



Research article

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**A revision and phylogenetic analysis of the millipede genus
Oxidus Cook, 1911 (Polydesmida, Paradoxosomatidae)**

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Abstract. The genus *Oxidus* Cook, 1911 is revised to contain five species, *O. avia* (Verhoeff, 1937), *O. gigas* (Attems, 1953), *O. gracilis* (C.L. Koch, 1847), *O. riukiaria* (Verhoeff, 1940), and “*species inquirenda*” *O. obtusus* (Takakuwa, 1942). A cosmopolitan species, *O. gracilis*, is widely found in temperate and sub-tropical regions over the world, but other species have limited distribution in restricted regions, e.g., *O. gigas* in northern Vietnam, *O. riukiaria* and *O. avia* in the Ryukyu Islands (Japan). Four species, *O. gracilis*, *O. riukiaria*, *O. avia* and *O. gigas*, are confirmed as different from each other in gonopod characters, coloration and body size. The status of the last species, *O. obtusus*, is still doubtful and requires examination of further fresh material. The phylogenetic relationships among species of *Oxidus* is analyzed using two fragments of the mitochondrial genes COI (Cytochrome c Oxidase subunit I) and 16S rRNA. Three species of *Oxidus* are clearly separated from each other; *O. gigas* and *O. gracilis* form a monophyletic sister group with *O. riukiaria*. The genus *Oxidus* is also monophyletic and more closely related to the genus *Tylopus* Jeekel, 1968 than to the genera *Sellanucheza* Enghoff, Golovatch & Nguyen, 2004 or *Kronopolites* Attems, 1914. In addition, an identification key to species of *Oxidus* is provided.

Keywords. Millipedes, Diplopoda, *Oxidus*, molecular phylogeny, revision.

Nguyen A.D., Korsós Z., Jang K.-H. & Hwang U.-W. 2017. A revision and phylogenetic analysis of the millipede genus *Oxidus* Cook, 1911 (Polydesmida, Paradoxosomatidae). *European Journal of Taxonomy* 293: 1–22. <https://doi.org/10.5852/ejt.2017.293>

Introduction

The genus *Oxidus* was proposed for the species *Fontaria gracilis* C.L. Koch, 1847 by Cook (1911). However, Attems (1914) also proposed a subgenus, *Kalorthomorpha*, of the genus *Orthomorpha* Bollman, 1893 for the same species. Later, *Kalorthomorpha* was elevated to full generic rank and considered to be a junior synonym of *Oxidus* (Jeekel 1968).

The taxonomic history of the genus *Oxidus* is relatively complicated. Takakuwa (1942a, b) did not accept the genus *Oxidus*, and he reported ten species under the genus *Orthomorpha* from South Korea: *Orthomorpha avia* Verhoeff, 1937, *O. circofera* Verhoeff, 1931 (= *Orthomorphella pekuensis* according to Hoffman (1963)), *O. circofera affinis* Verhoeff, 1936 (= *Orthomorphella pekuensis* according to Hoffman (1963)), *O. coarctata* (De Sausure, 1860), *O. gracilis*, *O. nordenskiöldi* (Attems, 1909) (= *Cawjeekelia nordenskiöldi* according to Jeekel (1988)), *O. pekuensis* (Karsch, 1881) (= *Orthomorphella pekuensis*), *O. riukiaria* Verhoeff, 1940, *O. cristata* Takakuwa, 1942 (listed as *species incertae sedis* by Nguyen & Sierwald (2013)) and *O. obtusus* Takakuwa, 1942. On the contrary, Chamberlin (1945) recognised the genus *Oxidus* from Java (Indonesia) with eight species, *Oxidus sequens* Chamberlin, 1945, *O. lamellifer* Chamberlin, 1945, *O. filarius* (Attems, 1932), *O. pygmaeus* (Pocock, 1894), *O. hydrobiologicus* (Attems, 1927), *O. annex* Chamberlin, 1945, *O. malabaricus* Chamberlin, 1945 and *O. pangrangus* Chamberlin, 1945. All those species were subsequently assigned to other genera (Jeekel 1963b; Nguyen & Sierwald 2013).

Chamberlin & Wang (1953) referred all *circofera*, *gracilis*, *pekuensis*, *obtusus* and *nordenskiöldi* to the genus *Oxidus*. However, Takakuwa (1954) listed only four species of *Oxidus*, *O. gracilis*, *O. avia*, *O. riukiaria* and *O. obtusus*, for the Japanese fauna. Wang (1957) added another species, *Oxidus* (*Varyomorpha*) *hsientienensis* Wang, 1957 (= *Nedyopus hsientienensis* according to Chen *et al.* (2006)) from Taiwan. Miyosi (1959) considered only five species *Oxidus gracilis*, *O. circofera*, *O. nordenskiöldi*, *O. cristatus* and *O. obtusus* from Japan.

Jeekel (1963a, 1968) accepted the validity of the genus *Oxidus*, and listed only five species and subspecies, *O. avia*, *O. gracilis*, *O. gracilis* ssp. *gigas* (Attems, 1953), *O. obtusus* and *O. riukiaria*. *O. gracilis* *gigas* was poorly described as *Kalorthomorpha gracilis* *gigas* from North Vietnam (Attems 1953), and recently raised to full species rank by Enghoff *et al.* (2004).

O. gracilis (C.L. Koch, 1847) is a cosmopolitan species; *O. obtusus* (Takakuwa, 1942) was known from South Korea, both *O. avia* (Verhoeff, 1937) and *O. riukiaria* (Verhoeff, 1940) were reported from South Japan, and lastly *O. gigas* (Attems, 1953) was recoded from North Vietnam. Except for *O. gracilis*, all the remaining species have had no further records since their establishment. This paper seeks to answer three questions based on morphological and molecular analyses. Morphological data is used to classify species and molecular data is used to examine their phylogeny.

- 1) How many species are actually in the genus *Oxidus*?
- 2) What are the relationships among species of *Oxidus*?
- 3) Does the genus *Oxidus* form a monophyletic group with closely related groups, e.g., *Tylopus* and *Sellanucheza*?

Material and methods

Taxon sampling, identification and DNA extraction

Examined material was collected by hand in northern Vietnam and southern Japan, or was borrowed from the collections in Kyungpook National University (KNU) (Korea) and the Hungarian Museum of Natural History (HNHM). All were examined under an Olympus SZX10 microscope with a drawing tube.

Total genomic DNA was extracted from leg tissue using the DNAeasy Blood & Tissue Kit (Qiagen TM).

DNA amplification and sequencing

Fragments of two mitochondrial genes, cytochrome c oxidase subunit I (COI) and 16S rRNA, were amplified using polymerase chain reaction (PCR). Universal primers LCO-1490 and HCO-2918 (Folmer *et al.* 1994) or COI-1F20 (5'-ACT CTA CTA ATC ATA AGG AT-3') and COI-1R19 (5'-TAA ACC TCC GGG TGA CCAA-3') were used to amplify a 680 bp fragment of the COI gene. Primer sets 16Sa (5'-CGC CTG TTT AHC AAA AAC AT-3') -16Sb (5'-CCG GTY TGA ACT CAR ATC CA-3') or 16S-1F19 (5'- CCG GTT TGA ACT CAG ATCA-3') and 16S-1R20 (5'-TGA CTG TTT AGC AAA GAC AT-3') were used to amplify a 550 bp fragment of the 16S rRNA gene. PCR conditions for amplification of the 16S rRNA gene were: an initial denaturation at 95°C for 2 min followed by 36 cycles of 95°C for 20 sec, 45°C for 40 sec and 72°C for 1 min, and a final extension at 72° for 5 min. The PCR conditions for the amplification of COI were: initial denaturation at 94°C for 5 min followed by 38 cycles of 94°C for 45 seconds, 42°C for 45 seconds, 72°C for 90 seconds, and a final extension at 72°C for 5 min. After thermal cycling, 2 µl PCR products were screened for potentially successful amplification of a fragment of 16S or COI through electrophoresis in 1% agarose-TBE 1X. The electrophoresis was performed at conditions of 100 mA and 120 V for 1 hour.

About 20 µl of successfully amplified PCR products were purified using ExosapIT or the QIAquick PCR Purification Kit (Qiagen Inc.). Purified PCR products were sequenced at Solgen, Inc (Korea) in an Applied Biosystems automatic sequencer (ABI3130 XL) using the same primer sets used for initial PCR.

Alignment and phylogenetic analysis

Each successful sequence was manually checked using BioEdit ver. 7.1 (Hall 1999) and confirmed by BLAST searches (Altschul *et al.* 1990). All confirmed sequences were aligned with MUSCLE (Edgar 2004). Ambiguous nucleotide sites and gaps were removed using MEGA ver. 6.0 (Tamura *et al.* 2013). The reliability of the alignment was estimated using distance estimation and model of p-distance. Nucleotide frequencies were statistically calculated using MEGA 6.0. These COI sequences were translated into amino acids for confirmation using transversion code in MEGA 6.0.

Model test was implemented in MEGA 6.0 to find the most appropriate maximum likelihood substitution model for COI and 16S. Models with the lowest Bayesian Information Criterion (BIC) scores were considered for describing the best substitution pattern for each gene. Codon positions included were: 1st + 2nd + 3rd + Noncoding. The selected model for COI was the Tamura-Nei model + G + I (Tamura & Nei 1993). For the combination of COI and 16S rRNA, the Hasegawa-Kishino-Yano model (Hasegawa *et al.* 1985) was used. Phylogenic trees were constructed using both maximum likelihood (ML) and Bayesian inference (BI) models. Maximum likelihood bootstrap analysis was conducted using MEGA 6.0 with 1000 replicates. A Bayesian inference (BI) tree was created using MrBayes ver. 3.1.2 (Huelsenbeck & Ronquist 2001) with 10 million generations, heating parameter of 0.06, and sampling every 1000 generations. All nucleotide sequences were deposited at GenBank. Collection localities, specimen –

Table 1. Details of specimens analysed for DNA, including GenBank numbers.

No.	Species	Locality	Voucher	16S rRNA	COI
1	<i>Oxidus gigas</i> (Attems, 1953)	Sapa, Lao Cai, Vietnam	IEBR-Myr 113	KX096912	KX096921
2	<i>Oxidus gigas</i> (Attems, 1953)	Duc Xuan, Ha Giang, Vietnam	IEBR-Myr 516	KX096917	KX096928
3	<i>Oxidus riukiaria</i> (Verhoeff, 1940)	Okinawa, Japan	IEBR-H500	KX096915	KX096926
4	<i>Oxidus riukiaria</i> (Verhoeff, 1940)	Okinawa, Japan	IEBR-H500J	KX096916	KX096927
5	<i>Oxidus gracilis</i> (C.L. Koch, 1847)	USA	GenBank	–	KR136012
6	<i>Oxidus gracilis</i> (C.L. Koch, 1847)	USA	IEBR-USA	KX096919	KX096931
7	<i>Oxidus gracilis</i> (C.L. Koch, 1847)	Okinawa, Japan	IEBR-HNHM 466	KX096913	KX096924
8	<i>Oxidus gracilis</i> (C.L. Koch, 1847)	Okinawa, Japan	IEBR-HNHM 471	KX096914	KX096925
9	<i>Tylopus roseiparaterga</i> Nguyen, 2012	Tam Dao, Vinh Phuc, Vietnam	IEBR-Myr 185A	KX096909	KX096923
10	<i>Tylopus crassipes</i> Golovatch, 1984	Sapa, Lao Cai, Vietnam	IEBR-Myr 92	KX096907	KX096920
11	<i>Kronopolites montanus</i> Golovatch, 2009	Pu Mat, Nghe An, Vietnam	IEBR-Myr 175	KX096908	KX096922
12	<i>Kronopolites</i> sp.	Pu Mat, Nghe An, Vietnam	IEBR-Myr 553	KX096911	KX096930
13	<i>Tonkinosoma jeekeli</i> Nguyen, 2011	Cuc Phuong, Ninh Binh, Vietnam	IEBR-Myr 545	KX096910	KX096929
14	<i>Sellanucheza grandis</i> (Golovatch, 1984)	Huong Son, Ha Tinh, Vietnam	IEBR-Myr 59	–	KR818293
15	<i>Sellanucheza grandis</i> (Golovatch, 1984)	Xuan Son, Phu Tho, Vietnam	IEBR-Myr 177	–	KR818296
16	<i>Sellanucheza hoffmani</i> Nguyen, 2011	Phong Nha, Quang Binh, Vietnam	IEBR-Myr 182	–	KR818298
17	<i>Tonkinosoma jeekeli</i> Nguyen, 2011	Cat Ba, Hai Phong, Vietnam	IEBR-Myr 203	–	KR818299
18	<i>Tonkinsoma flexipes</i> Jeekel, 1953	Cat Ba, Hai Phong, Vietnam	IEBR-Myr 207	–	KR818300

vouchers, and GenBank accession numbers are summarized in Table 1. Datasets are provided in Fasta files as supplementary data.

The COI dataset consisted of 525 bp sequences from 18 taxa. The 16S rRNA dataset included 466 bp sequences from 13 taxa. The combination of 16S and COI genes contained 991 bp sequences from 12 taxa (Table 2). The incongruence length difference (ILD) test showed congruence of genes COI and 16S. Therefore, the analyses were performed for gene COI and the combination of 16S and COI. Species of the genus *Tonkinosoma* Jeekel, 1953 (tribe Tonkinosomatini Jeekel, 1968) were employed as an outgroup.

Abbreviations used in text and figures

The terminology for the genus *Oxidus* follows that used by Likhitrakarn *et al.* (2010) for the genus *Tylopus*:

ca	=	cannula
co	=	coxa
pf	=	prefemorate
fe	=	femorate
l	=	postfemoral lamina
z	=	postfemoral spine
h	=	postfemoral process
sl	=	solenomere
sph	=	solenophore

Institutional abbreviations

All materials and voucher specimens are kept in:

IEBR	=	Institute of Ecology and Biological Resources, Hanoi, Vietnam
IPE	=	Institute of Phylogenomics and Evolution, Daegu, Korea
KNU	=	Kyungpook National University, Daegu, Korea
HNHM	=	Hungarian Natural History Museum, Budapest, Hungary

Results

Order Polydesmida Leach, 1813
Family Paradoxosomatidae Daday, 1889
Tribe Sulciferini Jeekel, 1968

Genus ***Oxidus*** Cook, 1911

Oxidus Cook, 1911: 628.

Kalorthomorpha Attems, 1914: 195 (proposed as subgenus of *Orthomorpha* Bollman, 1893; junior objective synonym of *Oxidus*, see Jeekel 1968: 71).

Oxidus – Brölemann 1916: 537.

Type species

Fontaria gracilis C.L. Koch, 1847, by original designation.

Diagnosis

The genus can be distinguished from other genera in the tribe Sulciferini by the combination of characters: paraterga being well-developed; metaterga with one or two rows of setae; legs without tarsal brushes; 5th sternum without modifications.

Gonopod relatively complicated; femorate weakly twisted, slightly or strongly enlarged distally, strongly grooved mesally, well demarcated laterally from postfemoral lamina *l* by a distinct sulcus; solenophore with two basal processes: long, highly elevated process *h* and short, spiniform/tuberculiform, sub-transverse process *z*; solenophore twisted or strongly spiral, with mesal well-developed lobule, but without lateral lobules; solenomere flagelliform, completely sheathed by solenophore.

Remarks

This genus is distributed in the northern part of the Oriental and the eastern part of the Palearctic regions. It is unlikely that the genus is of tropical origin because it is absent from the tropical regions in the Oriental (Jeekel 1963a). Its center of origin is more likely to be Japan (Jeekel 1968).

The genus *Oxidus* was placed in Sulciferini in view of the characters of the spiral solenophore completely sheathing the solenomere, the presence of a postfemoral demarcation and of postfemoral processes (Jeekel 1968). Golovatch & Enghoff (1993) recommended that the genus should be placed relatively close to the genus *Tylopus* Jeekel, 1968, rather than to other sulciferinine genera. The phylogeny of the genus *Oxidus* is discussed below.

***Oxidus gracilis* (C.L. Koch, 1847)**

Figs 1–2

Fontaria gracilis C.L. Koch, 1847: 142.

Fontaria gracilis – Koch C.L. 1863: 51, pl. 85, fig. 173.

Oxidus gracilis – Cook 1911: 628. — Jeekel 1963a: 20; Jeekel 1968: 71. — Nguyen & Sierwald 2013: 1292.

Orthomorpha (*Kalorthomorpha*) *gracilis* – Attems 1914: 196.



Fig. 1. *Oxidus gracilis* (C.L. Koch, 1847) from Okinawa Island, Japan. A. Entire body, length ca 23 mm. B–C. Segments 8–9–10. B. ♂. C. ♀. Scale bars = 1 mm. (photos by Z. Korsós)

Material examined

JAPAN: 1 ♂, Central Ryukyus, Okinawa-jima Island, Onna-son, Onna, Camp Hansen training area at Gate 28, 20 m a.s.l., 26°29'44" N, 127°51'36" E, 31 Aug. 2010, leg. Z. Korsós (IEBR-501); 3 ♂♂, 3 ♀♀, Central Ryukyus, Okinawa-jima, Okuni rindo forest trail, 26°43'55" N, 128°12'30" E, 25 Sep. 2012, leg. Z. Korsós (IEBR-H471); 3 ♂♂, 3 ♀♀, Central Ryukyus, Okinawa-jima, Nago city, Makiya, Makiya-notaki waterfall, 26°37'44" N, 128°02'42" E, 100 m, 2 Sep. 2012, leg. Z. Korsós (IEBR-H466).

USA: 13 ♂♂, 5 ♀♀, Florida, Columbia, Co., Ichetucknee St., PK, 3 May 1981 (IEBR-USA).

SOUTH KOREA: 4 ♀♀ (IPE-Myr 001), 1 ♂, Geoje Island, 8 Aug. 1970, leg. K.Y. Paik (IPE-Myr 003); 7 ♀♀, Geoje Island, 7 Aug. 1970, leg. K.Y. Paik (IPE-Myr 002); 1 ♂, Jeollanam-do, Gwangju, 6 Oct. 1962, leg. S.C.Kim (IPE-Myr 004); 4 ♂♂, Songnisan National Park, 24 Jul. 1961, leg. G.Y. Baek (IPE-Myr 005); 1 ♂, Gyeongsangbuk-do, Cheongsong-gun, Hyeondong-myeon, Wolmae-ri, 18 Jul. 1966, leg. Kim Jae Ho (IPE-Myr 007); 3 ♂♂, 3 ♀♀, leg. K.Y. Paik (IPE-Myr 009).

TAIWAN: 1 ♂, Miaoli County, Taian Township, Kuanwu, SE ridge of Mt. Yemagan, 24°31'13" N, 121°07'11" E, 1925 m, secondary mixed forest, 20 Oct. 2009, leg. L. Dányi and E. Lazányi (HNHM-T09-51); 1 ♂, 1 ♀, Taichung County, Heping Township, Dasyueshan Forest Recreation Areas, 10 Shsuehshan Rd., 7 May 2003, leg. Shining Wu (IEBR-Myr 550); 3 ♂♂, 1 ♀, Miaoli County, Taian Township, Dalu Forest Rf., West Feeder, Guanwu lodge, 12 Aug. 2002, leg. Chen Chao Chun (IEBR-Myr 551).

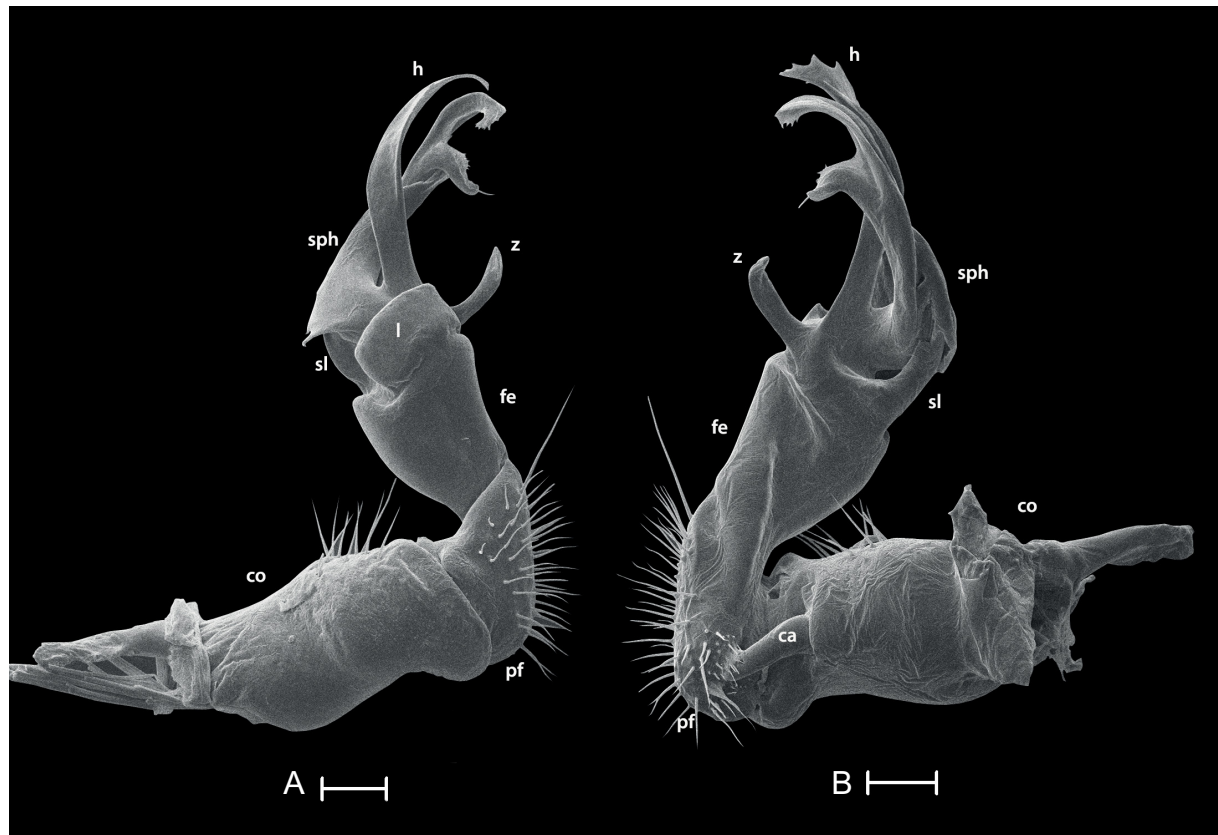


Fig. 2. Left gonopod of *Oxidus gracilis* (C.L. Koch, 1847), sample IEBR-USA. **A.** Lateral view. **B.** Mesal view. Scale bars = 100 µm.

Diagnosis

This species differs from its congeners in the gonopod femorite being strongly expanded distally, not cylindrically slender; postfemoral lamina *l* rectangular; spine *z* pointed tuberculiform; process *h* lamellar, slightly suberect, but bent upwards from midpart, serrated at distolateral part; both bases of process *h* and spine *z* clearly separated; mesal lobule of solenophore very well-developed, lamella-shaped, distinctly separated from tip of solenophore; tips of both mesal lobule and solenophore circularly emarginated.

DNA

For *Oxidus gracilis*, DNA data was collected for three genes (mitochondrial COI gene, nuclear 18S and 28S rRNA genes), but there is no data from the mitochondrial 16S rRNA. In this study, two fragments of COI and 16S were sequenced and deposited in GenBank (see Table 1).

Distribution

The species is widely distributed over the world, mainly in warm temperate regions, and associated with disturbed habitats (Nguyen & Sierwald 2013). Island records for the Ryukyu Archipelago, Japan: Tanega-shima, Nakano-shima, Akuseki-jima, Takara-jima, Amami-O-shima, Okinoerabu-shima, Yoron-to, Okinawa-jima, Senaga-jima, Miyagi-jima, Iheya-jima, Izena-jima, Ie-jima, Kudaka-jima, Tsuken-jima, Yakabi-jima, Tokashiki-jima, Tonaki-jima, Aguni-jima, Kume-jima, Miyako-jima, Ikema-jima, Ishigaki-jima, Iriomote-jima, Kohama-jima, Taketomi-jima, Aragusuku-jima, Hateruma-jima, Yonaguni-jima, Minami-Daito-jima.

Oxidus gigas (Attems, 1953)

Figs 3–4

Kalorthomorpha gracilis gigas Attems, 1953: 165, fig. 44.

Oxidus gracilis gigas – Jeekel 1963a: 20; Jeekel 1968: 60. — Golovatch 1983: 181.

Oxidus gracilis – Golovatch 1984: 54.

Oxidus gigas – Enghoff *et al.* 2004: 39. — Nguyen & Sierwald 2013: 1295.

Diagnosis

This species is particularly similar to *Oxidus gracilis* (C.L. Koch, 1847) in body appearance and gonopod conformation, but differs in larger size and in details of gonopod structure: process *h* narrow and long, pointed at the end and not serrated at distolateral margin; bases of process *h* and spine *z* less distinctly separated.

Material examined

VIETNAM: 1 ♂, 1 ♀, Lao Cai Prov., Sa Pa, Ham Rong Mts., limestone mountain, 1600 m, 16 Jul. 2006, Anh D. Nguyen, IEBR-113; 1 ♂, 6 ♀♀ (IEBR-Myr 516), 7 ♀♀ (IEBR-Myr 517), Ha Giang Prov., Bac Quang Distr., Duc Xuan Commune, Na Po village, limestone forest, under logs, 14–15 Apr. 2013, leg. Anh D. Nguyen; 1 ♂, 3 ♀♀, Vinh Phuc Prov., Tam Dao National Park, forest, 900–1000 m, Mar. 2012, leg. Hoang Long, IEBR-Myr 518; 2 ♂♂, 1 ♀, Lao Cai Prov., Co xan, in and around a cave, 1971, leg. Gy. Topál & I. Matskási, IEBR-H133.

Redescription

SIZE. Body length 31–34.5 mm (♂♂), 29.9–30.2 mm (♀♀); width of midbody pro- and metazonae 1.9–2.2 mm (♂), 2.2 mm (♀) and 2.8–3.2 mm (♂), 3.1–3.2 mm (♀), respectively.

COLORATION. Generally castaneous brown, but anterior half of metaterga and posterior margin of prozonae darkish brown. Posterior half of metaterga, anterior margin of prozonae and pleura castaneous brown; paraterga, sterna and legs brownish yellow.

HEAD. Slightly narrower than collum; labrum sparsely setose; epicranial suture distinct, dividing frons into two equal parts. Antennae long and slender, not claviform, reaching segment 3 if stretched posteriorly. Antennomere $1 < 7 < 2 = 3 = 4 = 5 = 6$ in length.

COLLUM. Subequal to, or slightly narrower than collum, trapeziform; surface shining and smooth, without rugosity; setae broken, but traces of two rows: 3+3 close to anterior margin and 1+1 in middle. Paraterga large, ear-shaped with broadly rounded laterocaudal corners; lateral side with a setiferous incision.

In width, segment $4 \leq 3 < 2 < 5-17$, thereafter gradually tapering towards telson. Prozonae and metaterga shining, smooth, without rugosity. Metaterga with two rows of setae: 2+2 close to anterior margin and



Fig. 3. *Oxidus gigas* (Attems, 1953) from Duc Xuan Commune, Ha Giang Prov., Vietnam. **A.** Entire body, length ca 34 mm. **B–C.** Segments 8–9–10. **B.** ♂. **C.** ♀. Scale bars = 1 mm. (photo by Anh Nguyen)

2+2 close to posterior margin. Transverse sulcus starting on metatergum 4, well developed, reaching base of paraterga on metaterga 5–19, beaded at bottom on some caudal segments. Stricture between pro- and metazonae clearly distinct, broad and striolate at bottom. Pleura shining, smooth; pleurosternal carinae completely absent or poorly developed only on pre-gonopodal segments.

PARATERGA. Well developed, lying subequal to or lower than metatergal surface. Calluses small, but obvious; lateral side with 2 setiferous incisions at $\frac{1}{3}$ and $\frac{2}{3}$ of its length. Anterior corner broadly rounded, but posterior corner acute, produced into a pointed projection on segments 15–17. Ozopores located behind second lateral incision of paraterga 5, 7, 9, 10, 12, 13, 15–19.

EPIPROCT. Long, but broadly truncated, dorsoventrally flattened, with two minute apical tubercles. Tip with four spinnerets. Hypoproct trapeziform, with two well separated, distolateral, setiferous knobs.

STERNA. Modestly setose, without modifications, but with a minute cone caudally near each coxa.

LEG. Long and slender, about 1.6–1.7 (♂), 1.4–1.5 (♀) times as long as midbody height. Tarsal brushes absent. Prefemora not swollen. Femora without modifications.

GONOPOD. Relatively complicated. Coxite cylindrical, long, subequal to femorite in length; distoventral part sparsely setose. Prefemorite densely setose, separated laterally from femorite by a distinct oblique sulcus. Femorite weakly twisted, grooved mesally, more or less slightly enlarged distally, without modifications and demarcated from postfemoral region by a lateral sulcus. Postfemoral region shorter than femorite, with a lateral sub-pentagonal lamina l; spine z pointed; process h long and pointed, not serrated laterally; bases of process h and spine z not separated. Solenophore strongly spiral; distomesal lobule well developed and distinctly separated from solenophore tip. Solenomere flagelliform, completely sheathed by solenophore.

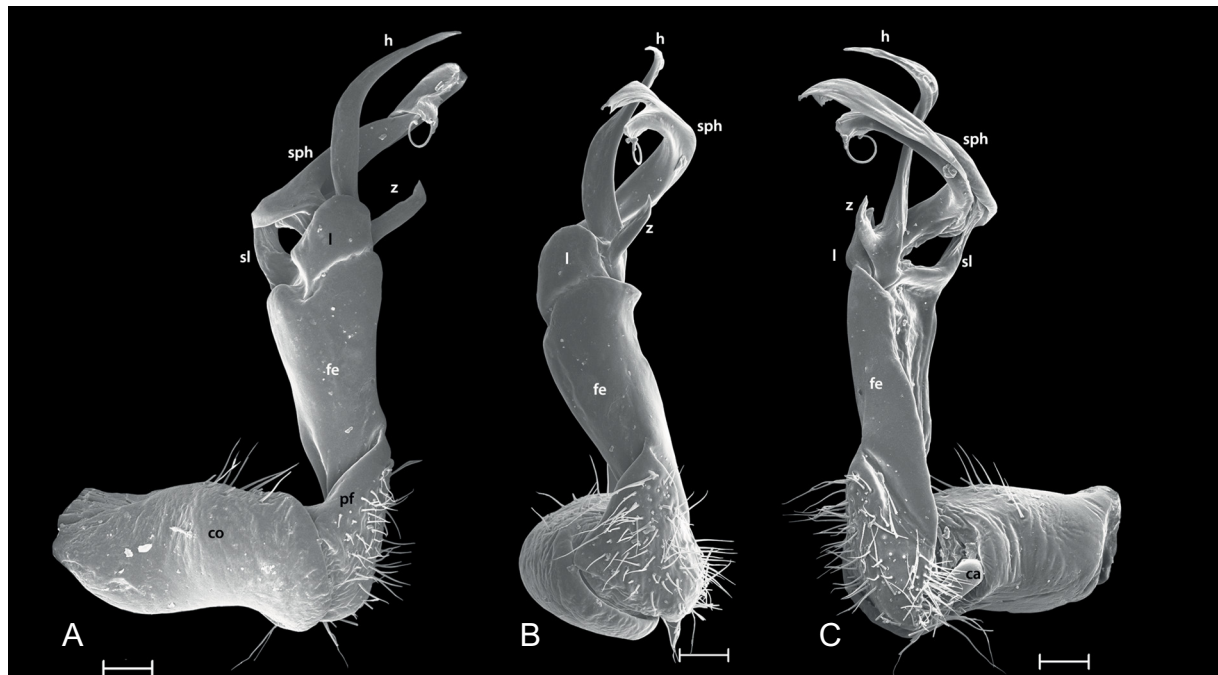


Fig. 4. Right gonopod of *Oxidus gigas* (Attems, 1953), from sample IEBR-Myr 113. **A.** Lateral view. **B.** Ventral view. **C.** Mesal view. The picture has been flipped horizontally. Scale bars = 1 μ m.

DNA

COI and 16S barcode data (partial) are deposited in GenBank (Table 1).

Habitats

All material was found under leaf-litter, logs and decaying wood.

Distribution

The species has only been recorded from northern Vietnam (Lao Cai, Ha Giang and Vinh Phuc Provinces).

Remarks

Attems (1953) proposed a new subspecies *Kalorthomorpha gracilis gigas* with a short note. Enghoff *et al.* (2004) raised this subspecies to full rank as *Oxidus gracilis*, and also showed minor differences in size and gonopod process *h*. The species is fairly similar to *O. gracilis*, but the molecular data has provided strong evidence to separate the species.

Golovatch (1984) misidentified *O. gracilis* in Vietnam (sample IEBR-H133), which is currently corrected as *O. gigas* in this paper.

Oxidus riukiaria (Verhoeff, 1940) Figs 5–7

Orthomorpha riukiaria Verhoeff, 1940: 139, figs 5–6.

Orthomorpha riukiaria – Takakuwa 1954: 39, fig. 34.

Oxidus riukiarius – Jeekel 1963a: 20.

Oxidus riukiaria – Nguyen & Sierwald 2013: 1295.

Material examined

JAPAN: 4 ♂♂, 4 ♀♀, Central Ryukyus, Okinawa-jima, Ogimi village, Nerome, above road no.58, 26°41'37" N, 128°06'43" E, 37 m a.s.l., 25 Sep. 2012, leg. Z. Korsós (IEBR-H470); 2 ♂♂, 2 ♀♀, Okinawa-jima, Yanbaru, Ogimi village, Nunha, Okuni rindo, 26°41'10.1" N, 128°09'02.7" E, 15 Sep. 2012, leg. Nakamura Y. (IEBR-H500); 2 ♂♂, Central Ryukyus, Okinawa Island, Yanbaru, Kunigami village, Cape Hedo, Ginama, broad-leaved evergreen forest, 176 m, 26°49'52" N, 128°16'20" E, 5 Apr. 2011, leg. Z. Korsós (HNHM-355); 1 ♂, Northern Ryukyus, Tokara group, Kuchino-shima Island, around Kuroshio-so guesthouse, 29°59'24" N, 129°55'21" E, 63 m, 14 Oct. 2012, leg. Z. Korsós (HNHM-477).

Other records (newly found ones underlined)

JAPAN: Northern Ryukyus, Tokara Group: Kuchino-shima Island; Central Ryukyus: Okinawa-jima, Kume-jima, Iheya-jima, Aguni-jima, Zamami-jima, Aka-jima, Geruma-jima (observations by Z. Korsós).

Diagnosis

Oxidus riukiaria can be distinguished from all other species of *Oxidus* by its smaller size (19.3–20.6 mm in length, width of pro- and metazonae 1.5–1.8 mm and 2.1–2.2 mm); gonopod coxae more or less stouter and longer than femorite; femorite narrow at base, and strongly expanded towards distal end; gonopod process *h* hook-like, larger and getting narrower towards pointed tip, spine *z* small, pointed spiniform; postfemoral lamina *l* triangularly rounded; mesal lobule not separated from solenophore tip, which is distinctly emarginated.

DNA

COI and 16S barcode data (partial) are deposited in GenBank (Table 1).

Remarks

This species was originally described by Verhoeff (1940) in the genus *Orthomorpha*, but was assigned to *Oxidus* by Jeekel (1963a). The taxonomic position of the species is also well supported by molecular data.

Distribution

South Japan (Ryukyu Islands).

Oxidus avia (Verhoeff, 1937)

Fig. 8A

Orthomorpha avia Verhoeff, 1937: 33, fig. 1.



Fig. 5. *Oxidus riukiaria* (Verhoeff, 1940) from Okinawa, Japan. A. Entire body, length ca 20 mm. B–C. Segments 8–9–10. B. ♂. C. ♀. Scale bars = 1 mm. (photos by Z. Korsós)

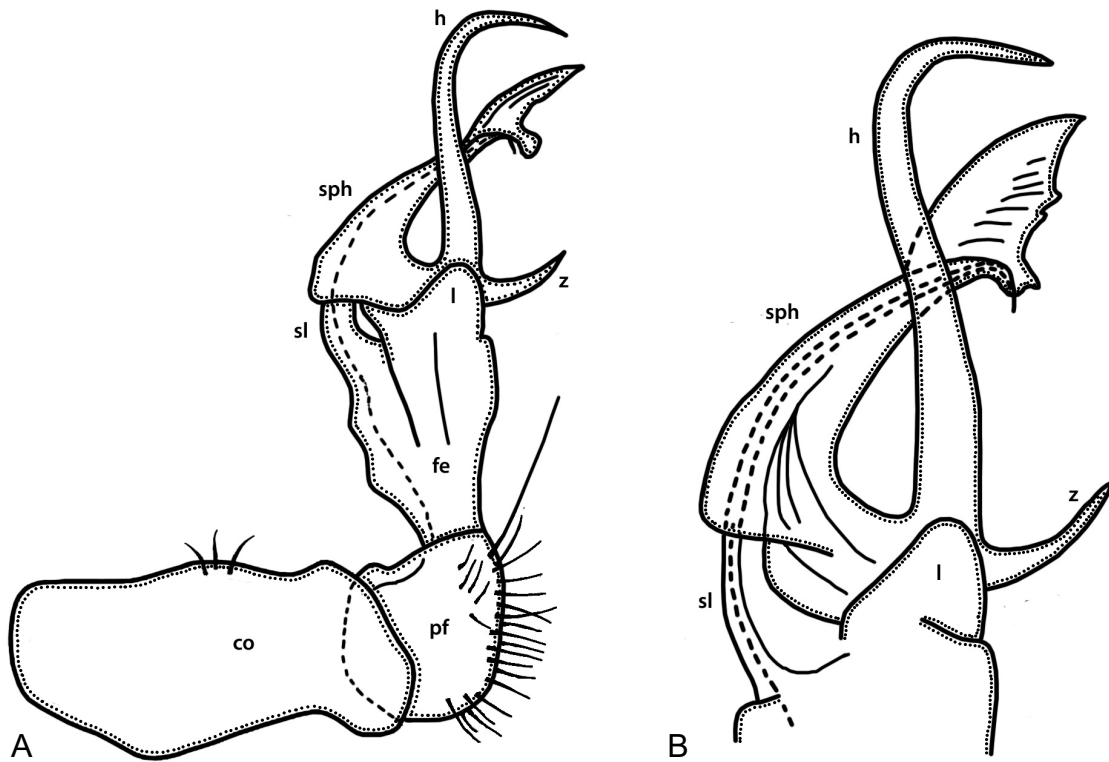


Fig. 6. Left gonopod of *Oxidus riukiaria* (Verhoeff, 1940). **A.** Lateral view. **B.** Postfemoral region, lateral view. Redrawn from Verhoeff 1940. No scale bars.

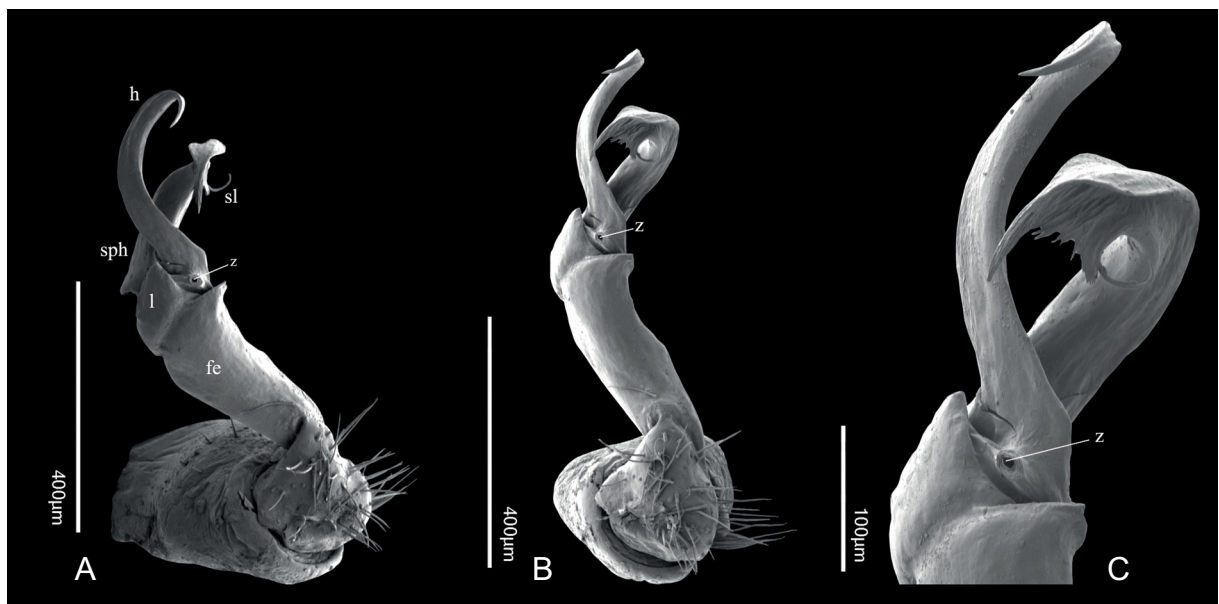


Fig. 7. Left gonopod of *Oxidus riukiaria* (Verhoeff, 1940), from sample IEBR-H470. **A.** Lateroventral view. **B.** Ventral view. **C.** Postfemoral region, ventral view. Note: z = spine z, but broken.

Oxidus avia – Takakuwa 1954: 38, fig. 33. — Jeekel 1963a: 20. — Nguyen & Sierwald 2013: 1295.

Diagnosis

This species can be separated from all other species of *Oxidus* by its much darker, almost uniformly dark brown body, slender base of gonopod solenophore, more clearly and obtusely emarginated posterior edge of telson; smaller postfemoral lamina *l*; stouter spine *z*; process *h* more erect, slender and pointed; the base of solenophore less protruded, more strongly rounded; mesal lobule distinctly separated from rounded solenophore tip.

Remarks

The species was originally described by Verhoeff (1937) under the genus *Orthomorpha*, then was assigned to the genus *Oxidus* by Takakuwa (1954).

Distribution

South Japan (Ryukyu Islands): Ishigaki-jima, Yonaguni-jima; Taiwan.

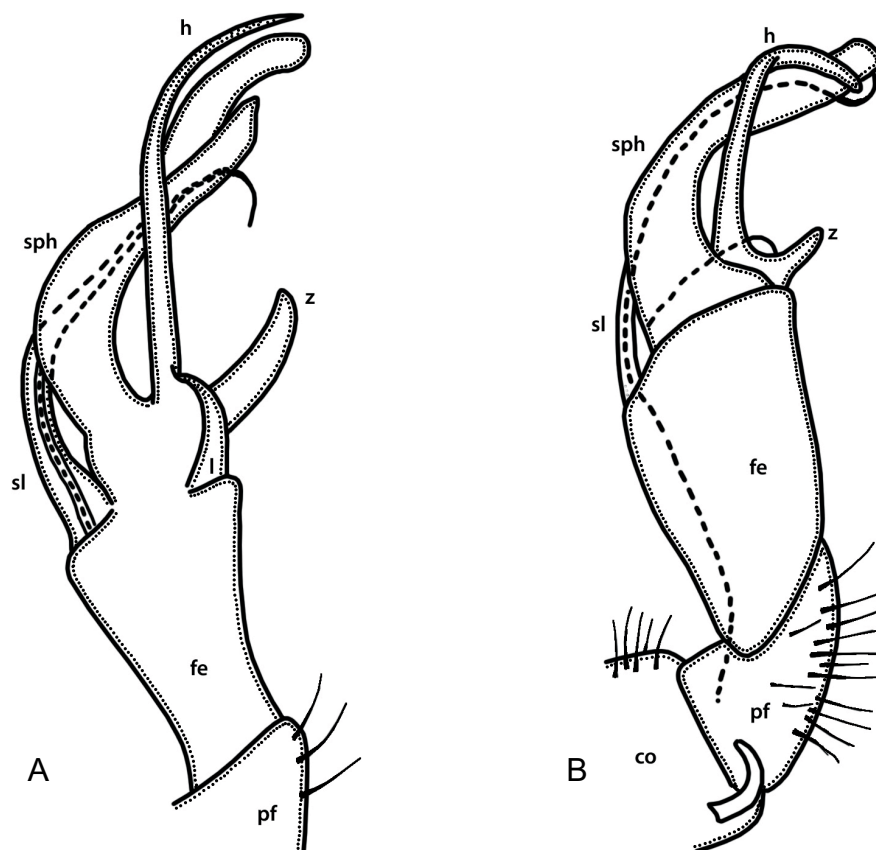


Fig. 8. **A.** Left gonopod of *Oxidus avia* (Verhoeff, 1937), lateral view. **B.** Right gonopod of *Oxidus obtusus* (Takakuwa, 1942), mesal view. Redrawn from Verhoeff 1937 and Takakuwa 1942a, respectively. No scale bars.

***Oxidus obtusus* (Takakuwa, 1942)**

Fig. 8B

Orthomorpha obtusa Takakuwa, 1942a: 363, 367, fig. 7.

Oxidus obtusus – Chamberlin & Wang 1953: 6. — Takakuwa 1954: 37. — Jeekel 1963a: 20. — Nguyen & Sierwald 2013: 1295.

Diagnosis

This species can be distinguished by its gonopod femorite being short, very broad gently distad; process *h* bent upward at tip, like a lamellar sickle; spine *z* small, obtuse; solenophore large, broad and simple, especially wide at its base; mesal lobule not separated from rounded solenophore tip (Takakuwa 1942a).

Remarks

This species was described from South Korea (Takakuwa 1942a). Later, Chamberlin & Wang (1953) reported two ♀♀ in the collection of the American Natural History Museum (New York) from Japan. It is possible that the locality was mislabelled because they stated that those specimens were collected and deposited by Dr. Takakuwa Y. Thus, it is believed that these two ♀♀ were collected from the same locality as the species holotype (South Korea).

Lim (2001) synonymized *O. obtusus* with *O. gracilis*. He argued that the two species differ only in the number of postfemoral branches, three in *O. obtusus* and four in *O. gracilis*; in his opinion, such a difference is minor and cannot be used to separate two species. With the two species *O. gracilis* and *O. gigas*, those characters are important for species delimitation. It is recommended, therefore, that *Oxidus obtusus* should be considered a valid species. Examination of further fresh material is needed to confirm that recommendation.

Distribution

South Korea.

Key to species of the genus *Oxidus* Cook, 1911

1. Gonopod femorite stout, more or less cylindrical. Spine *z* small and short, blunt spiniform ***O. obtusus*** (Takakuwa, 1942)
- Gonopod femorite strongly expanded distad, not cylindrical. Spine *z* long, pointed tuberculiform or spiniform 2
2. Postfemoral lamina *l* small, spiniform. Spine *z* stout, tip more or less rounded, tuberculiform. Base of solenophore slender; mesal lobule clearly separated from rounded tip of solenophore ***O. avia*** (Verhoeff, 1937)
- Postfemoral lamina *l* large, not spiniform. Spine *z* pointed spiniform. Base of solenophore not slender; tip of solenophore emarginated 3
3. Postfemoral lamina *l* triangular. Mesal lobule not separated from solenophore tip ***O. riukiaria*** (Verhoeff, 1940)
- Postfemoral lamina *l* not triangular. Mesal lobule clearly separated from solenophore tip 4
4. Postfemoral lamina *l* sub-pentagonal. Process *h* and spine *z* not separated clearly at base of postfemoral region. Process *h* pointed ***O. gigas*** (Attems, 1953)
- Postfemoral region *l* rectangular. Process *h* and spine *z* clearly separated at base of postfemoral region. Process *h* serrated along upper edge ***O. gracilis*** (C.L. Koch, 1847)

Table 2. Average base frequencies of two partial genes COI, 16S rRNA and the 16S-COI combination.

Genes	Average base frequencies				Length	Variable sites	Parsimony informative sites
	A	T	G	C			
COI	20.6	42.4	23.0	13.9	525	177	157
16S rRNA	31.9	35.7	24.0	8.4	466	199	161
16S-COI combination	26.0	39.1	23.3	11.6	991	367	297

Molecular phylogeny of the genus Oxidus Cook, 1911

DNA variations and distances

The aligned dataset for COI consisted of 525 bp sequences. The nucleotide frequencies for A, T, G, and C were 20.6%, 42.4%, 23.0%, and 13.9%, respectively. The GC content was 37%. The COI dataset contained 157 (29.9%) parsimony informative and 177 (33.7%) variable sites (see Table 2). The uncorrected p-distance between taxa ranged from 0.000 to 0.192. The overall p-distance was 0.146.

The aligned dataset for the 16S rRNA gene contained 466 bp sequences. The nucleotide frequencies of A, T, G, and C were 31.9%, 35.7%, 24.0%, and 8.4%, respectively. The GC content was 32.4%. The 16S rRNA dataset contained 161 (14.7%) parsimony informative and 199 (34.7%) variable sites (see Table 2). The uncorrected p-distance between the taxa ranged from 0.000 to 0.279. The overall p-distance was 0.169.

The aligned dataset for the combination of the 16S rRNA and COI genes contained 991 bp sequences. The nucleotide frequencies of A, T, G, and C were 26.0%, 39.1%, 23.3%, and 11.6%, respectively. The GC content was 34.9%. This combined dataset contained 297 (30.0%) parsimony informative and 367 (37.0%) variable sites (see Table 2). The uncorrected p-distance between the taxa ranged from 0.000 to 0.229. The overall p-distance was 0.157 (Table 3). Within the genus *Oxidus*, the three species compared had significant genetic distances: 0.106-0.113 between *O. gigas* and *O. gracilis*; 0.119-0.128 between *O. gigas* and *O. riukiaria*; and 0.103-0.106 between *O. gracilis* and *O. riukiaria*. The genetic distance between species of *Oxidus* and other genera ranged from 0.162 (between *O. riukiaria* and *Tylopus roseiparaterga*) to 0.229 (between *O. gigas* and *Kronopolites* sp.).

Phylogenetic analyses

Phylogenetic trees were reconstructed for the COI and 16S-COI combination using ML and BI. For ML-analysis, we considered clades with bootstrap values below 65%, between 65 and 89%, or more than 89% to be weakly supported, moderately supported, or strongly supported, respectively (Pimvichai *et al.* 2014). For BI-analysis, clades with a BI posterior probability less than 0.7 bpp, between 0.7 and 0.95 bpp, or more than 0.95 bpp are considered to be weakly supported, moderately supported, or strongly supported, respectively.

Based on a Bayesian Inference and Maximum Likelihood analysis of a 525 bp fragment of COI from 18 taxa, a phylogenetic tree was reconstructed (Fig. 9). The genus *Oxidus* formed a monophyletic clade. It was separated from its sister group (genus *Tylopus*) with an ML bootstrap value of < 65% and a BI posterior probability of 0.76 bpp. *Sellanucheza* is considered as the sister group of *Tylopus* plus *Oxidus*. The separation of *Sellanucheza* and (*Tylopus* plus *Oxidus*) is supported with an ML bootstrap value of <65% and a BI posterior probability of 0.98 bpp. Within *Oxidus*, the three species formed three clearly distinguished clades. However, *O. gigas* was more closely related to *O. gracilis*, with an ML bootstrap value of 74% and a BI posterior probability of 0.86 bpp. *O. riukiaria* was separated from the group of

Table 3. Uncorrected distance of the combination of COI and 16S rRNA genes calculated by MEGA 6.0. The genetic distances among species of *Oxidus* are highlighted in bold.

	Species / p-distance	1	2	3	4	5	6	7	8	9	10	11
1	<i>Tonkinosoma jeekeli</i> (545)											
2	<i>Kronopolites montanus</i> (175)	0.196										
3	<i>Kronopolites</i> sp. (IPE4)	0.196	0.010									
4	<i>Tylopus crassipes</i> (92)	0.201	0.225	0.220								
5	<i>Tylopus roseiparaterga</i> (185A)	0.205	0.207	0.203	0.119							
6	<i>Oxidus gigas</i> (516)	0.201	0.225	0.229	0.163	0.186						
7	<i>Oxidus gigas</i> (133)	0.203	0.224	0.223	0.172	0.187	0.040					
8	<i>Oxidus gracilis</i> (466)	0.188	0.205	0.201	0.166	0.172	0.113	0.108				
9	<i>Oxidus gracilis</i> (471)	0.190	0.205	0.201	0.169	0.172	0.113	0.106	0.002			
10	<i>Oxidus gracilis</i> (USA)	0.188	0.205	0.201	0.166	0.172	0.113	0.108	0.000	0.002		
11	<i>Oxidus riukiaria</i> (500)	0.186	0.208	0.208	0.166	0.170	0.128	0.125	0.106	0.104	0.106	
12	<i>Oxidus riukiaria</i> (500J)	0.186	0.204	0.206	0.169	0.162	0.128	0.119	0.104	0.103	0.104	0.037

O. gigas and *O. gracilis* with a well supported ML bootstrap value of 98% and a BI posterior probability of 1.00 bpp.

Based on BI and ML analysis of the 991bp combination of the genes 16S rRNA and COI, a phylogenetic tree of 12 taxa was constructed (Fig. 10). As in the COI tree, the genus *Oxidus* formed a monophyletic clade clearly separated from species of *Tylopus*, with a well supported ML bootstrap value of 100% and a BI posterior probability of 1.00 bpp. The three *Oxidus* species were also separated from each other. *O. riukiaria* formed a sister group with *O. gigas* plus *O. gracilis*, with an ML bootstrap value of 100% and a BI posterior probability of 1.00 bpp. The species *O. gigas* was separated from *O. gracilis* with a moderately supported ML bootstrap value of 83% and a BI posterior probability of 0.91 bpp.

Discussion

Both phylogenetic trees showed the monophyly of each *Oxidus* species (Figs 9–10). In other words, all three species are distinctly separated from each other (genetic distances: 0.106–0.113 between *O. gigas* and *O. gracilis*; 0.119–0.128 between *O. gigas* and *O. riukiaria*; and 0.103–0.106 between *O. gracilis* and *O. riukiaria*). The species *O. gigas* was originally proposed as a subspecies of *O. gracilis* (Attems, 1953). After being raised to full species rank, it is still closer to *O. gracilis* than to other species of *Oxidus*. This relationship is well supported by a monophyletic clade of the two species, *O. gracilis* and *O. gigas*. The other species, *O. riukiaria*, is considered as the sister species of *O. gracilis* plus *O. gigas*.

The genus *Oxidus* is placed in the Sulciferini with the typical characters of a spiral solenophore completely sheathing the solenomere, presence of postfemoral demarcation, and postfemoral processes (Jeekel 1968). Until now, no relationship analysis among sulciferine genera has been reported, except that Golovatch & Enghoff (1993) have reported a close relationship among species of *Tylopus* and *Oxidus gracilis*. These authors recommended that *Oxidus gracilis* or the genus *Oxidus* should be considered as the sister of genus *Tylopus* because both genera have similar gonopod characters, such as the presence of lamina *l*, process *h*, and spine *z*. Molecular analysis based on the COI gene and the combination of 16S and COI genes provides good supports for the recommendation of Golovatch & Enghoff (1993). In both phylogenetic trees, *Oxidus* and *Tylopus* formed a group with well supported bootstrap value and BI posterior probability (Figs 9–10).

The genus *Sellanucheza* Enghoff, Golovatch & Nguyen, 2004 was originally assigned to the tribe Tonkinosomatini (Enghoff *et al.* 2004). However, Golovatch (2013) suggested that it was better to place this genus in the tribe Sulciferini. His suggestion is well supported by the molecular data obtained from this study. The genus *Sellanucheza* is relatively close to the group of *Tylopus* + *Oxidus*, with a very high BI value (0.98 bpp) in the phylogenetic tree inferred from the 525 bp fragment of the COI gene (Fig. 10).

Conclusion

The genus *Oxidus* Cook, 1911 consists of five species, *O. avia*, *O. gigas*, *O. gracilis*, *O. riukiaria*, and a doubtful species, *O. obtusus*. While *O. gracilis* is a cosmopolitan species, widely found in temperate and sub-tropical regions over the world, the other species have limited distributions in particular regions: *O. gigas* in northern Vietnam, and *O. riukiaria* and *O. avia* in the Ryukyu Islands (Japan).

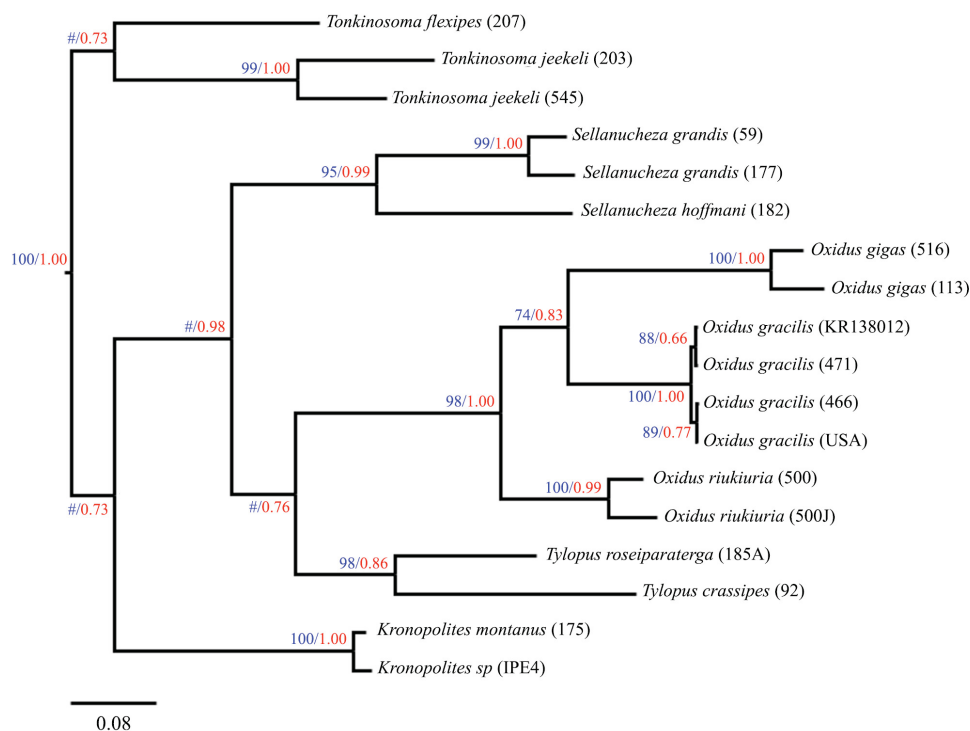


Fig. 9. Phylogenetic tree of the genus *Oxidus* and some closely related groups based on Maximum Likelihood and Bayesian Inference Analysis of a 525 bp fragment of the COI gene (# = a value less than 65%).

Oxidus is a monophyletic genus, and most closely related to the genus *Tylopus* based on the molecular analysis. All three species, *O. gigas*, *O. gracilis* and *O. riukiaria*, are distinctly separated and *O. gigas* is closer to *O. gracilis* than to *O. riukiaria*.

Acknowledgements

The authors thank the National Science Foundation of Korea for their support through the postdoctoral fellowship awarded to A.D. Nguyen, and Dr. Hsueh-Wen Chang (Sun Yat-Sen National University, Taiwan) for his kindly providing specimens of *Oxidus gracilis*. The second author would like to express his sincere gratitude to Prof. H. Ota and M. Toda (Tropical Biosphere Research Center, University of the Ryukyus, Okinawa) for inviting him to study millipedes in the Ryukyus. Two anonymous reviewers are acknowledged for their invaluable comments and suggestions to improve the paper.

This research is funded by the Vietnam National Foundation for Science and Technology Development (NAFOSTED) under grant number 106-NN.05-2015.22 to A.D. Nguyen, and the National Research Foundation of Korea (NRF), Ministry of Science, ICT & Future Planning under grant NRF-2012R1A2A2A01046531 to U.W. Hwang.

References

Altschul S.F., Madden T.L., Schäffer A.A., Zhang J., Zhang Z., Miller W. & Lipman D.J. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research* 25: 3389–3402.

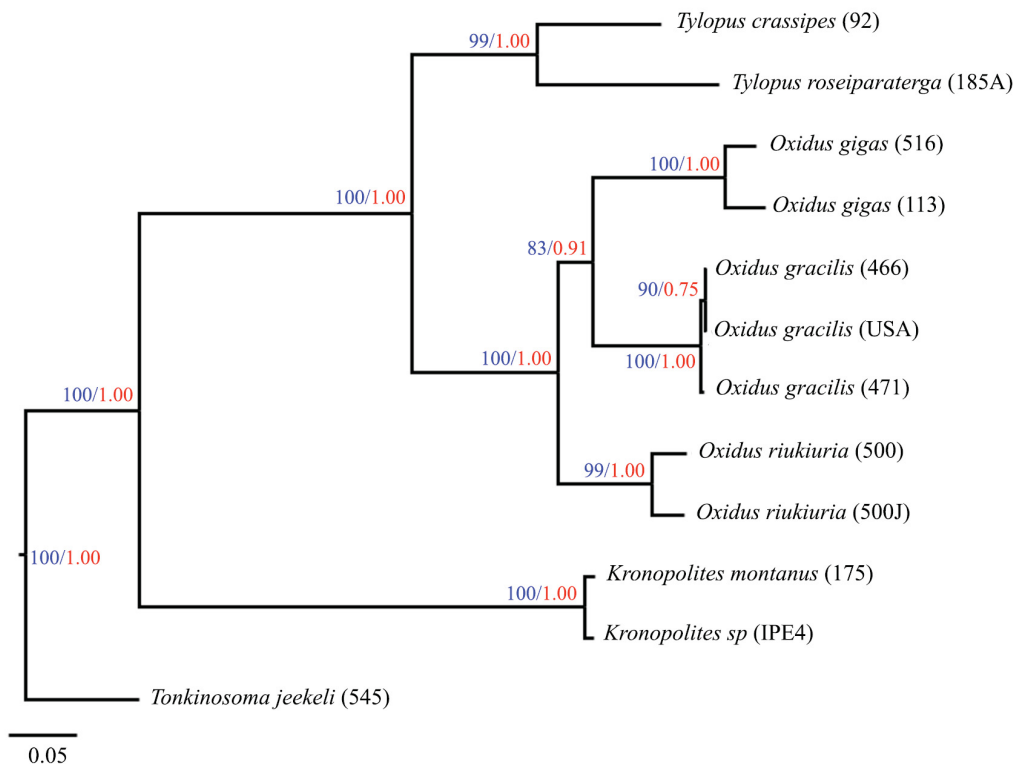


Fig. 10. Phylogenetic tree of the genus *Oxidus* and some closely related groups based on Maximum Likelihood and Bayesian Inference Analysis of a 991 bp fragment of the combination of 16S rRNA and COI genes.

- Attems C. 1914. Die indo-australischen Myriopoden. *Archiv für Naturgeschichte* 80A (4): 1–398.
- Attems C. 1953. *Myriopoden von Indochina: Expedition von C. Dawydoff (1938–1939)*. Mémoires du Muséum national d'Histoire naturelle, Série A – Zoologie 5 (3), Muséum national d'Histoire naturelle, Paris.
- Bollman C.H. 1893. The Myriapoda of North America. *Bulletin of the United States National Museum* 46: 1–210.
- Brölemann H.W. 1916. Essai de classification des Polydesmiens [Myriapodes]. *Annales de la Société entomologique de France* 84: 523–608. Available from <http://biodiversitylibrary.org/page/8559505> [accessed 6 Jan. 2017].
- Chamberlin R.V. 1945. On some diplopods from the Indo-Australian Archipelago. *American Museum Novitates* 1282: 1–43.
- Chamberlin R.V. & Wang Y.M. 1953. Records of millipeds (Diplopoda) from Japan and other Oriental areas, with descriptions of new genera and species. *American Museum Novitates* 1621: 1–13.
- Chen C.-C., Golovatch S.I. & Chang H.-W. 2006. The millipede tribe Sulciferini in Taiwan (Diplopoda: Polydesmida: Paradoxosomatidae). *Norwegian Journal of Entomology* 53 (2): 253–255.
- Cook O.F. 1911. The hothouse milliped as a new genus. *Proceedings of the United States National Museum* 40(1842): 625–631.
- Edgar R.C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* 32 (5): 1792–1797. <https://doi.org/10.1093/nar/gkh340>
- Enghoff H., Golovatch S.I. & Nguyen D.A. 2004. A review of the millipede fauna of Vietnam (Diplopoda). *Arthropoda Selecta* 13 (1/2): 29–43.
- Folmer O., Black M., Hoeh W., Lutz R. & Vrijenhoek R. 1994. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3: 294–299.
- Golovatch S.I. 1983. Millipedes (Diplopoda) of the fauna of Vietnam. In: Sokolov V.E. (ed.) *Fauna and Ecology of the Animals of Vietnam*: 178–186. “Nauka” Publishing, Moscow.
- Golovatch S.I. 1984. Contribution to the millipede fauna of Vietnam (Diplopoda), II. *Acta Zoologica Hungarica* 30 (1–2): 53–77.
- Golovatch S.I. 2013. On several new or poorly-known Oriental Paradoxosomatidae (Diplopoda: Polydesmida), XIII. *Arthropoda Selecta* 22 (1): 1–31.
- Golovatch S.I. & Enghoff H. 1993. Review of the genus *Tylopus*, with descriptions of new species from Thailand (Diplopoda, Polydesmida, Paradoxosomatidae). *Steenstrupia* 19 (3): 85–125.
- Hall T.A. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95–98.
- Hasegawa M., Kishino H. & Yano T. 1985. Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *Journal of Molecular Evolution* 22 (2): 160–174. <https://doi.org/10.1007/BF02101694>
- Hoffman R.L. 1963. A contribution to the knowledge of Asiatic strongylosomoid Diplopoda (Polydesmida: Strongylosomatidae). *The Annals and Magazine of Natural History*, Series 13 5 (58): 577–593. <https://doi.org/10.1080/00222936208651289>
- Huelsenbeck J.P. & Bollback J.P. 2001. Empirical and hierarchical Bayesian estimation of ancestral states. *Systematic Biology* 50: 351–366. <http://www.jstor.org/stable/3070928>

- Jeekel C.A.W. 1963a. Diplopoda of Guiana. *Studies on the Fauna of Suriname and other Guyanas* 4 (11): 1–157.
- Jeekel C.A.W. 1963b. Paradoxosomatidae from Borneo (Diplopoda, Polydesmida). *Tijdschrift voor Entomologie* 106 (6): 205–283.
- Jeekel C.A.W. 1968. *On the Classification and Geographical Distribution of the Family Paradoxosomatidae (Diplopoda, Polydesmida)*. PhD Thesis, University of Amsterdam, The Netherlands.
- Jeekel C.A.W. 1988. The generic position of *Orthomorpha bucharensis* Lohmander and *O. mumina* Gulicka, and the taxonomic status of *Hedinomorpha* Verhoeff (Diplopoda, Polydesmida, Paradoxosomatidae). *Bulletin Zoologisch Museum, Universiteit van Amsterdam* 11 (11): 97–104.
- Koch C.L. 1847. System der Myriapoden mit den Verzeichnissen und Berichtigungen zu Deutschlands Crustaceen, Myriapoden und Arachniden. In: Panzer G.W.F. & Herrich-Schäffer A. (eds) *Kritische Revision der Insectenfauna Deutschlands, III*: 1–196. Bändchen, Regensburg.
- Koch C.L. 1863. *Die Myriapoden 2. Getreu nach der Natur abgebildet und beschrieben*. Druck und Verlag von H.W. Schmidt, Halle.
- Likhitrakarn N., Golovatch S.I., Prateepasen R. & Panha S. 2010. Review of the genus *Tylopus* Jeekel, 1968, with descriptions of five new species from Thailand (Diplopoda, Polydesmida, Paradoxosomatidae). *ZooKeys* 72: 23–68. <https://doi.org/10.3897/zookeys.72.744>
- Lim K.Y. 2001. *Taxonomy of Millipedes (Arthropoda: Diplopoda) in Korea*. PhD dissertation, Chonbuk National University, Jeonju, Korea.
- Miyosi Y. 1959. *Über japanische Diplopoden*. Special Number, Arachnological Society of East Asia, Osaka.
- Nguyen A.D. & Sierwald P. 2013. A worldwide catalog of the family Paradoxosomatidae Daday, 1889 (Diplopoda: Polydesmida). *CheckList* 9: 1132–1353.
- Pimvichai P., Enghoff H. & Panha S. 2014. Molecular phylogeny of the *Thyropygus allevatus* group of giant millipedes and some closely related groups. *Molecular Phylogenetics and Evolution* 71: 170–183. <https://doi.org/10.1016/j.ympev.2013.11.006>
- Takakuwa Y. 1942a. Die Myriopoden aus Formosa, Philippinen u.s.w. *Transactions of the Natural History Society of Formosa* 32 (231): 359–367.
- Takakuwa Y. 1942b. Die *Orthomorpha*-Arten aus Japan. *Transactions of the Natural History Society of Formosa* 32 (224): 44–47.
- Takakuwa Y. 1954. *Diplopoden aus Japan und ihn angrenzenden Gebieten*. Japan Society for the Promotion of Science, Tokyo.
- Tamura K. & Nei M. 1993. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10 (3): 512–526.
- Tamura K., Stecher G., Peterson D., Filipski A. & Kumar S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30: 2725–2729.
- Verhoeff K.W. 1937. Zur Kenntnis ostasiatischer Diplopoden. II. *Zoologischer Anzeiger* 119 (1/2): 33–40.
- Verhoeff K.W. 1940. Zur Kenntnis ostasiatischer Diplopoden. V (Polydesmoidea). *Zoologischer Anzeiger* 131 (5/6): 129–145.

Wang Y.M. 1957. Serica 1g: Records of myriapods on Taiwan Islands (4). Six new polydesmids. *Quarterly Journal of the Taiwan Museum* 10 (3/4): 103–111.

Manuscript received: 26 May 2016

Manuscript accepted: 5 July 2016

Published on: 7 March 2017

Topic editor: Rudy Jocqué

Desk editor: Laurence Bénichou

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Zeitschrift/Journal: [European Journal of Taxonomy](#)

Jahr/Year: 2017

Band/Volume: [0293](#)

Autor(en)/Author(s): Nguyen Anh D., Korsos Zoltan, Jang Kuem-Hee, Hwang Ui-Wook

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