecta*, and *Nahublattella*, and the character is hardly assessable in the other species listed. Hence, it might be an autapomorphy of this grouping. This character is inconsistent with the assumed autapomorphies of subgroup 2.2.2.

Grouping D: Cryptocercus + (Anaplecta + (Nahublattella + (Supella? + (Euphyllodromia? + (Parcoblatta + (Nyctibora? + (Blaberus + Nauphoeta? + Blaptica? + Byrsotria?))))))

(125) Muscle s10 present: from subgenital plate to ejaculatory duct (BM: s10 absent) (126) Muscle r6 absent: from region R1c to region R1d (SG2.: r6 present) (127) Sclerites L1 and L2 far separated: articulation A2 absent (BM: L1 and L2 articulated in A2).

(127) is certainly the apomorphic state (6.2.4.), and the same is probably true of (125) (6.9.) and (126) (6.7.6., (K) in 7.5.). These characters are inconsistent with the assumed autapomorphies of subgroup 2.2.2. but consistent with (124) of grouping C. (s10 and r6 not investigated in *Supella, Euphyllodromia, Nyctibora, Nauphoeta, Blaptica,* and *Byrsotria.*) As regards (127), however, non-homology is suggested for *Cryptocercus, Anaplecta,* and the other species: *Cryptocercus* and *Nahublattella* have lost the right part of L2, which curves upwards and bears articulation A2 distally in the Blattarian ground-plan, but have retained L1. *Anaplecta* has retained the upcurved right part of L2 but has lost L1. Hence, A2 has possibly been lost in different ways. If this is true, A2 has been lost three times since *Anaplecta* and *Nahublattella* are clearly more closely related (autapomorphies of subgroup 2.2.3. in 7.4.).

Grouping E: Lamproblatta + (Anaplecta + (Nahublattella + (Supella + (Euphyllodromia + (Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica + Byrsotria))))))

(73) Tendon tre and muscles s8 and b4a,b absent (SG2.: tre, s8, and b4a,b present; BM: Presence of tre and s8 unclear, b4a,b present).

This is probably the apomorphic state (compare (I) in 7.5.; 6.7.5., 6.9.). To regard it as an autapomorphy of this grouping would be inconsistent with the many assumed autapomorphies of the subgroups 2.2.2. and 2.2.2.2.

Grouping F: (*Polyphaga* + *Ergaula*) + (*Anaplecta* + (*Nahublattella* + (*Supella* + (*Euphyllodromia* + (*Parcoblatta* + (*Nyctibora* + (*Blaberus* + *Nauphoeta* + *Blaptica* + *Byrsotria*))))))

(128) Region R1c probably fused with at least part of the broadly interconnected regions R1d and R1v: articulations A8 and A9 and membranous area 17 absent; sclerites R1M or R1N (SG2.2.: R1c separated from the broadly interconnected R1d and R1v: A8, A9, and 17 present; SG.2.: R1c separated from the narrowly interconnected R1d and R1v: A8, A9, and 17 present; BM: R1c connected with both R1d and R1v which are separated from each other: A8 and A9 absent but 17 present) (64) Muscle r3 absent: from region R1c to region R1v (BM: r3 present).

(128) is a less specific formulation of (63) as well as (74) (compare list in 7.4.) and is true of both the subgroups 2.2.2.2.2. and 2.2.3. (128) and (64) are certainly apomorphic – and the characters are probably intercorrelated (6.7.6.; compare (H) in 7.5.). The special kind of fusion, however, is rather different in *Polyphaga* + *Ergaula* (subgroup 2.2.2.2.2.) and in the other species (subgroup 2.2.3.); consequently, it is highly questionable whether the fusion, the resulting sclerites **R1M** and **R1N**, and the loss of **r3** are each homologous. If these states are still regarded as autapomorphies of this grouping, these characters would be inconsistent with the many assumed autapomorphies of the subgroups 2.2.2. and 2.2.2.

Grouping G: Tryonicus + Lamproblatta

(129) Apodeme age absent (BM: age present) (130) Extension R2m present (SG2.: R2m absent).

Both states are certainly apomorphic (6.7.4.). To regard them as autapomorphies of this grouping would be inconsistent with the many assumed autapomorphies of the subgroups 2.2.2. and 2.2.2.2.

Grouping H: Cryptocercus + (Polyphaga + Ergaula)

(131) Muscle s5 absent: from subgenital plate to left wall of genital pouch (BM: s5 present).

This state is certainly apomorphic (6.9.). To regard it as an autapomorphy of this grouping would be inconsistent with the assumed autapomorphies of subgroup 2.2.2.2.

Grouping J: Cryptocercus + Polyphaga

(132) Sclerite R1K present (BM: R1K absent).

The presence of a separate R1K is certainly apomorphic. However, the homology of the

R1K of these two species is uncertain since it is only indicated by a roughly similar position (6.7.6.). If this state is still regarded as an autapomorphy of this grouping, the character would be inconsistent with the many assumed autapomorphies of the Blattarian subgroups 2.2.2.2. and, if **R1K** is not assumed to have been lost secondarily in *Ergaula*, 2.2.2.2.

Grouping K: Cryptocercus + (Lamproblatta + Anaplecta)

(133) Muscle 14 reduced or absent: from sclerite L2 to region L41 (BM: 14 present and stout).

14 is certainly stout in the common ground-plan of Blattaria and Mantodea (6.2.1., 6.3.1.). Its reduction in *Cryptocercus* and its loss in *Lamproblatta* and *Anaplecta* are certainly apomorphic states (6.3.4.). To regard them as autapomorphies of the respective groupings would be inconsistent with the many assumed autapomorphies of the subgroups 2.2.2., 2.2.2.2., and 2.2.3. To regard the reduction of 14 as a synapomorphy of *Lamproblatta* and *Cryptocercus* (assuming parallel loss in *Anaplecta*) would be inconsistent with the assumed autapomorphies of subgroup 2.2.2.2. However, the possibility must be considered that 14 was strongly reduced in the ground-plan of subgroup 2.2.2. and has enlarged secondarily at the base of subgroup 2.2.2.2. (*Polyphaga* + *Ergaula*) where the hla-muscle 114 has been lost and 14 has acquired a new function in moving the hla-hook (6.3.4.). In this case the reduction in *Cryptocercus* and *Lamproblatta* would at least be homologous.

Grouping L: Anaplecta + (Supella? + (Euphyllodromia? + (Parcoblatta + (Nyctibora? + (Blaberus + Nauphoeta? + Blaptica? + Byrsotria?)))))

(109) Sclerite L1 absent (BM: L1 present) (110) Process(es) dca absent (SG2.: dca present)
(134) Pouch pne indistinct (BM: pne distinct) (25) Muscle 11 absent: from pouch pne to region L4d (BM: 11 present) (95) Region L4d absent (BM: L4d present).

These states are certainly all apomorphic (6.1.4., 6.3.4.), and they are probably all intercorrelated since (109), (110), (134), and (25) relate to reductions in the same area, and (95) relates to the opposite insertion area of muscle 11. L1, dca, a distinct pne, 11, and L4d are all clearly present in *Nahublattella*. In *Supella* and *Euphyllodromia* (109), (110), (134), and (25) have not been investigated. In *Nyctibora, Nauphoeta, Blaptica*, and *Byrsotria* (25) has not been investigated.

If these character states are regarded as autapomorphies of this grouping, the characters would be inconsistent with the assumed autapomorphies of subgroup 2.2.3.2. Since all states relate to the reduction of elements and are intercorrelated, the assumed autapomorphies of subgroup 2.2.3.2. are regarded as much more convincing: These include more complicated features, e.g. the **114**-translocation (**79**), the **L4K**-division (**84**), and the **12**-shift (**86**). Moreover, the presence of **L1** in another species of *Anaplecta* (McKittrick 1964) might suggest that (**109**) is not an autapomorphy of this grouping; however, since the phylogenetic position of this species within subgroup 2.2.3. is not known (the genus *Anaplecta* is not necessarily holophyletic), this argument is not of high value.

Arguments for other alternative groupings within Blattaria

Grouping M: (Archiblatta? + Periplaneta + Blatta + Deropeltis + Eurycotis) + Tryonicus? + (Anaplecta + (Nahublattella + (Supella? + (Euphyllodromia? + (Parcoblatta + (Nyctibora + (Blaberus + Nauphoeta + Blaptica? + Byrsotria?))))))

(135) Muscle s7 present: from subgenital plate to L2 (SG2. and BM: s7 absent) (24) Region L1a level (BM: L1a hood-shaped).

s7 (6.9.) has been regarded as a ground-plan muscle of Blattaria (7.3.) which has been lost in subgroup 2.2.2.2. (compare (56) in 7.4., with the reverse polarity assumption, and (L) in 7.5.). The anterior part of L1 (L1a) is hood-shaped in the common ground-plan of Blattaria and Mantodea (6.1.1.). In *Archiblatta, Periplaneta, Blatta, Deropeltis,* and *Eurycotis,* in both species of *Tryonicus,* and in *Nahublattella* L1a has become level (6.1.4.). In the other species listed this character has not been investigated (*Supella, Euphyllodromia*) or is not assessable for the complete loss of L1 (remaining species). *Lamproblatta* does not reveal (24) since it has a distinct vestige of the hood- or even of the plateau-shape (L1 bends into the dorsal **pne**-wall; 6.1.4.).

To regard (24) and (135) as autapomorphies of this grouping would be inconsistent with the assumed autapomorphies of subgroup 2.2. If *Cryptocercus* really has vestiges of s7, (135) would moreover be inconsistent with the assumed autapomorphies of subgroup 2.2.2.

Grouping N: (Archiblatta? + Periplaneta + Blatta + Deropeltis + Eurycotis) + (Tryonicus? + (Cryptocercus + (Polyphaga + Ergaula)))

(136) Tendon tre and muscle s8 present (SG2. and BM: tre and s8 absent).

The presence of **tre** (6.7.1., 6.7.5.) and **s8** (6.9.) has been regarded as a ground-plan feature of Blattaria (7.3.; compare (**73**) in 7.4., with the reverse polarity assumption, and (**I**) in 7.5.). (**s8** not investigated in *Archiblatta* and *Tryonicus*). If this state is regarded as an autapomorphy of this grouping, the character would be inconsistent with the assumed autapomorphies of the subgroups 2.2. and, if **tre** and **s8** are not assumed to have been lost secondarily in *Lamproblatta*, 2.2.2. and 2.2.2.2.

Grouping O: (Archiblatta? + Periplaneta + Blatta + Deropeltis + Eurycotis) + (Tryonicus? + (Lamproblatta + (Polyphaga + Ergaula)))

(137) Muscle r6 present: from region R1c to region R1d (SG2. and BM: r6 absent).

The presence of r6 (6.7.6.) has been regarded as a ground-plan feature of Blattaria (7.3.; compare (126) of grouping D, with the reverse polarity assumption, and (K) in 7.5.). If this state is regarded as an autapomorphy of this grouping, the character would be inconsistent with the assumed autapomorphies of the subgroups 2.2. and 2.2.2.

Grouping P: (Archiblatta + Periplaneta + Blatta + Deropeltis + Eurycotis) + (Tryonicus + (Cryptocercus + (Lamproblatta + (Polyphaga + Ergaula))))

(138) Groove rge present (SG2. and BM: rge absent).

The presence of rge (6.7.1., 6.7.6.) has been regarded as a ground-plan feature of Blattaria (7.3.; compare (75) in 7.4., with the reverse polarity assumption, and (J) in 7.5.). If this

state is regarded as an autapomorphy of this grouping, the character would be inconsistent with the assumed autapomorphies of subgroup 2.2.

Grouping Q: (Archiblatta + Periplaneta + Blatta + Deropeltis + Eurycotis) + (Lamproblatta + Cryptocercus)

(139) Region R1c separated from regions R1d and R1v: articulations A8 and A9 present (SG2. and BM: R1c connected with R1d and R1v: A8 and A9 absent).

The presence of this separation and of A8 and A9 (6.7.1., 6.7.6.) has been regarded as a ground-plan feature of Blattaria (7.3.; compare (128) of grouping F, with the reverse polarity assumption, and (H) in 7.5.). If this state is regarded as an autapomorphy of this grouping, the character would be inconsistent with the many assumed autapomorphies of the subgroups 2.2., 2.2.2., and 2.2.2.2.

Grouping R: (Archiblatta? + Periplaneta + Blatta + Deropeltis + Eurycotis) + (Tryonicus? + (Lamproblatta + (Polyphaga + Ergaula)) + (Anaplecta + (Supella? + (Euphyllodromia? + (Parcoblatta + (Nyctibora? + (Blaberus + Nauphoeta? + Blaptica? + Byrsotria?))))))

(25) Muscle 11 absent: from pouch pne to region L4d (BM: 11 present).

Muscle **11** has been found only in *Mantoida, Sphodromantis, Cryptocercus*, and *Nahublattella*, and since it is in the same relative position in all these species it has been regarded as homologous and as a muscle of the common ground-plan of Blattaria and Mantodea (6.1.1., 6.1.3., 6.1.4.). To regard the loss of **11** as an autapomorphy of this grouping would be inconsistent with the many assumed autapomorphies of the subgroups 2.2., 2.2.2. (since *Cryptocercus* is excluded), 2.2.3., and 2.2.3.2. (since *Nahublattella* is excluded). It is thus clearly suggested that **11** has been lost several times (or that the **11** of Mantodea, *Cryptocercus*, and *Nahublattella* are not homologous despite their similar positions).

7.7. Conclusions in terms of phylogeny

The phylogenetic ideas presented in 7.2.-7.4. are highly supported by many autapomorphies for the various subgroups. The inconsistent characters supporting the groupings listed in 7.6. can in most cases not compete with the clusters of autapomorphies given in 7.4., and this is due to various reasons:

- The (certainly) apomorphic character state relates to the loss or reduction of an element (such derivations are not as convincing in their role as possible autapomorphies as those relating to the presence of new elements): (126), (127), (73), (64), (129), (131), (133), (109), (110), (134), (25), (95).
- The (certainly or surmisedly) apomorphic character state is, if related as an autapomorphy to one of the groupings in 7.6., the only one suggesting the respective grouping, not supported by the distribution pattern of any other character and inconsistent with the distribution pattern of many other characters: (25) as related to grouping R, (122), (123), (124), (73), (131), (132), (133), (136), (137), (138), (139).

- For the (certainly) apomorphic character state the homology in the species concerned is questionable since the possibility of parallel evolution is revealed by other species having achieved the same apomorphic character state independently: (123).
- For the (certainly) apomorphic character state the homology in the species concerned is questionable since except for a formal correspondence the morphology of the respective elements is rather different: (127), (128), (132).
- The polarity of the character is unresolved or even suggested to be the reverse: (122), (135), (136), (137), (138), (139).
- The polarity of the character is suggested to be the reverse in a certain part of the phylogenetic tree, (i.e. the apomorphic character state has been secondarily reduced in the crucial species excluded): (133).

In my view, the only conceivable alternative resulting from the list in 7.6. is that supported by (124), (125), (126), and possibly (127): *Cryptocercus* might be the sister-group of subgroup 2.2.3. (Blattellidae and Blaberidae) and not of subgroup 2.2.2.2. (*Polyphaga*, *Ergaula*, *Lamproblatta*). And *Tryonicus* could well be the sister-group of *Cryptocercus* + subgroup 2.2.3.: The possibility of a close relation between *Tryonicus* and subgroup 2.2.3. has already been considered in 7.3., based on the similar morphology of the hla-hook ((35) and (36) in 7.4.). In *Cryptocercus* the hla-base has also shifted posteriad (fig.151), and the retractility and the large extension of the membranous base 30 of hla present in *Tryonicus* and subgroup 2.2.3. could well have been reduced in this species – in correlation with the shortening of hla (compare (M), (N) in 7.5.). However, in my view, the very similar reduction of sclerite L4K and of the nla-process and the shift of 12 in correlation with the plateau-like shape of the anterior face of the pne-pouch, the arguments suggesting that *Cryptocercus* belongs to subgroup 2.2.2., are somewhat more convincing ((42)-(46) in 7.4.).

Another problematical issue is the assumed phylogeny of subgroup 2.2.3.2.2. Apart from the fact that more species will have to be investigated in detail to get a really reliable result, some character states of members of the genus Blattella are somewhat in conflict with the hypothesis in 7.4. According to Mizukubo & Hirashima (1987), fig.41, there are a dla-lobe and a R4-sclerite (= RD1d) and possibly also a R5-sclerite (= RD2v) present in Blattella karnyi. (A muscle corresponding to r11 has not been found by these writers.) In Blattella germanica (Linné, 1767) I could also find sclerites which are certainly R4 and **R5.** According to McKittrick (1964), the females of *Blattella germanica* rotate their oothecae. These features suggest that *Blattella* belongs to subgroup 2.2.3.2.2.2.2. or, at least (if R4 and R5 are assumed to be secondarily reduced in *Parcoblatta*), to subgroup 2.2.3.2.2.2. (compare (115) and (117)-(120) in 7.4.). On the other hand, Blattella karnyi (not B. germanica) resembles Nahublattella in that the posterior part of sclerite L2 is branched, and each branch occupies a process. (The two branches of B. karnyi are LD2d and LD2v in Mizukubo & Hirashima, fig.41; those of *Nahublattella* are the sclerotisations of via and psa in fig.244, 245.) The morphology of this area would in *B. karnyi* be more primitive than in all species included in subgroup 2.2.3.2.2. (compare (94) in 7.4.). This might indicate that some of the apomorphic character states regarded as autapomorphies of the subgroups 2.2.3.2.2., 2.2.3.2.2.2., and 2.2.3.2.2.2.2. are cases of parallel evolution, or that **R4**, **R5**, and the rotation of the ootheca have developed earlier and have been

reduced again in various taxa belonging to subgroup 2.2.3.2.2. However, details of morphology of the *Blattella*-species are not yet investigated. Thus, there is compelling need for further investigations on the phallomeres of the various subgroups of Blattellidae to resolve these problems in terms of the evolution and polarity of characters.

The most parsimonious phylogenetic hypothesis resulting from the discussions in chapter 7 is shown in diagram 1 in 7.4. If the species investigated in this paper are true representatives of the Mantodean and Blattarian families and subfamilies they are usually assigned to (compare the systems of McKittrick 1964 and Beier 1968 given in chapter 2), the overall phylogeny of Mantodea and Blattaria is as follows:

In Mantodea, the basal dichotomy is between Mantoididae and all other families. The second dichotomy is between Chaeteessidae and the remaining families. In Blattaria, the basal dichotomy is between Blattinae + Polyzosteriinae and all other Blattaria. These remaining Blattaria form three groups: The first consists only of the rather isolated Tryonicinae. The second contains Cryptocercidae, Lamproblattinae, and Polyphaginae, the two latter taxa being especially closely related. The third group corresponds to Blattellidae + Blaberidae. Blattellidae are clearly paraphyletic, with Blaberidae being a rather subordinate subgroup. The earliest offshoot within Blattellidae (+ Blaberidae) are the Anaplectinae; the three subsequent offshoots are various taxa previously comprised in Plectopterinae. Blaberidae, Nyctiborinae, Blattellinae, and Ectobiinae form together a holophyletic group. Nyctiborinae and Blaberidae are possibly sister-groups.

As regards Blattaria, this phylogenetic hypothesis is in several repects very different from the system of McKittrick (1964):

- Tryonicinae are not related to Blattinae + Polyzosteriinae.
- Lamproblattinae are also not related to Blattinae + Polyzosteriinae but to Polyphaginae.
- Cryptocercidae are not the sister-group of Blattidae but probably of Polyphaginae + Lamproblattinae (or possibly of Blattellidae + Blaberidae).
- Blattellidae are paraphyletic since Blaberidae are one of their subgroups. (McKittrick has also expressed this idea in her phylogenetic trees – text figure 3 – but not in her system).
- Plectopterinae are paraphyletic.

This hypothesis is based almost exclusively on male postabdominal and genital morphology. Of course, there are still other character complexes which have proved to be useful in analysing Dictyopteran phylogeny, e.g. the morphology of the female genitalia, of the proventriculus (McKittrick 1964), or of the wings. The present knowledge on these character complexes has been revised in a phylogenetic approach in Klass (1995), and a study on the evolution of the ovipositor containing many new results has been completed more recently (Klass, in press). The many characters which are now reliably interpretable are consistent with the phylogenetic hypothesis presented here. Some characters, however, are still problematic, due to insufficent (in quantity and quality) data. To improve the data base for these character complexes, and also for the male genitalia, by detailed morphological investigations should be the major task of future work on Dictyopteran phylogeny.

7.8. Conclusions in terms of the side-reversal of the phallomere complex

Of the species discussed in this paper, *Nahublattella, Supella, Euphyllodromia* (Plectopterinae), *Blaberus, Byrsotria, Blaptica*, and *Nauphoeta* (Blaberidae) have side-reversed phallomeres, and this is certainly an apomorphic feature. According to Bohn (1987), side-reversal also occurs in some species of *Ectobius*. All these species belong to subgroup 2.2.3.2. If projected on the phylogenetic tree in 7.4., the distribution pattern of this feature is as follows:

The three basal offshoots within subgroup 2.2.3.2. give rise to species with side-reversed phallomeres (*Nahublattella, Supella, Euphyllodromia*). Subgroup 2.2.3.2.2.2.2., the sistergroup of *Euphyllodromia*, contains both normally orientated (*Parcoblatta, Nyctibora*, part of *Ectobius*) and side-reversed (*Blaberus, Byrsotria, Blaptica, Nauphoeta*, part of *Ectobius*) species. (*Ectobius* can be assigned to this subgroup since the females show the advanced rotation of the oothecae, (**117**) in 7.4.).

This distribution can be interpreted in two ways: (1) Side-reversal is a ground-plan feature of subgroup 2.2.3.2. *Nahublattella, Supella, Euphyllodromia*, the Blaberidae, and the respective species of *Ectobius* have retained this orientation. *Parcoblatta, Nyctibora*, and the other species of *Ectobius* have achieved their normal orientation by a second side-reversal (independently in the various taxa concerned). (2) In the ground-plan of subgroup 2.2.3.2. the phallomere complex is still normally orientated. The basal offshoots *Nahublattella, Supella*, and *Euphyllodromia*, and also the Blaberidae and the respective species of *Ectobius* have independently reversed the phallomere complex.

Alternative (1) is highly supported by the fact that the three basal offshoots of subgroup 2.2.3.2. are side-reversed. However, a definitive decision, whether (1) or (2) or any combination of these possibilities is true, is not possible at the present state of knowledge, and more species will have to be investigated. At least, it is strongly suggested that the orientation of the phallomere complex, side-reversed or normal, is not a very good criterion for phylogenetic conclusions.

7.9. Remarks on the procedure in the phylogenetic analysis and on character lists and character state matrices

Character lists, describing the characters, their states, and the assumed polarities, and character state matrices, describing the distribution of the character states over the taxa, have the function to present all the character states used and their distribution independently of any previous assumptions on phylogeny – as an objective basis for the phylogenetic analysis or as a starting-point for a computer-based cladistic analysis. The applicability of this method of presentation in the frame of an analysis concerned with a very complex type of character evolution, as it has been found in the male genitalia of Blattaria and Mantodea, is discussed here.

The procedure in the present phylogenetic analysis is mainly hierarchical: Mantodea and Blattaria have been, in the frame of the species investigated, regarded as sister-groups (Isoptera disregarded); this basic assumption is well-founded (Hennig 1969, Klass 1995).

Blattaria and Mantodea have then been reciprocally used as outgroups, and many features of the common ground-plan of Blattaria and Mantodea could be reconstructed. Then, in the discussions of phallomere evolution in 7.2. and 7.3., holophyletic subgroups have been established according to their hierarchy. It was begun with the search for apomorphic character states common to several species, permitting the delimitation of higher-ranked subgroups (subgroups 1.2., 2.1., and 2.2.). On this level, "apomorphic" relates to a comparison with features well-ascertained for the common ground-plan of Blattaria and Mantodea. These higher-ranked subgroups, if their holophyly could be well ascertained, were then split into more subordinate subgroups, again by searching apomorphic character states common to part of the species. At this level, "apomorphic" relates, if e.g. a Blattarian subgroup is under consideration, to a comparison either with the common ground-plan of Blattaria and Mantodea, or with the ground-plan of Blattaria, or with the ground-plan of any Blattarian subgroup superordinate to and including the subgroup under consideration. At last, in 7.6., the distribution of the states of the characters inconsistent with the majority has been discussed in terms of parsimony.

This hierarchical analysis has to be continuously accomplished with a procedure of reciprocal illumination: There has to be a mutual feedback between the characters used, also concerning their evidence in terms of phylogeny. This includes a continuous feedback to the delimitation of superordinate subgroups when working on subordinate subgroups, since an autapomorphy of a superordinate subgroup might be absent within a subgroup suggested to be subordinate to it, and whether a reversal has ocurred or whether the range of the superordinate subgroup has to be modified by removing the subordinate subgroup from it has to be discussed in terms of parsimony.

The feedback between characters and also the resulting preliminary assumptions on phylogenetic relationships can be necessary at various levels of the phylogenetic analysis: for the interpretation of morphology in terms of homology relations, for the assignment of a certain morphological condition present in certain species to a certain character state, as well as for recognising the polarity of character states within a certain subgroup (and, consequently, for the definition and formulation of characters and character states, too). Hence, in the present analysis, the assumptions and conclusions related to these issues and concerning certain subgroups are in many characters dependent on the distribution of apomorphic states of other characters regarded as autapomorphies of a subgroup superordinate to that under consideration. A character list and a matrix independent of previous reciprocal illumination and preliminary assumptions on phylogeny do not include this kind of feedback between characters (and their evidence) and are consequently incomplete or even highly misleading in some characters. The following examples shall illustrate this topic.

(1) Concern: Interpretation of morphology in terms of homology relations.

As discussed in 6.3.4., the fused sclerites L3 and L4K and the muscle l4 of *Ergaula capucina* resemble L3 and l14 of Blattellidae and Blaberidae, *Anaplecta* excluded. These l4 and l14 have been regarded as non-homologous, and the similar position of the anterior insertion of the muscle moving hla – l4 or l14 – is not a synapomorphy of these taxa. This hypothesis is only in part based on a homology analysis – using the criteria of relative

position and special structure – since it is not possible to identify the muscle of *Ergaula* reliably as the **14** by a morphological comparison alone. This identification also depends on a preliminary assumption of phylogenetic relationships between *Polyphaga* and *Ergaula* on the one hand and *Anaplecta* and the remainder of Blattellidae and Blaberidae on the other, and this assumption results from the distribution of the apomorphic states of other characters (autapomorphies of the subgroups 2.2.2.2.2 and 2.2.3. in 7.4.). Hence, many characters referring to the properties of **14** and **114** (e.g. (**58**) and (**79**) in 7.4.) would have to be regarded as not (reliably) assessable in *Ergaula* without preliminary assumptions on phylogeny.

(2) Concern: Assignment of a certain morphology to a certain character state.

The description of autapomorphy (27) of subgroup 2.2. (division of region L4l, see in 7.4.) is not valid for *Parcoblatta* since this species has the anterior part of region L4l completely lost (fig.268; compare sclerite L4U' of *Blaberus*, fig.299), and it is not a priori decidable if this loss was preceded by a division of L4l or not. That the condition in *Parcoblatta* has to be assigned to character state (27), or is derived from it, can only be recognised by regarding the evidence from the distribution of apomorphic states of other characters revealing the close relationship between *Parcoblatta* and *Anaplecta* and especially *Blaberus* (e.g. most autapomorphies of the superordinate subgroups 2.2.3. or 2.2.3.2. in 7.4.) – i.e. by practising reciprocal illumination between characters and after having made preliminary assumptions on phylogenetic relationships.

(3) Combination of concerns: Assignment of a certain morphology to a certain character state and recognition of the polarity.

In the common ground-plan of Blattaria and Mantodea the L2-sclerotisation within the **lve**-pouch (regions L2m, L2a, L2p), the **paa**-sclerotisation (region L2d), the **pda**-sclerotisation (posterior part of region L4l), and the region L4d are all firmly connected within one sclerite (e.g. *Mantoida*, fig.44-47). The apomorphic division of the left part of L2 and the named parts of L4 (= sclerite L4N in the ground-plan of subgroup 2.2.) is clearly different, and non-homologous, in *Lamproblatta* (resulting sclerites L2A+L4S and L2C+L4T, fig.178-180) and in *Nahublattella* (resulting sclerites L2D and L2E+L4N, fig.242-245; discussions in 6.2.4. and 6.3.4.). Non-homology can be recognised only by the different position of the L4d-region: In *Lamproblatta* L4d is connected with the L2-sclerotisation within lve (fig.178, 186); in *Nahublattella* L4d is connected with the sclerotisation of the insertion area of muscle 110, fig.244, 250); this is also the only property that can serve for a description of the difference in the formulation of the respective characters:

Character 1: Division in the left posterior part of main sclerite L2 and the associated parts of L4 which separates region L4d from the L2-sclerotisation within lve but does not separate L4d from the sclerotisation of the posterior insertion area of muscle 110 (division = articulation A10). Character states: (0) absent; (1) present (in *Nahublattella*); Polarity: 0>1.

Character 2: Division in the left posterior part of main sclerite L2 and the associated parts of L4 which separates region L4d from the sclerotisation of the posterior insertion

area of muscle **110** but does not separate **L4d** from the **L2**-sclerotisation within **lve**. Character states: (0) absent; (1) present (in *Lamproblatta*); Polarity: 0>1.

Nahublattella and the members of subgroup 2.2.3.2.2. (e.g. Blaberus, Parcoblatta) reveal many apomorphic features in common, and together they form the certainly holophyletic subgroup 2.2.3.2.: (79)-(89) in 7.4. The morphology near the left posterior end of L2 is in some species of subgroup 2.2.3.2.2. (e.g. Nyctibora) rather similar to Nahublattella and can easily be derived from it, and the division of L2 (articulation A10) is certainly homologous (fig.328b,h). However, in all members of subgroup 2.2.3.2.2. the L4d-region, and thus the only element yielding a criterion by which this special division of L2 can be recognised or charcterised, is absent: (95) in 7.4. (In the L2-division as present in subgroup 2.2.3.2., L4d has primitively also retained the connection with the common sclerotisation of paa and pda, and this would be a second criterion for recognising the difference to Lamproblatta, whose L2- and L4-division separates L4d from the sclerotisation of paa and pda; fig.329f, g. 6.3.4. However, this criterion can be applied neither to Nahublattella – this species shows a certainly autapomorphic division of sclerite L2E+L4N into a basal and a distal sclerite by the membranous ring 39, fig.244, which separates L4d from the sclerotisation of paa and pda – nor to subgroup 2.2.3.2.2. for the loss of L4d).

Hence, there is a first problem in the L2-division of subgroup 2.2.3.2.2. concerning the assignment of a certain morphology to a certain apomorphic character state: Without having used reciprocal illumination between characters previously, i.e. without having the preliminary assumption of the holophyly of the superordinate subgroup 2.2.3.2. resulting from the consideration of other characters, the characters 1 and 2 would have to be regarded as not assessable in those members of the subordinate subgroup 2.2.3.2.2. which show a L2-division (Table 1a). The assessment and the matrix entries of the characters 1 and 2 can only be proper if these two characters are considered in interdependence with other characters having apomorphic states common to *Nahublattella* and subgroup 2.2.3.2.2., i.e. if the probable holophyly of the superordinate subgroup 2.2.3.2. has been recognised previously (Table 1b).

Moreover, some members of subgroup 2.2.3.2.2. reveal a secondary fusion of L2D and L2E+L4N and a secondary loss of muscle 110. That these are reversals results clearly from the hierarchical analysis. Concerning the sclerotisations, the highly apomorphic character state achieved by this reversal conforms exactly with the state present in e.g. *Polyphaga* (all parts of L2 and L4N form together one sclerite), and it also conforms with the most plesiomorphic state within Blattaria and Mantodea (all parts of L2 and the posterior part of region L4I – L4N not yet differentiated as a separate sclerite – are contained in one sclerite). As regards 110, its absence does likewise correspond with the most plesiomorphic state within Blattaria and Mantodea.

Hence, there is a second problem in the L2-division of subgroup 2.2.3.2.2. concerning the recognition of the polarity and the definition of character states: In elaborating a character state matrix without having used reciprocal illumination between characters previously, the morphology of the sclerotisation and of **110** would have to be regarded as representing rather or most plesiomorphic states of the respective characters (Table 1a). Only the distribution of the states of other characters and their evidence in terms of phylogeny reveals

Table 1a,b: Character lists and character state matrices of the characters 1, 2, and 3; a) without regarding evidence of other characters in terms of phylogeny; b) under consideration of evidence of other characters in terms of phylogeny.

Matrices: 0 = most plesiomorphic state; 1 = apomorphic state derived from 0; 2 = apomorphic state derived from 1; / = character not investigated; ? = character not assessable; C = Character; Sp = Sphodromantis; Me = Metallyticus; Ch = Chaeteessa; Ma = Mantoida; Ar = Archiblatta; Eu = Eurycotis; Tp = Tryonicus parvus; Po = Polyphaga; Er = Ergaula capucina; Cr = Cryptocercus; La = Lamproblatta; An = Anaplecta; Na = Nahublattella; Su = Supella; Ep = Euphyllodromia; Pa = Parcoblatta; Ny = Nyctibora; Np = Nauphoeta; Bb = Blaberus; Bp = Blaptica; By = Byrsotria.

To the characters 1 and 2 the criterion of the connection of the **paa**-sclerotisation (region L2d) has been added in order to distinguish the described divisions from the division between the **paa**- and **pda**-sclerotisations as present in some Blattaria and Mantodea (compare (A) in 7.5.).

a)

Character 1: Division in the left posterior part of main sclerite L2 and the associated parts of L4 which separates region L4d as well as the sclerotisation of paa from the L2-sclerotisation within lve but does not separate L4d from the sclerotisation of the posterior insertion area of muscle 110 (= articulation A10). Character states: (0) absent; (1) present; Polarity: 0>1.

Character 2: Division in the left posterior part of main sclerite L2 and the associated parts of L4 which separates region L4d from the sclerotisation of paa and from sclerotisation of posterior insertion area of muscle 110 but does not separate L4d from the L2-sclerotisation within lve. Character states: (0) absent; (1) present; Polarity: 0>1.

Character 3: Presence of muscle 110. Character states: (0) absent; (1) present; Polarity: 0>1.

С	Sp	Me	Ch	Ma	Ar	Eu	Тр	Ро	Er	Cr	La	An	Na	Su	Ep	Ра	Ny	Np	Bb	Вр	Ву		
1	0	0	0	0	0	0	0	0	0	0	0	0	1	?	0	0	?	?	0	0	0		
2	0	0	0	0	0	0	0	0	0	0	1	0	0	?	0	0	?	?	0	0	0		
3	0	/	/	0	/	0	/	1	1	1	1	1	1	1	0	0	1	1	0	0	0		
														subgroup 2.2.3.2.2.									

b)

Character 1: Division in the left posterior part of main sclerite L2 and the associated parts of L4 which separates region L4d as well as the sclerotisation of paa from the L2-sclerotisation within lve but does not separate L4d from the sclerotisation of the posterior insertion area of muscle 110 (= articulation A10). Character states: (0) absent; (1) present; (2) secondarily absent; Polarity: 0>1>2. **Character 2:** Division in the left posterior part of main sclerite L2 and the associated parts of L4 which separates region L4d from the sclerotisation of paa and from the sclerotisation of the posterior insertion area of muscle 110 (= articulation A10). Character states: (0) absent; (1) present; (2) secondarily absent; Polarity: 0>1>2. **Character 2:** Division in the left posterior part of main sclerite L2 and the associated parts of L4 which separates region L4d from the sclerotisation of paa and from the sclerotisation of the posterior insertion area of muscle 110 but does not separate L4d from the L2-sclerotisation within lve. Character states: (0) absent; (1) present; (0) = 1.

Character 3: Presence of muscle **110**. Character states: (0) absent; (1) present; (2) secondarily absent; Polarity: 0>1>2.

С	Sp	Me	Ch	Ma	Ar	Eu	Тр	Ро	Er	Cr	La	An	Na	Su	Ep	Ра	Ny	Np	Bb	Вр	Ву
					_		_	_													
1	0	0	0	0	0	0	0	0	0	0	0	0	1	1	2	2	1	1	2	2	2
2	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
3	0	1	/	0	/	0	1	1	1	1	1	1	1	1	2	2	1	1	2	2	2
subgroup 2.2.3.2.2													2.								

that the absence of the division described in character 1 and the absence of 110 are plesiomorphic for Blattaria and Mantodea as a whole but apomorphic within subgroup 2.2.3.2. (discussion in 7.5. (Q), (R)). The character states achieved by these reversals can then be properly defined as highly apomorphic states (Table 1b).

As a consequence of the combined presence of the first and the second problem, in the character state matrix in Table 1a the items relating to the L2-divisions would suggest that these L2-divisions have originated independently in *Nahublattella* and in subgroup 2.2.3.2.2. and are non-homologous. In a computer-based cladistic analysis this would cause a misleading trend away from a holophyly of *Nahublattella* + subgroup 2.2.3.2.2. In the matrix in Table 1b this misleading impression is eliminated.

As a result, the assessment of homology relations, the definition of character states, the assignment of morphological conditions to certain character states, and the polarity assumptions, and hence also the respective entries of items into the matrix, can in some cases only be proper in dependence on a previous hierarchical analysis with reciprocal illumination and on the resulting preliminary assumptions in terms of phylogeny. It is, at least in the frame of the analysis presented here, not suitable to give a character list and a character state matrix with the characters considered independently of each other and of preliminary assumptions on phylogeny.

8. HOMOLOGY RELATIONS ACCORDING TO MIZUKUBO & HIRASHIMA (1987) AND GENERAL REMARKS ON THE ANALYSIS OF HOMOLOGY RELATIONS

The assumptions and procedures of Mizukubo & Hirashima

Mizukubo & Hirashima (1987) investigate the sclerites and the muscles of the phallomeres of *Periplaneta fuliginosa* (Blattidae / Blattinae), of 3 species of *Blattella* (Blattellidae / Blattellinae), and of *Opisthoplatia orientalis* (Blaberidae). Additionally, they use data from other writers concerning various species of Blattinae. The phallomeres of Blattinae are regarded as the most primitive. The results of the authors comprise: (1) Homologies of the phallomere elements of the different species. (2) Side-homologies of the elements of the left and the right halves of the phallomere complex. (3) A ground-plan for the sclerotisations of the phallomere complex of Blattaria, which is mainly based on the morphology of Blattinae.

As regards (1), the supposed homology relations are fundamentally different from those I assume for the respective close relatives *Eurycotis* (Blattidae), *Parcoblatta* (Blattellinae), and *Blaberus* (Blaberidae). For example, Mizukubo & Hirashima suppose that the hooks designated here as **hla** have developed from completely different elements in the three groups. (In my view these **hla** are strictly homologous.) Their opinions concerning the ground-plan of the Blattarian phallomeres are also completely contradictory of my results. The paper of Mizukubo & Hirashima must therefore be discussed in detail.

Mizukubo & Hirashima procede as follows:

- They divide both the left and the right side of the phallomere complex into 11 "subregions". The definition of "subregion" is: "The smallest and indivisible unit which

is a part of the bordered region in a plane and, in this entire region, possesses its own relative position determined by its relations with other surrounding subregions." (p.251). The relationships between sclerites and subregions are characterised on p.251: (A) Principally a single sclerite (often inclusive of the surrounding membrane) corresponds with a single subregion. (B) A single sclerite can spread over two or more subregions, or two or more subregions can participate in a single sclerite. (C) A subregion can be completely membranous.

- They do not explicitly say whether (B) and (C) are exclusively regarded as derived states, or if they can be already realised in the hypothetical ground-plan of Blattaria. However, the definition of the subregions makes sense only if the demarcation of "smallest indivisible units" obeys a uniform principle and this can only be the possession of an own sclerite according to (A). Hence, I interpret Mizukubo & Hirashima in the following way: A set of 11 subregions on each side, each subregion with one sclerite of its own, is regarded as the ground-plan pattern of Blattaria. (According to (A); (B) and (C) realised in derived states only.)
- They deduce the basic pattern of 11 subregions per side from the morphology of the various Blattinae: By considering several Blattinae and by combining their features, the dividing of the phallomere complex into subregions can be accomplished in a way that the relative positions of the subregions of the left side are a mirror-image of the relative positions of the subregions of the right side.
- In both *Blattella* and *Opisthoplatia* the dividing into subregions can be accomplished in a way that the subregions of the left side as well as those of the right side have the same relative positions as they have in Blattinae and in the hypothetical basic pattern. In their dividing procedure the authors assume losses or fusions for some sclerites (according to (B) and (C)).
- From these equal relative positions of the subregions they deduce homology relations between the subregions of the left and of the right side of the phallomere complex as well as between the subregions of the phallomeres of different species. Side-homologous subregions get the same names except for L (left) or R (right) in the first position.
- Concerning the closeness of the relations between neighboring subregions, they distinguish four categories which describe the closeness or intensity of the mutual relations between the respective sclerites: weak adjacency – adjacency – articulation – fusion. All the relations between all neighboring subregions together are the association pattern of the phallomere complex.
- They investigate the muscle insertions on the various subregions.
- In their homology analysis the authors largely neglect the musculature. If the course of a muscle is consistent with the homology assumptions deduced from the relative positions of the subregions, this is regarded as a confirmation. If there is inconsistency, the authors do not regard this as a matter of conflict.

The statements of Mizukubo & Hirashima include, or result in, the following assumptions regarding the ground-plan of Blattaria:

The left as well as the right side of the phallomere complex are provided with (exactly) 11 separate sclerites. (This results from the definition and characterisation of "subregion", compare above).

- The whole phallomere complex is bilaterally symmetrical (p.256). Consequently, all side-homologous subregions would have to be regarded as completely symmetrical, too.

The procedure and the argumentation of Mizukubo & Hirashima have some weak points, and many of their statements are in contradiction to my results. My critique concerns the topics discussed subsequently.

The negligence of the musculature as a reference frame for the homology analysis

Mizukubo & Hirashima base their homology analysis on the relative positions of the cuticular subregions to each other but largely neglect the musculature. Regarding their results, the courses of most muscles are inconsistent with the homology assumptions deduced from cuticular morphology. Referring to Matsuda (1976), they assume that, since the muscles develop independently of the exoskeleton, the insertions of muscles generally have a too large shifting potential in their evolution to be reliable landmarks in the analysis of homology relations. Matsuda (1976) regards the musculature as a valuable supplementary criterion only "when the structures within a relatively narrow range of species – within a family, or perhaps an order – are under study" (p.36). The question arises, therefore, how valuable the muscles are in homologising the phallomere elements of Blattaria and Mantodea and to what extent they deserve to be considered.

In *Eurycotis* and *Mantoida*, which are only distantly related, the cuticular elements of the phallomeres are quite similar in their principal arrangement. The arrangement of the musculature is to a large extent consistent with the homology relations resulting from the relative positions and special features of the cuticular elements: The main muscles of the phallomere complex have the same course (**12**, **13**, **14**, **16**, **r1**, **r2**, **r3**). In many cases the musculature can help in confirming homology assumptions. This is not inconsistent with the views of Mizukubo & Hirashima, but in my opinion it also suggests that one should not a priori regard the musculature as highly variable as these authors do.

In my view, (1) to regard the insertions a priori as conservative and (2) to assume shifts of insertions only if inconsistencies arise is the better approach. (1) In the comparison between Mantoida and Sphodromantis, the evolution of some sclerotisations could be reconstructed in detail, because the musculature was taken as an integral instrument of the homology analysis (compare L4 in 6.3.3.). In the comparison of distantly related species in which homologous cuticular elements show, apart from a somewhat similar position, hardly any similarities, the insertions of muscles can in my opinion be extremely valuable landmarks. For example, the insertions of 14, 12, and 114 suggest that sclerite L4K of Cryptocercus is homologous with a part of sclerite L4H of Eurycotis (L4n-region and anterior L4I-region). The insertion of I4 suggests (partial) homology for L4K of Ergaula and L4K of Cryptocercus, though L4K of Ergaula has shifted to the ventral hla-base and fused to sclerite L3 (compare in 6.3.4.). (2) On the other hand, of course, the muscle insertions have a certain shifting potential. By studying enough species, however, these shifts can often be "observed" step by step, and in many cases it becomes obvious whether the insertions of the muscles or the similarities in cuticular morphology are the better basis for homology assumptions. Drastic shifts of muscle insertions are e.g. that of 114 from the L4n-region (Eurycotis) to the L2a-region (Nahublattella) and that of l2 from sclerite

L1 (*Mantoida*) to the base of the **hla**-hook (*Nahublattella*). In these two cases, the morphology of *Anaplecta* reveals how these shifts have taken place (and that with high probability the insertions **have** shifted).

In my view the arrangement of the musculature is a very important element of the homology analysis. A simultaneous consideration and a mutual weighing of similarities in the cuticular elements and in the musculature – combined with the investigation of a larger sample of species – has proved most useful in this work. Moreover, in this kind of proceding, the consideration of the musculature has the advantage that the information about the shifts of insertions, the losses, divisions, fusions, or de-novo-formations of muscles can provide many autapomorphies – in addition to those gathered from cuticular morphology. In my view, a homology hypothesis on the Blattarian phallomere elements which accepts extensive inconsistencies in the arrangement of the musculature is not very convincing.

The division into 11 subregions per side

Mizukubo & Hirashima deduce the presence of a natural division into 11 subregions on each side of the phallomere complex from the morphology of various Blattinae. However, in my view their special kind of procedure is debatable. (The subsequently used terms of Mizukubo & Hirashima can be distinguished from mine by D or V in the second position.) Mizukubo & Hirashima assume that the sclerotisations comprised in **R1** in my terminology represent 7 subregions (compare *Eurycotis*, fig.74-78, 330g, 331e, 332e, and *Archiblatta*, fig.330f):

- **1. RD1d** essentially region **R1d** (sclerite **R1H**)
- 2. RD11 sclerotisation of process pra, part of region R1d
- 3. RD1m sclerotisation of spine sra, part of region R1d
- 4. RD1v essentially region R1v (sclerite R1G)
- **5. RD1vm** a ribbon-like sclerotisation connecting **R1H** and **R1G**; missing in *Eurycotis* but present in *Archiblatta* in the ventral wall of lobe **fda** (compare fig.330f and g)
- 6. RD2l essentially region R1c
- 7. RD2d essentially region R1t (with ridge pva)

The remaining subregions of the right phallomere are:

- 8. RD2v sclerite R2
- 9. RD3 sclerite R3
- 10. RVv sclerite L4G (region L4v on lobe vla)
- 11. RVd right part of sclerite L5 of *Periplaneta* (within ejaculatory duct, compare in 6.5.); RVd is supposed to have fused with its left counterpart LVd = left part of sclerite L5.

In my view, **R1** is in the common ground-plan of Blattaria and Mantodea either one undivided sclerite (more probable) or composed of three sclerites (**R1F**, **R1G**, **R1H**; separated by the articulations **A8** and **A9**; 6.7.1.). For the ground-plan of Blattaria I assumed the latter condition, which is still present in *Eurycotis*. For the subregions **RD11**, **RD1m**, **RD2d**, and **RD21** there is no indication that they have been separate

sclerites in the ground-plan of Blattaria. (The separation of **RD2d** and **RD2l = R1t** and **R1c** is realised as an apomorphic feature in some Mantodea and Blattellidae only, compare in 6.7.3., 6.7.4., 7.5.(**G**). **RD1l** and **RD1m** are separate sclerites in some Blattinae, but the outgroup comparison with Mantodea suggests that this is not a ground-plan feature of Blattaria.) As regards the ventral sclerotisation of the vla-lobe (**RVv = L4G**), it is not impossible that this is an element of the right half of the phallomere complex (according to Quadri 1940), but in my view this is not very probable (discussion in 3.1.).

The left complex is divided into the following subregions (compare *Eurycotis*, fig.65-69, 323e, 324e, 325e, and *Archiblatta*, fig.53-57, 323f, 324f, 325f):

- 1. LD1d left part of sclerite L1 (part of region L1a)
- 2. LD11 right part of sclerite L1 (parts of regions L1a and L1m)
- **3. LD1m** rightmost part of sclerite L1, near articulation A2 (part of region L1m)
- 4. LD1v sclerite L4F (posterior part of region L4c) + sclerotisation of paa (region L2d)
- 5. LD1vm essentially sclerite L2 (except region L2d)
- 6. LD21 posterior part of sclerite L4C of Archiblatta (posterior part of region L4l)
- 7. LD2d sclerite L3 (on hook hla)
- 8. LD2v sclerite L4D of *Archiblatta* (region L4n)
- 9. LD3 anterior part of sclerite L4C of Archiblatta (anterior part of region L4l)
- **10. LVv** sclerite L4E of *Archiblatta* (anterior part of region L4c)
- 11. LVd left part of sclerite L5 of *Periplaneta*

In 6.3.1. it has been shown that the sclerotisation of the L4I-region (LD2I and LD3) is undivided in the common ground-plan of Blattaria and Mantodea as well as in the groundplan of Blattaria, and that this situation is retained in *Archiblatta* (sclerite L4C) and *Eurycotis* (sclerite L4H). And there is no indication that L4I was present as two separate sclerites in still earlier times.

In some cases the division into subregions is based on apomorphic features of Blattinae (and Polyzosteriinae): L4F is a sclerite peculiar to these groups and certainly not a groundplan element of Blattaria. The L4n-region is only in Blattinae an isolated sclerite (L4D), not in *Eurycotis, Tryonicus*, or *Anaplecta*; the ground-plan situation of Blattaria, however, is unclear in this respect. The branching of the posterior part of L1 into several lobe-like extensions (LD1d, LD1m, LD1l) is a consequence of the posteriad expansion of L1 onto the dca-processes, and in this distinct form it is certainly an apomorphic state; that L1 is a fusion product of several previously isolated sclerites cannot be deduced fom this situation.

As a result, there are two principal reasons to refute (1) the division into 11 subregions in Blattinae as well as (2) the ascription of this division to the ground-plan of Blattaria:

- Neither the left nor the right side of the phallomere complex of *Periplaneta* (or other Blattinae or Polyzosteriinae) shows a priori a composition of exactly 11 subregions, nor does the Blattarian ground-plan show such a pattern. Most of the dividing into subregions is based either on apomorphic situations in a subgroup of Blattaria (Blattinae) or on arbitrary – and in my view wrong – assumptions on which sclerotisations were isolated from each other in the Blattarian ground-plan.

- Though the demarcation or identification of a ground-plan subregion has a clear theoretical background (indivisible unit = 1 sclerite), no uniform principle can be recognised in the practical application to extant species (analysis of Blattinae), let alone the attempt to come close to the definition or to explain discrepancies. It is not comprehensible why Mizukubo & Hirashima assume for some sclerites of Blattinae a contribution of several subregions and why for other sclerites they do not. The dividing procedure seems to aim to have subregions with corresponding relative positions on the left and on the right side. So the division of the L4I-region (into LD2I and LD3) results in having – like on the right side (RD2I and RD3) – one subregion for the sclerotisation in the anterior ventral wall (LD3 and RD3) and one for the sclerotisation in the lateral edge of the phallomere (LD2I and RD2I). (According to my results, only the division on the right side is a ground-plan feature: articulation A3.)

The argumentation concerning homology assumptions

Mizukubo & Hirashima mainly make use of the first criterion of homology (relative positions). However, the specific procedure of the dividing into subregions described above makes the homology assumptions questionable: From the fact that the left as well as the right side **can** be (in a largely arbitrary way) divided into 11 areas having the same relative positions cannot be decuced that these areas are side-homologous because of their equal relative positions (circular argumentation). Assumptions of homology would only be justified, if (1) these areas have specific features in common (i.e. if there are **similar structures** in the same relative positions, e.g. similar sclerites, muscle insertions, processes, apodemes, etc.), or if (2) an equal arrangement on both sides results from a uniform principle of dividing. The same critique applies to the homology assumptions that concern the comparison of different species: Again, the surface of the phallomeres is divided largely arbitrarily into subregions with equal relative positions, and the subregions are then supposed to be homologous because their relative positions are equal.

Moreover, the reliability of the homology hypothesis becomes further diminished by the fact that neither the side-homologies nor the homologies between different species are supported by similarities in the arrangement of the musculature or in the intensities of the mutual relations between the subregions / sclerites:

- Side-homologies: Related to the side-homologies assumed by Mizukubo & Hirashima, the musculature of *Periplaneta* is completely different in the left and in the right half of the phallomere complex (of 20 phallomere muscles only two are a pair, Mizukubo & Hirashima, fig.6). As regards the principal relative positions of the subregions, the association patterns of the left and of the right side of *Periplaneta* are very similar. This simply results from the fact that the two halves of the phallomere complex have been arbitrarily divided into subregions which are in the same relative positions. However, the subregions of the left and of the right side supposed to be homologous hardly have any intensity of the mutual relations in common (Mizukubo & Hirashima, fig.2).
- Homologies between species: In *Periplaneta* and *Blattella germanica*, of 14 or 7, respectively, intrinsic muscles of the left half of the phallomere complex only 2 have

The special morphology of the supposedly homologous subregions (e.g. position in a pouch, formation of a process) is not considered at all.

The symmetry of the phallomere complex in the Blattarian ground-plan

The investigations and conclusions of Mizukubo & Hirashima are restricted to Blattaria; Mantodea are not mentioned at all. With their statement "We cannot detect proto-types of the genitalia indirectly on evidence obtained from other insect groups." (p.250) the authors deprive the phallomeres of Mantodea of any value to contribute to the reconstruction of the ground-plan of the Blattarian phallomeres. However, in the reconstruction of the ground-plan of any group an outgroup comparison can be very useful. In the case of the Blattarian phallomeres the consideration of the Mantodean phallomeres was of great value for the determination of the polarities of characters within Blattaria (and within Mantodea). Since Mizukubo & Hirashima neglect Mantodea, the statement "We believe that, at the period of the formation of the order, the early Blattaria had symmetrical genitalia" has no foundation at all. According to this statement, the asymmetry of the Blattarian and the Mantodean phallomeres is a case of parallel evolution. However, my results clearly suggest that the very special kind of asymmetry present in Blattaria and Mantodea is homologous and a feature of their common ground-plan.

Mizukubo & Hirashima recognise the side-reversed similarities in the phallomeres of Blattellidae (*Blattella*) and Blaberidae (*Opisthoplatia*). However, they do not assume homology for these similarities but parallel evolution due to similar selective pressure. Hence, they assume completely symmetrical phallomeres even for the last common ancestor of Blattellidae and Blaberidae. These opinions are refuted:

- Since extreme asymmetry had already been established in the common ground-plan of Blattaria and Mantodea, it must have been present in the common ancestors of Blattellidae and Blaberidae, too.
- The similarities of the left complexes of Blaberidae and the more derived Blattellidae (*Nyctibora*, *Parcoblatta*) are so detailed and peculiar that the probability for parallel evolution is in my view infinitely small; side-reversal is substantially ascertained by my results (compare in 6.13.).
- That a reversal of the left-right asymmetry must be considered as a possible evolutionary pathway is clearly demonstrated by those species of *Ectobius* (Ectobinae) having sidereversed phallomeres (compare in 6.13.).

9. HOMOLOGY RELATIONS ACCORDING TO GRANDCOLAS (1994) AND THE PHYLOGENETIC POSITION OF *CRYPTOCERCUS*

Apart from other morphological studies, Grandcolas & Deleporte (1992) and Grandcolas (1994) investigate the phallomere sclerites of some Blattaria. The latter paper contains nearly all the information given in the former, and also some additional data, and will be referred to in the following discussions.

Grandcolas (1994) investigated the phallomeres of *Periplaneta americana* (Blattinae), *Cryptocercus punctulatus*, and several Polyphaginae and proposes a homology hypothesis for the phallomere sclerites. He finds many synapomorphies suggesting *Cryptocercus* to be a subordinate taxon of Polyphaginae. However, his homology hypothesis is very different from my homology assumptions for *Archiblatta* (Blattinae), *Eurycotis* (Polyzosteriinae), *Polyphaga, Ergaula* (Polyphaginae), and *Cryptocercus*, and these discrepancies and the resulting assignment of *Cryptocercus* have to be discussed.

9.1. Discussion of the homology relations assumed by Grandcolas

The data base of Grandcolas

Grandcolas gives data on phallomere morphology in figures showing the cuticular phallomere elements of *Periplaneta americana* (fig.1), *Heterogamodes ursina* (fig.3), *Therea petiveriana* (fig.5), and *Cryptocercus punctulatus* (fig.6), and in sketches showing the principal sclerite pattern in Blattinae (fig.2) and Polyphaginae (fig.4). He terms the sclerites in the same manner as McKittrick (1964), but due to different homology assumptions the names of the sclerites are in many cases different, too. Some differences result from Grandcolas' assumptions on side-homologies, which are expressed by giving side-homologous sclerites the same names (except for L or R in the first position to name the side). The sclerite terminology of Grandcolas is rather different from mine, and table 2 gives the synonymy and the homology assumptions. To distinguish them from mine the terms of Grandcolas will be provided throughout with *.

Table 2: Synonymy of the sclerite terms of Grandcolas (1994) and those used in this paper.

Terms of Grandcolas are provided with *. Somewhat questionable synonymies are provided with ?. L1 of *Periplaneta* (2nd column): In his fig.2 Grandcolas 1994 assigns L1 to L2*, but it is not clear whether he assigns it to L2d* or to L2v*.

	Periplaneta Archiblatta	Cryptocercus	Therea	Heterogamodes
L1*	L5	L1	L1	L1
L2d*	L4C and L1?	L3	L4N?	L4N?
L2v*	L2 and L1?	L2	L2	L2
L3d*	L3	part of L4N	L4K?	L3
L3v*	L4D	part of L4N	L3?	L4K or L4M?
vp*	L4G	L4G	L7?	
N*		R2	L8	L8
R2*	R2 and R1F	R1F	R2	R2
R3d*	R1G and R1H	R1J	R1M	R1M
R3v*	R3	R3	R3	R3

In Grandcolas' fig.3 and 5 showing *Heterogamodes* and *Therea* at least some of the apomorphies listed in 7.4. can be recognised, and these permit the integration of these species into my phylogenetic hypothesis. *Therea*, fig.5, shows five of these apomorphies:

(48) L8 = N* is present. (55) L2 = L2v* and the lve-pouch almost reach the left edge of the left complex. (62) R2 = R2* and R3 = R3v* are fused. (63) R1M = R3d* is present. R2 = R2* is so broad that R3 = R3v* is for most of its breadth confluent with it (compare in 7.3., subgroup 2.2.2.2.2., and fig.330m). Hence, *Therea* can be assigned to subgroup 2.2.2.2. (*Polyphaga + Ergaula + Lamproblatta*) by (48) and (55), to subgroup 2.2.2.2.2. (*Polyphaga + Ergaula + Lamproblatta*) by (48) and (55), to subgroup 2.2.2.2.2. (*Polyphaga + Ergaula*) by (62) and (63), and to subgroup 2.2.2.2.2. (*Ergaula*) by the breadth of R2, and Grandcolas is probably right in assuming a close relation between *Ergaula* and *Therea*. *Heterogamodes*, fig.3, shows at least (48) L8 = N* and (63) R1M = R3d* and can be assigned to subgroup 2.2.2.2.2. (*Polyphaga + Ergaula*). These assignments permit treating *Therea* and *Heterogamodes* – independently of the assumptions of Grandcolas – as true representatives of subgroup 2.2.2.2.2. ("Polyphaginae") in the following discussions and to assume that at least all autapomorphies of the subgroups 2.2., 2.2.2., and 2.2.2.2. are also present in these species (if there have not occurred secondary changes). Also, the morphology of the phallomere sclerites can be expected to be at least similar to *Polyphaga* and *Ergaula*.

The way Grandcolas uses his terminology in Polyphaginae (excluding *Cryptocercus*) and his homology assumptions between Polyphaginae and *Cryptocercus* or *Periplaneta* can only be inferred from the figures showing *Therea* and *Heterogamodes*. The terminology applied to these two species can largely be transferred to the Polyphaginae I have studied, *Polyphaga* and *Ergaula*, since for most phallomere elements the homology relations between *Therea I Heterogamodes* and *Polyphaga I Ergaula* are quite evident; in some other cases, however, problems arise. In combination with Grandcolas' figures on *Periplaneta* and *Cryptocercus*, this transfer allows the comparison and discussion of the homology relations which are assumed for Polyphaginae (in general), Blattinae, and *Cryptocercus* by Grandcolas and by me. *Polyphaga* and *Ergaula* (alone), *Periplaneta*, and *Cryptocercus* can be compared independently of this transfer, since the homology relations between these species have been discussed in chapter 6. In the following discussions (A)-(F) the phallomere morphology of *Therea*, *Heterogamodes*, *Polyphaga*, and *Ergaula* (designated as Polyphaginae) will be compared with that of *Cryptocercus* and Blattinae (and, in part, Polyzosteriinae, which are closely related to Blattinae).

Sclerite L1* sensu Grandcolas and the genital opening (A)

Grandcolas names the sclerite next to the genital opening L1*. In *Cryptocercus* and *Therea* and probably also in *Heterogamodes* L1* is sclerite L1 (compare fig.3, 5, 6 of Grandcolas and fig.151). As regards the homology of L1 = L1* of these three species, I agree with Grandcolas. However, if L1* of *Heterogamodes* really is the homologue of the L1 = L1* of *Cryptocercus*, *Polyphaga*, and *Ergaula*, the opening concerned would not be the genital but the phallomere-gland opening (compare black arrow in fig.3 of Grandcolas and P in fig.106, 121, 153). In *Cryptocercus*, *Polyphaga*, and *Ergaula*, and *Ergaula* the genital opening is much more ventrally: the ejaculatory duct (D in fig.122, 151) opens into the **lve**-pouch, next to sclerite L2 = L2v*. This relation resulted clearly from own investigations of the internal anatomy.

In *Periplaneta* $L1^*$ is sclerite L5, which is situated inside the true ejaculatory duct (compare in 6.5.). Since the assumption of Grandcolas that this L5 is homologous with

L1 of Polyphaginae and *Cryptocercus* is only based on the similar position next to the genital opening and since the genital opening has been misidentified in Polyphaginae and *Cryptocercus*, the homology of these sclerites is no longer supported. McKittrick (1964) assumes homology for the L1 of Polyphaginae, *Cryptocercus*, and Blattinae (as I do), and this assumption is confirmed by the similar morphology of the sclerites, by a similar position relative to other sclerites, by similar muscle insertions, and by a position next to the phallomere-gland opening (discussion in 6.1.).

Sclerites L2d*, L3d*, and L3v* sensu Grandcolas (B)

As regards L2d*, L3d*, and L3v* of *Therea* and *Heterogamodes*, neither the muscles nor the exact morphology and relative position of the sclerites are shown in fig.3 and 5 of Grandcolas, and an exact homologisation with the sclerites of *Polyphaga*, *Ergaula*, *Cryptocercus*, and *Periplaneta* is, therefore, not possible.

L3v* of *Periplaneta* is sclerite L4D (L4n-region, fig.325f). L3v* of *Heterogamodes* might correspond to either L4M or L4K of *Polyphaga* and *Ergaula* (fig.325k). However, neither L4M nor L4K nor any other sclerite of *Polyphaga* and *Ergaula* is strictly homologous with L4D (discussion in 6.3.4.). Hence, the sclerites L3v* of *Heterogamodes* and *Periplaneta* are certainly not homologous. L3v* of *Therea* is possibly homologous with L3 (on the hla-hook) of *Ergaula*, *Polyphaga*, and *Periplaneta*.

L2d* of *Periplaneta* is sclerite L4C (L4I- and L4d-regions, fig.325f). L2d* of *Therea* and *Heterogamodes* probably correspond to L4N (on the pda-process) of *Polyphaga* and *Ergaula* (fig.325k). L4N, however, is not strictly homologous with L4C but only with the posterior part of L4C (discussion in 6.3.4.).

L3d* of *Periplaneta* and *Heterogamodes* – I agree with this homology assumption – correspond to sclerite L3 of Blattinae, *Polyphaga*, and *Ergaula* (on the hla-hook in fig.53, 117). L3d* of *Therea* occupies a shallow bulge (not a long hook as hla is), and hla is hence supposed to be reduced; however, the long and somewhat hook-like process, whose sclerotisation is designated L3v*, is in my view more likely to be hla. I suppose that L3d* of *Therea* is sclerite L4K, which is on a shallow bulge like in *Ergaula* (fig.326d). In *Cryptocercus* (fig.6 of Grandcolas, fig.150, 151) the sclerites are designated as follows: L3* (Grandcolas probably assumes a fusion of L3v* and L3d*) is L4N. Hence, L4N and pda of *Cryptocercus* are regarded as the homologues of L3 and hla of *Polyphaga* and *Ergaula* (fig.117) and Blattinae (fig.53). The hla-hook of the other species is thus supposed to be quite reduced in *Cryptocercus* (as Grandcolas also supposes for *Therea*, which assumption, however, is probably not true).

L2d* is L3. Hence, L3 and hla of *Cryptocercus* are regarded as the homologues of L4N and pda of *Polyphaga* and *Ergaula* (fig.117) and of L4C and pda of Blattinae (fig.53).

In my view, hla and L3 of *Cryptocercus* are homologous with hla and L3 of the other species (discussion in 6.4.3.), and pda and L4N of *Cryptocercus* are homologous with pda and L4N (or the posterior part of L4C, respectively) of the other species (discussion in 6.3.4.). These relations are clearly demonstrated by the muscles inserting on these elements (compare e.g. 114 in *Eurycotis* and *Cryptocercus*, fig.72, 157) and by the relative positions of the respective elements (compare the dorsoventral arrangement of the posterior

part of L4C, pda, L3, and hla in *Archiblatta* and of L4N, pda, L3, and hla in *Cryptocercus*, fig.65, 150). As a result, Grandcolas has certainly misidentified L2d* and L3* in *Cryptocercus* (as compared with Blattinae, *Polyphaga*, *Ergaula*, and probably *Heterogamodes*).

As regards *Cryptocercus* and *Therea*, the sclerites L3* (or L3v* and L3d*) as well as the L2d* are probably also not homologous (compare the interpretation of the *Therea*-sclerites above), since in *Therea* L3v* is ventral to L2d*, whereas in *Cryptocercus* L3* is dorsal to L2d*. No argument is given to explain this difference.

Sclerite R2* sensu Grandcolas (C)

As regards *Periplaneta*, Grandcolas follows McKittrick (1964) in the definition and demarcation of $\mathbf{R2^*}$. This $\mathbf{R2^*}$ includes sclerite $\mathbf{R2}$ but additionally the regions $\mathbf{R1t}$ and $\mathbf{R1c}$ (= sclerite $\mathbf{R1F}$). The two sclerites of this $\mathbf{R2^*}$, $\mathbf{R2}$ and $\mathbf{R1F}$, are dorsoventrally articulated with each other in A6 (fig.75, 76). The homology relations which Grandcolas assumes concerning the $\mathbf{R2^*}$ -sclerotisations are in some respects not completely clear. This is in part due to incomplete information about which sclerotisations are assigned to $\mathbf{R2^*}$ and to the incompleteness of the figure showing the phallomeres of *Periplaneta*.

In fig.1 showing *Periplaneta*, Grandcolas labels that sclerite $R2^*$ which I designate R2 in Blattinae / Polyzosteriinae (fig.75-77). However, **R1F** is not contained in this figure. Since in McKittrick **R1F** is assigned to **R2*** (McKittrick 1964, fig.108: the central and the right parts of the slerite termed **R2**), **R1F** is probably part of **R2*** in the terminology of Grandcolas.

For Polyphaginae and *Cryptocercus* Grandcolas evidently assumes a fusion of the two sclerites of **R2*** (**R2**, **R1F**) and a concomitant loss of articulation **A6**. This is indicated by one of the supposed synapomorphies of *Cryptocercus* and Polyphaginae: "Sclerite **R2*** with two tubercles, which are not articulated dorso-ventrally" (p.151). (The only articulation within McKittrick's **R2** is **A6** between my sclerites **R2** and **R1F**, fig.75, 76, which is a ground-plan feature of Blattaria). In fig.3 and 5 showing *Heterogamodes* and *Therea*, Grandcolas labels that sclerite **R2*** which I designate **R2** in *Polyphaga* and *Ergaula* (fig.135-137); the entire sclerite **R1M** is designated **R3d***, without any contribution of **R2***. In fig.6 showing *Cryptocercus*, Grandcolas labels that sclerite **R2** which I designate **R3d***, without any contribution of **R2***. In fig.6 showing *Cryptocercus*, Grandcolas labels that sclerite **R2** is designated **N*** (see below in (**D**)). Hence, **R1F = R2*** of *Cryptocercus* and **R2 = R2*** of Polyphaginae are regarded as the results of this fusion and as strictly homologous. I cannot agree with these homology assumptions:

- That R1F of Cryptocercus has developed by a fusion of R1F and R2 of Periplaneta is certainly wrong: In Cryptocercus R1F and R2 take the same relative positions as R1F and R2 do in Periplaneta (and in Eurycotis, compare fig.75 and 161 and in 6.7.4., 6.7.6.), and these sclerites are certainly strictly homologous. Articulation A6 is in Cryptocercus as well-developed as in Eurycotis.

- That R2 of Polyphaginae has developed by a fusion of R1F and R2 of *Periplaneta* is, in my view, also wrong: In *Polyphaga* and *Ergaula* the sclerotisation homologous with R1F of *Periplaneta* (regions R1c and R1t) is contained in the anterior part of sclerite R1M (compare fig.332e and i), and the R2-sclerites of these species are strictly

homologous (fig.332e,i). Since the morphology of the respective part of the right phallomere of *Therea* and *Heterogamodes* is similar to *Polyphaga* and *Ergaula*, the same relations are assumed for these species.

- The resulting assumption that **R1F** of *Cryptocercus* is homologous with **R2** of Polyphaginae is also refuted.

Sclerite N* sensu Grandcolas (D)

Sclerite N* of both *Therea* and *Heterogamodes* certainly corresponds to L8 of *Polyphaga* and *Ergaula* (fig.117), which is situated on the rightmost part of the left complex, close to the right phallomere. According to Grandcolas, a sclerite N* is also present in *Cryptocercus*. From its relative position shown in Grandcolas' fig.6 results that this N* is R2 (fig.161-163): it articulates with both R1F = R2* and the left posterior end of R3 = R3v* (articulations A6 and A7), and its general shape and position also fit. However, in my opinion (compare in (C)) this R2 = N* of *Cryptocercus* is not homologous with L8 of *Polyphaga* and *Ergaula* but with R2 of *Polyphaga*, *Ergaula*, *Periplaneta*, and *Eurycotis* (fig.75, 135, 161; compare in 6.7.4.), and L8 is missing in *Cryptocercus*.

The vp*-lobe (ventral phallomere) sensu Grandcolas and its sclerotisation (E)

In *Periplaneta*, *Therea*, and *Cryptocercus* Grandcolas (fig.1, 5, 6) designates a sclerotised lobe in the median ventral wall of the phallomere complex as the ventral phallomere **vp*** (**vla**-lobe in my terminology). As regards *Cryptocercus* and *Periplaneta* I agree with him: The lobe is the true **vla** with sclerite **L4G** in its ventral wall (fig.63, 148). In *Therea*, however, since this species is closely related to *Ergaula*, the lobe concerned can be regarded as homologous with the **lba**-lobe of *Polyphaga* and *Ergaula*, which bears sclerite **L7** in its ventral wall (fig.115).

The figures showing the general phallomere structure of Blattinae and Polyphaginae (Grandcolas' fig.2, 4) furthermore reveal that Grandcolas assumes homology for L7 of Polyphaginae (including *Polyphaga*, *Ergaula*, and *Therea*) and L4G of *Cryptocercus* and Blattinae / Polyzosteriinae (the white posteromedian sclerites in these figures). In my opinion this assumption is wrong: In 6.2. and 6.3. the area belonging to the ventral phallomere or vla-lobe of *Polyphaga* has been identified, and the sclerotisation homologous with L4G of the other species has proved to be contained in L4M (fig.325e,f,h,k); the position of the ventral insertion of muscle l6a, the position of the vla-lobe relative to the lve-pouch, and the **lobe** corresponds only to the rightmost part of the vla-lobe of the other species, and that L7 is a new sclerite of *Lamproblatta* + *Polyphaga* + *Ergaula* (and probably of at least some other Polyphaginae).

Side-homologies according to Grandcolas (F)

In *Cryptocercus*, Polyphaginae, and Blattinae Grandcolas assumes side-homologies for the sclerites of the left half and of the right half of the phallomere complex. The only argument is that an identical number of sclerites with similar form and position were recognisable on each side of the ejaculatory duct opening (p.146).

I cannot agree with these assumptions: The number of sclerites is not identical on both sides (compare in Grandcolas' fig.2, 4), and the shapes and relative positions of the sclerites supposed to be side-homologous are far from being similar on both sides (compare e.g. L2v*/L2d* and R2* in fig.1, 3, 5, 6 of Grandcolas). Furthermore, the position of the genital opening, defined as the center of symmetry of these supposed side-homologies, has been identified incorrectly in Polyphaginae and *Cryptocercus* (compare (A)).

Generally, a superficial correspondence in the number, arrangement, and shapes of the sclerites on the left and on the right side could well indicate side-homologies, but the muscles should be investigated in terms of confirmation or contradiction. The musculature, however, does not at all support the side-homologies assumed by Grandcolas.

9.2. The phylogenetic position of Cryptocercus

Cryptocercus punctulatus, Polyphaga aegyptiaca, and *Ergaula capensis* are, apart from Blattinae, the only species investigated in both Grandcolas (1994) and this paper. According to my results *Ergaula* and *Polyphaga* are more closely related, according to Grandcolas *Ergaula* and *Cryptocercus* are more closely related.

Grandcolas lists many autapomorphies suggesting the holophyly of various groupings of Polyphagidae. The autapomorphies of all groupings containing *Cryptocercus* will be discussed subsequently according to their hierarchy, focused on the question whether the features listed provide arguments to include *Cryptocercus* in the respective grouping. The first three groupings include *Cryptocercus*, *Ergaula*, and *Polyphaga* and are not contradictory of my results. The fourth grouping includes *Cryptocercus* and *Ergaula* but not *Polyphaga* and is directly in conflict with my results. If this latter grouping – with or without *Crytocercus* – proves to be holophyletic, the last two groupings subordinate to it are also in conflict with my results.

Many of these autapomorphies relate to those phallomere sclerites for which Grandcolas' homology assumptions for *Cryptocercus* and Polyphaginae have been refuted in 9.1. (A)-(E), and they are in my opinion not valid; they will be commented with "misidentification", and the letter of the respective discussion in 9.1. will be added for reference. The autapomorphies are numbered like in Grandcolas (no numbers used in the first two groupings). From the quotations references like "in male genitalia" will be omitted. The autapomorphies concerned with tibial, head, "paraproct" (= subanal lobe), or female genital morphology have been reinvestigated. The autapomorphies 10, 16, and 17 of Grandcolas have been omitted since they refer to characters of the wings, which are completely absent in *Cryptocercus*.

Polyphagidae (including Cryptocercus)

- "Sclerite $L2v^*$ with the form of an arch invaginated in ventro-posterior direction." An arch-shaped $L2 = L2v^*$ extending along an invagination (lve-pouch) is a feature of the common ground-plan of Blattaria and Mantodea (6.2.1., 7.1.).
- "Sclerite L1* with thick-lipped edges." This probably refers to the plateau-like anterior face of the pne-pouch and of L1, and this is probably a synapomorphy of the respective

species (compare in 7.3., subgroup 2.2.2.; secondary reduction has been assumed for *Lamproblatta*).

- "Sclerite L1* turned on the ejaculatory duct and its opening."
- "Apical apodema of sclerite L1* curved around the ejaculatory duct opening."
- These two features probably refer to the hood-like shape of sclerite L1 and to its close relation to the opening of the phallomere-gland (= "ejaculatory duct": compare in (A)). Both features, however, are present in the common ground-plan of Blattaria and Mantodea (6.1.1., 7.1.).
- "Sclerite **R2*** with two tubercles, which are not articulated dorso-ventrally." Misidentification (C).
- "Female paraprocts with a membranous area in their sub-basal and internal parts." According to fig.13 of Grandcolas (= fig.334a in this paper) this refers to a membranous area at the median base of the subanal lobes (similar to Y in fig.321c). However, a membranous area taking the same position is also present in the females of e.g. *Periplaneta* (fig.334e), *Deropeltis* (fig.334d), and *Lamproblatta* (fig.334c) and is certainly not an autapomorphy of Polyphagidae or Polyphagidae + *Cryptocercus*.
- "Straight, long and narrow paratergites." According to my own investigations (Klass, in press: te and tg in fig.15, 16), the fused paratergites of the abdominal segments 8 and 9 of the females are, as compared with *Periplaneta*, somewhat lengthened and narrowed in *Cryptocercus* but not in *Polyphaga*. Apart from this, a slight change of the proportions of sclerite elements is in my view not very convincing as an autapomorphy.
- "L1* pourvu d'une dilatation basale" (Grandcolas & Deleporte 1992). This feature probably relates to the transverse expansion of L1 at its posterior margin, which continues towards both sides into the extensions L1l and L1m (fig.120, 153, 323i,l). However, a similar expansion, with at least one extension L1m, is also present in e.g. *Mantoida* (fig.49, 323d), and this is certainly a feature of the common ground-plan of Blattaria and Mantodea (6.1.1.). The extensions or regions L1l and L1r are also not restricted to *Cryptocercus* and Polyphagidae (compare e.g. *Tryonicus angustus*, fig.107, 323h, and *Nahublattella*, fig.243, 244, 323n).

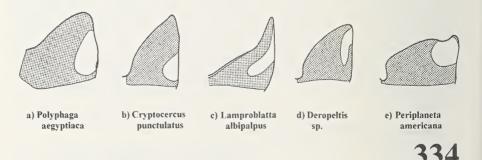


Fig.334: Paraprocts of female Blattaria. – Ventral view of left subanal lobe; posterior \uparrow , anterior \downarrow , median \rightarrow . Sclerotised areas (paraproct) are stippled, membranous areas are white. Fig.334a according to Grandcolas (1994).

Polyphaginae (including Cryptocercus)

- "Hook sclerite L3d* directed internally and posteriorly." Misidentification (B).
- "Tubercles of the sclerite **R2**^{*} fused together." Misidentification (C).
- "Sclerite L3v* plate-like." Misidentification (B).
- "Spermatheque des femelles nettement bifide" (Grandcolas & Deleporte 1992). The polarity of this character (spermatheca bifid or unbranched) is unclear, but the bifid condition is certainly not a synapomorphy of *Cryptocercus* und Polyphaginae since it is also present in e.g. Blattinae, Polyzosteriinae, *Lamproblatta*, and *Mastotermes* (McKittrick 1964).

Cryptocercus + Therea + Eucorydia + Ergaula + Polyphaga + Eupolyphaga + Anisogamia

- 1 "Sclerite **R2*** with the fore tubercle showing a sharp outer apophysis." Misidentification (**C**).
- 2 "Neoformation N*, right to L1*, presenting a ventral loop." Misidentification (D).

Cryptocercus + Therea + Eucorydia + Ergaula

- 9 "Inner apophysis of sclerite L2d* less sharp." Misidentification (B).
- 11 "Presence of an expanded and warty area on the inner basal part of the anterior arch." In the female genitalia, the left and right second valvifers are narrowly connected with each other at their anterior margins by a median transverse bridge (anterior arch of McKittrick 1964). Grandcolas probably refers to a posteriad expansion of the sclerotisation of the second valvifers towards the bases of the second and third valves, which is lateral to this transverse bridge (compare Klass, in press: fig.2, 3). "warty" might refer to the small and thick setae in this area. The expansions as well as the bristles are present in *Cryptocercus*, but also in e.g. *Sphodromantis, Lamproblatta*, and *Eurycotis* (own investigations). If I have understood this autapomorphy correctly, it has to be refuted.
- 12 "Apical spur lacking on the outer caudal margin of the fore tibiae." I have investigated the spurs of the fore tibiae in *Polyphaga*, *Ergaula capucina*, *Cryptocercus*, *Lamproblatta*, and *Deropeltis* (fig.335a-e). All these species have 5 apical spurs, whose bases are either outside or inside the sclerotisation of the tibia. These apical spurs can be homologised one by one, if the slightly curved row of spurs z, y, x is taken as a landmark. The apical spur at the distal end of this row, which is always outside the tibial sclerotisation, has been arbitrarily termed 1. *Ergaula, Polyphaga, Lamproblatta*, and *Deropeltis* correspond in their sets of apical spurs: Two adjacent spurs at the inner caudal margin of the fore tibiae are outside the tibial sclerotisation (1,5), three other spurs at the outer caudal margin are inside the tibial sclerotisation (2,3,4). Only in *Cryptocercus* spur **5** is inside the tibial sclerotisation. Hence, the apical spurs of *Cryptocercus* and *Ergaula* do not show any special situation in common differing from the other species. The autapomorphy is refuted.
- 13 "Neoformation N* adjacent to L1 horizontal." Misidentification (D).
- 14 "Spermatheca sclerite vertical." Grandcolas probably refers to the orientation of the spermathecal plate of the female genitalia (McKittrick 1964). This sclerite, which is

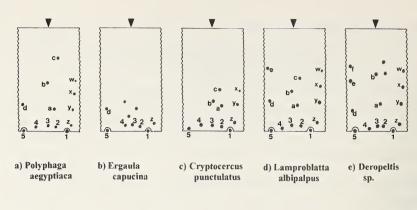


Fig.335: Spurs on fore-tibiae of Blattaria. – The sclerotisation and the spurs of the left fore-tibia are shown; basal \uparrow , distal \downarrow . The area bordered by straight or undulate lines is the sclerotisation of the tibia. This sclerotisation is cut lengthwise along the ventral = inner edge of the tibia (undulate lines) and unfolded. Black arrows mark the dorsal = outer edge of the tibia. Black dots represent the bases of spurs. Most spurs are labelled with numbers (apical spurs) or small letters – according to the homology relations assumed. Some apical spurs have their base outside the tibial sclerotisation.

vestigial in *Cryptocercus*, has a vertical orientation also in *Lamproblatta* and *Sphodromantis*, and the posterior main part of the sclerite of Blattinae and Polyzosteriinae is vertical, too. (These sclerotisations lie within the posterior wall of the bulge containing the spermathecal opening (compare Klass, in press: fig.2, 3). Thus, the vertical orientation is certainly not an autapomorphy of this grouping.

15 "Sclerite **R2*** with a hind tubercle large and rounded." Misidentification (C).

Cryptocercus + Therea + Eucorydia

- 25 "Fore tubercle of R2* very small." Misidentification (C).
- 26 "Hind tubercle of R2* fused with R3v*." Misidentification (C). Moreover, R1F (= R2*) of *Cryptocercus* is in no place fused with R3 (= R3v*). (According to fig.6 of Grandcolas the articulation A3, fig.163, is probably regarded as the point of "fusion").
- 27 "Frontal maculae of circular outline." These frontal maculae are more or less clearly demarcated cuticular areas median to the antennal bases. I have investigated them in the following species (from externally only): In *Ergaula capucina* they are clearly demarcated and like in Grandcolas, fig.16 drop-shaped. In *Deropeltis* and *Polyphaga* they are nearly circular. In *Cryptocercus* and *Lamproblatta* no maculae could be found. According to this distribution of the character states the autapomorphy is refuted.
- 28 "Postclypeus little or even not rounded." It is not clear whether this feature refers to (1) the bulging of the postclypeus or to (2) the arch-like course of its anterior margin (= sutura epistomalis). According to (1), this feature would be like in e.g. *Periplaneta*



or *Sphodromantis*, whose clypei are hardly bulged. According to (2), I could not find a sutura epistomalis in *Cryptocercus*. In both cases the autapomorphy has to be refuted.

- 29 "Arch of L2v* horizontal." "Arch of L2v*" is probably the part of L2 within the lvepouch. However, the orientation of L2 in *Cryptocercus* is not or hardly different from that in *Polyphaga* or *Mantoida*. If, however, the lack of an upcurving of the right parts of L2 and lve is referred to (6.2.1., 6.2.4.), this feature, if really present in the three species, would be derived. However, according to fig.5 of Grandcolas, in *Therea* L1 and the right end of L2 are still in contact (articulation A2), whereas in *Cryptocercus* the loss of this contact A2 and the loss of the right part of L2 (upcurved in other Blattaria) are probably intercorrelated. Hence, the levelness of the right part of L2 would probably not be homologous in *Therea* and *Cryptocercus*.
- 30 "Basis of inner apophysis of L2d* widened." Misidentification (B).
- 31 "Neoformation N* protruding." Misidentification (D).
- 32 "L3v* as a narrow plate in dorso-caudal position." Misidentification (B).

Cryptocercus + Therea

- 34 "L3d* very shortened." Misidentification (B).
- 35 "Neoformation N* as a rod." Misidentification (D).

Conclusions

All surmised synapomorphies suggesting that *Ergaula*, *Eucorydia*, or *Therea* are more closely related to *Cryptocercus* than to *Polyphaga* are not valid or at least (only 29) questionable. On the other hand, in 7.3. many apomorphies have been listed which clearly suggest that at least *Ergaula* (and *Lamproblatta*) is more closely related to *Polyphaga* than to *Cryptocercus* (autapomorphies of the subgroups 2.2.2.2. and 2.2.2.2.2. in 7.4.). That *Therea* and *Eucorydia* are true members of Polyphaginae and that they are closely related to *Ergaula* is not questioned or even confirmed in the case of *Therea*, which shares at least one synapomorphy with *Ergaula* and several synapomorphies with *Ergaula* and *Polyphaga* (compare in 9.1.).

The synapomorphies of Grandcolas suggesting *Cryptocercus* to be a member of Polyphaginae or Polyphagidae are all not valid either. The only exception is the plateaulike anterior face of sclerite L1. However, *Cryptocercus* is probably closely related to Polyphaginae (autapomorphies of subgroup 2.2.2. in 7.4., but compare in 7.7.), and, if *Lamproblatta* is also included, *Cryptocercus* might well be assigned to the Polyphaginae sensu Grandcolas (representing the basalmost offshoot).

As regards the various other groups usually assigned to Polyphagidae (Holocompsinae, Euthyrrhaphinae, Latindiinae, and Tiviinae in Grandcolas, and some others), hardly anything is known about the morphology of their male and female genitalia, and their phylogenetic relationships are still open to question.

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APPENDIX

Synonymy of the terminology of the phallomere elements

LaGreca: Mantodea

LaGreca (1954) has introduced special terms for the phallomere elements of Mantodea (left column, taken directly from LaGreca 1954, p.27). Some of these are used for formative elements as well as for sclerites situated inside or upon them. In the following table the synonymy with the terms used in the present paper (right column) is given.

Fallomero dorsale di sinistra (fs, not fv)	Left complex minus vla-lobe and sclerite L4A
Lamina dorsale (ld)	Sclerite L4B
Lamina ventrale (lv)	Pouch lve and sclerite L2 (mainly region L2a)
Processo apicale (pa)	Process paa and region L2d
Apofisi falloide (af)	Process afa and sclerite L1B
Processo anteriore (pn)	Pouch pne and sclerite L1 or L1A
Lobo membranoso (lo)	Process loa
Fallomero dorsale di destra (fd)	Lobe fda and region R1d
Corpo del fallomero (fd)	Lobe fda and region R1d
Braccio mediale del fallomero (bm)	Leftmost part of lobe fda and region R1d
Apodema anteriore (an)	Sclerite R3 including apodeme age
Processo ventrale sclerificato (pv)	Tooth / ridge pva and region R1t
Piastra ventrale (pi)	Tooth / ridge pia and region R1v
Area sensoria (as)	(not treated in the present paper)
Fallomero ventrale (fv)	Lobe vla and remaining ventral wall of left complex and sclerite L4A
Processo articolare (ar)	The part of sclerite L4A near articulation A1
Lobo mediale (lm)	Rightmost part of lobe vla and sclerite L4A
Processo distale (pd)	Process pda and pertaining parts of L4 or L4A
Pene (p)	Lobe(s) goa next to genital opening

McKittrick: Blattaria

Synonymy is given for the terms used in the present paper and those used in McKittrick 1964, fig.106-126. Since my results concerning the homologies of the sclerites are different from those of McKittrick, the synonymy is different in the various subgroups, and some representative groups are selected. This synonymy is also valid for many taxonomic papers in which the terminology of McKittrick has been used (e.g. Roth 1974).

Blaberidae Supella		Blattellinae Nyctiborinae		Anaplecta	
McKittrick	present paper	McKittrick	present paper	McKittrick	present paper
Ll	R3, R2, R1T, R4	L2vm	L2 (inside lve: L2D)	L1	L1 (?)
L2vm	L2 (inside lve: L2D)	L2d (virga)	L2 (on via: L2E+L4N)	L2v	L4K
L2d (virga)	L2 (on via: L2E+L4N)	L2	L4U	L2vm	L2 (inside lve)
R2	L3 (on hla), L4U	L3	L3	L2d	L2 (on paa), L4N
		R2	R2, R1S, R1P	L3	L3
		R3	R3	R2	R2, R1N, L4G
				R3	R3
Nahublattelle	2	Cryptocercus		Blattinae,	
<i>Nahublattelle</i> Lophoblatta	-	Cryptocercus		Blattinae, Polyzosteriir	iae
	-	<i>Cryptocercus</i> McKittrick	present paper		nae present paper
Lophoblatta			present paper	Polyzosteriir	
Lophoblatta McKittrick	present paper	McKittrick		Polyzosteriin McKittrick	present paper
Lophoblatta McKittrick L1	present paper R2, R1N, R3	McKittrick L1	LI	Polyzosteriin McKittrick L1	present paper
Lophoblatta McKittrick L1 L2vm	present paper R2, R1N, R3 L2 (inside Ive: L2D)	McKittrick L1 L2v	L1 L2 (inside lve)	Polyzosteriin McKittrick L1 L2v	present paper L1 L2
Lophoblatta McKittrick L1 L2vm L2d	present paper R2, R1N, R3 L2 (inside lve: L2D) L1	McKittrick L1 L2v L2d	L1 L2 (inside lve) L2 (on paa)	Polyzosteriin McKittrick L1 L2v L2d	present paper L1 L2 L4C,D,E; L4H
Lophoblatta McKittrick L1 L2vm L2d R1	present paper R2, R1N, R3 L2 (inside Ive: L2D) L1 L2E+L4N	McKittrick L1 L2v L2d L3	L1 L2 (inside lve) L2 (on paa) L3	Polyzosteriin McKittrick L1 L2v L2d L3	present paper L1 L2 L4C,D,E; L4H L3

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