

Diagnostic Key for the Neotropical Alticine Genera

(Coleoptera: Chrysomelidae: Alticinae)*

By Gerhard Scherer

Zoologische Staatssammlung München
(with 42 figures)

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* Translation from the German with complementary notes and additions. Title of the German edition (1962) Bestimmungsschlüssel der neotropischen Alticinen-Genera. – Ent. Arb. Mus. Frey 13 (2): 497–607. Not included in this translation are the descriptions of *Meraaltica*, *Meraaltica punctata*, and the list of Clark's species of Monoplatini.

Preface to the English Translation of the Key to Neotropical Alticinae Genera (1962)

At the end of the 1950's I had started to compile a listing of the genera of neotropical Alticinae. This was a difficult task which took a lot of time, and would so again for anyone attempting to duplicate this. A reviser would have one benefit, however, that is of knowing what genera are now available.

At that earlier time, I failed to include two genera: *Apleuraltica* and *Procalus*. *Procalus* had been described in the subfamily Galerucinae. *Terpnochlorus* was not yet discovered in the neotropics. *Ayalaia* was not included because I knew it only from its description. Several genera published by Bechyné in "Beiträge zur Neotropischen Fauna" (1959) were not included because Bechyné's paper, privately published and not generally publicized, was unknown to me. Besides these genera, 36 new genera have been described in the mean time. The oriental genus *Nonarthra* must be added to the neotropical fauna, because of the introduction of a single species into Panama (Balsbaugh, 1982).

For all his work with this translation, I would like to thank my friend and colleague, Dr. Edward U. Balsbaugh, Jr., North Dakota State University, currently the best authority for the nearctic and adjacent neotropical chrysomelid fauna. Now translated into English, my key will have a larger circle of readers. I also hope that a reviser will emerge, for there is a decided need for a revision. For the genera described since 1962, I am providing only a alphabetical list noting genotype, morphological characters and placement in the classification. Most of these genera I know only from the literature.

Gerhard Scherer

Munich, December 1, 1981

This paper is an English translation of the original German by Gerhard Scherer which was first published in the Entomologischen Arbeiten aus dem Museum Frey, Tutzing, Vol. 13, Issue 2, 1 October 1962. In as much as it is the only synoptic modern work dealing with the Neotropic Alticinae, it is of considerable value in spite of having been written 20 years ago. Many taxonomic changes have occurred in this fauna during these intervening years. We have made no corrections in factual content to include such changes, but we do accept responsibility for having inadvertently introduced errors. Likewise we have employed a rather liberal interpretation of Scherer's style so that the translation is as consistent as possible with American taxonomic phraseology. We also gratefully acknowledge the tremendous assistance of Dr. James L. Krysan, U.S. Department of Agriculture, Northern Grain Insect Research Laboratory, Brookings, S. D., who arranged for both initial colloquial translation by Dr. Wayne M. Frericks, and for the typing of the manuscript.

Edward U. Balsbaugh, Jr.

Entomology Department, North Dakota State University

Eric H. Smith

ORKIN Pest Control, Atlanta, Georgia

Genera not included in my key (1962)

Terpnochlorus, *Nonarthra*, and *Procalus* are included in the text.

Andiroba Bechyné & Bechyné, 1965

Monotypic: *Andiroba egleri* Bech. & Bech. (Para/Amapá)

Close to *Brasilaphthona*; anterior coxal cavities closed; pronotum very convex, nearly spherical; femora of 4 anterior legs robust.

Anelytropleurica Bechyné & Bechyné, 1964

Monotypic: *Anelytropleurica anthicina* Bech. & Bech. (Brazil: Mato Grosso)

Close to *Syphraea*; without elytropleural carina; length 2.3–2.8 mm.

Apleuraltica Bechyné, 1956

Monotypic: *Apleuraltica curculionoides* Bechyné (Guadeloupe)

Close to *Aedmon*; antennae short, apically strongly delated; sides of pronotum not margined; cylindrical; apterous.

Araguaenia Bechyné, 1968

Monotypic: *Araguaenia microtricha* Bechyné (Venezuela)

Close to *Cacoscelis*; antebasal impression on pronotum weak and indistinct, but lateral folds distinct; punctures on elytra confused with longitudinal tendency.

Archilactica Bechyné & Bechyné, 1975

Monotypic: *Archilactica gabonioides* Bech. & Bech. (Venezuela)

Close to *Monomacra*; antebasal impression on pronotum angulate at sides.

Asphaerina Bechyné, 1963

Monotypic: *Asphaera fallax* Clark (Brazil)

Close to *Asphaera*; in the middle of the prosternum a protuberance (♂) epipleura and elytropleura large.

Ayalaia Bechyné & Bechyné, 1960

Genotype: *Ayalaia salvadorensis* Bech. & Bech. (El Salvador)

Close to *Syphraea*, *Parasyphraea*, *Resistenciana*; shape somewhat resembles *Monomacra* and *Varicoxa*. Different from these genera by its triangular frons which is covered with setae and bent downward when viewed laterally. Two species, both from El Salvador.

Benficana Bechyné & Bechyné, 1961

Monotypic: *Benficana longigena* Bech. & Bech. (Brazil: Pará)

Close to *Phyllotrupes*; interantennal ridge atrophied; ridges of genae evidently high and long; 3rd segment of front tarsus dilated; antennae very slender.

Bonfilsus Scherer 1967

Monotypic: *Aedmon subpubescens* Bechyné, 1956 (Guadeloupe)

Should have its place in the key for the Monoplatini between couplet 49 and 50 (*Pleurochroma* and *Allochroma*). Inner margin of posterior tibial apex definitely drawn out into a tooth. Antennal calli small; nearly no frontal lines visible; seen from the side frons not bent against vertex; interantennal carina deviding to the sides in front of antennae.

Carecuruna Bechyné & Bechyné, 1965

Monotypic: *Carecuruna egleri* Bech. & Bech. (Brazil)

Close to *Monoplatus* (*Sphaeronychus*)

Carminaltica Bechyné & Bechyné, 1961

Genotype: *Carminaltica uniformis* Bech. & Bech. (Brazil: Pará)

Close to *Monoplatus*: elytra reddish carmine transparent; 2 species, both Brazil: Pará.

Chalatenanganya Bechyné & Bechyné, 1963

Monotypic: *Chalatenanganya quadrifida* Bech. & Bech. (El Salvador)

Close to *Chlamophora*; anterior coxal cavities closed; antebasal impression on pronotum straight and bent laterally to base and fading; frontal margin of frons in male with 4 strong teeth; front tibiae scarcely flattened.

Chrysogramma Jacoby, 1885

Acrocyum Jacoby, 1885. – Bechyné. 1963, *Iheringia-Zoologia* 31: 38 (syn.)

Cuyabasa Bechyné, 1959

Genotype: *Wanderbiltiana minor* Bechyné (Brazil)

Close to *Phenrica* and *Disonycha*; last segment of hind-tarsus globose, swollen; head and pronotum as in *Phenrica* and *Disonycha*.

Diacacoscelis Bechyné, 1968

Genotype: *Cacoscelis lucens* Erichson

Close to *Cacoscelis*; tibiae dilated toward apex, without an anteapical emargination; labrum with numerous (10–18) punctures; elytra punctures irregular.

Egleraltica Bechyné & Bechyné, 1965

Genotype: *Egleraltica jariensis* Bech. (Brazil)

Close to *Systema*; pronotum with antebasal impression; punctures on elytra in regular rows.

Enneomacra Bechyné & Bechyné, 1961

Monotypic: *Lactica acutangula* Weise. (Brazil: Amazonas, Pará)

Close to *Monomacra*; intercoxal plate of prosternum large and prelongated over coxae; antebasal impression of pronotum only indicated at sides.

Huarinillasa Bechyné, 1959

Monotypic: *Huarinillasa kuscheli* Bechyné (Bolivia)

Close to *Sangaria*; without antebasal depression on pronotum; side margins of pronotum rounded.

Lacpatica Bechyné & Bechyné, 1977 (Chevrolat i. l.)

Genotype: *Monomacra macula* F. (Braz.: Guanabara, Min. Ger., S. P., Mato Grosso)

Close to *Paralactica*; Intercoxal plate of prosternum prolonged over coxae; antennae short; interantennal carina straight; black discal spots on elytra.

Lacpaticoides Bechyné & Bechyné, 1977

Genotype: *Monomacra nicodema* Bechyné (Brazil)

Close to *Paralactica*; in "Group 4"; fifth abdominal segment as large as preceding 3 segments together; antennae long; interantennal carina knob-shaped and arched when seen laterally; intercoxal plate of prosternum short. Color yellowish brown.

Maritubana Bechyné & Bechyné, 1961

Monotypic: *Maritubana neothonia* Bech. & Bech. (Brazil: Pará)

Close to *Neothona*; without interantennal ridge.

Maturacaita Bechyné & Bechyné, 1977

Monotypic: *Maturacaita terapoda* Bech. & Bech. (Braz.: Amazonas)

Close to *Paralactica*; intercoxal plate of prosternum prolonged over hindmargin of coxae and having on its hindedge a perpendiculate longitudinal lamella.

Miritius Bechyné & Bechyné, 1965

Monotypic: *Miritius egleri* Bech. & Bech. (Brazil.: Pará)

Close to *Hypolampsis*; antennits 3–6 long, 7–11 short.

Neoacanthobioides Bechyné & Bechyné, 1976

Genotype: *Longitarsus homotarsus* Bech. & Bech. (Venezuela)

Close to *Longitarsus*; elytral punctures in regular rows.

Neocacoscelis Bechyné, 1968

Monotypic: *Cacoscelis filiformis* F.

Close to *Cacoscelis*; pronotum with longitudinal folds near base; two median setife-

rous punctures on labrum much closer to one another than to the others; punctures on elytra confused, only the ones beside the scutellum and sidemargin in regular rows.

Neodiphaulaca Bechyné & Bechyné, 1975

Genotype: *Diphaltica itapiranga* Bech. (Brazil: S. Catarina, Paraná)

Close to *Diphaulaca*; punctures on elytra confused.

Oreinodera Bechyné & Bechyné, 1963

Monotypic: *Oreinodera aptera* Bech. & Bech. (San Salvador)

Apterous; antebasal impression on pronotum shallow and shortened laterally, not extending to base; scutellum small but distinct; punctures on elytra in rows; no humeral calli.

Phenrica subgenus **Orthophenrica** Bechyné & Bechyné, 1966

Genotype: *Asphaera quadrimaculata* Clark (Brazil: Pará, Amazonas)

Labrum with 6 setiferous punctures, the inner pair accompanied by 1–3 (rarely more) small punctures (but not in subgenus *Phenrica*); usually longer than 6 mm (Subgen. *Phenrica* usually smaller); intercoxal plate not as narrow as in subgenus *Phenrica*.

Palmaraltica Bechyné, 1959

Genotype: *Palmaraltica heteronycha* Bech. (Bolivia)

Close to *Asphaera*; upper surface pubescent with golden-yellow setae. 2 species: 1 Bolivia, 1 Venezuela.

Paracacoscellis Bechyné & Bechyné, 1961

Monotypic: *Cacoscelis tibialis* Jacoby

Close to *Cacoscelis*; punctures on elytra in numerous rows.

Paralacticoides Bechyné & Bechyné, 1977

Genotype: *Paralacticoides bicarinata* Bech. & Bech. (Brazil: Mato Grosso)

Close to *Paralactica*; intercoxal plate of prosternum rooflike or bulgelike; upper surface unicolorous.

Parchicola Bechyné & Bechyné, 1975

Genotype: *Monomacra yena* Bech. (Southern Brazil)

Close to *Monomacra*.

Podaltica Bechyné & Bechyné, 1963

Monotypic: *Podaltica harrietta* Bech. & Bech. (San Salvador)

Close to *Monomacra*; legs very long, especially hind femora which extend above elytra and are flat on its inner side and thus with sharp edges.

Protosilapha Bechyné, 1973

Genotype: *Haltica pallens* Blanchard (Chile)
Close to *Psilapha*. Two species, both from Chile.

Rosalactica Bechyné & Bechyné, 1977

Genotype: *Lactica maculicollis* Jacoby
Close to *Paralactica*; intercoxal plate of prosternum keel-like; upper surface, especially elytra pink-colored.

Strabala subgenus **Isostrabala** Bechyné & Bechyné, 1975

Genotype: *Lactica brasiliensis* Bryant (Brazil)
Elytra oval (subgen. *Strabala* rounded); sides with longitudinal interval convex (subgen. *Strabala* regularly convex).

Styrepitrix Bechyné & Bechyné, 1963

Monotypic: *Styrepitrix boqueronica* Bech. & Bech. (San Salvador)
Close to *Epitrix* and *Acallepitrix*, but with normal epipleura without an excavation near hind femora; pronotal base straight.

Temnocrepis Bechyné & Bechyné, 1963

Monotypic: *Temnocrepis trifiniensis* Bech. & Bech. (San Salvador)
Apterous; scutellum nearly unrecognizable; pronotal antebasal impression distinctly limited laterally; punctures on elytra in regular rows, humeral calli smooth, not distinct.

Trifinicola Bechyné & Bechyné, 1963

Monotypic: *Trifinicola freundi* Bech. & Bech. (San Salvador)
Similar to *Cyrsilus* but with an antebasal impression on pronotum; epipleura not impressed near hind femora; punctures on elytra in regular rows; front angles of pronotum broadly rounded; distinct antebasal impression on pronotum; upper surface of pronotum with fine and sparse punctures.

Triphaltica Bechyné, 1968

Genotype: *Diphaltica ruderalis* Bechyné (Brazil)
Close to *Wittmeraltica* but antennae filiform; setae of setiferous punctures short; pronotal margins without setiferous punctures (except the one of the front angles); *Wittmeraltica* has moniliform antennae; setae of setiferous punctures on elytra very long; margins of pronotum with setiferous punctures.

Vilhenaltica Bechyné & Bechyné, 1964

Genotype: *Vilhenaltica dibolosoma* Bech. & Bech. (Brazil)
Close to *Monomacra*; head like *Dinaltica*. Two species, both Brazil.

Yumaphthona Bechyné & Bechyné, 1976

Monotypic: *Brasilaphthona doria* Bech. (Brazil)

Close to *Sanariana*; antennae short, extending only to middle of elytra; longer in *Sanariana*.

Preface of the German Edition (1962)

This paper comprises a diagnostic key for the South and Central American genera of the Alticinae. At the same time, it serves to summarize the numerous genera of this subfamily. Unfortunately, its arrangement does not reflect natural relationships. The generic diagnoses are adapted only to the South and Central American species and are not necessarily appropriate for other areas. Therefore descriptions of some genera are listed several times. The genus *Ayalaia* Bechyné, 1960* which is unknown to me, is not considered, and the four species of *Hemiglyptus* G. Horn from Chile (in the Junk-Schenkling Catalog) are now in the genus *Psilpha* Clark.

I am deeply grateful to Dr. G. Frey who enabled me to visit London for several weeks to study the rich material of the British Museum (Nat. Hist.). Without this London sojourn, a complete overview of the neotropical genera of Alticinae would not have been possible. My sincerest thanks are also directed toward the ladies and gentleman of the British Museum (Nat. Hist.) and the Commonwealth Institute of Entomology. Above all, Dr. E. B. Britton, Miss. C. M. F. von Hayek, Mr. J. Balfour Browne, Mr. G. E. Bryant, Mr. Pope, and Mr. R. T. Thompson are to be very heartily thanked. I especially would like to thank my dear colleague, Dr. B. J. Selman, of the Commonwealth Institute. My thanks also go to the Director of the Zoological Collection of Bavaria, Dr. W. Forster, for so much assistance as well as for many fruitful discussions, and to Dr. Gisela Mauermayer and Dr. Ina Leuthold for procuring literature which was sometimes difficult to obtain. Mr. E. Diller of the Zoological Collection of Bavaria is to be heartily thanked for preparing the figure of *Omototus morosus* Clk. Finally, my thanks go to those who are not named who supported me by word and deed.

Division of groups

- 1 (12) Antennae eleven-segmented.
- 2 (5) Apical segment of posterior tarsi more or less globosely swollen (Fig. 1b).
- 3 (4) Elytra with nine continuously punctate striae (marginal and short-juxtascutellar striae not counted).

Group 1 p. 9. (Monoplatini)

* See "Genera not included in my key (1962)" p.

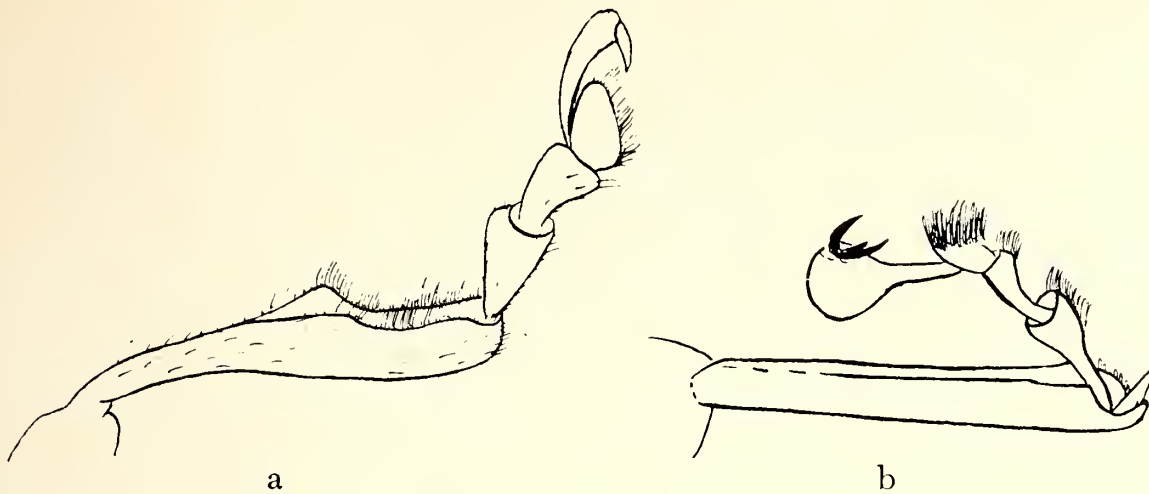


Fig. 1. Right hind leg of:
a) *Blepharida rhois* Rog.,
b) *Hypolampsis maculicollis* (Jac.)

- 4 (3) Elytra impunctate, or elytra confusingly punctate
Group 2 p. 28 (Oedionychini)
- 5 (2) Apical segment of posterior tarsi not globosely swollen.
- 6 (7) Posterior tibiae with a somewhat rounded preapical dorsal tooth, distally followed by a curved, crested, ciliated emargination (Fig. 1a):
Group 3 p. 32
- 7 (6) Posterior tibiae without ciliated emargination:
- 8 (9) Eyes emarginate and quite kidney-sharped: Fig. 21a):
Group 4 p. 34
- 9 (8) Eyes not or only moderately emarginated (Fig. 21b):
- 10 (11) Pronotum with a prebasal transverse impression:
Group 5 p. 36
- 11 (10) Pronotal base without such a transverse impression:
Group 6 p. 70
- 12 (1) Antennae 9 or 10-segmented:
Group 7 p. 84

Group 1 MONOPLATINI

The common characteristics of the Monoplatini are the closed anterior coxal cavities, punctate-striate elytra, and the globosely swollen apical segments on the posterior legs.

In his monograph of this group (Cat. Halt., I, 1860), Clark used the form of the palpi for subdivision into genera. Since the variations in the palpi are restricted to sexual di-

morphism and have little to do with orthogenesis, the systematics of the Monoplatini became so complicated that hardly anything more has been done with it since the appearance of Clark's catalog. The uselessness of the palpi for subdivision into systematic units was already shown by the fact that Clark repeatedly described species in different genera in one and the same paper.

The Coleopterorum Catalogus of Junk-Schenkling (1940) produced further confusion in the systematics of the Monoplatini. Csiki – without knowing the material – combined genera and this diagnoses was repeated in other references. However, since Clark chose the palpi for differentiation, the confusion within the Monoplatini became even greater.

It was very difficult for me to find common characters and to choose a genotype from the multitude of similar species which fit Clark's description.

However, within the Monoplatini, there are correlated morphological characters which permit the separation of genera. It is sometimes difficult to express such general characters in words, i. e., arched, shorter, or more slender; characters which occur through entire series of species, and which allow these groups to be recognized at first glance.

Unfortunately, in this process, some genera must be synonymized (e. g., the genus *Homammatus* Clark). In this case *Euphenges* had priarily in the same paper; also the head formation had already been introduced in the literature as a characterization for *Euphenges*. Unfortunately, I had no choice but to propose some new genera, an activity which I tried to keep at a minimum since the task of systematics is first to create order; that is, not only to recognize a systematic unit, but also to employ it as an organizing link. Thus, the genus *Rhinotmetus* Clark is the only genus which is once again divided because the metallic forms have a somewhat different head formation. Within the entire Monoplatini, besides the development of the metallic elytra, or of the entire insect, there are further common characters.

In this paper, I am adding at the end of the Monoplatini (p.) a list of Clark's species which were available to me and which I could study. There is still ambiguity in the genera *Hypolampsis* and *Physimerus*. I considered these 2 genera to be one and the same at first, but then separated them when the type material was no longer available to me. I am sure that one or the other is a synonym. Also, among others, one or two species are synonymous with *Euphenges rufotestaceus* (Clark). This species shows variation concerning the emargination on the upper side of the posterior tibia within the same population. This species is extremely variable and Clark described it as *Allochroma* from Rio de Janeiro (also as *Hypantherus turgidus*) and as *rufotestaceus* from Brazil without further details; one group in the British Museum came from Santa Catarina, and in the Frey Museum there are 44 examples from Marcapata in Peru. *Hypolampsis pumilio* Clark was the male of *Hypolampsis inaequalis*, but in the British Museum there also males of *inaequalis* as paratypes. Remarks on synonym which are cited in this list could be expanded.

Key to the Divisions

- 1 (4) Elytra with humeral calli and 9 rows of punctures (not counting the marginal and short juxtascutellar puncture rows).
- 2 (3) Pronotal and prebasal transverse furrow very distinct, laterally limited by distinct longitudinal grooves.
Division A (p. 11)
- 3 (2) Pronotum without a prebasal transverse furrow, or with a more or less distinct prebasal transverse furrow not limited laterally by longitudinal grooves.
Division B (p. 11)
- 4 (1) Elytra without humeral calli and only 7 rows of punctures (not counting marginal and short juxtascutellar rows of punctures).
Division C (p. 27)

Division A

Prebasal thoracic transverse furrow very distinct and laterally bounded by longitudinal grooves. Anterior angles of prothorax somewhat tooth-like externally; sides straight, a little angled at middle, thin edged. Antennal calli fused together; ocular sulci very close to inner edge of eyes.

Elytra somewhat broader than pronotum; humeral and basal calli distinct, shallow postbasal transverse impression.

Posterior femora reach almost to apices of elytra.

Monoplatus Clark 1860
= *Sphaeronychus* DeJean in litteris
(*Metriotes* Clark 1860)

Genotype: *Monoplatus nigripes* Clark

Ca. 35 species: Distribution South and Central America.

Division B

- 1 (2) Elytra with a deep, median impression on either side of suture: impression extends in an attenuated form to between humeral and basal calli (Fig. 2); metasternum with a coneshaped structure; see also fig 18:

Meraaltica Scherer 1962

Genotype: *Euphenges lemoeides* Clark 1860 (amazon)

- 2 (1) Elytra without a median impression on either side of suture; metasternum without any cone.
- 3 (8) Head and prothorax very rugose, latter with several protuberances, at least one protuberance on each side of the midlinenear anterior edge.

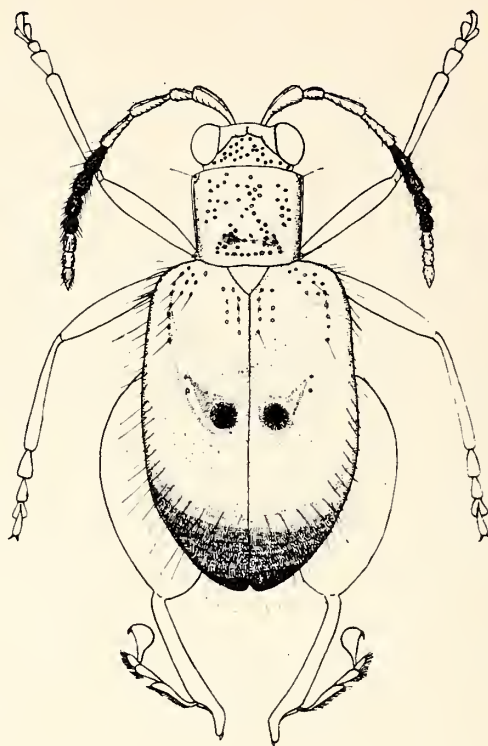


Fig. 2. Habitus of *Meraaltica lemoeides* (Clark)
(Magnification 10×)

- 4 (5) Apical antennal segment very short, usually shorter than wide:

Brachyscelis Germar 1834

(= *Imatium* Clark 1860 syn.)

(= *Leptotrichus* Clark 1860 syn.)

Genotype: *Brachyscelis vellerea* Germar (Brazil: Rio de Janeiro; Paraguay).

- 5 (4) Apical antennal segments not at all shortened.

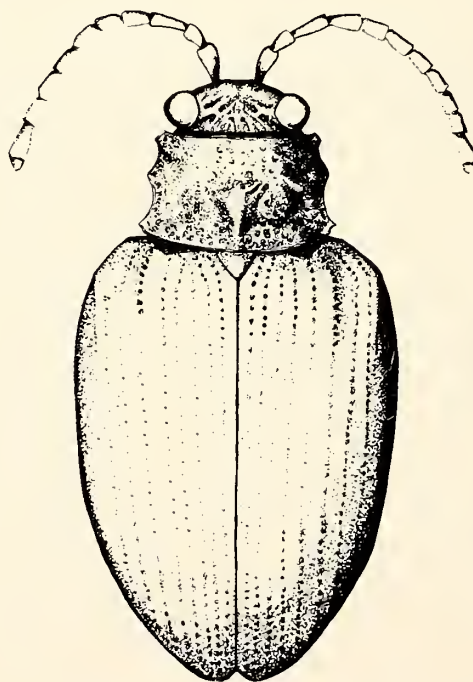


Fig. 3. Habitus of *Omototus morosus* Clark

- 6 (7) Prothoracic sides somewhat slanted toward lateral margins; elytra without postbasal transverse depression (Fig. 3):

Omototus Clark 1860

Lectogenotype: *Omototus morosus* Clark 1860 (Cayenne)

- 7 (6) Prothoracic sides straight; elytra with distinct postbasal transverse impression:

Homotyphus Clark 1860

(= *Sesquityphus* Bechyné 1959 syn.)

Genotype: *Homotyphus holosericeus* Clark 1860 (Brazil: Rio de Janeiro).

Homotyphus lacunosus could well be regarded as the most typical representative of this genus because Clark based his generic diagnosis entirely on this species. Nonetheless, Bechyné (Ent. Arb. Mus. Frey 9 [2], 1958, p. 706) proposed his genus *Sesquityphus* on this species, and *H. holosericeus* Clark as the type for the genus *Homotyphus*. *Homotyphus lacunosus* Clark has beneath the humeral calli the elytral pleura completely atrophied. The epipleura are here separated from the elytral disk only by a row of dots. The elytral pleural ridge first begins shortly behind this area. However, there are all sorts of intermediate forms. Among others, *Homotyphus antillarum* Bechyné 1956, *H. roseobrunneus* Bechyné 1959 and *H. aureonotatus* Bechyné 1959 should also remain in the genus *Sesquityphus*.

- 8 (3) Prothorax without protuberances on either side of midline near anterior edge; can have a prebasal transverse depression:

- 9 (10) Antennae similar to Cerambycidae, always significantly longer than body; frons flexed from dorsum of head at more than a right angle and concave; antennal calli expanded like a bladder. Body narrow and longitudinal.

Loxoprosopus Guer. 1829–1844

Genotype: *Loxoprosopus ceramboides* Guer. (Brazil: Minas Gerais).

10 species: 5 Brazil, 1 Bolivia, 3 Equador, 1 British Guiana.

- 10 (9) Antennae not cerambycid-like or longer than body; frons not so far flexed from the dorsum of the head; antennal calli not so much expanded:

- 11 (56) Pronotal margins straight, slightly curved or arched, never angled laterally:

- 12 (13) Pronotum very constricted at anterior edge; connecting to head very close to eyes closely behind eyes (Fig. 4):

Behind antennal calli there usually is a callosity whose posterior end continues in an arch from a posterior edge of one eye to the other; behind this callosity is a medial longitudinal ridge over vertex:

Rhinotmetus Clark 1860

Genotype: *Oedionychus leptcephalus* Perty 1832 (Brazil: Rio de Janeiro, Minas Gerais).

- 13 (12) Prothorax not as closely attached to head behind eyes; anterior angles of prothorax always pointing outward.

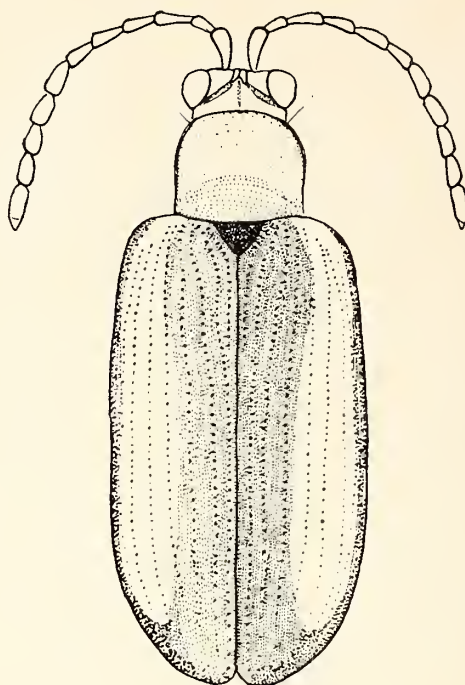


Fig. 4. Habitus of *Rhinotmetus marginatus* Clark
(Magnification 9×)

- 14 (15) Thorax black with broad yellow margins; elytra black, usually with yellow markings; head also usually yellow black:
Flat longitudinal forms, always shagreened, dull, and setose; elytra always more than twice as long as wide; without any definite postbasal transverse depression on elytra. **Cerichrestus** Clark 1860
Genotype: *Cerichrestus balyi* Clark (Brazil: Rio de Janeiro; Santa Catharina).
Uniform morphological characters for this genus are hard to find, so that only the yellow coloration on the thoracic sides must suffice. This yellow coloration is coupled with a uniform habitus however: flat, thin, shagreened. The head shape, usually a very usable character in the Alticinae, is very variable here.
- 15 (14) Always differently colored than above:
- 16 (19) Antennal calli very elongated posteriorly in the direction of posterior margin of eyes; vertex with a median longitudinal keel:
Convex, oval; thoracic sides usually straight; a postbasal transverse impression present on elytra:
- 17 (18) Genae and anterior frons long, frons with a long longitudinal ridge:
Euphenges Clark 1860
= *Homammatus* Clark 1860 syn.
Genotype: *Euphenges sericeus* Clark (Brazil)
- 18 (17) Genae and frons short, frons triangularly expanded, without a longitudinal ridge; thickly setose: **Apalotrius** Clark 1860
Monotypic: *Apalotrius pubescens* Clark (Brazil: Amazon)

- 19 (16) Antennal calli not elongated posteriorly, vertex without a median longitudinal keel:
- 20 (39) Inner margin of apex of posterior bibiae not extended in a tooth shape (Fig. 5–9):

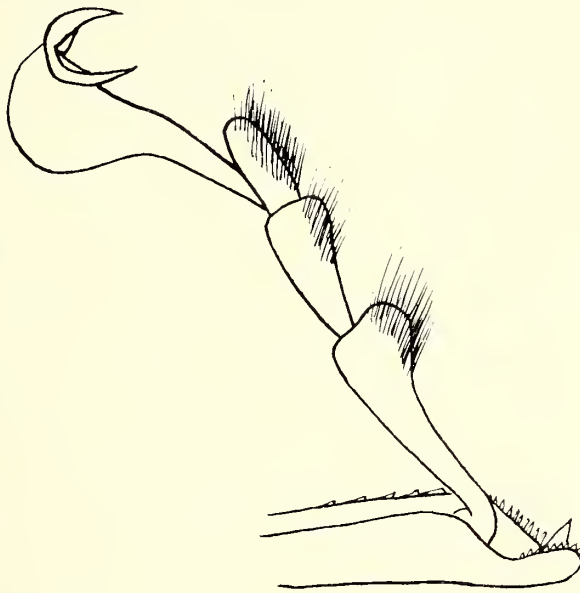


Fig. 5. End of the right posterior tibia of *Atyphus furcipes* Clark
(Magnification 40×)

- 21 (30) Metallic, or with a transverse setose band extending over elytra at midpoint; if with band, then elytral apices yellow and setose. Relatively large insects, from 4.5–9 mm in length.
- 22 (27) The entire insect is metallic or with a transverse setose band on the elytra.
- 23 (26) Elytra without transverse setose band.
- 24 (25) Elytral punctation very reduced, especially evident near elytral suture, in distinct postbasal transverse impression, and in impression between humeral and basal calli; very convex forms (ca. 4.5 mm long).

Palopoda Erichson 1847

Genotype: *Palopoda tersa* Erichson (Peru)

2 species; 1 Peru, 1 Bolivia.

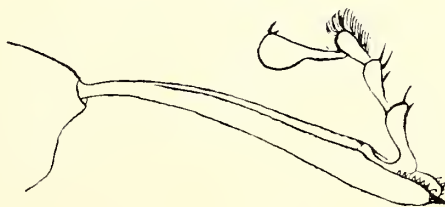


Fig. 6. Hind leg of *Palopoda parcepunctata* Bech.

- 25 (24) Elytral punctation uniformly distinct, interstrial spaces convex: a distinct postbasal transverse impression; very convex shaped (length 6–8 mm):

Atyphus Clark 1860

Monotypic: *Atyphus furcipes* Clark (Columbia).

This is exactly the same as the 4 *Atyphus* spp. described by Clark, but which seemingly did not fit properly there. *Atyphus carbonarius*, *A. flaviventris*, and *A. vittatus* belong in *Physimerus*; indeed, the latter is even a synonym of *Physimerus vittatus*. Bechyné (Ent. Arb. Mus. Frey, 9 [2], 1958, p. 705) previously established this genus with the species *furcipes* Clark.

- 26 (23) Elytra with transverse setose band.

Elytra metallic or non-metallic, with a distinct postbasal transverse impression and a median yellow setose band; elytral apices also setose, and many times edges and elytral border are setose so that median band and setose apices are continues with margins to elytral suture.

Frons with a distinct transverse impression on anterior half, so that posterior half is quite sloped toward frons. Posterior swollen margin of frons simulates a transverse carina, Genae short. A very short longitudinal carina between antennae. Antennal calli large, sending off a pointed process anteriorly between antennal sockets and frontal longitudinal ridges; posteriorly, they are defined by a groove which extends obliquely to posterior rim of eyes.

Prothorax thickly setose, prothorax is at least half as wide as it is long; sides ahead of center somewhat tapered to anterior angles; base has a slight transverse impression.

Inner margin of apex of posterior bibiae nearly rectangular and with fine teeth (Fig. 7 and 9b):

Bellacincta Scherer 1962

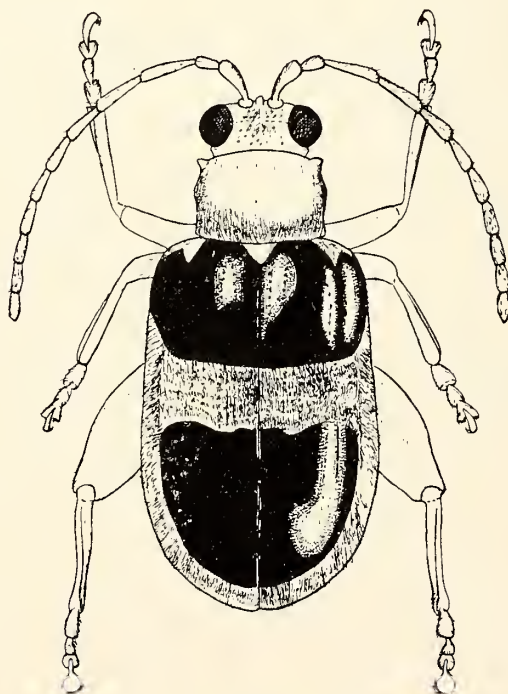


Fig. 7. Habitus of *Bellacincta rufolimbata* (Jac.)
(Magnification 7×)

Genotype: *Omototus rufolimbatus* Jacoby (Peru, Brazil: Acré). A subspecies of *rufolimbatus* (*chaparensis* Bech.) is known from Bolivia: another species (*humeronotatus* Clark) is known from Brazil (Island of St. Paul).

- 27 (22) Only elytra metallic; without a distinct postbasal transverse depression on elytra.
- 28 (29) Pronotum without prebasal transverse impression; anterior angles fixed; posterior tibia somewhat balanced before apex and also before tarsal joint, with both marginal edges unconnected (Fig. 8a):

Eupeges Clark 1860

Monotypic: *Eupeges praeclara* Clark (Brazil: Amazon).

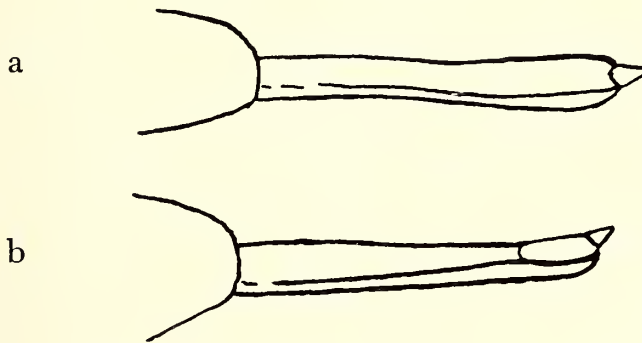


Fig. 8. Posterior tibia of
a) *Eupeges praeclara* Clk.
b) *Hydmosyne inclyta* Clk.

- 29 (28) Pronotum with prebasal transverse impression; anterior angles simply shaped like a right angle, only somewhat compressed; posterior tibiae not balanced near apex before tarsal joint, but both edges somewhat connected (Fig. 8b):

Hydmosyne Clark 1860

Genotype: *Hydmosyne inclyta* Clark

- 30 (21) Forms without metallic coloring, and without a setose transverse band on elytra:

- 31 (32) Forms about 5 mm in size; elytra always pitch brown to black with yellow or gray lateral borders; elytral suture can be colored similarly:

Pronotal prebasal impression occurs only on each side of center, but very distinct, pronounced and extends forward usually setae in whirls in these basal impressions accentuate impression. Thorax strongly convex.

No definite postbasal transverse impression on elytra; Elytra significantly wider than prothorax.

Frons triangular and forming posteriorly a short longitudinal ridge between antennae; antennal calli give rise to pointed processes anteriorly between these frontal longitudinal ridge and antennal attachments – usually a lancetshaped split lies between these processes.

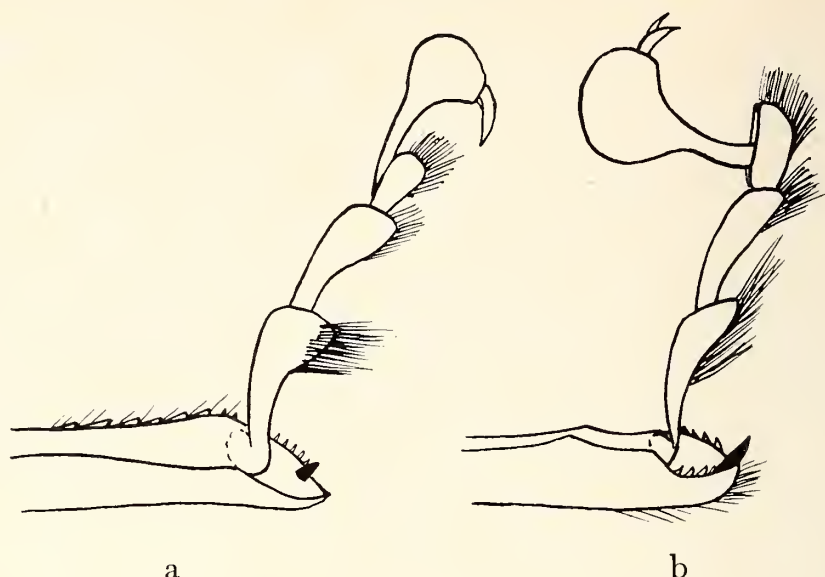


Fig. 9. Apices of right posterior tibia of
 a) *Chaparena chacoensis* (Bowd.)
 b) *Bellacincta rufolimbata* (Jac.)
 (Magnification 40×)

Head black or black with yellow, elytra dark pitch brown to black with yellow or gray margins, elytral suture can also be similarly colored.

Chaparena Bechyné 1959

Genotype: *Metriotes chacoensis* Bowd. (Bolivia).

5 species which are distributed in Peru and Bolivia.

This genus is very similar to *Cerichrestus* Clark. In the latter, the thorax has a yellow lateral margin, and is less convex, and the prebasal thoracic impression extends over the entire base, but is somewhat deeper laterally.

- 32 (31) Elytra without yellow lateral margins; prebasal transverse impression occupies entire base – This transverse impression can also be very weakly formed or entirely absent: Usually small forms, ca. 3 mm; elytra significantly wider than prothorax; postbasal transverse impression in elytra usually very distinct:

- 33 (34) Antennal segments (5–8) very shortened and thickened, last 3 joints again slender, shortened joints only about as long as broad (Fig. 10):

Pronotal prebasal transverse impression distinct, postbasal transverse impression on elytra also pronounced.

Surface smooth and shiny, but contains relatively long setae not too close together. Chestnut brown, ca. 3 mm.

Calipeges Clark 1860

Monotypic: *Calipeges crispus* Clark (Brazil: Pará).

- 34 (33) All antennal segments up to segment 4 shortened or small:

- 35 (36) Antennae short, extending only to elytral base, all antennal segments up to segment 4 very short; lateral margin of prothorax very wide, extending ven-

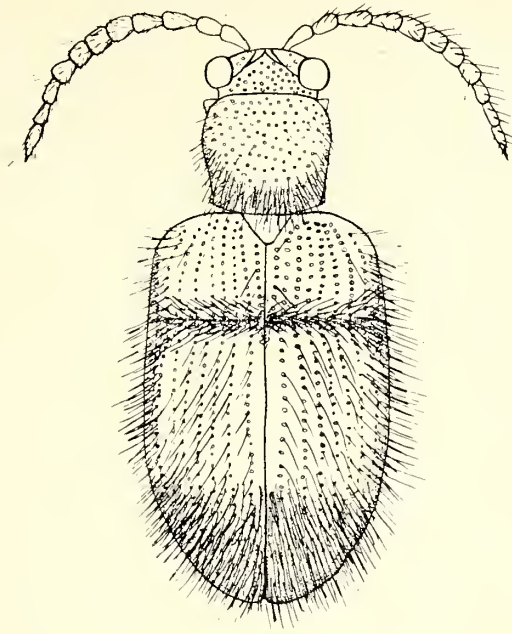


Fig. 10. Habitus of *Calipeges crispus* Clark
(Magnification 15×)

trally and not visible when viewed from above, prothorax thus appears cylindrical; basal transverse impression definite; with a smooth, non-sculptured longitudinal strip along middle near anterior margin.

Elytra very wide, significantly wider than prothorax and relatively short, length-width ratio approximately 3:2; postbasal transverse depression distinctly pronounced.

Genae and frons very long, a longitudinal frontal ridge only suspected.

Distigmoptera Blake 1943*

Genotype: *Haltica pilosa* Illiger (USA from Hudson Bay south to Mexico). 14 species with their primary distribution in the USA, of which one species is from Canada, one from Mexico, two from Costa Rica, and I have a new species from Brazil (Mato Grosso).

- 36 (35) Antennae longer, extending beyond base of elytra, all segments significantly longer than wide; flat, thickly setose.

Prothorax not so strongly convex as in preceding genus, lateral margins easily visible when viewed from above; basal transverse impression distinct, uniformly punctate and setose.

Elytra considerably wider than prothorax, relatively long and thin, length to width ratio about 2:1; a postbasal transverse impression usually very distinct.

- 37 (38) Basal calli in front of postbasal transverse impression on elytra distinct, elytra behind postbasal transverse impression once again somewhat convex, a true

* Balsbaugh, E. U. 1969: *Pseudolampsis* (Coleoptera, Chrysomelidae, Alticinae): Distribution and Synonymy. – The Coleopterists Bulletin 23: 16–18.

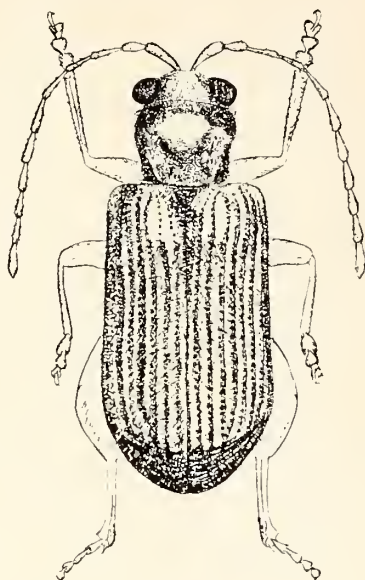


Fig. 11. Habitus of *Hypolampus balyi* Clark
(Magnification 13×)

apical flattening begins on last third, while lateral margins first begin to bend in toward apex, so that elytra thus appear cylinder-shaped (Fig. 11):

Hypolampus Clark 1860
= *Hylodromus* Clark 1860 syn.
= *Coelocephalus* Clark 1860 syn.
= *Poebates* Clark 1860 syn.

Genotype: *Hypolampus balyi* Clark (Brazil: Rio de Janeiro).

The genus *Hylodromus* was proposed because of the widened antennal segments 3, 4, 5, and 6, but it is a typical representative of *Hypolampus*. *Coelocephalus* is differentiated only by the longer antennae, and I cannot recognize a separate genus in *Poebates*.

- 38 (37) Basal calli in front of postbasal transverse impression on elytra well developed; elytra moderately tapered from transverse impression to apices; elytra also narrowed laterally before middle, appear wedge-shaped.

Aedmon Clark 1860
= *Hadropoda* Suffr. syn.

Genotype: *Aedmon sericellum* Clark (Puerto Rico)

Clark described a species from Puerto Rico, Bechyné described 3 from Guadeloupe (*Hadropoda heikertingeri*, *H. orsodacnina*, *H. adumbrata*), Suffrian one from Cuba (*H. ferrugineus*); the *Hadropoda* species of Blake from the Caribbean area are unknown to me.

- 39 (20) Inner margin of posterior tibial apex definitely drawn out into a tooth (Figs. 12–14):
- 40 (41) Round to oval form of coccinellid-like habitus, compact form, convex: Prothorax basally twice as wide as long, sides straight and converging anteriorly; base as wide as elytra, no prebasal transverse impression.

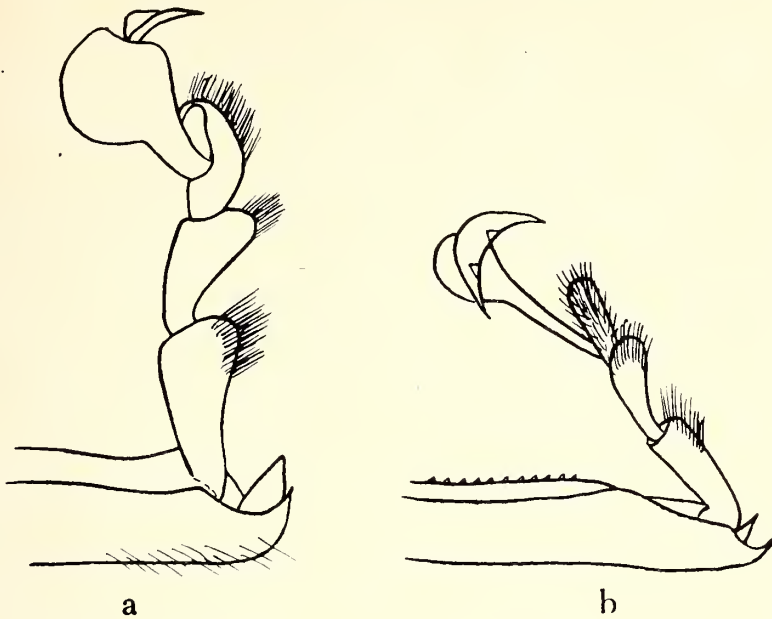


Fig. 12. Apex of right posterior tibia of
 a) *Sparnus chaparensis* Bech.
 b) *Anerapa aeneipennis* (Bowd.)
 (Magnification 40×)

Elytra very convex, without any postbasal transverse impression, without basal calli.

Antennal calli nearly horizontally delimited posteriorly, ocular sulci extend laterally from thence to posterior margin of eye; frons triangular and only moderately flexed toward antennal calli.

Antennae extend to elytra base, third antennal segment longest, subsequent segments shortened and thicker.

Sparnus Clark 1860

= *Cyrton* Clark 1860 syn.

Genotype: *Sparnus globosus* Clark (Brazil: Amazon).

41 (40) Not round to oval, not of coccinellid-like habitus; prothorax at base not twice as wide as long:

42 (45) Nearly glabrous forms; elytra metallic; head and prothorax yellow or reddish-brown:

Pronotal prebasal transverse impression usually extending laterally somewhat anteriorly, antennal calli large, separated from each other by a deep slit, a process extends anteriorly from each frontal elevation between antennal insertion and longitudinal interantennal carina, which limitation to clypeal carina is sometimes very indistinct, frons triangular, lateral margin swollen so that it appears as if from the short carina a short longitudinal carina were drawn off on either side and extended anteriorly; space between "anteriorly extended carina" and anterior margin of eyes just as wide, that there is space for first antennal segment, genae therefore short.

- 43 (44) Without postbasal transverse impression on elytra:

Elytra not especially convex, appearing flat from sides. Frons triangular, plain, distinctly angled against antennal calli when viewed from side.

Thorax transversely oriented, margin narrow, somewhat transversely compressed in front of base. Length ca. 6 mm.

Gethosynus Clark 1860

Monotypic: *Gethosynus sanguinicollis* Clark (Brazil: Espirito Santo).

- 44 (43) With distinct postbasal transverse impression on elytra:

Elytra not strikingly arched, flat when observed from side.

Frons short and triangular, posterior edge simulates an anteriorly extending transverse carina, frons not excessively bent against antennal calli.

Thorax transversely orientated, very thinly margined, distinctly compressed transversely in front of base, this compression tends forward laterally. Length ca. 5–6 mm.

Anerapa Scherer 1962

Genotype: *Chaparena yungarum* Bechyné

Two species: *Chaparena yungarum* Bechyné and *Metriotes aeneipennis* Bowd. (Both Bolivia).

- 45 (42) Non-metallic colored forms, in part smooth and nearly glabrous or also thickly setose and dull:

- 46 (51) Smooth, nearly glabrous, shining forms:

- 47 (50) Without definite postbasal transverse impression on elytra:

- 48 (49) Elytra somewhat narrowed posteriorly, uniformly tapering to margins, elytra evenly convex from base to center when viewed from side.

Frons lies in nearly same plane as postantennal calli, frons with longitudinal and transverse carinae; antennal calli delimited horizontally posteriorly, to some extent they grade into frons without anterior interruption.

Thorax transverse, not constricted at base, sides somewhat flat and nearly parallel.

Bright yellow with some red to pitch brown ornamentation. Length 6–9 mm.

Exartematopus Clark 1860

Genotype: *Exartematopus scutellaris* Clark (Brazil).

In reference to the above mentioned species, a subspecies is known from Santa Catharina, and another species is known from Espirito Santo.

- 49 (48) Elytra parallel, markedly sloping on edges, elytra flat when viewed from side.

Prothorax more than 1.5 times as wide as long, sides somewhat parallel at first, and in front of the center sides inclined toward anterior angles; base not transversely impressed; upper surface shiny and very finely punctate. Posterior margins of antennal calli extend obliquely to posterior margin of eye, barely defined on sides and anteriorly give off a long process; interantennal ridge very long and thin and extending to anterior half via a transverse carina.

Antennal segments 3, 4, and 5 long, following segments short. An emargination before apex on outside of posterior tibiae not always demonstrable with certainty. 3.2–4.5 mm in size with relatively flat, oval, compact form.

Pleurochroma Clark 1860

Genotype: *Pleurochroma balteatum* Clark 1860 (Brazil: Amazon).

- 50 (47) With definite postbasal transverse impression on elytra: Thorax usually strikingly transverse, often finely, rarely coarsely punctate, but never rugose. External antennal segments usually shortened.

Allochroma Clark 1860

Genotype: *Allochroma coccineus* Clark (Brazil: Rio de Janeiro).

I would also include the monotypic genus *Cleophes* Jacoby, 1886 from Panama, close to or even within the genus *Allochroma* which I know only from the description.

- 51 (46) Thickly setose, non-lustrous forms:

- 52 (53) Pronotum nearly as broad as elytra base, sides converging anteriorly, pronotal prebasal transverse impression scarcely noticeable:

Frons triangularly arched without longitudinal and transverse carinae, only moderately bent toward antennal calli; antennal calli defined posteriorly by a furrow, running obliquely to posterior margin of eyes.

Elytra without postbasal transverse impression, moderately convex, narrowed posteriorly. Brownish, often olive green forms.

Phylacticus Clark 1860

Genotype: *Phylacticus ustulatus* Clark (Cayenne)

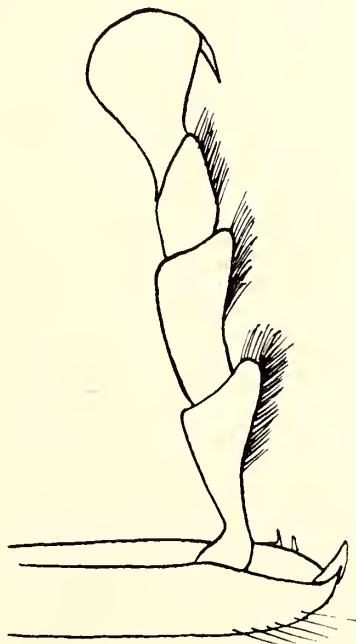


Fig. 13. Apex of right hind tibia of *Phylacticus ustulatus* Clark (Magnification 40×)

There is an example of *ustulatus* from Costa Rica (Schild-Burgdorf Collection, Costa Rica, Tucurrique) in the Frey Museum which completely corresponds with the type.

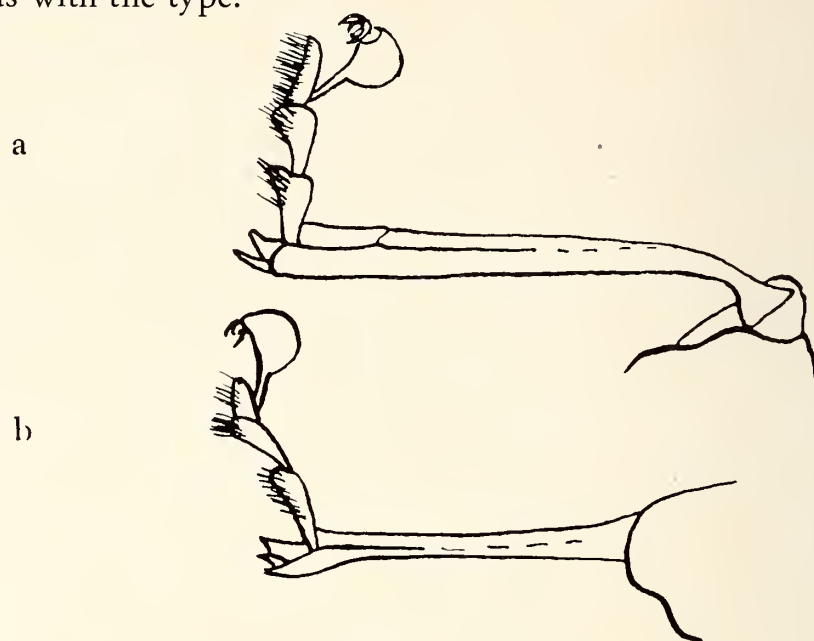


Fig. 14. Posterior leg of
 a) *Physimerus flaviventris* (Clk.)
 b) *Physimerus juvencus* (Clk.)
 (Magnification 30×)

53 (52) Pronotum narrower than elytral base, elytra parallel, not narrowed posteriorly.

54 (55) All antennal segments markedly longer than wide.

Elytral wider than prothorax and without definite postbasal transverse impression.

Sides of prothorax nearly straight and parallel in front of center, at first somewhat slanted near anterior angles. Surface relatively slightly arched, prebasal transverse furrow laterally displaced forward.

Antennal calli transverse, giving rise anteriorly between interantennal ridge and base of the antenna, a pointed process, often transformed into interantennal ridge; laterally, delimitation of antennal calli usually signified only by a lack of punctures; a more or less distinct side line on frons runs above antennal calli to posterior margin of eye, posterior boundary nearly horizontal. Interantennal ridge short, short triangular frons plain and somewhat obtusely angled against postantennal swellings, posterior margin somewhat swollen resembling a transverse carina running forward on each side. Greater than 4 mm in size; head yellow to reddish brown, partially darkened like pronotum; elytra yellow, pitch brown or black, sometimes with yellow flecks or stripes.

Physimerus Clark 1860

= *Thrasygoeus* Clark 1860 syn.

Genotype: *Physimerus vittatus* Clark (Brazil: Rio de Janeiro)

- 55 (54) Antennal segments 6–11, very much shortened, shorter or no longer than wide. Pronotum approximately 1.5 times as wide as long. Sides straight and nearly parallel; base distinctly transversely impressed; closely punctate and with short setae.

Furrow sutural groove which delimits antennal calli posteriorly runs obliquely to posterior margin of eye, antennal calli not defined laterally and give rise to a long process anteriorly; frons very short, swollen on sides (like transverse carinae), with a marked transverse impression on anterior edge.

Antennal segments 3, 4, and 5 long, with following segments very short, as long as wide (or shorter), broad external edge of posterior leg behind center with a blunt tooth from which arises a jagged edge.

About 6 mm in size, relatively highly arched.

Hypantherus Clark 1860

Genotype: *Hypantherus concolor* Clark (Brazil: Amazon)

- 56 (11) Prothorax with a bulge on lateral margin somewhat ahead of center (Fig. 16):

- 57 (58) Relatively broadly convex forms; antennae very short, primarily with outer segments very shortened:

Head and prothorax distinctly and densely punctate; frons triangular with a very short longitudinal carina between antennae; frons somewhat angled from antennal calli; antennal calli difficult to recognize in punctures; frontal sutures run obliquely to posterior margin of eye.

Prothorax thin, about as wide as eyes; transverse impressions in front of base.

Elytra extremely convex with a postbasal transverse impression. Proximal end of hind leg prolonged into a tooth shape.

Panchrestus Clark 1860

Genotype: *Panchrestus pulcher* Clark (Brazil: Amazon)

- 58 (57) Relatively thin flat forms; outer antennal segments not markedly shortened.

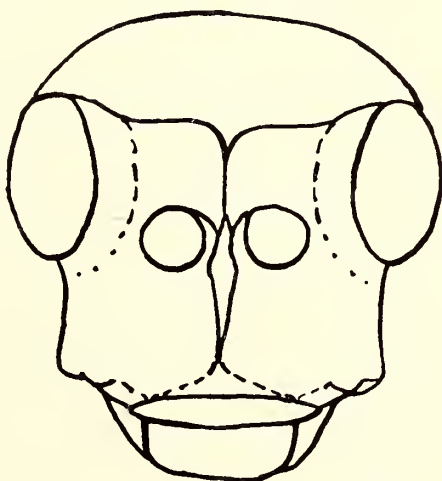


Fig. 15. Anterior view of the head of *Tetragnotes subanchoralis* Clark (Magnification 30×)

- 59 (22) Genae long, significantly longer than first antennal segment broad, so that head has an angular appearance; a longitudinal carina runs over smooth frons; frons and antennal calli approximately in one plane (Fig. 15).

Pronotum very constricted on anterior margin, head inserted into prothorax to eyes, with eyes clearly projecting beyond it at anterior margin; on either side of midline, about midway longitudinally, a more or less pronounced protuberance; pronotum not transverse, base with transverse impression.

- 60 (61) Metasternum raised on either side in a prominent tubercle above coxae; underside of femora forming a sharp edge; tibiae strongly bowed:

Roicus Clark 1860

(= *Sterneugonia* Bechyné 1959 syn.)

Genotype: *Roicus sexmaculatus* Clark (Amazon)

2 species (1 Amazon, 1 Bolivia)

- 61 (60) Metasternum not raised on either side in a prominent tubercle above coxae; underside of femora more rounded; tibiae not bowed (Fig. 16).

Tetragonotes Clark 1860

Genotype: *Tetragonotes elegans* Clark (Brazil: Rio de Janeiro).

- 62 (59) Genae short, but long enough for easy insertion of first antennal segment. Pronotum distinctly transverse, not so closely attached to head at anterior edge, always wider than eyes; prebasal transverse impression; no protuberances about halfway along the prothorax on either side of the midline. Frons short with longitudinal and transverse carinae, in nearly same plane as antennal calli.

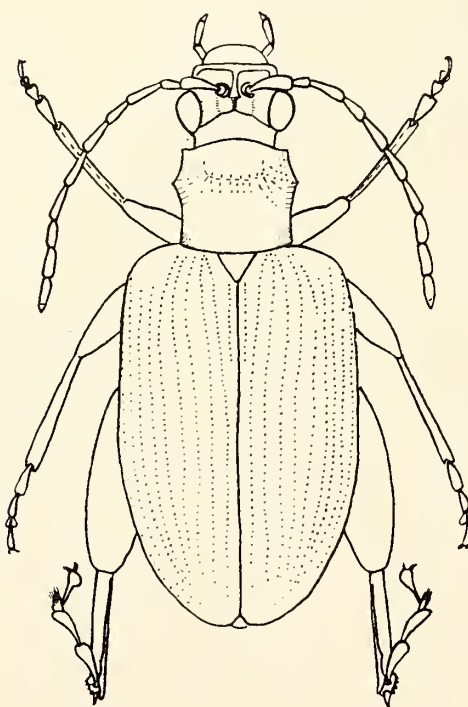


Fig. 16. Habitus of *Tetragonotes elegans* Clark
(Magnification 9×)

- 63 (64) No postbasal depression on elytra: innerproximal end of posterior tibia expanded to a definite tooth; antennae do not extend to apices of elytra:

Octogonotes Drapiez 1819

Genotype: *Octogonotes banoni* Drapiez.

- 64 (63) Postbasal transverse impression on elytra distinct; proximal end of posterior tibia rounded; antennae reach nearly to elytra apices:

Zeteticus Harold 1875

(*Peribleptus* Clark 1860)

Genotype: *Peribleptus laevigatus* Clark (Brazil, Rio de Janeiro)

Division C

Antennal calli absent; only a hint of lateral lines on the frons; frons and vertex sparsely punctate; frons flat and relatively wide between antennae, separated from frons above by a V-shaped impression; eyes not very strongly convex but large.

Pronotum large, evenly convex; ratio of length to width, 3:5; sides rounded and converging anteriorly.

Elytra without humeral and basal calli; seven continuous striae rows of punctures, marginal and juxtascutellar punctures not counted; seventh row (external) reduced to a few punctures and disappears at center; intervals convex; elytra thinner in area of posterior femora when examined from side.

Posterior femora very thick; tarsi not attached to posterior tibiae exactly at apex; upper edge of posterior tibiae with a sharp angle on either side just before the apex, posterior end of tibia transversely truncated:

Ulrica Scherer 1962

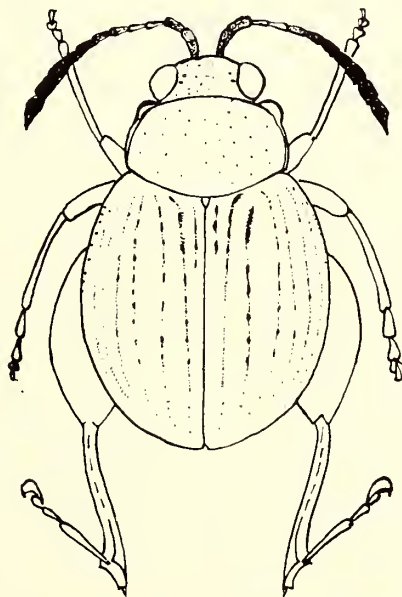


Fig. 17. Habitus of *Ulrica minutus* (Jacoby)
(Magnification 20×)

Monotypic: *Sparnus minutus* Jacoby (Bolivia) (Fig. 17)

Sparnus minutus has nothing in common with the genus *Sparnus*. *Sparnus* has antennal calli as well as humeral calli, and the rest of the body plan is entirely different.

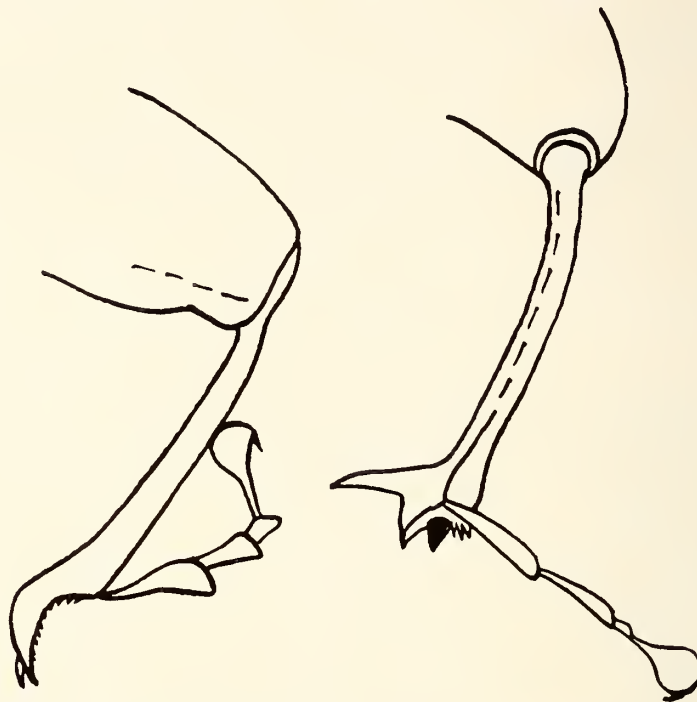


Fig. 18. Posterior tibia of
a) *Meraaltica lemoeides* (Clk.)
b) *Meraaltica punctata* Scherer

Group 2

The apical segment of the posterior tarsus is more or less globose in shape; the elytra are either impunctatum or confusingly punctate: The formerly large genus *Oedionychus* was separated into numerous new genera by Bechyné in several monographs. The establishment of these genera presented great difficulties which also is apparent in this key. The shape of epipleura, for example, is a very uncertain character. Punctations of the head, as in the case of *Kuschelina*, also occurs in other genera. The placement of many species within these genera is also uncertain. In spite of this, an attempt will be made to summarize the genera which are difficult to separate in a table.

- 1 (12) Posterior femur not overly thickened, transverse diameter about $\frac{1}{2}$ – $\frac{2}{3}$ length of tibia, apical segment of posterior tarsus swollen but not globose; basal tarsal segment of hind leg approximately $\frac{1}{3}$ as long as tibia.
- 2 (7) Epipleura easily visible when viewed from side:
- 3 (4) Frons and transverse carina on frons white, rest of head dark pitch-brown to black.

Anterior angles of pronotum (Fig. 19) extend forward with margins close to eyes; no prebasal transverse furrow. Rectangular antennal calli separated by a fine suture; frontal longitudinal carina between antennae very narrow, transverse carinae wider; frons separated from vertex by a fine horizontal row of punctation. Elytra without basal calli; brown with yellow markings, or yellow with dark markings, also metallic with and without markings, also brown and unmarked. Length over 5 mm.

Homophoeta Erichson 1847

Genotype: *Chrysomela albicollis* Fabricius 1787

About 50 species, distributed over all of South and Central America.

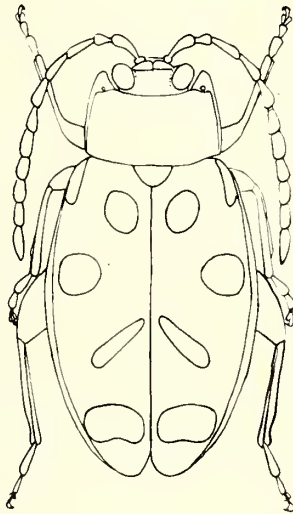


Fig. 19. Habitus of *Homophoeta albicollis* (F.)
(Magnification 6×)

- 4 (3) Frons and clypeal transverse carina colored like rest of head; anterior angles of pronotum also projecting forward near eyes; margins wide apart; prebasal transverse impression absent.

Elytra a little wider than pronotum at base; basal calli weakly developed.

Antennal calli triangular, more or less well separated, their anterior points lying between antennae and frontal longitudinal carina (Fig. 20).

- 5 (6) Anterior margin of metasternum prominently thickened.

Aspicela Clark 1865

Aspicela Dejean in litteris

Clark gave no individual genus diagnosis, so that one might almost want to accept Harold (Coleopt., Vol. XV, 1876, p. 95) as author.

Genotype: *Altica unipunctata* Latr.

About 12 species, whose distribution is limited to Columbia and Equador.

- 6 (5) Anterior margin of metasternum not raised or swollen:

Asphaera Chevrolat 1843

(*Litosonycha* Clark 1865)

Omophoita Chevrolat, 1837 sensu Bechyně

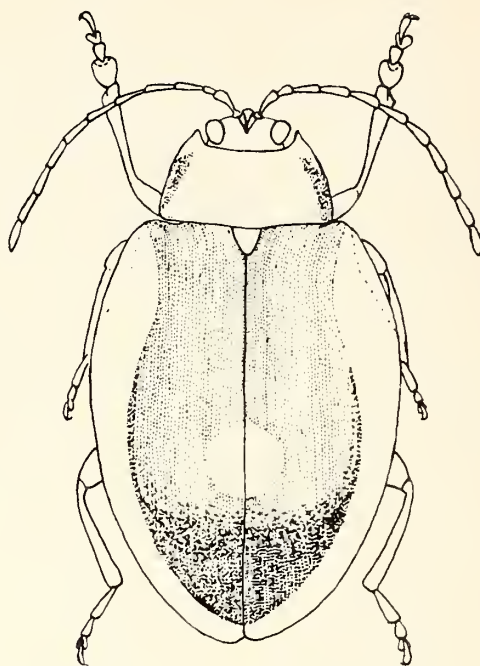


Fig. 20. Habitus of *Aspicela unipunctata* Latr.
(Magnification 4×)

Genotype: *Asphaera auripennis* Harold 1876 (Brazil: Rio de Janeiro).

More than 130 species distributed over all of South and Central America. Two species, *lustrans* Crotch and *abdominalis* Chev., extend into the USA.

- 7 (2) Epipleura shape obliquely inward and upward not visible from side.
- 8 (11) Frons not highly convex between Antennae:
- 9 (10) Lateral margins of pronotum strongly rounded; elytral pleura narrow; elytra rounded, relatively broad and short (6:7):

Pleurasphaera Bechyné 1958

Monotypic: *Asphaera decipiens* Clark 1865

Brazil: Rio de Janeiro, Minas Gerais; Argentina; Misiones: Loreto.

- 10 (9) Lateral margins of pronotum not strongly rounded; elytra parallel, relatively narrow and long (3:5); epipleura fully developed only in anterior third:

Longasphaera Bechyné 1955

Monotypic: *Longasphaera hispicornis* Bechyné

Brazil: Minas Gerais.

- 11 (8) Frons highly convex between antennae appearing as a large tubercle; lateral margins of the pronotum weakly rounded; elytral pleura wide.

Rhynchasphaera Bechyné 1955

Genotype: *Rhynchasphaera orophila* Bechyné 1955

3 species: (1 Bolivia, 2 Peru).

- 12 (1) Posterior femora very much thickened, its transverse diameter approximately equal to the tibial length; apical hind tarsal segment enlarged and globosely swollen; basal tarsal joint of posterior leg only about $\frac{1}{5}$ as long as tibia:

- 13 (28) Without pronotal prebasal transverse impression, in contrast to condition found in *Philopona*; if ambiguous, see No. 26:
- 14 (15) Antennae clavate, club formed of 4 markedly widened apical segments:
Pyxidaltica Bechyné 1956
 Monotypic: *Oedionychus variegatus* Jacoby 1880
 Brazil: S. Caterina, Parana, S. Paulo, Rio de Janeiro, Minas Gerais.
- 15 (14) Antennae filiform, not noticeably widened apically:
- 16 (17) Strongly convex forms – when viewed from side, height greater than half their length.
 Large forms, smallest measure 6 mm; strong elytral punctation:
Paranaita Bechyné 1955
 Genotype: *Oedionychus opimus* Germar
 15 species with numerous subspecies (Argentina, Paraguay, and adjacent Brazil):
- 17 (16) Less strongly convex forms – when viewed from side, their height is much less than half their length, only about $\frac{1}{3}$ of length:
- 18 (21) Epipleura not visible when viewed from side:
- 19 (20) Epipleura horizontal, not visible from side; elytral pleura wide and grooved in dorsal view.
Walterianella Bechyné 1955
 Genotype: *Oedionychus interruptovitattus* Jacoby (Rio Grande do Sul, Santa Catarina).
 More than 100 species with a distribution over all of South and Central America.
- 20 (19) Epipleura slanted upward and inward, not visible from side; therefore, elytral pleura not groove-shaped, but directed obliquely downward:
Alagoasa Bechyné 1955
 Genotype: *Oedionychus libentinus* Germar (Minas Gerais)
 About 150 species with several subspecies, whose distribution extends over all of South and Central America.
- 21 (18) Epipleura slanted downward, visible from side; elytral pleura visually narrow; if wider then, still not groove-shaped.
- 22 (25) Pronotum strongly convex, wide lateral margin sharply delimited and bent toward pronotum at a 90° angle; epipleura visible laterally at elytral apices.
- 23 (24) Metepisternae not tapered posteriorly but parallel-sided; metepisternae narrowing posteriorly in all other Oedionychini.
Nycteronychis Bechyné 1955
 Monotypic: *Oedionychus trivittatus* Baly (Brazil: Rio de Janeiro, S. Paulo).
- 24 (22) Very similar to previous genus, but with metepisternae narrowing posteriorly:
Araoua Bechyné 1955

Genotype: *Altica umbratica* Olivier (French Guiana)

A second species is known from Rio de Janeiro, whose position in this genus is questionable.

- 25 (22) Pronotum relatively flat, only weakly convex, wide pronotal margin not bent back against prothorax or bent back only a little:
- 26 (27) Epipleura wide and visible from side; sometimes, with only a slight indication of prebasal transverse impression on pronotum. Pronotum shortest in center, anterior margin appears to be cut out for head with apices extended anteriorly:

Wanderbiltiana Bechyné 1955

Genotype: *Oedionychus nitidus* Fabricius

Ca. 30 species: 1 Argentina, 1 Paraguay, 1 Mato Grosso, all others are limited to southeastern Brazil (S. Catarina, Paraná, S. Paulo, Rio de Janeiro, Espirito Santo).

- 27 (26) Epipleura relatively narrow; anterior and posterior margins of pronotum nearly parallel, somewhat longer in center; pronotum appears as a transversely arched rectangle except with anterior corners directed obliquely forward:

Kuschelina Bechyné 1951

Genotype: *Oedyonychus adjunctus* Jacoby (Bolivia).

More than 30 species: 1 Uruguay and southern Brazil; 4 Paraguay; 1 Paraguay and Argentina; 1 Paraguay and Brazil (Rio Grande do Sul, S. Catarina); 4 Rio Grande do Sul, 5 Santa Catarina, 1 Rio de Janeiro, 1 Pernambuco, 1 Paraíba, 2 Brazil without further details; 1 Argentina and Chile; 1 Bolivia; 3 Mexico; 1 Texas; 7 others, USA.

- 28 (13) Distinct prebasal transverse impression on prothorax similar so that in *Philopona*; in contrast to *Philopona*, intercoxal plate wide:
- 29 (30) Smaller 3–7 mm (usually around 5 mm) relatively narrow beetles, very flat. Epipleura visible from side; intercoxal plate plain or convex.

Capraita Bechyné 1957

Genotype: *Oedionychus sexmaculatus* Illiger (USA)

Ca. 60 species: South, Central, and North America.

- 30 (29) Larger, over 8 mm, more rounded and markedly convex; epipleura strongly slanted posteriorly, not visible from side; medially intercoxal plate with a sharp longitudinal keel which gives off lateral grooves:

Callangaltica Bechyné 1958

Monotypic: *Oedionychus batesi* Baly 1859 (Peru)

Group 3

On the outer edge of the posterior tibia there is a short stout tooth followed by a ciliated emargination.

- 1 (6) Small beetles from 1.5–3.5 mm. in length:

- 2 (5) Straight furrows run from antennal sockets over frons on either side to posterior margin of eye, their anterior margins simulate ridge-like antennal calli; this ridge-like structure uninterrupted to medial margin of eye.

Pronotum from above with visible side margins, moderately convex with frontal and hind angles:

- 3 (4) Inner edge of posterior tibia not elongate; small (1.5 –3.5 mm), ovate, black or metallic beetles:

Elytra base not wider than pronotum; elytra slightly convex, usually punctures in very exact longitudinal striae.

Pronotum usually punctate; sides slightly rounded and somewhat convergent anteriorly; anterior angles beveled; pronotal base medially somewhat widened posteriorly, therefore slightly doubly sinuate.

Chaetochema Stephens 1831

= *Halticops* Brethes 1928 syn.

Genotype: *Altica hortensis* Geoffroy (Palearctic)

Ca. 80 neotropic species, but distributed world-wide.

- 4 (3) Inner edge of posterior tibia elongated at apex, set with small teeth, and extends beyond posterior tibial spine inserted on lower edge of outer side. Small, 1.8–3.5 mm in length, round to oval; convex brown or black or brown flecked with black, or vice versa:

Elytral base not wider than pronotum; elytra strongly convex; punctuation varies from scarcely punctate to confused punctate to regular rows of punctures.

Pronotum smooth to strongly punctate; sides nearly straight and markedly converging anteriorly; anterior angles beveled; pronotal base widened posteriorly at middle, therefore markedly double sinuate.

Heikertingerella Csiki 1940

Genotype: *Homophyla adusta* Harold (Peru).

Ca. 70 species: Brazil to Central America.

- 5 (2) Head without frontal lines; pronotum cylindrical, margins not visible from above, constricted in front and on base, without forming front and hind angles. Elytra with very narrow lines of punctures:

Terpnochlorus Fairmaire 1904

Genotype: *Terpnochlorus perierei* Fairmaire (Madagascar)

3 species: 1 Madagascar, 1 Centralafrica, 1 Venezuela.

- 6 (1) Larger beetles, always longer than 4 mm in size; forms resembling *Chrysomela*; almost always yellow with dark pitch brown or reddish brown markings, rarely vice versa:

Frontal sutures curved, extending upward on either side of the antennal insertions (= inner and posterior limits of antennal calli); pronotal anterior edge at same level as posterior margin of eyes, with a row of pronounced punctures; this row curved toward lateral margin at an angle for half pronotal length; another row of punctures to either side at base; row of punctures on anterior

edge sometimes reduced to a single row of weak punctures and not angled toward lateral margin; rows of punctures on base sometimes completely absent. Elytral base as wide as that of pronotum; punctuation with few exceptions occurs in single rows:

- 7 (8) Frons without a striking transverse impression just before anterior margin:

Blepharida Rogers 1856

Genotype: *Chrysomela rhois* Forst. (USA to Mexico). 2 Palearctic species; 1 Nearctic species; ca. 20 neotropical species: 15 Mexico, 1 Guatemala, 1 Haiti, 1 Cuba, and Puerto Rico, 1 French Guiana, 1 Chile (?); ca. 40 African species; the number of Indo-Australian species is unclear (= ? *Ophrida*).

This distribution was the situation when the key was first published in 1962. The situation is still unclear. Bechyné recognized saw *Blepharida* as a Tribe and split them into several genera. In my opinion *Blepharida* includes one Nearctic species and many species in Africa. It is still too early for me to form an opinion about its distribution in the Neotropical and Oriental Region. It seems that many of the Central and South American species belong to the following genus *Notozona*.

- 8 (7) Frons with a conspicuous transverse impression just before anterior margin creating a swollen edge:

Notozona Clark 1865

Genotype: *Chrysomela bifasciata* Olivier (Guyenne)

22 species: 9 Central America; 5 Mexico, 2 Guatemala, 1 Nicaragua, 1 Honduras; 1 Venezuela; 3 Cayenne; 1 Peru (?); 4 Brazil without further details, 1 Amazon; 1 Bahia; 1 Pará; 1 S. Paulo, E. Santo, and Minas Gerais.

These two last genera could easily be confused.

Group 4

Eyes conspicuously emarginate and kidney-shaped:

- 1 (2) Emargination of eyes relatively slight; ca. 4–5 mm long, shagreened, very narrow flat forms:

Heikertingeria Csiki
(details p. 62)

- 2 (1) Eyes distinctly emarginate, wider forms, round to round-oval;

- 3 (6) Oval forms, elytra with parallel sides:

- 4 (5) Insects over 4 mm in size, yellow-brown with pitch brown markings on elytra:

Pronotum more than twice as wide as long; sides straight and converging anteriorly; anterior corners act as if attached; a slight transverse impression in front of base, not limited by sides.

Elytra with humeral and basal calli; confusely punctate. Frontal longitudinal and transverse carinae together T-shaped, highly convex and relatively broad; adjacent antennal calli give off pointed processes anteriorly which usually run into frontal longitudinal carina without interruption; space separating antennal calli continues to frontal longitudinal carina (Fig. 21a).

Very closely related to *Phenrica* Bech. (p. 67). *Phenrica* with only slightly kidney-shaped eyes.

Nephrica Harold 1877
(*Cyclophysa* Baly)

Genotype: *Nephrica kirschi* Harold

Ca. 4 species: 1 Amazon, 1 Bolivia, 1 Peru, 1 Peru and Amazon.

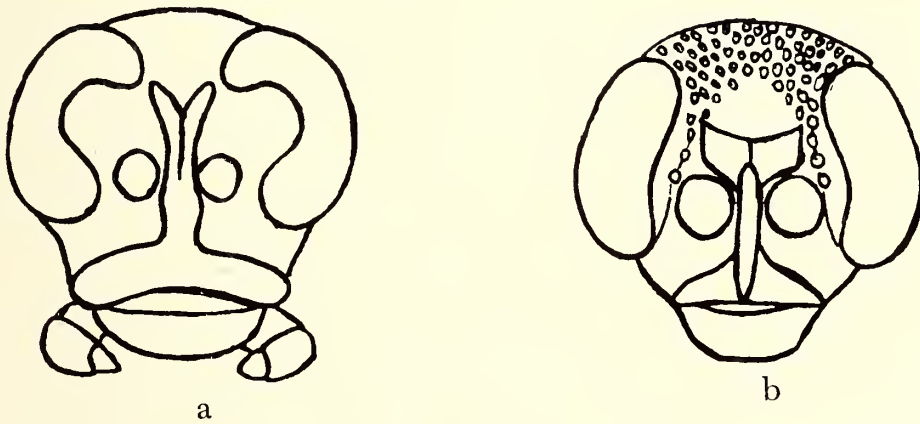


Fig. 21. Anterior view of the heads of
a) *Nephrica kirschi* Har.
b) *Phenrica aequinoctialiformis* Bech.
(Magnification a) 20×, b) 32×)

- 5 (4) About 6 mm in size, reddish-brown with a dark pitch-brown vertex:

Pronotum twice as wide as long, sides straight and converging anteriorly, anterior angles oblique; distinct prebasal transverse impression delimited on sides by longitudinal grooves.

Elytra scarcely punctate; humeral calli and medium-sized basal calli.

Frontal longitudinal carinae very long with shorter t-shaped transverse carinae; antennal calli moderate (Fig. 22).

Paralactica Bechyné 1961

Monotypic: *Haltica capitata* Illiger (Amazon)

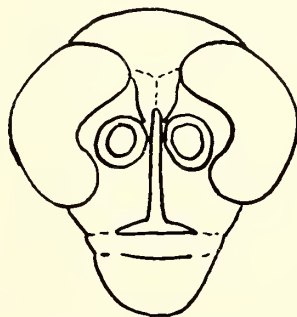


Fig. 22. Anterior view of the head of *Paralactica capitata* (Illiger)
(Magnification 20×)

- 6 (3) Round to round oval forms, sides of elytra rounded:
5.5–6 mm in size, yellow-brown.
Pronotum similar to that of previous genera, also twice as wide as long, but prebasal transverse impressions can be absent.
Elytra strongly convex; confusely punctate; basal calli only weakly developed.
- 7 (8) No pronotal prebasal transverse impression; behaves like a *Cassida* because antennae consist of strongly developed closely-spaced cylindrical segments:
Sophraenella Jacoby 1904
Monotypic: *Sophraenella fulva** Jacoby (Amazonas: Pará)
- 8 (7) With a very indistinct pronotal prebasal transverse impression clearly delimited by sharp longitudinal folds only on sides. **Pedilia** Clark 1865
Genotype: *Pedilia rufa* Clark (Pará)
3 species: 1 Pará, 1 Bahia, 1 Peru

Group 5

With a pronotal prebasal transverse impression:

- 1 (54) Pronotal transverse prebasal impression extends to lateral margins or runs into posterior angles, not delimited laterally by any longitudinal folds:
- 2 (5) Elytra without humeral or basal calli, scutellum absent.
- 3 (4) Pronotal margins diverge anteriorly to thickened oblique anterior angles; elytra with 9 continuous rows of punctures, with short juxtascutellar row fully developed in spite of absence of a scutellum, striae punctures likewise absent at humeral calli; all punctural striae even and distinct continuous to elytral apices.
Antennae thickened apically and extend over first quarter of elytra.
1.8–2 mm long, reddish-brown:
Exaudita Bechyné 1955
Monotypic: *Exaudita hilaria* Bechyné (Brazil: Santa Catharina)
- 4 (3) Pronotal margins diverge anteriorly to thickened anterior angles; elytra very indistinctly punctured, punctation distinct only near base; a deep sutural line very characteristic. Antennae filiform, long.
1.5–1.6 mm long, red-brown, elytra pitch brown:
Aulonodera Champion 1918
Monotypic: *Aulonodera darwini* Champ. (Chile)
- 5 (2) Elytra with humeral calli:
- 6 (23) Elytral punctural striae in 9 (withour marginal and short juxtascutellar rows) complete single or double rows.

- 7 (8) Antennal calli behind with broad callosity which also can be regarded as a posterior elongation of antennal calli to posterior margin of eye.

Pronotal anterior angles obliquely flattened, appearing somewhat tooth-like laterally; sides diverge markedly anteriorly; base constricted and transversely impressed; many times, with a tubercle in anterior half on either side of center.

Elytral base significantly wider than pronotal base; basal calli weak; punctation occurs in more or less orderly longitudinal striae up to double rows, frequently with confused punctation near suture.

Frons triangular, bent at a right angle to frontal calli when viewed laterally; a longitudinal frontal carina, if present, always very weakly developed.

Anterior coxal cavities closed posteriorly.

Very narrow beetles, 4–6 mm in length, yellow-colored, some with a red sutural border on elytra, or a red longitudinal band on either elytron (Fig. 23).

Nasigona Jacoby 1902

Genotype: *Nasigona pallida* Jacoby (Peru, Bolivia).

4 species: 1 Bolivia, 1 Peru, 1 Bolivia and Peru, 1 Cayenne.

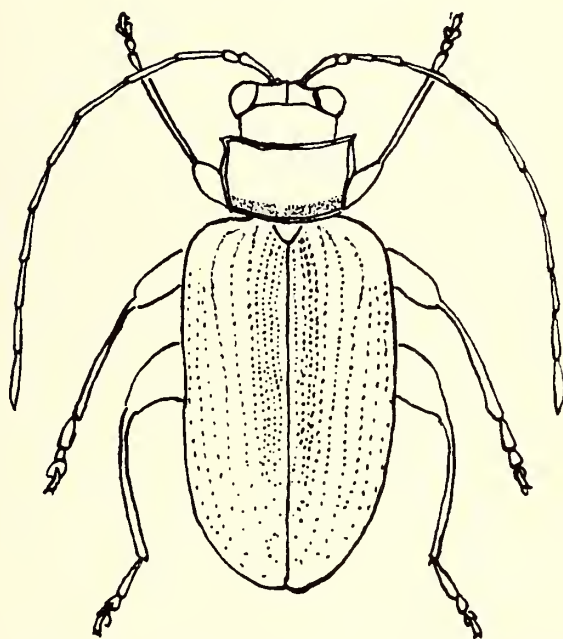


Fig. 23. Habitus of *Nasigona pallida* Jac.
(Magnification 10×)

- 8 (7) No other callosities occurring behind antennal calli:
- 9 (12) Pronotum strongly punctate:
- 10 (11) Pronotum with a few scattered, broad, shallow punctures (Fig. 24):
- Pronotal margins nearly straight; margins appear to diverge anteriorly, but actually with equal distance between anterior and posterior angles; base somewhat expanded at center, therefore doubly sinuous; prebasal transverse groove not distinct and therefore designated as a transverse impression; ratio of length to width in all species approximately 22:29.

Elytra long and parallel, width to length ratio 10:17; humeral and basal calli distinct; nine distinct punctural striae.

Antennal calli triangular; frons sutures extend to posterior margins of eyes; frontal longitudinal carina very thin and sharp. Antennae extend to middle of elytra.

Setose all over, although sometimes sparsely so.

2.6–3 mm in length, yellow or red, or pitch-brown beetles.

Leptophysa Baly 1877

= *Pseudoepitrix* Jacoby 1885 syn.

= *Goianinha* Bechyné 1957 syn.

Genotype: *Leptophysa batesii* Baly (Pará) (Fig. 24).

Leptophysa batesii Baly = *Systema deyrollei* Baly syn. 13 species, which are distributed over Central America and the islands of the Caribbean area, besides 4 species whose distribution lies in Argentina and all of Brazil.

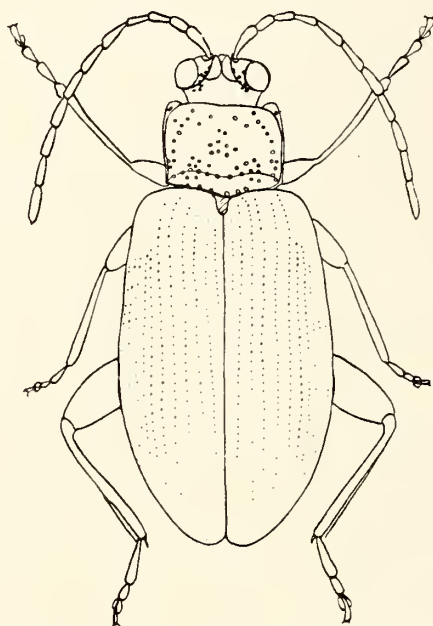


Fig. 24. Habitus of *Leptophysa batesii* Baly
(Magnification 15×)

- 11 (10) Pronotum evenly and strongly punctate.

Sides of pronotum rounded, convergent anteriorly; anterior angles rounded, but not slanted; upper surface moderately convex; prebasal transverse impression indistinct.

Frontal suture distinct, running to posterior margin of eyes, frontal longitudinal and transverse carinae flat and relatively wide.

Length 2.6 mm, dark brown with a dull sheen.

Apraea Baly 1877

Monotypic: *Apraea jansoni* Baly (Jamaica).

- 12 (9) Pronotum smooth or very finely punctate:
 13 (18) Anterior angles of pronotum not angulate:
 14 (17) Elytral striae consist partially or entirely of double rows of punctures:
 15 (16) Pronotal anterior angles obtusely bent toward anterior margin; sides with narrow margins; base constricted; sometimes an indistinct prebasal transverse impression, deepest medially; base may be depressed medially, which may simulate a widening posteriorly.

Elytral base significantly wider than pronotal base; basal calli distinct.

Antennal calli separated from each other by a groove, laterally they extend to inner margins of eyes, posteriorly separated by a perpendicular transverse groove; at times, this separation very indistinct with only the antennal calli separating suture visible; frons triangular.

Coxal cavities open posteriorly.

2.0–4.5 mm in length, yellow-brown, elytra with pitch-brown markings.

Orodes Jacoby 1891

Genotype: *Orodes nigropictus* Jacoby (Panama)

4 species: 1 Rio Grande do Sul, 1 Brazil without further details, 1 Venezuela, 1 Panama.

- 16 (15) Pronotal prebasal transverse groove very indistinct, usually only indicated by a punctate impression on either side of middle; sides regularly rounded.
 Elytral base slightly wider than pronotal base, short juxtascutellar row of punctures irregular and in part arranged in a double row, which can also occur in continuous row behind middle; basal calli very moderately formed.

Antennal calli large, separated from one another by a groove, posteriorly delimited by a marked, step-like transverse furrow which extends on either side to the inner margin of the eye; with no true lateral delimitation of antennal calli, but with only a flat indentation on inner margin of eye; frons closely bent against antennal calli; narrow longitudinal carina limited to space between antennae; robust antennae (males) extend beyond middle of elytra.

Length 3.5–4.2 mm; reddish brown; legs and antennae black; elongate oval.

Calliphron Jacoby 1891

Monotypic: *Calliphron ferrugineum* Jacoby (Panama).

- 17 (14) Elytral punctation in 9 very fine regular longitudinal rows: Pronotal prebasal transverse groove very distinct and definite; sides straight and diverging anteriorly; base constricted.

Elytral base significantly wider than that of pronotum; basal calli very strongly developed.

A 4 mm, metallic-blue colored species of the genus.

Chorodecta Harold 1875

The typical representatives of this genus appear under No. 27.

- 18 (13) Anterior angles of pronotum distinctly angulate:
 19 (22) Pronotal base dilated from center posteriorly, therefore doubly sinuous.
 20 (21) Elytra without post-basal transverse impression, basal calli weakly developed:
 Pronotal prebasal transverse impression sharply pronounced; lateral margin very narrow, sides nearly straight and parallel; prothorax relatively broad, greatest length (along middle) in proportion to width equals 19:29, laterally equals 9:29.

Elytral base hardly wider than pronotal base; elytra very convex.

Characteristic antennal calli absent, instead with ledge-shaped structures in front of straight frontal sutures which extend from posterior end of frontal longitudinal carina to posterior margin of eye; these antennal calli-like areas not delimited either anterior or laterally, thus extend to inner margins of eyes; frontal longitudinal and transverse carinae present. Antennae filiform and extending nearly to middle of elytra.

Anterior coxal cavities closed posteriorly.

Dodericrepa Bechyné 1951

Genotype: *Crepidodera aenescens* Boheman

3 species: Brasil: 1 Rio de Janeiro, 1 Mato Grosso, 1 S. Paulo.

- 21 (20) Elytra with post basal transverse impression, extremely convex basal calli:
 Pronotal prebasal transverse groove sharply pronounced, opening into lateral margin near posterior angles; a small fovea may occur near posterior corners over which transverse furrow passes; very convex; sides nearly straight and diverging anteriorly; greatest length (along middle) in proportion to width equals 26:31, on lateral margin, 16:31 (Genotype!)

Elytral base wider than pronotal base.

Antennal calli convex, well separated, only occasionally poorly separated from one another. Frontal longitudinal carina between antennae thin, frons long, triangular, and bent back against antennal calli. Antenna extend somewhat over middle of elytra; first segment very long, as long as subsequent 2 or 3 taken together.

Anterior coxal cavities open posteriorly.

2–3 mm long, usually brown, but also metallic-colored.

Exoceras Jacoby 1891

Genotype: *Exoceras facialis* Jac. (Panama).

10 species: 1 Guadeloupe, 1 Costa Rica, 1 Panama, 2 Venezuela, 1 Bolivia, 2 Rio de Janeiro, 1 Santa Catharina, 1 Paraná.

- 22 (19) Pronotal base **not** dilated from middle posteriorly, **not** doubly sinuate:
 Pronotal base constricted, sides nearly straight, diverging anteriorly; prebasal transverse furrow distinct opening to base near posterior angles (Fig. 25).
 Elytral base significantly wider than pronotal base; humeral and basal calli well defined.

Antennal calli triangular structures touching each other, horizontally limited behind, delineation in part somewhat indistinct. The lateral lines on frons, which run to posterior margin of eyes, are indicated by shallow depressions. Frons with longitudinal and transverse carinae. Antennae do not quite extend to middle of elytra. Anterior coxal cavities closed posteriorly; claws with basal appendages.

- a (b) Posterior femora do not extend to elytral apices, elytra not noticeably narrowed posteriorly, quite convex and decline near apices; intervals increase at posterior half of elytra.
2.8.–3.3 mm in length, yellowish-red anterior body and metallic elytra, also entirely metallic (Fig. 25).

Piobuckia Bechyné 1956

Genotype *Piobuckia promecosomoides* Bechyné (Rio Grande do Sul)
3 species: 1 Rio Grande do Sul, 1 S. Paulo, 1 Bolivia (in the case of the latter, I doubt the relationship to this genus).

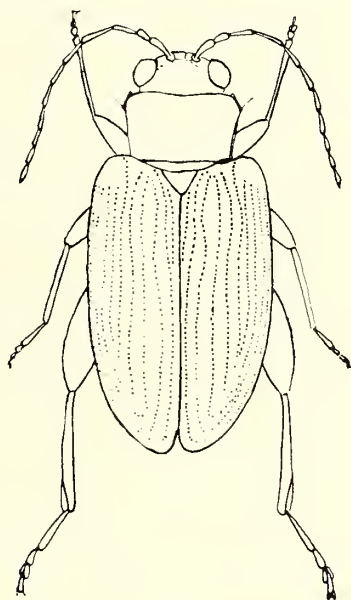


Fig. 25. Habitus of *Piobuckia promecosomoides* Bech. (♀)
(Magnification 10×)

- b' (a) Posterior femora extend nearly to apices of elytra or beyond them; elytra attenuated and flattened posteriorly; intervals decrease posteriorly:
c (d) Ventral surface of posterior femora with a bulge; tibiae markedly bowed.
4.8 mm in length; reddish yellow with dark violet elytra:

Tenosis Clark 1865

Monotypic: *Tenosis purpureipennis* Clark (Rio de Janeiro)

- d (c) Surface of posterior femora without any bulge; tibiae not markedly bowed (Fig. 26).

2.8–3.5 mm in length; anterior body reddish; antennae, legs, underside, and elytra black, also entire beetle black or metallic in color, or entire beetle brown, in part with black elytral base and margins:

Glenidion Clark 1860

= *Sangaria* Harold 1876 syn.

(*Ptinomorpha* Harold)

Genotype: *Glenidion rubronotatum* Clark

ca. 12 species: Bolivia, Columbia, Peru, northern Brazil, Venezuela, and Central America.

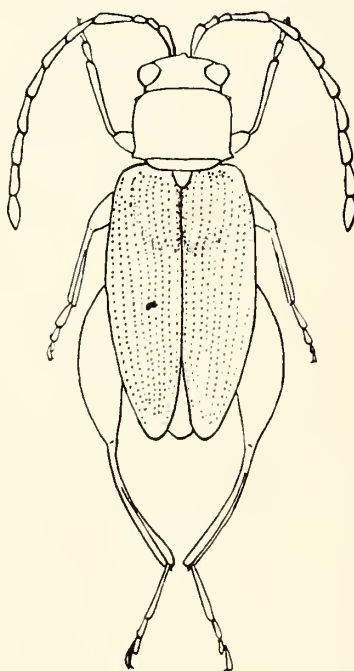


Fig. 26. Habitus of *Glenidion haagi* (Harold)
(Magnification 10×)

- 23 (6) Elytral punctation confusely, sometimes with a tendency to form longitudinal rows:
- 24 (35) Frontal sutures absent or very indistinct.
- 25 (28) Pronotal anterior angles definitely beveled:
- 26 (27) Pronotal lateral margins nearly parallel and very thin edged; prebasal transverse impression shallow and extending to lateral margins:

Elytra punctate with irregular narrow single or double rows, basal calli present.

Vertex relatively coarsely punctate-punctation can also extend in front of antennal sockets; antennal calli not delimited, recognizable only by absence of reticulation, separated from each other by a fine suture, forming a single surface anteriorly with frons. Antennae do not extend to middle of elytra.

Anterior coxal cavities open posteriorly.

Small, narrow, 2–3 mm long, light to dark pitch brown beetles.

Neothona Bechyné 1955

Genotype: *Neothona prima* Bechyné

12 species: 1 Rio Grande de Sul with a subspecies in S. Paulo, 2 Rio de Janeiro, 2 Mato Grosso, 2 Bolivia, 2 Venezuela, 1 Jamaica, 1 Trinidad.

- 27 (26) Pronotal lateral margins straight and diverging anteriorly; base constricted and distinctly transversely impressed.

Elytral base significantly wider than pronotal base; basal calli very strongly developed; punctation confused, with a tendency to form longitudinal rows.

Vertex with scattered strong punctures; antennal calli separated from one another by a strong suture, not delimited to the sides, limited behind by a fine line located on antennal calli themselves, i e., does not extend to inner margin of eyes; frons bent against antennal calli; frons very wide; antennae extend to first fourth of elytra.

Anterior coxal cavities open posteriorly.

2.4–4 mm long, relatively strongly convex short forms; entirely metallic green or blue, or black with a bronze sheen.

Chorodecta Harold 1875

Genotype: *Chorodecta coarctata* Harold (Colombia)

4 species: 1 Colombia, 1 Peru, 2 Bolivia.

This genus, because of the anterior angles of the prothorax, is listed again under No. 34.

- 28 (25) Pronotal anterior angles not beveled:

- 29 (30) Head coarsely punctate, especially near inner and posterior margins of eyes: Pronotal lateral margins nearly parallel and straight; length to width ratio 5:6, which at first glance produces a square impression; prebasal transverse impression smooth.

Elytral base significantly wider than pronotal base; basal calli hardly developed at all; punctations very fine and confused.

Antennal calli distinct, somewhat rectangular, close to one another; frontal longitudinal and transverse carinae wide, together T-shaped; when viewed from the side, frons only slightly bent toward antennal calli. Antennae do not extend to middle of elytra.

Anterior coxal cavities open posteriorly.

4.5–7 mm long, black, elytra with yellow longitudinal or transverse bands.

Agasicles Jacoby 1904

Genotype: *Agasicles vittata* Jacoby

3 species: 1 Bahia, 1 Amazon, 1 Brazil, Argentina, and Peru.

- 30 (29) Head not markedly punctate.

- 31 (32) Anterior coxal cavities closed posteriorly:

Pronotal sides usually more strongly rounded in anterior half, which simula-

tes a constriction of base, base somewhat wider than anterior margin; prebasal transverse impression weak.

Elytral base significantly wider than pronotal base; confusely punctate with a tendency to longitudinal rows; basal calli indistinct.

Antennal calli separated by longitudinal suture with no delimitation posteriorly and laterally; frontal suture completely absent; besides a frontal longitudinal carina, usually with a transverse carina; antennae extend to middle of elytra.

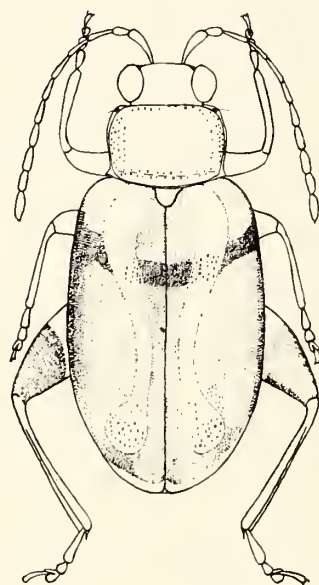


Fig. 27. Habitus of *Systema s-littera* (L.)
(Magnification 10×)

3–6 mm beetles, usually yellowish-brown, also green to black; usually a dark longitudinal stripe present on yellow elytral background (Fig. 27)

Systema Melsheimer 1847

= *Prasona* Baly 1861 syn.

Genotype: *Systema blanda* Melsh. (USA).

More than 70 species, of which 19 are Nearctic.

Previously Jacoby (Biol. Cent.-Amer. Col VI, 1, 1880–92 [1884] p. 321) doubted the validity of the genus *Prasona* but proposed the following distinction: Antennae, more robust, especially in the males, and the third antennal segment longer than the fourth. The antennae are more robust because the beetles are bigger and the other character too is not consistent. One can be misled by the green color in *Prasona*, and think that it is a common characteristic of the species before him.

- 32 (31) Anterior coxal cavities open posteriorly:
- 33 (34) Pronotum relatively wide, nearly twice as wide as long; Anterior and posterior margins about equally wide; sides evenly rounded; prebasal transverse impression shallow.

Antennal calli poorly defined; lateral frontal sutures usually very indistinct, area between them and internal margin of eye marked by punctures; frons relatively wide between antennae; viewed from front, inner margins of eyes converge posteriorly.

Elytral base little wider than pronotal base; punctation confused with a tendency toward longitudinal rows; basal calli scarcely developed.

2–3 mm long, usually metallic colored beetles, but also species with black elytra and yellow anterior bodies.

Genaphthona Bechyné 1956

Genotype: *Aphthona jessia* Bechyné

Ca. 13 neotropical species.

- 34 (33) Pronotal margins straight and diverging anteriorly; base constricted and distinctly transversely impressed.

Elytral base significantly wider than pronotal base; basal calli very prominently developed; punctations confused with a tendency toward longitudinal rows.

2.4–4 mm, relatively convex short forms; entirely metallic green or blue, or black with a bronze sheen.

Chorodecta Harold 1875

Genotype: *Chorodecta coarctata* Harold (Columbia)

4 species: 2 Bolivia, 1 Peru, 1 Columbia

This genus also keys out to No. 27 because of the variability of the anterior angles of the pronotum.

- 35 (24) Head with distinct frontal sutures:

- 36 (39) Antennal calli transverse and extending to inner margin of eyes; at most with a slight impression in front of inner margins of eyes, behind limited horizontally:

- 37 (38) Anterior coxal cavities open posteriorly:

Pronotum evenly convex; anterior and posterior margins appear equally wide, but with base wider; sides evenly rounded and running at anterior angles somewhat curved toward anterior margin, at anterior angles – curvature between forms, a right to an acute angle.

Elytral base somewhat wider than pronotal base; basal calli very weakly developed (more developed in yellow forms); punctation confused, and only in yellow forms with a tendency towards longitudinal rows (with other differences in antennal calli).

Frons short with a T-shaped longitudinal carina; antennal calli extend to inner margin of eyes, with somewhat more dense punctation near inner margin of eyes.

Forms up to 18 mm in size; head and thorax usually yellowish-brown to reddish-brown; elytra usually metallic green or blue, but also entirely yellow.

Cacoscelis Chevr. 1843

Genotype: *Altica marginata* Fabricius

More than 30 species distributed from Brazil and Paraguay to Mexico.

38 (37) Anterior coxal cavities closed behind:

Pronotal anterior and posterior margins appear equally wide, but with base wider; sides anterior to middle nearly straight and diverging, then curved to anterior angles; anterior angles obtuse, not rounded as in preceding genus – anterior setal pore entirely in the angle; prebasal transverse impression distinct.

Elytral base somewhat wider than pronotal base; basal calli moderately developed; punctation confused with a tendency toward longitudinal and double row formation.

Frontal longitudinal carina well developed, transverse carina weakly developed. Antennae extend nearly to middle of elytra.

Tibia with a longitudinal carina on back side.

Yellowish-brown, elytra metallic green, apices of tibiae and tarsi blackened (Fig. 28).

Itapiranga Bechyné 1956

Monotypic: *Itapiranga bicolor* Bechyné (Brazil: Rio Grande do Sul).

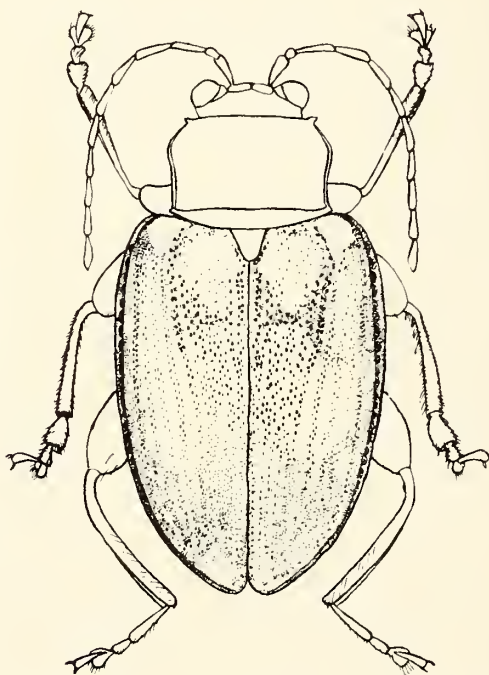


Fig. 28. Habitus of *Itapiranga bicolor* Bech. (Magnification 8×).

39 (36) Antennal calli laterally delimited by lateral sutures of frons – their posterior delimitation not horizontal to inner margin of eyes.

40 (49) Antennal calli transverse, touching on their shortest sides; also rounded to angular forms lying side by side.

- 41 (42) Internal margin of eye somewhat emarginate, eyes definitely kidney-shaped; inner margins of eyes strikingly divergent anteriorly:

Pronotum somewhat narrower than elytra; base somewhat beveled in area of humeral calli; base slightly impressed; sides slightly rounded and very narrowly margined, simply curved toward anterior margin at anterior angles, sometimes anterior angles pointed tooth-like toward sides.

Elytra long, parallel, and usually only slightly punctate.

Frons very wide, antennal calli restricted to middle, round to angular-round, lying next to each other, frontal longitudinal carina penetrating between them from in front; frontal longitudinal and transverse carinae together T- or Y-shaped.

Anterior coxal cavities open posteriorly.

4–9 mm long, elongated oval, elytra usually yellow with longitudinal black stripes (also vice versa) or also entire beetle metallic colored; more rarely, transverse stripes or both combined.

Disonycha Chevrolat 1844

Genotype: *Crioceris collata* Fabricius

More than 170 species, distributed from North to South America.

- 42 (41) Inner margin of eye scarcely emarginate; eyes ovate:

- 43 (46) Pronotal sides markedly rounded in front of middle so base appears constricted, but with base wider than anterior margin; if ambiguous, specimen longer than 5 mm:

- 44 (45) Small, usually 2.5–3 mm, sometimes 4.8 mm long forms; usually yellowish-brown, but also metallically colored:

Pronotal posterior angles protrude somewhat toward side; sides very narrowly margined and curved toward anterior margin at anterior angles; prebasal transverse impression smooth.

Elytral base somewhat wider than pronotal base; basal calli present; punctuation confused, sometimes hardly punctate at all, also a tendency toward longitudinal rows.

Antennal calli transverse; frontal carina weak and forming a T-shaped structure with longitudinal carina. Antennae reach nearly to middle of elytra.

First tarsal segment of hind leg longer than subsequent segments taken together, slightly longer than a third of tibial length. Anterior coxal cavities vary from open to closed.

Varicoxa Bechyné 1955

Genotype: *Systema ustulata* Harold

16 species: 2 Santa Catarina with a race of each in Rio de Janeiro and Rio Grande do Sul, 1 Paraná, 1 S. Paulo, 2 Rio de Janeiro; 2 Venezuela, 1 Peru, 1 Colombia and Central America, 1 Colombia and Venezuela, 1 Costa Rica, 2 Guatemala.

- 45 (44) Relatively large beetles, 5–7 mm long, brown or green with black markings, usually longitudinal stripes on the elytra:
 Pronotal posterior angles project to sides only slightly or not at all; anterior angles appear “stuck on” or as laterally projecting teeth; prebasal transverse impression usually smooth.
 Elytral base somewhat narrower than pronotal base; basal calli not developed; punctation confused with a tendency toward longitudinal rows.
 Contiguous antennal calli form an anterior process which blends into an arched, turculate frons between antennae; frons dilated and triangular anteriorly; antennae extend nearly to middle of elytra.
 Metatarsus of posterior legs of normal morphology; anterior coxal cavities closed posteriorly.

Disonychodes Bechyné 1955

Genotype: *Systema exclamationis* Boheman 1859

1 species with 3 races: Argentina, Paraguay, and southern Brazil.

- 46 (43) Pronotal sides evenly rounded, base only a little wider than anterior margin:
 47 (48) Small, 2–3 mm, usually metallic beetles, but also species with black elytra and yellow anterior bodies; bodies short and oval:
 Pronotum relatively wide, nearly twice as wide as long; prebasal transverse furrow shallow.
 Elytral base somewhat narrower than pronotal base; punctation confused with tendency toward longitudinal rows; basal calli barely developed.
 Lateral sutures of frons sometimes very indefinite; antennal calli in part poorly defined; area between inner margin of eye and lateral sutures of frons punctate; frons relatively wide between antennae; inner margins of eyes converge posteriorly when viewed from front.
 Anterior coxal cavities are open posteriorly.

Genaphthona Bechyné 1956

Genotype: *Aphthona jessia* Bechyné

Ca. 13 Neotropic species.

- 48 (47) Larger beetles, 3.5–8 mm long, metallic blue, green, or even black; elongated oval:
 Pronotal sides of pronotum rounded; the base somewhat wider than anterior margin, prebasal transverse furrow runs through a smooth pit to lateral border.
 Elytral base somewhat wider than pronotal base; basal callus weak; punctures in very irregular close longitudinal rows.
 Relatively large, triangular to rounded antennal calli; lateral sutures of frons run as strong furrows to posterior margin of eyes; longitudinal and transverse carinae of frons T-shaped.

Altica Fabricius 1775

Genotype: *Chrysomela oleracea* L (Europe)

More than 50 Neotropic species, otherwise distributed over the entire earth.

The genera *Conococha*, *Lysathia*, *Macrohaltica*, and *Deuteraltica* which were proposed by Bechyné (Bechyné: Contrib. to Neotrop. Fauna, I, 4, 1959, p. 303–305. – Pesquisas, Zoologia, nr. 6, 1960, p. 52, 53 and 55) are here united in the genus *Altica*. I have somewhat modified the following key from that of Bechyné (1960).

1 (2) Wingless forms:

Conococha Bechyné 1958

Genotype: *Altica blancasi* Bechyné

2 species are known from Peru. *Conococha parva* Bech. belongs to the genus *Syphraea*.

2 (1) Winged forms:

3 (4) Labrum with several setiferous punctures without definite location and number. Bodies over 5 mm:

Macrohaltica Bechyné 1959

Genotype: *Altica plicata* Erichson

4 (3) Labrum with 6 setiferous punctures (3 on either side) arranged in a transverse row:

5 (6) Antennae and legs robust, 4 anterior femora also thickened, antennal segments 2–4 progressively longer, first segment approximately as long as 2 subsequent ones together. Anterior coxae separated:

Lysathia Bechyné 1959

Genotype: *Altica flavipes* Boheman

6 (5) Legs and antennae slender, 4th antennal segment as long as 3rd and as long as 1st and 2nd taken together. Anterior coxae contiguous:

Deuteraltica Bechyné 1960

Genotype: *Altica longicornis* Jacoby

49 (40) Antennal calli upright, acute- or blunt-angled triangles; longitudinally contiguous.

50 (51) Pronotal base nearly as wide as elytral base; vertex rugosely punctate, many times with an unstippled spot in center of vertex:

Pronotal sides evenly rounded; surface reticulate and punctate.

Elytra reticulate and confusedly punctate; basal calli weakly developed.

Frontal longitudinal carina thin and long, transverse carina less well developed; genae very long.

Length 4–10 mm; metallic elytra and red anterior body.

Phrynocephala Baly 1861

= *Hemiphrynus* G. Horn 1889 syn.

Genotype: *Phrynocephala pulchella* Baly

9 species, whose distribution, with 1 exception (Paraguay), lies in Central America.

G. Horn (Trans. Amer. Ent. Soc. XVI, 1889, p. 200) proposed the genus *Hemiphrynus* with *pulchella* as the type species on the basis that these beetles have wide, deeply furrowed, sharpened posterior tibiae. However, the tibiae of *Phrynocephala* are similar but thinner.

50 (51) Pronotal base always noticeably more narrow than elytral base; vertex smooth, at most finely punctate, but never rugosely so.

52 (53) Anterior coxal cavities closed posteriorly:

Anterior pronotal angles seem more attached and less acute; angled somewhat to outside; sides diverge anteriorly in front to middle and then inflect toward anterior angles; base transversely impressed, impression ending in a basin-shaped area on posterior angles; pronotum relatively short, length to width ratio approximately 5:9.

Elytral base significantly wider than pronotal base; basal calli strikingly arched; very sparsely and weakly confusedly punctate; sparsely, finely setose near apices.

Area between antennal calli and inner margins of eyes deeply impressed, antennal socket gives a swollen delimitation; frons between antennae thin and flat, dilated anteriorly, plane; when viewed from side, frons flexed against antennal calli at an obtuse to a right angle; longitudinal carina absent; antennae extend to middle of elytra.

5–6 mm long, yellow-brown to reddish-brown beetles, usually with blackish tibiae and tarsi (Fig. 29).

***Teresopolisia* Bechyné 1956**

Genotype: *Teresopolisia callosa* Bechyné

5 species, which are distributed over the states of Rio de Janeiro and S. Paulo.

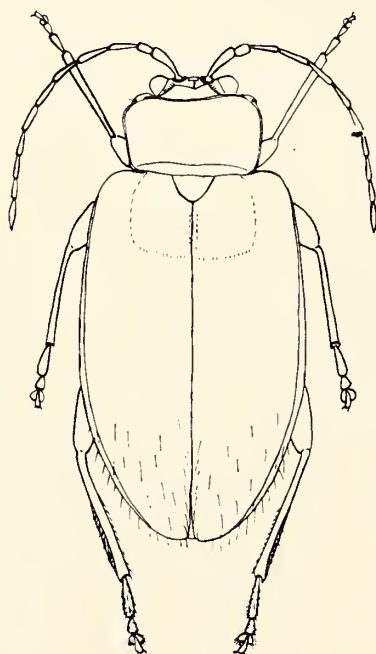


Fig. 29. Habitus of *Teresopolisia callosa* Bech.
(Magnification 7×)

- 52 (53) Anterior coxal cavities open posteriorly:
 4–8 mm long, slender, relatively flat forms; head and prothorax brown; elytra usually metallic green of blue:
 Thorax narrow, usually only $\frac{1}{4}$ as wide as long; sides diverging straight from base to just in front of middle, then inflecting inward to anterior angles; prebasal transverse impression shallow.
 Elytral base significantly wider than pronotal base; elytra parallel; basal calli moderately developed; punctation completely confused.
 Frontal longitudinal carina long; antennae extend beyond middle of elytra.
 First tarsal segment of hind legs longer than succeeding segments combined and $\frac{1}{3}$ as long as tibia.

Acanthonycha Jacoby 1891

Genotype: *Pelonia elegantula* Jacoby

More than 60 species: Distributed over all of Brazil, Bolivia, Peru, Colombia, Venezuela, and Central America.

- 54 (1) Pronotal prebasal transverse impression delimited laterally by longitudinal folds, or extends to base or fades near lateral margins:
- 53 (80) Pronotal base widens posteriorly in center, therefore doubly sinuous:
- 56 (77) Elytral punctation in 9 longitudinal rows, not counting marginal and short juxtascutellar rows of punctures:
- 57 (68) Pronotal anterior angles not beveled:
- 58 (65) Basal calli on elytra very distinct:
- 59 (62) Antennal calli as narrow ridges diverging obliquely posteriorly, not contiguous, frontal longitudinal carina penetrates between them at anterior end:
- 60 (61) Sides of pronotum smooth, evenly rounded; prothorax $1.5\times$ wider than long. Elytral base as wide as pronotal base; basal calli well developed, rows of punctures on basal calli somewhat faded.
 Small ridge-shaped antennal calli lie in front of straight frontal sutures which extend to posterior margins of eyes; frons and vertex smooth, frontal longitudinal carina and transverse carina T-shaped; antennae extend only to elytral base:
 Anterior tibiae of males strikingly dilated; anterior coxal cavities closed posteriorly.

2.2 mm pitch-brown forms with a leaden sheen and reddish-brown legs.

Yungaltica Bechyné 1959

Monotypic: *Yungaltica schindleri* Bechyné (Bolivia).

- 61 (60) Pronotal sides straight to somewhat rounded; Pronotum only $\frac{1}{5}$ as wide as long, therefore appearing square; prebasal transverse suture delimited at sides by a sharp longitudinal fold, but runs in a reduced form from there to lateral margin.

Elytral base significantly wider than pronotal base; basal calli well developed; intervals at times extremely convex, especially that portion extending posteriorly from humeral calli, elytra relatively long and slender.

Sharp frontal furrows extend from posterior end of frontal longitudinal carina to posterior margin of eyes; the area in front of furrows simulates ridge-like antennal calli; these 2 frontal furrows sometimes include a swollen elevation at their contact point.

Anterior coxal cavities closed posteriorly.

Ca. 2 mm long, metallic forms.

Diosyphraea Bechyné 1959

Genotype: *Diosyphraea heikertingeri* Bechyné

Ca. 9 species: Bolivia and Peru.

- 62 (59) Antennal calli relatively wide, contiguous, frontal longitudinal carina penetrates between them only at their anterior end, lateral lines of frons deep, extending to posterior margin of eyes:

Pronotal sides rounded; elytral base hardly wider than pronotal base; basal calli well marked, rows of punctures somewhat faded on them.

Anterior coxal cavities closed posteriorly; anterior tibiae dilated in males.

- 63 (64) Ca. 4.5 mm, pitch brown forms with reddish-brown extremities. Frontal longitudinal carina bears 2 tooth-like processis anteriorly which extend beyond labrum.

Marcapatia Bechyné 1958

Genotype: *Crepidodera longicornis* Jacoby (Peru)

Crepidodera inflatipes Bechyné from Santa Caterina also belongs to this previously monotypic genus.

- 64 (63) Ca. 3.5–6 mm, yellowish-brown to reddish-brown forms, some with dark markings. Frontal longitudinal carina with no processes anteriorly to extend beyond labrum:

Psilapha Clark 1865

Genotype: *Psilapha flava* Clark (Colombia).

7 species, which may in part (???) another genus. 4 Chile, 1 Colombia, 1 Peru (latter 2 with valid generic characters), 1 Argentina.

- 65 (58) Basal calli on elytra very poorly developed, hard to recognize:

- 66 (67) Prebasal transverse impression not extending laterally beyond longitudinal folds to lateral margins:

Prothoracic sides rounded; prothorax relatively wide, length to width ratio 3:5; upper surface smooth.

Elytral base hardly wider than pronotal base.

Antennal calli diverging posteriorly, usually not separated much anteriorly by frons, frontal sutures extending to posterior margin of eyes. Filiform antennae not extending to middle of elytra.

Anterior coxal cavities closed posteriorly.

3–4 mm, reddish-brown beetles.

Stegnea Baly 1879

Genotype: *Stegnea nigripes* Baly (Peru).

Some Neotropic species described under *Crepidodera* and under *Asiolestia* belong in this previously monotypic genus. Cf. Bechyné: Beitr. Z. neotrop. Fauna (Contr. to Neotrop. Fauna), I, 4, 1959, p. 319.

Stegnea nigripes Baly 1879 = *Crepidodera alwina* Bechyné 1955 syn.

- 67 (66) Pronotal prebasal transverse impression extends laterally over longitudinal folds to lateral margins:

Pronotum very convex; sides evenly rounded and converging somewhat anteriorly, length to width ratio approximately 5:6.

Elytral base as wide as pronotal base.

Area simulating antennal calli extends laterally to inner margin of eyes and many times consists of a shallow impression so that antennal calli may only be simulated. Frontal longitudinal carina narrow, usually with a transverse carina; antennae extend approximately to middle of elytra.

Usually small forms, 2.5–6 mm long, oval arched, compact beetles, many times a yellow-red head and thorax with metallic elytra, also entirely metallic, rarely entirely brown beetles.

A few *Syphraea* species have nine striae of punctures on elytra. The majority of species are confusely punctate with a tendency toward longitudinal striae.

Syphraea Baly 1876

Genotype: *Syphraea pretiosa* Baly

More than 100 species, distributed over South and Central America.

- 68 (57) Pronotal anterior angles definitely beveled:

- 69 (70) Elytra densely setose:

Pronotal sides slightly rounded; pronotum usually distinctly punctate:

Elytral base only a little wider than pronotal base; pilosity occurs at least on sides and at apices; basal calli, if present, only moderately developed.

Characteristic antennal calli, as in following 3 genera, not present, but only simulated by anterior edge of straight frontal sutures; frontal longitudinal and transverse carinae T-shaped. Antennae usually, extend over first third of elytra.

Anterior coxal cavities closed posteriorly.

Small, 2–3 mm, elongated oval, brown to black beetles with light colored setose.

Epitrix Foudras 1859

Genotype: *Epitrix atropae* Foudras (Europe)

Distributed over the entire earth.

- 70 (69) Elytra not densely setose:

Oval small forms, 1.5–3 mm in length. Pronotum wide and extremely convex, sides slightly rounded to straight. Elytral base no wider than pronotal base, elytra strongly convex, characteristic antennal calli absent, simulated by anterior edge of distinct frontal suture.

- 71 (74) Frontal longitudinal and transverse carinae together T-shaped; basal calli on elytra barely developed:

- 72 (73) Pronotum smooth or microscopically finely reticulated, thus appearing dull, but not punctate; pronotal prebasal transverse impression always very distinct.

Elytra markedly convex and rounded, not tapered posteriorly.

Margaridisa Bechyné 1958

Genotype: *Crepidodera flavescens* Baly 1876

8 species: 2 Santa Catarina, 1 Mato Grosso, 2 Venezuela, 1 Nicaragua, 1 Mexico.

- 73 (72) Pronotum distinctly punctate, many times with somewhat elongated punctures; prebasal transverse impression may be only a very weak indentation between distinct longitudinal folds, or may be completely absent.

Elytra taper posteriorly.

Small, 1.8–2.2 mm long, dark pitch brown to black beetles with reddish-brown extremities.

Minotula Weise 1924

Genotype: *Minotula nitens* Weise

3 species endemic to the Juan Fernandez Islands.

- 74 (71) Frontal transverse carina absent, longitudinal carina thickened; basal calli on elytra distinct:

- 75 (76) Frontal longitudinal carina wide between antennae, tapered anteriorly; pronotum barely punctate:

Hermenegilda Bechyné 1958

Genotype: *Hermenegilda clypeata* Bechyné

2 species: 1 Rio de Janeiro, 1 S. Paulo.

- 76 (75) Frontal longitudinal carina with a triangular dilatation anteriorly; prothorax distinctly punctate; often metallic forms:

Acallepitrix Bechyné 1959

Genotype: *Chalcoides erichsoni* Jacoby

Ca. 40 species distributed in South and Central America.

- 77 (56) Elytra confusely punctate, sometimes with a tendency toward longitudinal rows, also very narrow longitudinal rows occur, but always more than 9:

- 78 (79) Pronotal prebasal transverse impression weak, but laterally delimited by very distinct longitudinal folds; which cut through the impression.

Elytra very wide and without margin; epipleura very closely applied internally and above so that elytral pleura are very sharp when viewed from below; elytra very reminiscent of Cassidinae; antennal calli very definite, only somewhat fused with one another and with frontal longitudinal carina:

Pronotal anterior angles rounded, somewhat tooth-like, projecting to sides; sides slightly rounded, converging anteriorly; relatively wide, length to width ratio 5:9.

Elytral base as wide as pronotal base; basal calli extremely weak; punctations occurs in narrow longitudinal rows.

Lateral lines of frons deep and extending to posterior margin of eyes; antennal calli well separated at sides and posteriorly; frontal longitudinal carina tapers somewhat anteriorly, a transverse carina absent. Antennae extend to first third of elytra.

Anterior coxal cavities open posteriorly; prosternal process narrow.

Somewhat over 5 mm long, rounded oval beetles of a yellowish-brown color with pitch brown spots on head, pronotum, and elytra.

Parecynovia Bechyné 1958

Monotypic: *Parecynovia expulsa* Bechyné (Rio Grande do Sul).

- 79 (78) Pronotal prebasal transverse impression sharp and distinct, laterally delimited by longitudinal folds, beyond which transverse impression continues to lateral margin; elytra strongly convex and distinctly margined, epipleura remaining nearly horizontal; antennal calli absent, they are simulated by anterior margin of frontal suture; lateral frontal sutures absent:

Pronotum strongly convex; sides evenly rounded and somewhat convergent anteriorly; anterior corners only rarely definitely beveled, but anterior setal pores lie on sidemargin of anterior angles if angle not pronounced (viewed from behind); length to width ratio approximately 5:6.

Elytral base as wide as pronotal base; punctation confused with a tendency toward longitudinal rows (only a few species with 9 rows of punctures); basal calli only moderately developed.

Area which resembles antennal calli, situated in front of frontal sutures, extends laterally to internal margin of eyes and some times has a shallow indentation so that antennal calli simulated; frontal longitudinal carina narrow, and usually with a transverse carina; antennae extend to approximately middle of elytra.

Anterior coxal cavities open posteriorly; prosternal process margined and wide between coxae.

Usually small (2.5–6 mm long), oval, convex, compact beetles, many times yellowish-red head and thorax with metallic elytra, also entirely metallic beetles, rarely entirely brown beetles.

Syphraea Baly 1876

Genotype: *Syphraea pretiosa* Baly

More than 100 species, distributed throughout South and Central America.

- 80 (55) Pronotal base not widened posteriorly in center, therefore not doubly sinuous:

- 81 (88) Antennae as long or longer than the body:

- 82 (83) Pronotal anterior angles definitely beveled; sides nearly straight and parallel; width to length ratio approximately 7:5; prebasal transverse impression not very definite.

Elytral base little wider than pronotal base, elytra somewhat dilated posteriorly, basal calli distinct; elytra relatively long. Antennal calli contiguous, together forming an elevated equilateral triangle whose one apex lies in front of posterior end of frontal longitudinal carina; another callosity located behind antennal calli; frontal suture barely present; frontal longitudinal carina reduced to a tubercle-like elevation between antennae; frons triangular and completely smooth. Segment 1 of antennae very long, significantly longer than segments 2+3 taken together.

Anterior coxal cavities closed posteriorly.

2.5–3 mm long, relatively narrow yellowish-brown beetles.

Stenophyma Baly 1877

Genotype: *Stenophyma elegans* Baly (Brazil without further details). Another species is also known from Brazil (Pará: Amazon, Taracua, and Belem).

- 83 (82) Pronotal anterior angles not beveled:

- 84 (85) Very compact, convex, round-oval forms; disregarded posterior femora, reminiscent of *Chrysomela*:

Frons wide, more than $1.5 \times$ transverse diameter of eye; anterior margin of frons swollen, the frons itself flat and relatively wide between the antennae; additional carinae extend from the interantennal longitudinal carina back over the rounded antennal calli. Frontal sutures not distinct. Antennae slender, extend to apices of elytra.

Pronotum large and wide, sides rounded, and somewhat convergent; width to length ratio 9:5; with a short transverse impression in front of center of base.

Elytral base no wider than pronotal base; humeral calli normal, basal calli moderately developed; fine punctations (genotype) arranged in 9 more or less orderly continuous longitudinal rows visible only when viewed from side – when viewed from above, punctation appears confused.

Anterior coxal cavities closed posteriorly; posterior femora and tibiae very long, former extending beyond apices of elytra:

Yellowish-red with metallic or black elytra; length of genotype 7.2 mm.

Caloscelis Clark 1865

Genotype: *Caloscelis azureipennis* Clark (Pará)

3 species: Brazil, Peru, Mexico.

- 85 (84) Forms with a slender habitus:

86 (87) Elytra with 9 continuous rows of punctures:

Pronotal sides somewhat more rounded on anterior half so that base appears somewhat constricted; width to length ratio approximately 3:2; prebasal transverse impressions very distinct and deep, ending laterally shortly before angles; upper surface smooth.

Elytral base somewhat wider than pronotal base; basal calli moderate; elytral punctuation very distinct.

Antennal calli very markedly convex, posteriorly delimited almost horizontal by furrow which extends to posterior margin of eyes; lateral limits of antennal calli forms only an indentation, triangular frons very much bent against antennal calli; antennae extend to elytral apices; first antennal segments significantly longer and more robust than outer ones.

Anterior coxal cavities open posteriorly.

3.6 mm long, yellowish-red head and thorax, metallic blue or pitch-colored elytra.

Mesodera Jacoby 1885

Genotype: *Mesodera fulvicollis* Jacoby (Guatemala)

2 species: 1 Guatemala, 1 Mexico.

87 (86) Elytra confusely punctate:

Pronotal sides nearly straight; pronotum relatively short and wide, twice as wide as long; pronotal prebasal transverse impression indistinct, laterally bending forward so that when viewed from behind it resembles a very flat V. Elytral base hardly broader than pronotal base; basal calli moderately developed, humeral calli well-developed.

Antennal calli not very well separated, adjacent to each other; lateral lines of frons very weak; eyes large, somewhat kidney-shaped; frons triangular, longitudinal carina only hinted at; antennae long and slender, extending beyond elytra.

Anterior coxal cavities open posteriorly; posterior femora markedly thickened.

4.3 mm long; yellow with 2 black dots on each elytron.

Megasus Jacoby 1884

Monotypic: *Megasus bimaculatus* Jacoby (Guatemala).

88 (81) Antennae extend, at most, beyond middle of elytra:

89 (104) Nine continuous rows of punctures on elytra, not counting marginal and short juxtascutellar rows:

90 (91) Pronotal anterior angles definitely beveled; elytra densely setose:

Pronotal sides rounded; upper surface in part markedly punctate but also smooth; prebasal transverse impression very distinct.

Elytral base wider than pronotal base; basal calli moderate; anterior margin of frontal sutures marked as antennal calli by punctuation in front of internal

margin of eyes, therefore forming drop-shaped structures which give off processes to posterior margins of eyes; frontal longitudinal and transverse carinae T-shaped.

Anterior coxal cavities open posteriorly, prosternal process relatively wide. 1.8–3.8 mm long forms; usually yellow anterior bodies and metallic or black elytra, also entirely metallic forms.

Trichaltica Harold 1876

Genotype: *Trichaltica denticollis* Harold.

More than 30 species, distributed from Argentina to the USA. Brazil: Rio Grande do Sul 1, Santa Catarina 2, Rio de Janeiro 3, Rio de Janeiro and Minas Gerais 1, Goias 1, Mato Grosso 1 and 1 race, Rio Grande de Norte 3 and 1 race, Amazon 1; Argentina 3; Uruguay 1; Bolivia 4; Ecuador 1; Peru 1; Colombia 1; Venezuela 1; Colombia and Panama 1; Central America 2; USA 1.

- 91 (90) Pronotal anterior angles not angled; elytra with no setae or only a few setae:
- 92 (97) Anterior coxal cavities closed posteriorly:
- 93 (96) Pronotum very finely punctate or impunctated:
- 94 (95) Pronotal sides markedly rounded, somewhat convergent anteriorly but convergence not evident to naked eye; prebasal transverse suture distinct and opening into base near posterior angles.

Elytral base noticeably wider than pronotal base; basal and humeral calli distinct; single rows of punctures distorted on posterior half.

Relatively narrow antennal calli laterally delimited by very deep lateral frontal suture; eyes not very large, appear nearly hemispherical laterally; frontal longitudinal carina narrow, and a transverse carina scarcely present. Posterior femora not markedly thickened.

Reddish-brown, posterior half of elytra black; antennae to segments 2 & 3, tarsi, tibiae, femora in part black; length 4.4 mm.

Eugoniola Csiki 1940
(*Eugonia* Baly 1879)

Monotypic: *Eugonia dimidiatipennis* Baly (Peru)

- 95 (94) Pronotal sides slightly rounded, somewhat convergent anteriorly; prebasal transverse impression not very distinct.

Elytral base noticeably wider than pronotal base; basal calli moderate; continuous rows of punctures some times somewhat distorted.

Antennal calli large, adjacent, separated by a groove; lateral frontal sutures extend to posterior margin of eyes; frontal longitudinal carina narrow, with transverse carina; antennae extend beyond middle of elytra.

Posterior femora relatively slender.

Color yellow, elytral suture black, in part black dots on the thorax and

black vertex; legs and underside usually black and frequently mouth parts black; length 4–5.5 mm.

Iphitrea Baly 1864

Genotype: *Iphitrea limbata* Baly

8 species: 1 Bolivia, 1 Bolivia and Peru, 1 Peru, 2 Colombia, 1 Colombia and Venezuela, 1 Venezuela, 1 Central America.

96 (93) Pronotum coarsely punctate:

Pronotum somewhat more rounded on anterior half so that base appears constricted, but actually somewhat wider; pronotum nearly twice as wide as long (32:17); prebasal transverse suture very indistinct with delimiting longitudinal folds somewhat more definite.

Elytral base somewhat wider than pronotal base; basal and humeral calli definite; longitudinal rows very distinct.

Antennal calli scarcely defined, partially separated by a fine groove, but easily recognized as callosities; frontal sutures absent; frons tightly bent against frontal calli; head coarsely punctate like pronotum; antennal segments 1–5 of males much enlarged.

2.5–2.6 mm long; pitch-black colored, legs (posterior femora excepted) and antennal segments 1–5 yellow-brown. At first glance, reminiscent of *Epi-trix*.

Suetes Jacoby 1891

Monotypic: *Suetes niger* Jacoby (Mexico)

97 (92) Anterior coxal cavities open posteriorly:

98 (99) Pronotal anterior angles not projecting tooth-like to sides:

A yellowish-brown species, somewhat more than 8 mm long, legs and antennae black.

Pronotal lateral margins somewhat offset, slightly rounded; prebasal transverse furrow distinct but not deep, width to length ratio 3:2.

Elytral base significantly wider than pronotal base; elytra narrow and long, exactly twice as long as wide; basal calli moderate; longitudinal punctures sometimes scattered outside rows.

Antennal calli adjacent, with a bow-shaped callosity behind; lateral delimitation of antennal calli indicated only by a slight indentation; frontal longitudinal carina limited to a tubercle between antennae; frons indented transversely.

Aphanocera Jacoby 1884

Monotypic: *Aphanocera fulveola* Jacoby (Panama).

99 (98) Pronotal anterior angles projecting tooth-like to sides:

100 (101) Pronotal lateral margin flat to flange-like: elytral punctation occurs in double rows, usually on a shagreened surface, sometimes interval convex in a ledge form; basal calli not always definite; sides sometimes partially round-

ed, also dilated posteriorly, but also nearly parallel; base little wider than pronotal base.

Pronotal prebasal transverse impression usually very distinct and sharply pronounced; but also several species with weaker transverse furrows; marginated sides slightly rounded and convergent anteriorly.

Antennal calli large, limited laterally only by an indentation, very distinctly separated from each other posteriorly; frontal longitudinal carina with transverse carina.

Anterior coxal cavities just open posteriorly.

Usually 6–7 mm (also 4 mm) long and metallic blue; also species with red anterior bodies, entirely brown or even metallic green.

Chlamophora Chevrolat 1843

Genotype: *Chlamophora clypeata* Clark (Brazil: Rio de Janeiro)

Ca. 20 species, which all have their distribution in southern Brazil and in part in Argentina; the one species described by Jacoby from Panama must belong in another genus.

- 101 (100) Pronotal lateral margin not flat or flange-like:

- 102 (103) Antennal calli relatively narrow, diverge posteriorly and defined laterally by lateral frontal sutures; lateral frontal sutures may also be indicated only by a row of fine punctures; usually, no transverse carina developed on triangular frons, longitudinal carina disappears anteriorly. Antennae extend to anterior half of elytra.

Pronotal sides slightly rounded and converging very little anteriorly; prebasal transverse impression very distinct and sharp, with very robust lateral longitudinal folds.

Elytral base somewhat broader than pronotal base, basal calli very distinct; 9 continuous rows of punctures much reduced in some species.

Anterior coxal cavities open posteriorly.

2.5–5 mm beetles, yellow anterior bodies, with metallic elytra or even entirely metallic.

Diphaulaca Chevrolat 1849

Genotype: *Altica aulica* Olivier (Mexico to Colombia and Guyana).

More than 50 species: South and Central America.

- 103 (102) Antennal calli relatively wide, nearly horizontally delimited posteriorly; lateral frontal sutures absent, instead with a flat trough; frontal longitudinal carina disappears anteriorly on triangular frons, a transverse carina rarely present.

Pronotal sides slightly rounded, somewhat convergent anteriorly; prebasal transverse impression very distinct.

Elytral base scarcely wider than pronotal base; punctation occurs in 9 double rows or more rarely in single rows; basal calli distinct, scattered fine setae primarily on apical curvature.

Anterior coxal cavities open posteriorly.

3.5–5 mm long forms; yellow anterior bodies and metallic elytra (Fig. 30):

Plectotetra Baly 1862

Genotype: *Plectotetra clarki* Baly

16 species, distributed in Central America.

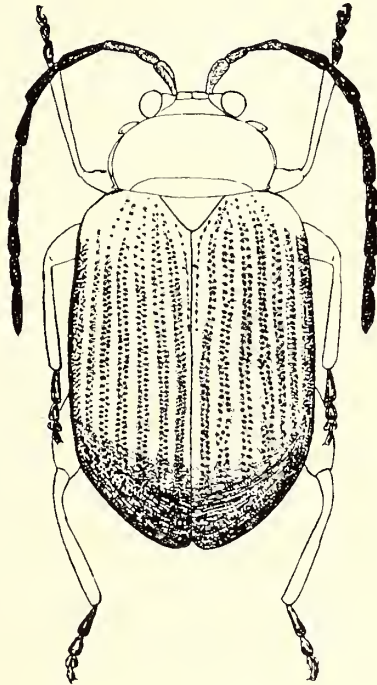


Fig. 30. Habitus of *Plectotetra clarki* Baly
(Magnification 8×)

104 (89) Elytra confusely punctate, sometimes with a tendency toward longitudinal rows:

105 (106) Elytra coarsely shagreened, 4 longitudinal carinae* on each elytron; characteristic humeral calli absent, instead 2 external longitudinal carinae fuse in this area:

Pronotal anterior angles rectangular; sides moderately rounded; upper surface finely shagreened, prebasal transverse impression sharp.

Elytral base no wider than pronotal base, but elytra much broader at their midpoint.

* Longitudinal carinae on the elytra are found in the genus *Hemilactica* Blake, 1927, which in nature is unknown to me. However, according to the description, the elytra apparently are not shagreened. Several longitudinal ridges on elytra are mentioned. Prebasal transverse impression is not very defined, except at its ends and in the middle. Head coarsely punctate. Apparently, this genus is limited to Cuba.

Genotype: *Hemilactica pulchella* Blake. 3–5 mm long; yellowish to reddish brown with pitch-brown spots. 2 species Cuba.

Antennal calli diverge somewhat posteriorly, with sharp lateral boundaries which extend as definite lateral frontal sutures to posterior margin of eyes; frontal longitudinal carina extends over frons, with no transverse carina. Antennae extend beyond middle of elytra.

Anterior coxal cavities open posteriorly.

7–8 mm long, metallic green.

Rhypetra Baly 1875

Monotypic: *Rhypetra costata* Baly (Colombia, Bolivia).

106 (105) Elytra always without these 4 longitudinal ridges:

107 (110) Elytra, and usually all of beetle, shagreened. Another shagreened genus occurs under No. 116 which differs from *Heikertingeria* only in the differently-shaped prebasal thoracic transverse impression (deep and distinct), and from *Lactina* in the more slender form:

108 (109) Relatively narrow and flat forms 4–5 mm long; head and thorax usually yellowish-brown to red; elytra uniformly dark or light with dark longitudinal vittae which may be interrupted into spots:

Pronotal anterior angles rounded off to anterior margin or cut short; sides slightly rounded, somewhat converging anteriorly; base relatively slightly compressed transversely – this transverse indentation limited at sides near posterior angles by a sharp longitudinal fold which runs obliquely forward to middle of pronotum; pronotum finely reticulated, therefore dull.

Elytral base scarcely wider than pronotum; basal calli only very weakly developed; confusely punctate with relatively wide, but very shallow punctures which may bear a seta.

Inner margin of eye emarginate, eyes appear kidney-shaped; closely adjacent antennal calli well defined by a fine reticulation, but many times poorly defined laterally; a longitudinal ridge extends over vertex behind antennal calli; characteristic lateral frontal sutures absent; frontal longitudinal and transverse carinae T-shaped; antennae extend beyond middle of elytra; segments 2 and 3 many times about same length.

Anterior coxal cavities open posteriorly. **Heikertingeria** Csiki 1940
(*Pelonia* Clark)

Genotype: *Pelonia nigripennis* Clark (Amazon, Peru).

14 species: Brazil, Paraguay, Bolivia, Peru, Panama, Guatemala. Most of the species are concentrated in the Amazon region.

109 (108) Relatively wide, roundish-ovate, convex forms, 6–8 mm in length, metallic green or blue in color:

Pronotal anterior angles pointed anteriorly or projecting somewhat to side; sides very slightly rounded; prebasal transverse furrow sharp.

Elytral base somewhat wider than pronotal base; basal calli moderately developed, shagreened, densely and confusedly punctate; many times with short setae.

Frontal longitudinal carina penetrates between antennal calli from front, lateral boundaries of calli form deep lateral frontal lines which extend to posterior margins of eyes, calli well defined posteriorly. Frontal longitudinal carina relatively wide without a transverse carina. Eyes not kidney-shaped. Antennae extend approximately to middle of elytra; third antennal segment significantly longer than second.

Anterior coxal cavities open posteriorly.

Lactina Harold 1875

Genotype: *Lactina murina* Harold (Ecuador)

14 species: 8 Colombia and Venezuela, 1 Peru and Bolivia, 1 Peru, 2 Ecuador.

- 110 (107) Elytra not strikingly shagreened:
 - 111 (116) Pronotum definitely constricted at base:
 - 112 (115) Anterior coxal cavities closed posteriorly:
 - 113 (114) Pronotum very narrow, width to length ratio 7:6; anterior edge meets head very close behind eyes so that eyes extend beyond sides; prebasal transverse impression weak with bordering folds sharp and curved to outside (Fig. 31):
- Elytral base wider than pronotal base; weak basal calli, punctation confused with tendency toward longitudinal and double rows; elytra gradually tapering posteriorly.

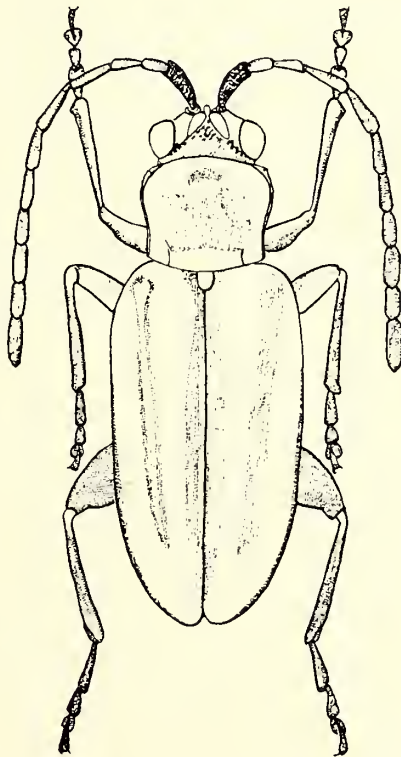


Fig. 31. Habitus of *Grammicopterus flavescens* Blanch.
(Magnification 10×)

Antennal calli distinctly delimited, drop-shaped, their tips directed back and out, e. e., they diverge posteriorly; deep lateral frontal sutures, which serve as lateral boundaries, extend to posterior margin of eyes and have relatively strong punctures; behind antennal calli lies an equally punctured callosity; frontal longitudinal carina sharp, transverse carina moderate; frons bent back against antennal calli (viewed from side). Antennae extend beyond elytra.

Yellowish-brown with pitch-colored frons and vertex, a pitch-colored spot on pronotum, pitch-colored elytral suture and margin, as well as a pitch-colored longitudinal band on each elytron, and blackings on the legs.

Grammicopterus Blanch. 1851

Monotypic: *Grammicopterus flavescens* Blanch. (Chile)

- 114 (113) Very similar to preceding genus, pronotum also narrow, width to length ratio 8:6–10:7; in the case of latter ratio, anterior angles extend beyond eyes (e. g., *B. consularis* Har., but this species not very typical of genus). Clearest difference shown in prebasal transverse impression of pronotum, here well pronounced and with lateral boundary bent toward inside:

Elytra parallel and not narrowed posteriorly.

Genotype without a characteristic frontal longitudinal carina but with a pointed process which extends anteriorly from the antennal calli; antennal calli not drop-shaped, lie close to each other but separated by a suture and bounded externally by lateral frontal sutures.

5.5–6.5 mm long beetles, yellow with dark spots on elytra or also with violet elytra.

Iphitroides Jacoby 1891

Genotype: *Iphitroides quadrimaculata* Jacoby

5 species: 4 Mexico, 1 Colombia

- 115 (112) Anterior coxal cavities open posteriorly:
 Pronotal prebasal transverse impression not very sharply defined, especially its lateral boundary; sides nearly straight; width to length ratio 9:6. Elytral base wider than pronotal base; average basal calli, confused punctures with a tendency toward longitudinal rows.
 Antennal calli close together separated by a suture, laterally and posteriorly, boundaries not always distinct, with a small callosity behind antennal calli; anteriorly, antennal calli give rise to a pointed process besides a relatively wide frontal longitudinal carina. Antennae extend to middle of elytra. 4.5–6 mm long; yellow head and pronotum, latter in part with pitch-brown spots; elytra dark with yellow margin and suture.

Caeporis Clark 1865

Genotype: *Caeporis marginata* Clark (Argentina, Brazil)

2 species: Argentina and southern Brazil.

- 116 (111) Pronotum not constricted at base:

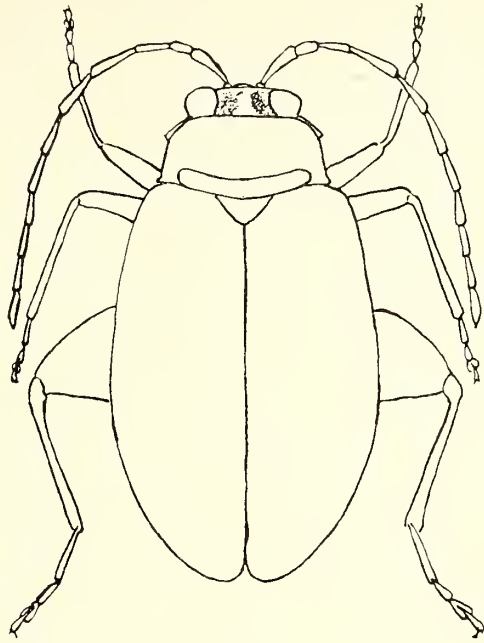


Fig. 32. Habitus of *Coroicona jacobyi* (Baly)
(Magnification 12×)

- 117 (126) Pronotal sides straight and markedly converging anteriorly:
 118 (119) Pronotal anterior angles pointed anteriorly; upper surface smooth; prebasal transverse impression very distinct; width to length ratio of pronotum 7:5: Elytral base significantly wider than pronotal base; punctation strong and confused, with a tendency toward longitudinal rows; strongly convex; basal calli weak.

Frontal sutures extend in a straight line from posterior end of frontal longitudinal carina to posterior margin of eyes, enclosing antennal calli; antennal calli small and diverging posteriorly; frontal longitudinal carina short, transverse carina not developed. Antennae not extending as far as middle of elytra.

Anterior coxal cavities open posteriorly.

Length 5 mm; legs black, head and thorax reddish-brown, elytra yellow.

Idaltica Bechyné 1955

Monotypic: *Idaltica boliviana* Bechyné (Bolivia)

- 119 (118) Pronotal anterior angles beveled; prebasal transverse impression also distinct; pronotum very wide and short, width to length ratio 10:5: Elytral base little wider than pronotal base; basal calli moderate. Antennal calli indistinct, contiguous; antennae usually extend beyond middle of elytra.

Anterior coxal cavities open posteriorly.

Beetles usually yellow, sometimes with black coloring on head, thorax, or elytra, also metallic-colored species; sometimes also with black coloring on femora and tibiae, also extremities entirely black (Fig. 32).

- 120 (121) Pronotum and elytra always reticulated, appearing dull; frontal longitudinal carina between antennae usually wider than one antennal socket; ocular sulci very deep, markedly widening above frons, thus vertex constricted (Fig. 32).

Coroicona Bechyné 1956

Genotype: *Lactica jacobyi* Baly 1879

9 species: Rio de Janeiro, Amazon, Peru.

- 121 (120) Pronotum and elytra not reticulated, smooth and shiny:
- 122 (123) Ocular sulci, as in preceding genus, very deep, widening very much above frons, so that frons and vertex present an elevated Y-like appearance:

Dinaltica Bechyné 1956

Genotype: *Lactica fromonti* Duvivier 1884 (Rio de Janeiro)

Ca. 25. species: Rio Grande do Sul 1, Santa Catarina 2, Parana 1, S. Paulo 4, Rio de Janeiro 11, Espirito Santo 1, Amazon 3, Bolivia 2, Peru 1.

- 123 (122) Ocular sulci obscure, only a single puncture remaining at posterior margin of eye:

- 124 (125) Posterior tibiae with a straight lower margin when viewed from side:

Lactica Erichson 1847

(*Monomacra* Dejean in litteris)

Genotype: *Lactica melaleuca* Erichson (Peru)

More than 120 species, with 4 of them in North America.

- 125 (124) Posterior tibiae with lower edge curved when viewed from side (almost tooth-shaped in males):

Utingaltica Bechyné 1961

Genotype: *Haltica calcarata* Illiger (Amazon)

4 species: 1 Amazon, 1 Peru, 1 Amazon and Peru, 1 Bolivia.

- 126 (127) Pronotal sides rounded or straight, but not noticeably convergent anteriorly:

- 127 (134) Coarse punctures on and beside lateral frontal sutures:

- 128 (131) Pronotum never twice as wide as long:

Yellowish- to reddish-brown roundish-ovate beetles, only a few species with metallic elytra; punctate lateral frontal sutures sometimes severely constrict vertex, although vertex itself, at least a middle strip, always free of punctures.

Pronotal lateral sides always moderately rounded; pronotal prebasal transverse impression not very distinct and many times weakly defined, not delimited laterally by longitudinal folds, but simply deflected toward base, sometimes slight perpendicular pits present.

Elytral base not wider than pronotal base; basal calli absent or only weakly developed; punctures confused, at times very weak, in part with a tendency toward longitudinal rows.

Antennal calli rounded adjacent structures; course punctures in front of eyes; antennae in part extend to middle of elytra.

Anterior coxal cavities open posteriorly.

- 129 (130) Frons very wide, eyes small, transverse diameter of an eye two or more times width of frons.

Yellowish- to reddish-brown beetles, usually with black tibiae and tarsi, rarely black heads; length 3–5 mm:

Strabala Chevrolat 1837

Genotype: *Altica ferruginea* Olivier

Ca. 23 species, distributed from southern Brazil to the southern USA.

- 130 (129) Frons less broad; eyes large, transverse diameter of eye at most width of the frons.

Yellowish- to reddish-brown beetles, only a few species with metallic elytra, tibiae and tarsi often black, often heads black, length 2.5–3 mm.

Parasyphraea Bechyné 1959

Genotype: *Monomacra nigriceps* Boheman

Ca. 17 species: Southern Brazil, Bolivia, Peru, Colombia.

- 131 (128) Pronotum usually twice as wide as long, appearing very short:

Pronotal anterior angles appear offset and many times project laterally like small teeth; sides almost straight, a little rounded; prebasal transverse impression usually very weak, definite lateral folds not present.

Basal calli on elytra usually weak, but always definite; if punctation present, very weak and confused.

Antennal calli rectangular, adjacent, many times contiguous; eyes more or less kidney shaped.

- 132 (133) Bodies flattened, equipped with a pronotal protuberance on either side near posterior angles.

Head and pronotum yellow with pitch-brown spots, elytra metallic blue with yellow margins; length 6.5 mm.

Balzanica Bechyné 1959

Monotypic: *Balzanica cacoscelina* Bechyné (Bolivia)

- 133 (132) Bodies normally convex, pronotum without protuberances:

Pronotum always yellow, head often black; elytra usually with yellow spots or bands on a dark background; also entirely yellow or rarely darkly metallic; length over 4 mm.

Phenrica Bechyné 1957

Genotype: *Disonycha austriaca* Schauf.

Ca. 70 species which are distributed over South and Central America.

- 134 (127) No coarse punctures present on or beside lateral frontal sutures; if character doubtful, then elytra erect pubescence:

- 135 (136) Small, narrow, 2.6–3.8 mm long beetles, with very weak basal calli on elytra and indistinct antennal calli; yellow with 2 black spots on elytra, also thorax and head may be darkened as well as leg segments and base and apices of antennae:

Prothorax relatively narrow with a length-to-width ratio of 6:7; sides smoothly rounded; prebasal transverse impression distinct, thin, and bounded at sides by a longitudinal fold; upper surface smooth and shiny. Elytral base little wider than pronotal base, punctation very weak and in very narrow longitudinal rows; in all species elytra are yellow with one black spot on each humeral callus which extends inwardly backwards over middle and another black spot behind middle which extends to just in front of elytral apices; apices themselves may likewise be black.

Antennal calli very indistinct; frontal sutures absent; interantennal longitudinal carina wide and dilated anteriorly; with a strong puncture near posterior margin of eye; antennae not extending to middle of elytra.

Anterior coxal cavities open posteriorly. **Cornulactica** Bechyné 1955

Genotype: *Cornulactica jacobyi* Bechyné

7 species: 1 Argentina and Santa Catarina, 1 Rio Grande do Sul, 1 Pará, 1 Bahia, 1 Amazon, 1 Venezuela, 1 Mexico and Guatemala.

- 136 (135) Elytra with distinct basal calli; antennal calli distinct; 4 black spots are missing from elytra:

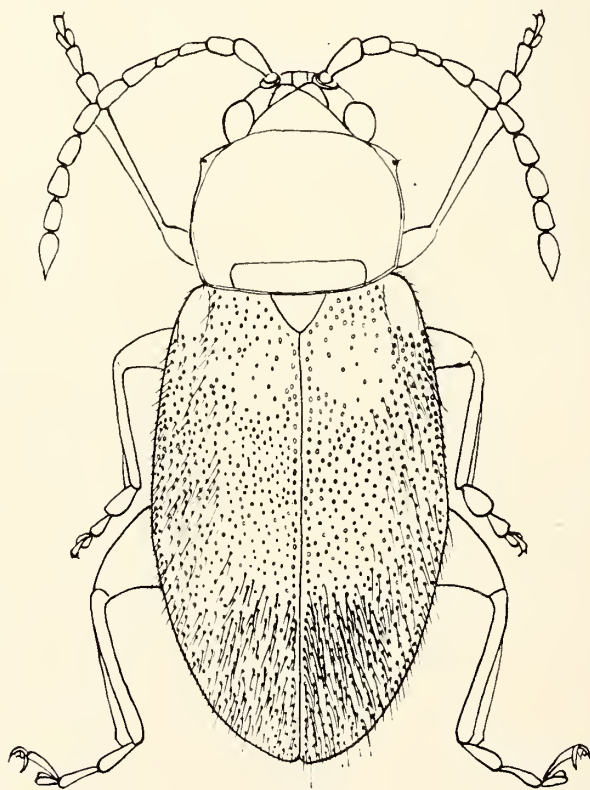


Fig. 33. Habitus of *Wittmeraltica muriensis* Bech.
(Magnification 15×)

- 137 (138) Elytra noticeably pubescent; elongated oval forms (Fig. 33):
 3.7 mm long; luminous yellow-red with black tibiae and tarsi, anterior angles of pronotum beveled; sides at first diverging from back to front and then inflecting toward anterior angles; prebasal transverse impression very definite and bounded laterally by longitudinal folds.
 Elytral base scarcely wider than pronotal base; basal calli distinct; confusedly and strongly punctate; with erect setae.
 Antennal calli large, diverging posteriorly; lateral frontal sutures very close to internal margin of eyes; interantennal longitudinal carina long; antennae extend to first third of elytra.
 Anterior coxal cavities open posteriorly.
- Wittmeraltica** Bechyné 1956
- Monotypic: *Wittmeraltica muriensis* Bechyné (Rio de Janeiro)
- 138 (137) Elytra not noticeably pubescent; roundish-oval forms:
- 139 (140) Antennal calli poorly separated, at least posteriorly; frons and vertex punctate; anterior angles of pronotum rounded off toward the anterior edge; with an interantennal longitudinal carina:
 Pronotal sides evenly rounded; pronotum twice, or nearly twice, as wide as long.
 Elytral base scarcely wider than pronotal base; confusedly punctate; basal calli distinct:
 Antennal calli rounded, poorly defined posteriorly and laterally; interantennal longitudinal carina and transverse carina T-shaped; antennae extend close to middle of elytra.
 Anterior coxal cavities open posteriorly.
 Approximately 5 mm long; all species reddish-yellow with blue or blue-green elytra.
- Resistenciana** Bechyné 1956
- Genotype: *Lactica clara* Harold 1875 (Colombia)
 5 species: 1 Panama, 1 Colombia, 1 Peru, 1 Bolivia, 1 Brazil (Chaco) and Paraguay.
- 140 (139) Antennal calli well separated; frons and vertex impunctate; anterior angles of pronotum projecting tooth-like to sides; only a longitudinal interantennal carina:
 Pronotal sides somewhat rounded; prebasal transverse impression very definite.
 Elytral base somewhat wider than pronotal base; elytral punctuation confused with definite tendency toward longitudinal rows (the few small brown species punctate in more or less regular longitudinal rows); basal calli distinct.

Antennal calli diverging posteriorly, appearing as small, distinctly shaped diamond-shaped structures, well limited to all sides, deep lateral frontal sutures extend to posterior edges of eyes; interantennal longitudinal carina fades out anteriorly; antennae extend nearly to middle of elytra.

Anterior coxal cavities open posteriorly.

2.8–4.8 mm long, usually blue or metallic green, also with yellow anterior bodies or even entirely yellow.

Diphaltica Barber 1941

Genotype: *Diphaulaca nitida* Jacoby 1884 (Central America)

Ca. 16 species and subspecies: 1 Santa Catarina with 2 races in the Amazon area and 1 in Minas Gerais, 4 Rio de Janeiro, 3 Bolivia, 3 Peru, 2 Central America.

Group 6

Pronotal base without any transverse impression:

Division A

Apical spur of posterior tibiae wide, bifurcated on apex, overall shape ovate; head deeply sunken in prothorax, scarcely visible from above.

- 1 (2) Eyes widely separated at top of inner margin of eyes:

Dibolia Latreille 1829

Genotype: *Haltica cryptocephala* Koch 1803 (Europe)

4 Central American species, 3 Nearctic species, ca. 30 Palearctic species, 17 African species.

- 2 (1) Eyes so close together at top of inner margin of eyes that only a very thin strip left between them:

Usually small 3–4.5 mm long beetles of yellowish-brown coloration with pitch-brown stripes or spots, or vice versa.

Megistops Boheman 1859

Genotype: *Megistops quadrinotatus* Boheman (Ecuador)

24 species: 11 Central American species; 2 Ecuador, 1 Bolivia; 1 Bolivia and Argentina; 1 Paraguay; 6 Brazil; 1 Venezuela; 1 presumed from South America.

Division B

Apical spur of posterior tibiae with one tip, not divided:

- 1 (10) Pronotum dilated posteriorly from middle, therefore doubly sinuous:

- 2 (7) Elytral punctures in 9 continuous longitudinal rows, excluding marginal and short juxtascutellar punctures:

3 (4) Elytra without basal calli and without prominent humeral calli:

Pronotal sides markedly convergent anteriorly; base of elytra just as wide as pronotal base; elytra very convex. Head less marked, very flat; eyes less arched; hardly any lateral frontal sutures. Posterior femora greatly thickened; posterior tibiae sharp-edged, margins serrate at apices; apical spur noticeably long (Fig. 34). 1.2–1.8 mm black beetles of very compact somewhat ovate shaped; legs and antennae more clear.

Pseudodibolia Jacoby 1891

= *Monotalla* Bechyné 1956 syn.

Genotype: *Pseudodibolia picea* Jacoby (Mexico, Costa Rica)

3 species: (1 Mexico, Costa Rica, 1 Guadeloupe, 1 Grenada).

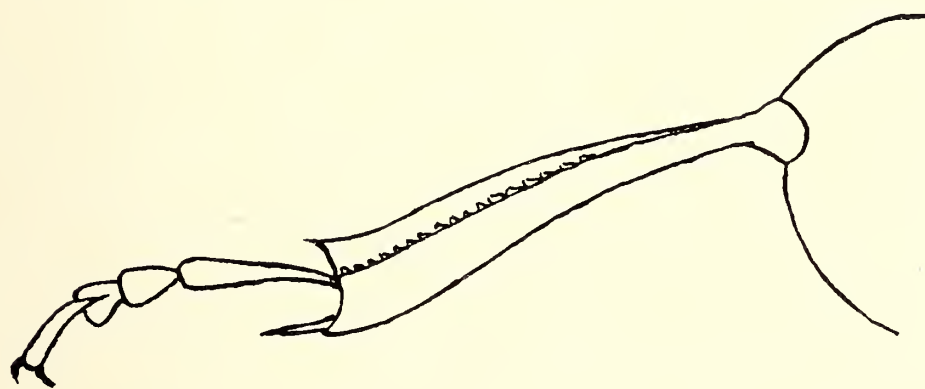


Fig. 34. Hind leg of *Pseudodibolia guadeloupensis* (Bech.)
(Magnification 80×)

4 (3) Elytra with basal calli, and well-developed humeral calli:

5 (6) Pronotal anterior angles definitely beveled, sides slightly rounded, subparallel; upper surface convex, only the base is somewhat flatter which almost gives an illusion of an indistinct transverse impression:

Very distinct frontal sutures, running in a straight line from posterior end of interantennal longitudinal carina to posterior margin of eye; characteristic antennal calli not developed; short interantennal longitudinal carina lying between antennae widens anteriorly into a triangular frons.

1.4–1.5 mm long, yellow-brown

Guadeloupena Bechyné 1956

Monotypic: *Guadeloupena caduma* Bechyné (Guadeloupe).

6 (5) Pronotal anterior angles rounded, not beveled; sides rounded, converging anteriorly, upper surface coarsely punctate, evenly convex, base not convex which gives illusion of a transverse impression.

Frontal sutures distinct, extending to posterior margins of eyes; interantennal and transverse carinae flat and relatively wide.

Length 2.6 mm, dark pitch-brown with a dull sheen.

Apraea Baly 1877*

Monotypic: *Apraea jansoni* Baly (Jamaica)

- 7 (2) Elytra confusedly punctate, or with a tendency toward longitudinal rows:
- 8 (9) Posterior tibial apices elongated on inner side to a tooth-like process so that at first glance posterior tibial apices appears doubly-toothed; marginal edges of posterior tibiae equipped with fine teeth at apex; all tibia of males very distinctly dilated at their apices.**

Pronotal sides slightly rounded and converge anteriorly; length (in middle) to width ratio 7:10, on sides ratio only 4.5:10.

Elytra finely punctate with a tendency toward longitudinal rows, basal calli absent, humeral calli well developed; elytral base somewhat wider than pronotal base.

With a sharp, nearly straight frontal suture on either side which extends from posterior margin of interantennal longitudinal carina to posterior margin of eyes; above these frontal sutures, i. e. in right angle included in them, lie 2 antennal-calli-like structures; frons with longitudinal carina and a weaker transverse carina; antennae extend somewhat beyond middle of elytra.

This genus is very closely related to *Heikertingerella* Csiki, but it lacks the apical tooth and related ciliated emargination of the hind leg. 4–4.6 mm long, wide oval, very compact, metallic green.

Doloresa Bechyné 1955

Monotypic: *Doloresa metallica* Bechyné (Venezuela)

- 9 (8) Apices of posterior tibiae not specialized; upper side of posterior tibiae not laterally bounded by 2 marginal borders, but formed by a single median keel-like protuberance.

* This genus must be very similar to the genus *Glyptobregma* Blake which is unknown to me. According to the description, *Glyptobregma* also has deep frontal sutures, a coarsely punctate pronotum, the base of the thorax is widened posteriorly in the middle, the strong longitudinal punctation of the elytra, and anterior coxal cavities opening posteriorly. Genotype of *Glyptobregma* is *Glyptobregma portoricense* Blake (Puerto Rico).
6 species: 5 Cuba, 1 Puerto Rico.

** The genus *Argopistes* Motsch. has at the apex of the hind tibia a similar structure (4 Palearctic, 17 Indo-Australian, 9 African and 2 Nearctic species [1 Florida, 1 Mexico]), 2 species from the Caribbean area are known, one from Cuba and one from Puerto Rico. This genus is immediately recognizable, however, from its coccinellid-like round form. Pronotal base very tight to elytra base. The pronotal sides converge anteriorly much more, the pronotal base is more than twice as wide as it is long, while in *Doloresa* it is never more than 1.5 × as wide. Also the elytra are completely rounded, and humeral calli are only weakly indicated. The epipleura are very closely appressed internally so that the elytral edge takes on a knife-like appearance. In *Doloresa*, the epipleura are positioned more or less horizontally and visible from the sides. The genotype is *Argopistes biplagiatus* Motsch. (East Siberia to Japan and China).

Pronotal sides nearly straight and converging anteriorly; length to width ratio in middle 4:8.5, and on sides, 3:8.5.

Elytral base little wider than pronotal base; basal calli absent, humeral calli well developed; elytra very convex.

Frontal sutures absent; antennal calli flat roundish structures; interantennal longitudinal carina somewhat widened in front of antennae and bounded at its anterior end by a transverse carina; antennae extend approximately to middle of elytra.

Forms about 4.5 mm in size, yellow, often with dark markings on elytra.

Ptocadica Harold 1876

Genotype: *Ptocadica straminea* Harold (Panama, Colombia, Peru).

Another species is known from Costa Rica.

- 10 (1) Pronotal base not dilated from middle posteriorly, therefore not doubly sinuous:
- 11 (44) Pronotal anterior angles not beveled:
- 12 (35) Elongate, inconsistent forms, not chrysomelid or coccinellid-like habitus:
- 13 (14) Beetles incapable of flight, with abbreviated elytra which leave very large pygidium and following tergites uncovered (Fig. 35):

Elytra often somewhat wrinkled and covered with scattered setae, not very convex.

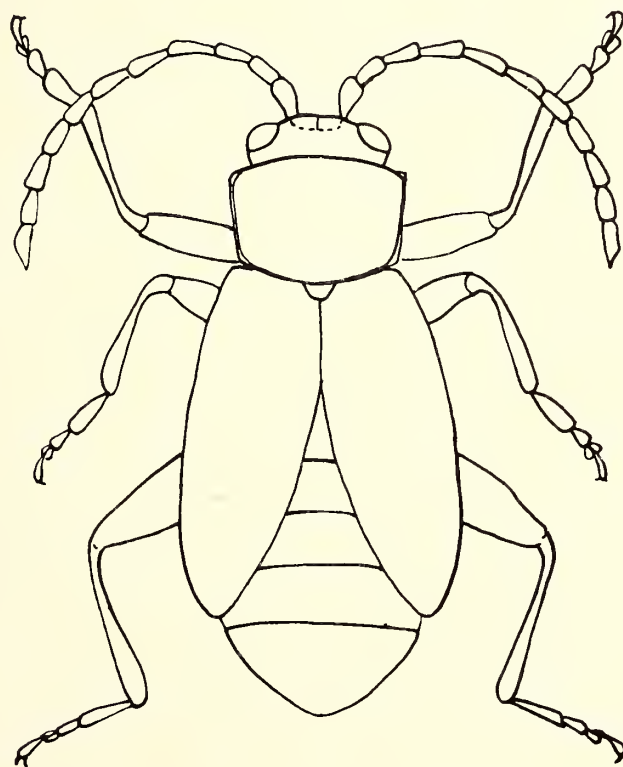


Fig. 35. Habitus of *Forsterita boliviensis* Bech.
(Magnification 15×)

Pronotum flat, sides diverge somewhat anteriorly; upper surface almost smooth or shagreened to wrinkled; frons very wide; antennal calli relatively large, approximate but separated from each other by a prominent fissure; no definite frontal sutures; interantennal longitudinal and transverse carinae relatively flat; antennae robust.

3–3.5 mm long, metallicallly-colored forms.

Forsterita Bechyné 1959

Genotype: *Forsterita boliviensis* Bechyné

3 species: 1 Bolivia, 2 Peru.

This genus is very closely related to those genera recorded in the catalogs as Galerucinae: *Sjoestedtina* Weise from Africa (Kilimmanjaro), *Khasia* Jacoby from India (Nilgiri Hills and Khasi Hills), and *Marseulia* Joannis from Syria.

- 14 (13) Elytra of normal length, not shortened:
- 15 (34) Posterior tarsus with first segment of normal length, no longer than half of tibia.
- 16 (17) Antennal calli poorly defined posteriorly, frontal sutures indistinctly marked:**** ****

*** The genus *Nasidia* Harold, 1876, which is unknown to me in nature and is related to the next genus (*Oxygona*), also has poorly defined antennal calli. The middle antennal segments are the shortest (the fifth the shortest, and the eight the longest). The interantennal longitudinal carina is short and arched like a bow, and does not subdivide into anteriorly diverging branches bounding the posterior part of the clypeus, as in *Oxygona*. There is a pit-like depression having a few punctures on the middle of the frons above the antennal calli extending to the eyes. Anterior corners of the pronotum not projecting tooth-like to the sides. The pronotum is flattened near on base with an indication of a longitudinal keel in the middle. Upper surface finely punctate: Elytra significantly wider than the pronotum; definitely and closely punctate. The upper surface of the tibiae is flattened and furnished with a sharp longitudinal keel (similar to that in *Itapiranga*). Front tarsal segment of posterior legs as long as the remaining tarsal segments, anterior coxal cavities closed, 7 mm long; elongate oval; moderately convex; rust red; underside and femora yellow; femora at the tips and on the outer margin, dark-brown; tibiae, tarsi, and antennae, blackish.

Monotypic: *Nasidia haagi* Harold (Colombia).

**** According to its description the genus *Prosplecesta* Weise, 1921, belongs next to *Nasidia* Har., but differs significantly because of the frontal form and the wide body structure which resembles that of a wide *Phyllodecta*. The antennal calli are distinguished only by their smoothness from the remaining wide frons. Antennae filiform, extending to the humeral calli of the elytra; of the short segments 2–6, the second is somewhat smaller than the remaining ones, and the following longer segments are all the same length. Thorax wide, less rounded posteriorly, markedly rounded anteriorly, anterior angles fully rounded. Elytra only a little wider than the pronotum, less closely, but extremely, finely punctate; suture edges separated by an indent strip; regularly two rows of punctures on the outer edges, the remainder fairly diffuse. Anterior coxal cavities closed; tibiae with a fine central ridge on their back sides; first tarsal segment as long as the following two segments together. 3.5 mm long; reddish-brown, shiny; antennae from segment 4, tips of the femora, tibiae, and tarsi black.

Monotypic: *Prosplecesta rufula* Weise (Amazon: Manaos).

Frons usually more or less punctate, head and pronotum black or dark metallic, elytra either same or with yellow markings; Posterior femora very thickened.

Phyllotreta Chevrolat 1837*

Genotype: *Chrysomela brassicae* F. (Europe)

15 Neotropic species (Central America), otherwise distributed over the entire world.

17 (16) Antennal calli also well defined posteriorly:

18 (19) Pronotal anterior angles directed laterally and tooth-like galericine-like (Fig. 36).

Phyllotrupes Hope 1840

Oxygona Chevrolat 1847

(*Oxygonus* Clark 1865)

Genotype: *Crioceris cyanipennis* Fabricius

Ca. 30 species, distributed in South and Central America.

19 (18) Pronotal anterior angles not laterally directed and tooth-like:

20 (29) Pronotal sides moderately rounded so that base appears same width as anterior edge, base actually somewhat wider:

21 (22) Antennae usually extend to elytral apices or beyond; antennal calli, adjacent, and upright size, bounded posteriorly by a triangular callosity which confines wrinkled or punctate vertex:

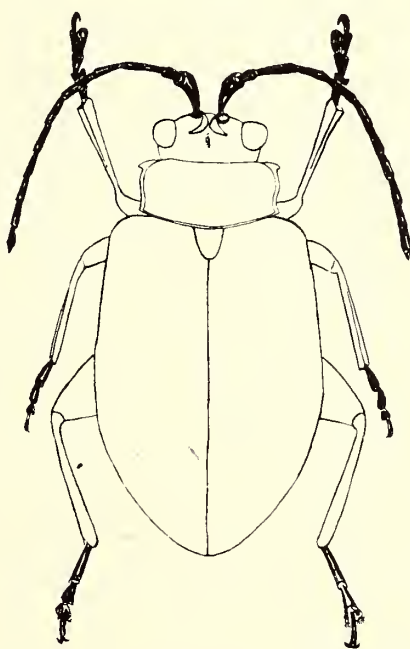


Fig. 36. Habitus of *Phyllotrupes acutangula* (Chevr.)
(Magnification 5×)

* White R. E. 1970: Authorship of the Genus *Phyllotreta* (Chrysomelidae: Coleoptera). – Proc. Soc. Wash. 72 (3): 388–389.

Antennal calli give off processes anteriorly which usually change without interruption into interantennal space, frons triangular in front of antennae with no longitudinal and transverse carina and sharply bent against upper frons when viewed from side.

Pronotal margin usually sulcus-like, pronotum $1.5 \times$ as wide as long.

Elytra wide, somewhat wider posteriorly, confusedly and relatively coarsely punctate.

Internal edge of apex of posterior tibia cut off nearly rectangularly and with very fine teeth. Anterior coxal cavities open posteriorly.

3–6 mm forms, brown or brown with metallic spots, also entirely metallically colored (Fig. 37):

Ocnoselis Erichson 1847

Genotype: *Ocnoselis cyanoptera* Erichson (Peru).

Ca. 14 Neotropic species (6 Peru, 3 Bolivia, 2 Colombia, 1 Venezuela, 2 Argentina [Prov. Yujuy]).

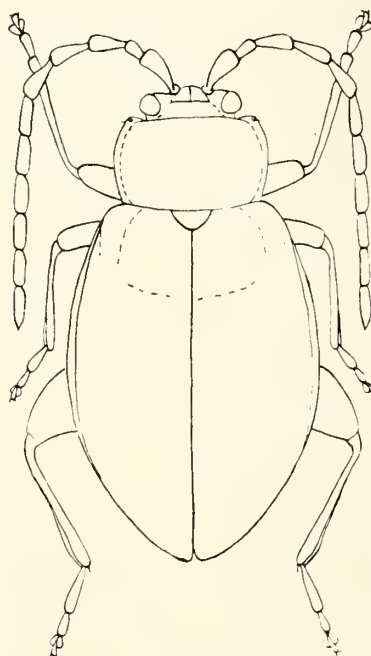


Fig. 37. Habitus of *Ocnoselis cyanoptera* Er.
(Magnification $7 \times$)

- 22 (21) Antennae usually significantly shorter, extending at most to middle of elytra, antennal calli always transverse and never bounded posteriorly by any callosity:
- 23 (26) Anterior coxal cavities closed posteriorly:
- 24 (25) Pronotum moderately arched and relatively smooth; anterior edge somewhat more constricted than base:

Antennal calli lancet-shaped, clearly defined, bounded laterally by lateral frontal sutures which extend to posterior margin of eyes; with a relatively

wide fissure between antennal calli, fissure as wide as interantennal longitudinal carina between antennae; latter carina spreads out somewhat anteriorly, and transverse carina only weakly developed.

Antennae long; individual segments long, length at least 4 times diameter.

Elytra parallel, without distinct basal calli; punctation confused.

3.5–5.5 mm long beetles; head black, thorax yellow, elytra yellow with black markings.

Pseudogona Jacoby 1885

Genotype: *Pseudogona panamensis* Jacoby

2 species are known from Panama.

- 25 (24) Pronotum evenly convex and prominently punctated; sides rounded off at anterior angles and bent in toward anterior margin; with an indentation on lateral margins near posterior angles:

Antennal calli bounded posteriorly by a nearly horizontal but only slightly curved frontal suture, which extends to posterior margin of eyes; lateral frontal sutures absent so that antennal calli not defined laterally but extend to inner margin of eye; eyes very small; frons very wide, more than twice as wide as transverse diameter of eye; interantennal ridge tubercle-shaped between antennae, antennae reach only to base of elytra; outer 5–6 segments nearly round.

Elytra with 9 continuous rows of punctures (not counting marginal and juxtasutellar rows of punctures); basal calli entirely absent; humeral calli only very weakly indicated; wingless.

Small 2–2.3 mm long beetles, bronze-colored with reddish-brown legs and antennae.

Docemina Champion 1918

Monotypic: *Docemina crassipes* Champ. (Tierra del Fuego)

- 26 (23) Anterior coxal cavities open posteriorly:

- 27 (28) Antennal calli very distinctly margined posteriorly, posterior limiting furrow runs in a straight line to posterior margin of eye; a characteristic lateral frontal suture absent, with an indentation in its place; antennae robust, segments short, their diameter being more than half their length.

Beetles about 4 mm in size; head and pronotum black, a black transverse band on posterior half of elytra; elytra confusedly punctate.

Babiohaltica Bechyné 1955

Monotypic: *Babiohaltica corumbana* Bechyné (Brazil: Mato Grosso; Argentina: Buenos Aires).

- 28 (27) The posterior marginal sutures of the distinct antennal calli run obliquely to lateral frontal suture, latter extends to posterior margin of eye; antennal calli usually formed as thin ridges; a characteristic interantennal longitudinal carina absent, frons wide and dilated anteriorly, antennae slender, diameter of segments less than half the length.

Elytra confusedly punctate with a tendency toward longitudinal rows up to 9 continuous longitudinal rows.

Usually small 2–4 mm beetles, brown, but also metallic-colored:

Brasilaphthona Bechyné 1958

Genotype: *Aphthona verticalis* Baly

Ca. 45 species which are distributed over South and Central America.

- 29 (20) Pronotal sides more rounded on anterior half so that base appears constricted, base actually somewhat wider than anterior margin:
- 30 (31) Posterior limiting furrow of antennal calli confined only to antennal calli themselves, not extending to internal or posterior margins of eye; otherwise frontal sutures absent or very indistinct, antennal calli also laterally bounded. Beetles about 5 mm in size, yellowish-brown to reddish-brown colored; antennae tend to hypertrophy (Fig. 38):

Corynothona Bechyné 1956

= *Baialtica* Bechyné syn.

= *Bryanthaltica* Bechyné syn.

Genotype: *Systema antennata* Jacoby (Amazon, Pará)

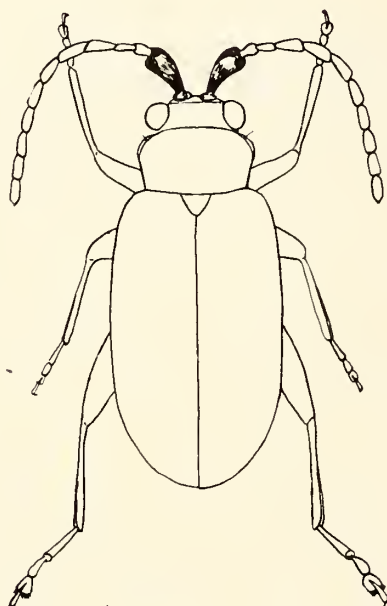


Fig. 38. Habitus of *Corynothona antennata* (Jac.) (♂)
(Magnification 7×)

According to the description (Ent. Arb. 7, 1956, p. 978), *Baialtica* is differentiated from *Corynothona* in having open anterior coxal cavities, and in the morphology of the antennae. However, *Baialtica* also has closed anterior coxal cavities, except that the epimers are very thin, so that they are scarcely visible. *Bryanthaltica* has no hypertrophied antennal segments, while in *Corynothona*, the first segment is hypertrophied, and in *Baialtica* other antennal segments are strongly developed.

- 31 (30) Antennal calli convex and well defined, bounded at sides by a definite lateral frontal suture:
- 32 (33) Antennal calli transverse, kidney-shaped, lateral frontal suture lies very close to inner margin of eye; posteriorly calli bounded by a deep furrow so that transition to vertex occurs in steps, separated from each other by a deep, but very narrow furrow; antennae long and extend to or somewhat beyond middle of elytra, or even to apices of elytra or beyond.
2–4.5 mm beetles, head and thorax usually yellowish-brown, elytra usually metallic in color or with metallic spots: **Lupraea** Jacoby 1885
(*Palaeothona* Jacoby, 1885)*
Genotype: *Lupraea longicornis* Jacoby
Ca. 29 species: Southern USA and Central America; 2 species in Amazon region.
- 33 (32) Antennal calli transverse, rectangular, deep lateral frontal suture not as close to internal margin of eye as in preceding genus, but nearly as far removed from inner margin of eye as width of antennal calli, bounded posteriorly by a fine, less deep furrow so that transition to vertex not as step-like; separated from each other by a wide, deep fissure.
Antennae long, extending to or somewhat beyond middle of elytra; antennae of males tend to have lateral extensions from the 3rd segment on and may even be dentate.
3–4 mm beetles, usually brown in color, elytra in part with black markings, also entirely black.
Sanariana Bechyné 1955
Genotype: *Sanariana rubra* Bechyné
5 species (1 Argentina; 2 Brazil; 1 Santa Catarina, 1 Rio de Janeiro; 2 Bolivia: of these, one with a race in Argentina).
- 34 (15) First tarsal segment of posterior leg as long as or longer than half the tibia. Antennal calli usually poorly defined. Anterior coxal cavities open posteriorly: 1–3.5 mm long, usually 2 mm long beetles, yellowish-brown, brown or black colored, a few species flightless without humeral calli:
Longitarsus Latreille *apud* Berthold 1827
Genotype: *Chrysomela atricilla* L. (Europe)
Ca. 50 Neotropic species, otherwise distributed over the entire earth.
- 35 (12) Consistently round-oval forms of *Chrysomela* or coccinellid-like habitus (Figs. 39–40):
- 36 (43) Very convex and *Chrysomela*-like habitus:
- 37 (40) Antennal calli touching each other; anterior coxal cavities open posteriorly: Anterior angles of pronotum acute and pointed forward; yellowish- to red-

* Bechyné, Pesquisas 1960 Zoologia, No. 6, p. 23.

dish brown, some with black markings; fresh material from one species showing green color; 6.6–15 mm long.

- 38 (39) Frons in front of antennae very short, characteristically consisting only of a transverse carina; larger beetles, 9–15 mm long, more roundish and very convex (Fig. 39): a (b) Claws single:

Crimissa Stal 1858

(*Strongylotarsa* Dej. nom. nud.)

Genotype: *Crimissa cruralis* Stal

5 species: distributed from southern Brazil, Paraguay to Panama.

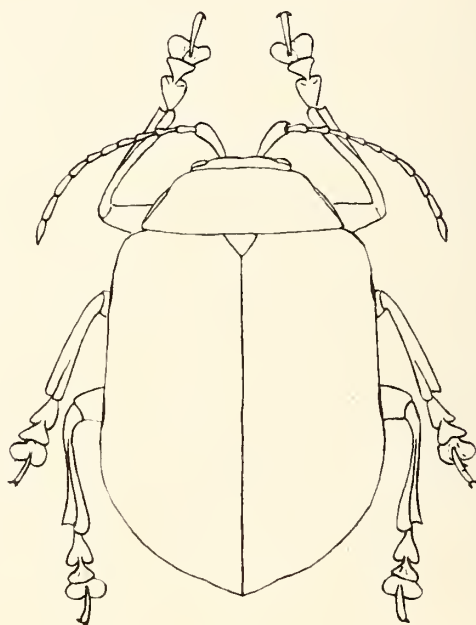


Fig. 39. Habitus of *Crimissa cruralis* Stal
(Magnification 3×)

- b (a) Claws split:

Elithia Chap. 1875

Genotype: *Elithia grossa* Chap.

5 species: 1 Brazil, 1 Chile, 1 Ecuador, 1 Cayenne, Peru, and Panama.

- 38 (39) Frons in front of antennae triangular, sides of this triangle ridgelike with frontal margin as a broad base; not so long, 6.6–10 mm, not so rounded and not as convex: Claws split.

Procalus Clark 1865*

Genotype: *Coelomera mutans* Blanchard

2 species: 1 Chile, 1 Chile, Bolivia, Brazil, Argentina

* **Procalus** Clark 1865, Ann. Mag. N. H. (3) 16: 257, 260 (Galerucinae) Laboissière 1932, Mem. Mus. Hist. Nat. Belgique, horst. ser., 4 (4): 147 (Alticinae)
Willcox 1971, Col. Cat. Suppl. (Ed. Sec.) 78: 695

- 40 (37) Antennal calli widely separated; frons in front of antennae poorly differentiated from remaining frons; anterior coxal cavities closed posteriorly:
Pronotal anterior angles somewhat pointed; sides rounded; slight impressions, such as in the *Blepharida*, can sometimes be recognized on both sides of base.

Elytral base no wider than pronotal base; 9 rows of punctures, but also with very narrow rows, punctures sometimes faded.

Frons wide in front of antennae.

- 41 (42) A transverse impression on anterior margin on frons in front of antenna so that anterior margin appears swollen:

Anterior tibiae characteristic of genus but have undergone variation so that genus also is mentioned in group 3. Posterior tibiae usually with a ciliated emargination near apex.

Yellowish-brown with reddish-brown to pitch brown markings; 6–10 mm long; related to *Blepharida*:

Notozona Clark 1865

Genotype: *Chrysomela bifasciata* Olivier (Cayenne)

22 species: 9 Central America: 5 Mexico, 2 Guatemala, 1 Nicaragua, 1 Honduras; 1 Venezuela; 3 Cayenne; 1 Peru (?); 4 Brazil without further data; 1 Amazon; 1 Bahia, 1 Pará; 1 S. Paulo, E. Santo, and Minas Gerais.

- 42 (41) No transverse impression on anterior margin of frons in front of antennae; posterior tibia not emarginate; yellowish-brown with pitch brown spots; 5–8 mm long; with exception of tibiae, very reminiscent of *Blepharida*:

Acrocyum Jacoby 1885

Genotype: *Acrocyum dorsalis* Jacoby

4 species: Central America

- 43 (36) Less convex, not **Chrysomela**-like (Fig. 40); coccinellid-like habitus, oval circumference, elytra moderately convex overall; no basal calli:

- a (b) Wide interantennal longitudinal carina which runs anteriorly on both sides to a thin transverse carina; remaining frons in front of antenna once again transverse-carina-like on either side behind transverse carina; antennal calli transverse, laterally distinct, or only poorly defined. Antennae short, apical segments all wider than long.

Sophaena Baly 1865

= *Axiotheata* Duponchel 1842 syn.

= *Cyrtosphaerus* Clark 1865 syn.

Genotype: *Sophaena ornata* Baly (Amazon)

6 species: 3 Amazon, 2 Cayenne, 1 Peru.

Duponchel (Dict. Univ. II, 1842, p. 390) indeed gave a short description, but there are no valid species, Clark published his genus *Cyrtosphaerus* 8 months later.

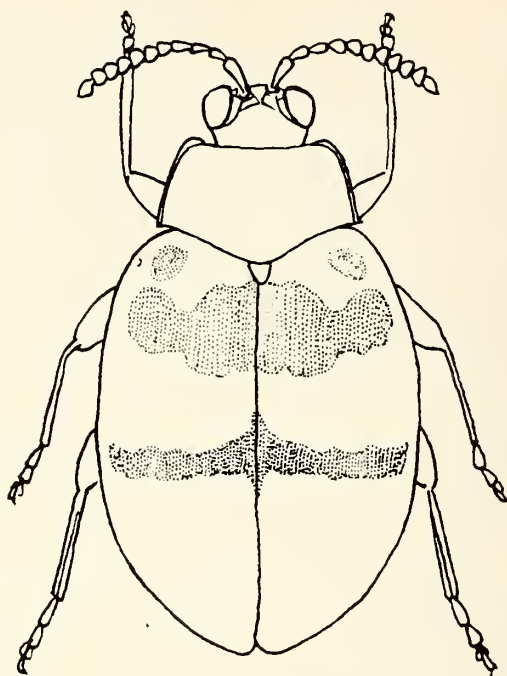


Fig. 40. Habitus of *Sophraena peruviana* (Har.)
(Magnification 8×)

- b (a) Wide interantennal longitudinal carina, bounded anteriorly by an equally wide transverse carina; antennal calli transverse, laterally bounded by a very deep lateral frontal suture with prominent punctures. Antennae extend to first half of elytra, all segments strikingly longer than wide. 4–6 mm long and usually yellowish-brown with dark spots.

Chrysogramma Jacoby 1885

Genotype: *Chrysogramma octomaculata* Jacoby (Guatemala)

5 species: 1 Guatemala, 4 Mexico.

- 44 (11) Pronotal anterior angles definitely beveled (Fig. 41 and 42):
- 45 (48) Elytra confusedly punctate:
Broadly ovate, relatively strongly convex with wide prothorax, definite humeral and basal calli on elytra.
- 46 (47) Pronotum not punctate or very finely punctate; frontal sutures extend in large arches to posterior margins of eyes and do not touch, but rather run parallel to them however at a very close interval; these arches severely constrict vertex, with setiferous punctures located beneath them; length 2–4 mm (Fig. 41):

Homoschema Blake 1950

Genotype: *Homoschema ornatum* Blake

Ca. 18 Central American species.

- 47 (46) Pronotum coarsely confusedly punctured; frontal sutures end at inner margin of eyes where posterior margin begins, setiferous punctures above this point; length 2–3 mm.

Gioia Bechyné 1955

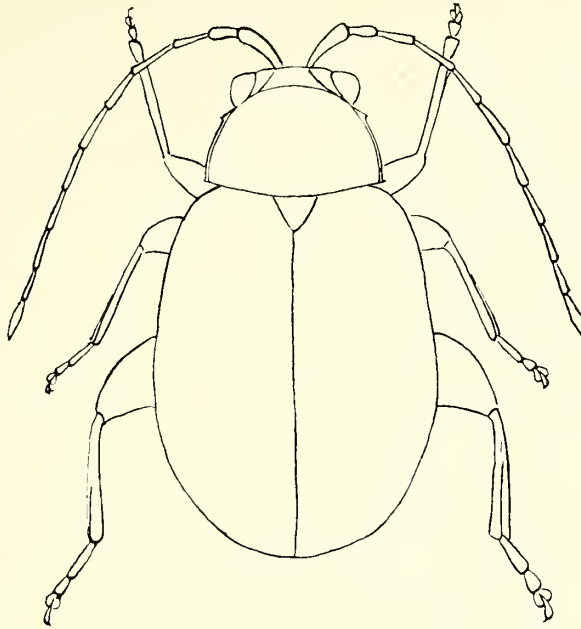


Fig. 41. Habitus of *Homoschema jamaicense* Blake
(Magnification 20×)

Genotype: *Gioia wittmeri* Bechyné

8 species: 2 Rio de Janeiro, 3 Venezuela, 3 Guadeloupe.

This genus very much resembles the African genus *Monodaltica* Bech. from which it is difficult to separate except for the fact that the apical tooth of the posterior tibiae is bifurcated in *Monodaltica*.

- 48 (45) Elytral punctures in 9 continuous longitudinal rows – not counting marginal and short juxtascutellar punctation:

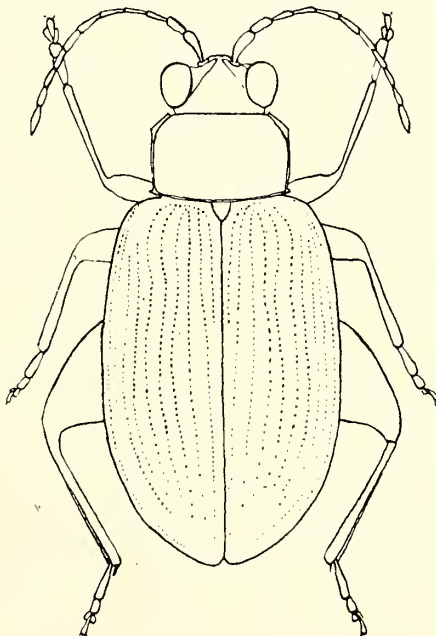


Fig. 42. Habitus of *Cyrsylus recticollis* Jac.
(Magnification 15×)

- 49 (50) Anterior coxal cavities open posteriorly; elytral base somewhat wider than pronotal base; basal calli on elytra, if present, very modestly developed; antennal calli distinct, lateral frontal sutures rather distant from internal margin of eyes and extend to posterior margin of eyes; claws single:

1.5–2 mm long forms, many times brown, but also metallic-colored.

Centralaphthona Bechyné 1960

Genotype: *Aphthona deyrollei* Baly

25 Neotropic species: 21 species are scattered over the region from Colombia to Mexico and the Islands of the Caribbean area: 1 Paraguay, 1 Bolivia, 1 Venezuela, 1 Brazil: Rio Grande do Norte.

- 50 (49) Anterior coxal cavities closed posteriorly; elytral base definitely wider than pronotal base; pronotum gives impression that sides of base diverge anteriorly; basal calli on elytra very definitely pronounced; antennal calli distinctly bounded by frontal and lateral sutures; lateral frontal sutures very close to inner margin of eyes and extend to posterior margin of eyes. Claws split.

2.5–3.5 mm long forms, brown to black in color, but also metallic colored (Fig. 42):

Cyrsylus Jacoby 1891

Genotype: *Cyrsylus recticollis* Jacoby

10 species: 9 Central America, 1 Brazil (S. Paulo and Pará) and Bolivia.

Group 7

- 1 (2) Antennae with 10 segments:

Body elliptical; antennae widely separated; pronotum without indentations; elytra with regular rows of punctures; posterior tibiae recessed as a groove on posterior part on upper side, margin of excavation often furnished with teeth; posterior tarsi not attached on apex, but on dorsal side of the tibia. A little before apex (sometimes at one half tibial length): first tarsal segment of posterior leg approximately half as long as tibia; anterior coxal cavities closed posteriorly. (Compare with Fig. 1 in key to alticine genera of Africa:

Scherer, Ent. Arb. Mus. Frey, 12, 1961, p. 254.)

Psylliodes Latreille apud Berthold 1827

Genotype: *Chrysomela chrysocephala* L. (Europe)

13 Nearctic species which extend in part into the Neotropic region (2 to Guatemala, 4 Mexico): 82 Palearctic species; 2 African species; 42 Indo-Australian species.

- 2 (1) Antennae 9-segmented; shape oval:

Nonarthra Baly 1862

Genotype: *Nonarthra variabilis* Baly (from NE Afghanistan till Taiwan). An oriental genus from which *N. cyaneum* Baly was discovered in Panama (Balsbaugh 1981, The Pan-Pacific Entomologist no. 814: 411–414).

References

- Bechyné, J. 1951: Chrysomeloidea Americains nouveaux ou peu connus (Coleoptera). – Rev. Chil. Ent., 1: 75–112.
- — 1955: Reise des Herrn Georg Frey in Südamerika: Alticidae (Col. Phytophaga). – Ent. Arb. Mus. Frey, 6: 74–266.
- — 1956: Beiträge zur Kenntnis der neotropischen Alticiden und Galeruciden. – Ent. Arb. Mus. Frey, 7: 965–1071.
- — 1956: Über die Alticiden-Sammlung Heikertinger (Col. Phytophaga). – Ent. Arb. Mus. Frey 7 (2): 577–598.
- — 1957: Alticides neotropicaux de la Collection du Museo Civico di Storia Naturale «Giacomo Doria» di Genova (Coleopt. Chrysomelidae). – Ann. Mus. Civ. Stor. Nat. Genova, 69: 51–74.
- — 1957: Voyage de M. le Dr. A. Roman au Brésil (1914–1915). Eumolpides, Galerucides et Alticides (Col. Phytophaga). – Ark. Zool. II: 133–152.
- — 1958: Notizen zu den neotropischen Chrysomeloidea (Col. Phytophaga). – Ent. Arb. Mus. Frey, 9: 478–706.
- — 1959: Beiträge zur Kenntnis der Alticidenfauna Boliviens. Coleopt. Phytoph. – Beitr. z. neotrop. Fauna 1: 269–388, Jena 1959.
- — 1963: Notes sur quelques Chrysomeloidea néotropicaux nouveaux ou peu connus (Col. Phytophaga). – Bull. mens. Soc. linn. Lyon 32 (8): 235–239.
- — 1968: Contribution à la faune du Congo (Brazaville). Mission A. Villiers et A. Descarpentries. LXXXI. Coléoptères Alticidae. – Bull. Inst. fr. Afr. noire Sér. A, 30 (4): 1687–1728; 6 figs.
- — and B. Springlova de Bechyné, 1960: Beiträge zur Kenntnis der salvadorenischen Chrysomeloidea (Col. Phytoph.). – Pesquisas Zoologia 6: 5–73.
- — 1961: Notas sobre Chrysomeloidea Neotropicais. – Bol. Mus. Paraense Emilio Goeldi, 33: 1–50.
- — 1961: Notas sobre Chrysomeloidea neotropicais II. – Bolm. Mus. Paraense Emilio Goeldi Zool. no. 37: 1–93; 9 figs.
- — 1963: Beitrag zur Kenntnis der Salvadorenischen Chrysomeloidea (Col. Phytophaga) 1. Fortsetzung. – Iheringia Zool. no. 31: 1–79.
- — 1964: Notes sur quelques Chrysomeloidea néotropicaux (Coleoptera Phytophaga). – Revta Fac. Agron. Univ. cent. Venez. 3 (3): 69–123; 27 figs.
- — 1965: Notes sur les Chrysomeloidea capturés par le Dr. W. A. Egler au Rio Jari (Brésil: Pará/Amapá) en 1961 (Col. Phytophaga). – Bol. Mus. Paraense Emilio Goeldi Zool. no. 53: 1–44; 9 figs.
- — 1966: Evidenz der bisher bekannten Phenrica-Arten (Col. Phytophaga, Alticidae). – Ent. Tidskr. 87 (3–4): 142–170; 16 figs.
- — 1973: Notas sobre algunos Phytophaga de origen Paleantartico (Coleoptera). – Revta chil. Ent. 7: 25–30; 4 figs.
- — 1975: Notas sobre la serie filetica de Monomacra y sus formas convergentes (Col. Phytophaga, Alticidae). – Rev. Fac. Agron. (Maracay) 8 (4): 25–140.
- — 1976: Phytophages (Coléoptères) récoltés en Guyane Française par la Mission du Muséum National d'Histoire Naturelle (1). – Ann. Soc. ent. Fr. (N. S.) 12 (4): 527–556.
- — 1977: Zur Phylogenie einiger neotropischer Alticiden (Col. Phytophaga). – Studies on Neotropical Fauna and Environment 12: 81–145.
- Blake, D. H. 1942–1943: New Species of the Genus Hadropoda Suffrian from the West Indies. – Bull. Mus. comp. Zool. Harv. College, Cambridge, 92: 413–441.
- — 1943: The generic position of Hypolampsis pilosa (Illiger) and some related new species (Coleoptera: Halticidae). – Proc. Ent. Soc. Wash., 45: 207–221.
- — 1947: A new genus of flea beetles from the West Indies. – J. Wash. Acad. Sci., 37: 92–95.
- — 1950: A new genus of flea-beetles from the West Indies. – Psyche 57: 10–25.

- — 1951: Synonymies and new species of Flea-Beetles (Coleoptera, Chrysomelidae). – Proc. Ent. Soc. Wash., 53: 138–146.
- — 1952: Six news Species of Megistops with keys to the known species (Coleoptera). – Psyche, 59: 1–12.
- — 1953: Eight new Neotropical chrysomelid beetles (Coleoptera). – J. Wash. Acad. Sci., 43: 232–237.
- — 1953: The Chrysomelid Beetles of the Genus Strabala Chevrolat. – Proc. U.S. Nat. Mus., 103: 121–134.
- — 1955: Revision of the vittate Species of the Chrysomelid Beetle Genus Disonycha from the Americas South of the United States. – Proc. U.S. Nat. Mus., 104: 1–86.
- Clark, H. 1860: Catalogue of Halticidae in the Collection of the British Museum. – London.
- — 1865: An Examination of the Halticidae of South America. – J. Ent., 2: 375–412.
- Harold, E. v. 1875: Beiträge zur Kenntnis der Halticae oedipodes. – Col. Hefte 13: 1–26.
- Heikertinger, F. 1924: Die Halticinengenera der Palearktis und Nearktis. – Koleopt. Rdsch., 11: 25–70.
- Horn, G. H. 1889: A synopsis of the Halticinae of Boreal America. – Trans. Amer. Ent. Soc., 16: 163–320.
- Jacoby, M. 1880–1892: Biologia Centrali-Americana. Insecta, Coleoptera, 6, Phytophaga.
- Lacordaire, Th. & Chapuis, F. 1875: Genera des Coléoptères. – Paris.
- Scherer, G. 1960: Beitrag zur Kenntnis der Alticidenfauna Brasiliens (Col. Phytoph.). – Ent. Arb. Mus. Frey, 11: 180–272.
- — 1961: Bestimmungsschlüssel der Alticiden-Genera Afrikas (Col. Phytoph.). – Ent. Arb. Mus. Frey, 12: 251–288.
- — 1967: Alticinae aus Guadeloupe (Col. Chrysomelidae). – Ann. Soc. ent. Fr. (N. S.), 3 (1): 215–220.

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Author's Address:

Dr. Gerhard Scherer

Zoologische Staatssammlung

Maria-Ward-Straße 1b

D-8000 München 19

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Zoologisch-Botanische Datenbank/Zoological-Botanical Database

Digitale Literatur/Digital Literature

Zeitschrift/Journal: [Entomologische Arbeiten Museum G. Frey](#)

Jahr/Year: 1983

Band/Volume: [31-32](#)

Autor(en)/Author(s): Scherer Gerhard

Artikel/Article: [Diagnostic Key for the Neotropical Alticine Genera \(Coleoptera: Chrysomelidae: Alticinae\). 1-89](#)