

On the Staphylinidae of the Greek island Lesbos II, with supplementary notes on the fauna of Samos and Chios (Coleoptera: Staphylinidae)

V. ASSING

Abstract

A field trip to the Greek island Lesbos conducted in spring 2016 yielded a total of 1640 specimens belonging to 169 species of Staphylinidae. Nine new species are described and illustrated: *Myllaena lesbia* ASSING sp.n., *Aloconota (Aloconota) aegaea* ASSING sp.n. (also recorded from Samos), and *A. (A.) lesbia* ASSING sp.n. of the Aleocharinae, *Sunius potti* ASSING sp.n. of the Paederinae, *Bryaxis lesbius* BRACHAT sp.n., *Bythinus simplicipalpis* BRACHAT sp.n., *Protamaurops assingi* BRACHAT sp.n., and *Tychus lesbius* BRACHAT sp.n. of the Pselaphinae, and *Stenichnus (Stenichnus) lesbius* MEYBOHM sp.n. of the Scydmaeninae. The new species of Paederinae, Pselaphinae, and Scydmaeninae are most likely endemic to Lesbos. Evidence suggests that *Quedius henroti* COIFFAIT, 1970, previously considered an endemic of Lesbos, is in fact distributed in the Pelopónnisos rather than in Lesbos. A comprehensive list of the named and unnamed species currently known from Lesbos is provided. Numerous described species are reported from the island for the first time, eight of them represent first records from Greece. The known staphylinid fauna of Lesbos currently includes 199 species, 184 named and 15 unnamed, unidentified, or tentatively identified. Nine (5 %) of the named species and at least two of the unnamed Pselaphinae and Scydmaeninae are hypothesized to be endemic to the island. Thus, despite greater overall diversity and a generally similar composition, the fauna of Lesbos includes fewer island endemics than that of the geographically close and significantly smaller island Samos. As was to be expected, the fauna of Lesbos displays close affiliations with that of the adjacent Turkish mainland. Several species reach their westernmost distribution limit in this island (some of them also in Chios and Samos). As many as six myrmecophilous species have been recorded from Lesbos, five of them associated with *Messor* spp. and one with *Cataglyphis nodus* (BRULLÉ, 1833). In an appendix, *Ocalea brachyptera* FAGEL, 1971 is reported from Chios, the first record of this species from Greece. *Aloconota (Aloconota) samia* ASSING sp.n. (Samos) of the Aleocharinae and *Stenichnus (Stenichnus) chius* MEYBOHM sp.n. (Chios) of the Scydmaeninae are described and illustrated.

Key words: Coleoptera, Staphylinidae, Palaearctic Region, East Mediterranean, Greece, Aegean Islands, Lesbos, Samos, Chios, taxonomy, new species, diversity, endemism, myrmecophily, new records, species list.

Introduction

With an area of 1636 km², Lesbos is the largest of the North Aegean islands. It is separated from western Anatolia (Çanakkale, Balıkesir, and Izmir provinces) by a distance of only 9 km in the north and 15 km in the east. The closest major North Aegean islands are Chios (48 km to the south) and Limnos (74 km to the northwest); Samos is some 120 km farther south. Like Chios and Samos, Lesbos has been separated from the Anatolian mainland only since the end of the last glacial period (TRIANTIS & MYLONAS 2009). The major mountain ranges are Lepetimnos (highest peak at 968 m) in the north, Olympos (967 m) in the south, and Ordimnos (Profitis Ilias at 799 m) in the west. Lesbos has a remarkable diversity of natural and semi-natural habitats, with extensive forests (oak, pine, chestnut), phrygana and other shrubland, rivers, streams, extensive lagunes, and salines.

According to SCHÜLKE & SMETANA (2015), the staphylinid fauna of Greece is currently composed of 3160 species and subspecies. The Staphylinidae (exclusive of the Pselaphinae and

Scydmaeninae) of Lesbos were already studied little more than a decade ago (ASSING 2005b), based on 334 specimens collected during a field trip conducted by Arved Lompe (Nienburg) and Heinrich Meybohm (Großhansdorf). This material was composed of more than 70 species, among them two species, *Rugilus lesbius* ASSING, 2005 and *Oxypoda lesbia* ASSING, 2005, believed to be endemic at that time. However, in the meantime the former has been recorded also from Samos and West Turkey, and the latter has been reported from West Turkey, the Greek mainland, Crete, Rhodos, and Chios.

Aside from the species reported by ASSING (2005b), there are only scattered literature records of Staphylinidae from Lesbos. Three island-endemic species of *Tychus* LEACH, 1817 (Pselaphinae) were described in BESUCHET & SABELLA (2012) and SABELLA et al. (2012). COIFFAIT (1970) reported six species. However, two of these records are doubtful because they may be based on misidentifications (see Tab. 1). Another one, *Quedius henroti* COIFFAIT, 1970, which was newly described in that paper and which is still listed as endemic to Lesbos in SCHÜLKE & SMETANA (2015), is of uncertain status for other reasons. According to the original description, this species is allied to *Quedius semiaeneus* (STEPHENS, 1833) and *Q. nitipennis* (STEPHENS, 1833), and the unique male holotype is 5 mm long and labelled “Mt Olimbos, 1200 m, Ile des Lesbos (Grèce), [...] Cerruti et Henrot leg.” (COIFFAIT 1970). First, a species matching the description has never been collected again in Lesbos, despite extensive search. Second, a species allied to *Quedius semiaeneus* and *Q. nitipennis* is highly unlikely to be endemic to Lesbos. Finally, the altitude of Oros Olympos in Lesbos is only 967 m and no mountain in the whole island even comes close to an altitude of 1200 m (see above). In consequence, it can be concluded that the holotype was probably collected elsewhere; aside from Lesbos, COIFFAIT (1970) reports Staphylinidae from Cyprus, Corfu, and the Pelopónnisos. In fact, there is a male from the Pelopónnisos (Messinia, Oros Kiparissias, 440 m, 8.VI.2007, leg. Giachino & Vailati) in my collection, which matches the original description. These observations suggest that *Q. henroti* is distributed in the Pelopónnisos and absent from Lesbos. According to the respective curators in charge, the holotype is neither in the Coiffait collection (Paris) nor in the Cerruti collection (Rome) (Taghavian, e-mail 17 May, 2016; Piattella, e-mail 20 May, 2016). The Henrot collection was looked for, but not found by the curators of the Muséum National d’Histoire Naturelle in Paris, although it is definitely deposited there (Taghavian, e-mail 27 May, 2016).

In order to further examine the Staphylinidae of Lesbos, a field trip was conducted in March, 2016, together with Andreas Hetzel (Hildesheim), a specialist of Carabidae. The present paper continues an ongoing project aiming at an exploration of the Staphylinidae of the East Mediterranean islands. Aside from the preliminary study of Lesbos (ASSING 2005b), the faunas of Cyprus, Crete, Rhodos, Samos, Chios, and Karpathos were examined earlier (ASSING 2013a–b, 2015a–c, 2016a, ASSING & WUNDERLE 2001). For overviews of species numbers, numbers of endemic species, and other island-related aspects see these articles.

Volker Brachat (Geretsried) and Heinrich Meybohm (Großhansdorf) provided additional records of Pselaphinae and Scydmaeninae, respectively, collected during a field trip to Lesbos in 2005.

The descriptions of the Pselaphinae and Scydmaeninae are authored exclusively by Volker Brachat and Heinrich Meybohm, respectively.

Material and methods

The material treated in this study is deposited in the following collections:

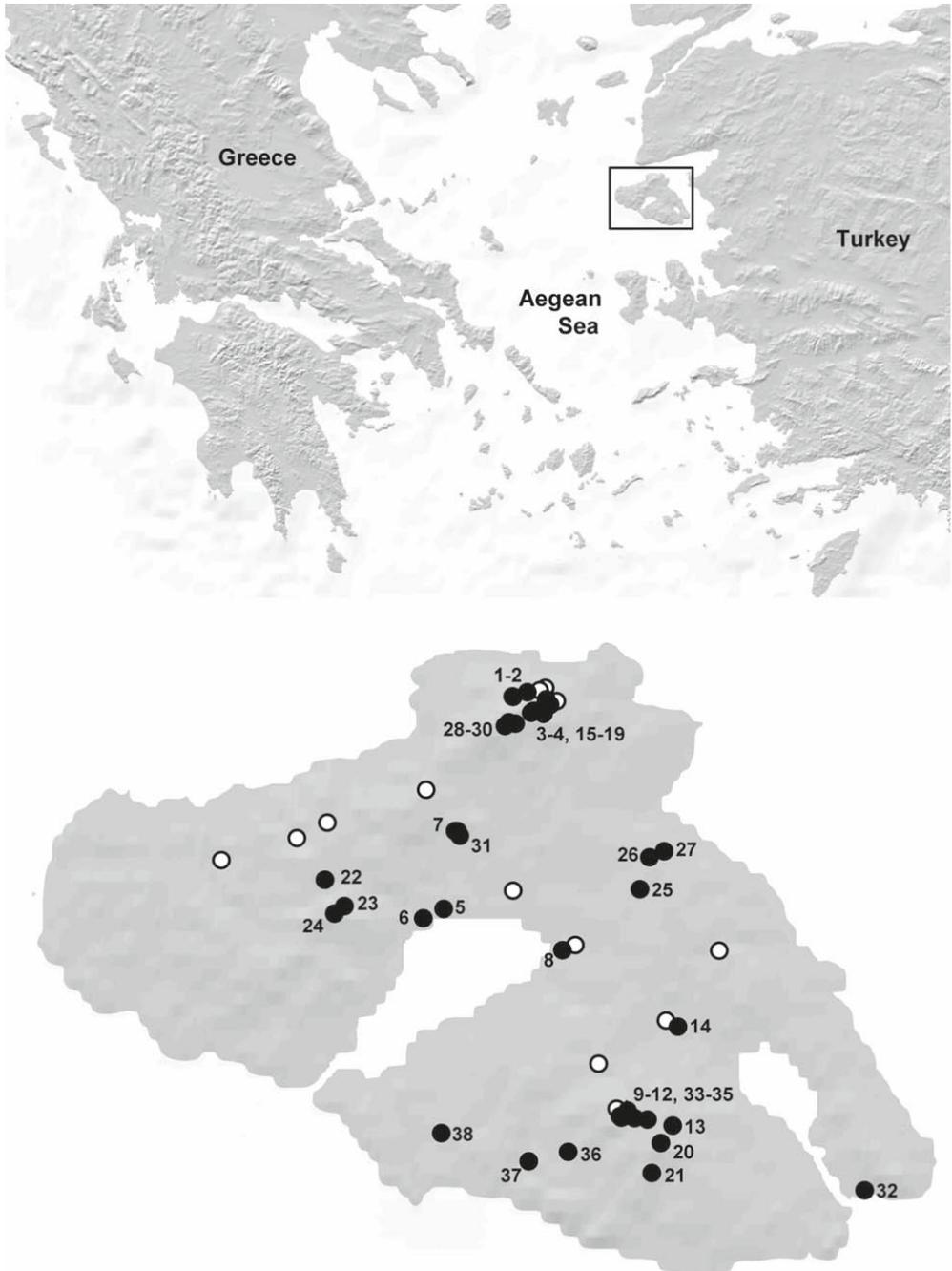


Fig. 1: Geographic position of Lesbos (above) and sample plots of the 2005 (white circles) and 2016 (black circles) field trips (below). Plots where no Staphylinidae were found and sample numbers of the 2005 sample plots are omitted.

cAss	author's private collection
cBra	private collection Volker Brachat, Geretsried
cMey	private collection Heinrich Meybohm, Großhansdorf
MNB	Museum für Naturkunde, Berlin (J. Frisch, J. Willers)

The Pselaphinae and Scydmaeninae are all deposited in cBra and cMey, respectively. Reference material of the remaining species is deposited in MNB and cAss.

The morphological studies were conducted using a Stemi SV 11 microscope (Zeiss Germany) and a Jenalab compound microscope (Carl Zeiss Jena). The images of the Aleocharinae and Paederinae were created using a digital camera (Nikon Coolpix 995) and a photographing device constructed by Arved Lompe (Nienburg) and CombineZ software. The remaining photographs were provided by Heinrich Meybohm. The maps were created using MapCreator 2.0 (primap) software.

Body length was measured from the anterior margin of the mandibles (in resting position) to the abdominal apex, the length of the forebody from the anterior margin of the mandibles (in resting position) to the posterior margin of the elytra, head length along the middle from the anterior margin of the clypeus (without ante-clypeus) (Aleocharinae) or from the anterior margin of the frons (Paederinae, Pselaphinae) to the posterior constriction of the head, elytral length at the suture from the apex of the scutellum to the posterior margin of the elytra, and the length of (the median lobe of) the aedeagus from the apex of the ventral process to the base of the aedeagal capsule, except for the Pselaphinae for which the total length of the aedeagus is indicated. The "parameral" side (i.e., the side where the sperm duct enters) is referred to as the ventral, the opposite side as the dorsal aspect.

Results

General results

During the field trip in 2016 various habitats across most of the island were sampled (Fig. 1). In all, 1640 specimens belonging to 169 species were collected (Tab. 1). Including the Pselaphinae and Scydmaeninae collected in 2005 and communicated to me by Volker Brachat and Heinrich Meybohm, respectively, as well as previous literature records (except for three doubtful records), the currently known staphylinid fauna of Lesbos is composed of 199 species, 175 of them previously described and positively identified, nine newly described (three species of the Aleocharinae, one of the Paederinae, four of the Pselaphinae, and one of the Scydmaeninae), and 15 undescribed, unidentified, or tentatively identified (Tab. 1). Thus, 85 % of all the species currently known from Lesbos were collected during the field trip in 2016 and the number of species recorded from Lesbos has more than doubled. The unidentified species are either represented exclusively by females or belong to unrevised species groups (e.g., *Hydrosmecta* spp.). The undescribed species belong to the Aleocharinae, Pselaphinae and Scydmaeninae. Eight of the positively identified species are reported from Greece for the first time.

Compared to other islands examined earlier, Lesbos hosts a remarkable diversity. The number of species currently known from the island clearly exceeds that of Samos and Rhodos, which can be explained by the geological history (previous connection to the Anatolian mainland) of Lesbos, its vicinity to the Turkish coast, its relatively large size, and particularly its habitat diversity. On the other hand, the number of island endemics is relatively low. In all, Lesbos hosts at least eleven endemic species (nine named and at least two unnamed), one of the Paederinae, seven of the Pselaphinae, and three of the Scydmaeninae (one described and two unnamed). The status of two additional, newly described species (*Myllaena lesbia*, *Aloconota lesbia*) is uncertain; they are currently known only from Lesbos, but they are fully winged and ripicolous, suggesting that

they may be more widespread. Similarly, the status of the probably undescribed species of *Zoosetha* MULSANT & REY, 1874 is unclear. In many respects, the fauna of Lesbos resembles that of Samos, an island characterized by a similar habitat diversity, but distinctly smaller, situated farther to the south, and featuring two significantly higher mountain ranges. The two latter differences may explain why the diversity of island endemics is greater in Samos (18 species) than in Lesbos, despite lower overall diversity (Samos: approximately 140 species).

The non-endemic fauna is composed of species that are widespread in the East Mediterranean, in Southeast Europe, Turkey, or in large parts of the West Palaearctic Region, as well as of species that have more restricted distributions. Several species are confined to the Aegean region; they are known from some North Aegean islands and West Anatolia, the distributions of some of them extending northwards to Bulgaria, e.g., *Faronus distinctus*, *Anaulacaspis nigrina*, *Dinusa smyrnensis*, *Leptusa samia*, *Myrmecopora convexula* (a myrmecophile associated with *Messor* sp.), *Notothecta pisidica*, *Aploderus lydicus*, *Medon lydicus*, and *Rugilus lesbius*. Some species are more or less widespread across Anatolia and partly the Middle East or Middle Asia, and have their western distribution limit in Lesbos (some of them also recorded from Chios and/or Samos), e.g. *Metopsia assingi*, *Phloeocharis longipennis*, *Cordalia anatolica*, *Cypha tenebricosa*, *Pella cinctipennis*, *Pseudomedon dido*, *Quedius fissus*, and *Q. job*. *Tribatus creticus* is an example of an East Mediterranean species reaching its northern distribution limit in Lesbos.

The diversity of myrmecophilous Staphylinidae is remarkably rich in Lesbos. Five of the six species are associated with *Messor* spp.: *Dinusa smyrnensis*, *Myrmecopora convexula*, *Notothecta pisidica*, *Stenus hospes* (apparently a facultative myrmecophile; see discussion in ASSING (2015c)), and *Tribatus creticus*. *Piochardia reitteri* is associated with *Cataglyphis nodus* (BRULLÉ, 1833). Despite thorough search of numerous *Tetramorium* nests, however, no myrmecophilous species of *Astenus* DEJEAN, 1833 was recorded. *Astenus procerus* was found with a species of *Tetramorium* MAYR, 1855, but does not represent an obligatory myrmecophile.

Gaps in the staphylinid fauna are not as evident as was observed, for instance, for the much more isolated island Karpathos (ASSING 2016a). On the other hand, the absence of some species is remarkable. An example is *Xantholinus chiosicus* ASSING, 2015, a species recorded from Chios, Samos, and Karpathos, but apparently missing in Lesbos. Also, some common species of the Aegean region (e.g., *Omalium rugatum*, *Othius lapidicola*) were strikingly rare in Lesbos.

Tab. 1: Staphylinidae recorded from Lesbos. The material collected in March 2016 and the Pselaphinae collected by Arved Lompe and Heinrich Meybohm in spring 2005 are listed in the samples column; the number of specimens is given in parentheses behind the sample number. In the previous records column, only articles containing primary records are listed, except for one species of Pselaphinae which is listed for Lesbos in the Palaearctic Catalogue (SCHÜLKE & SMETANA 2015), although primary records with specified localities were never published. The records of three species by COIFFAIT (1970) are considered highly doubtful and omitted from the list (see doubtful records below). The species are listed by subfamily and sorted alphabetically within subfamilies. Species that have been recorded exclusively from Lesbos and, based on available evidence, are most likely endemic to the island are marked with an asterisk.

Sample data 2016 (all leg. Assing & Hetzel): **1**: 7 km ESE Mithimna, W Argennos, 39°21'23"N, 26°15'18"E, 440 m, stream valley, litter of *Quercus ilex* and *Platanus* sifted, 18.III.; **1a**: same data, but pasture, under stones and in sheep dung; **1b**: same data, but floated from gravel; **1c**: same data as sample 1, but 21.III.; **2**: 7 km ESE Mithimna, 39°21'33"N, 26°16'05"E, 390 m, stream valley, debris near stream sifted, 18.III.; **3**: Oros Lepetimnos, 39°20'43"N, 26°16'16"E, 870 m, north slope near large rocks, litter of small trees, fern, grass, and herb roots sifted, 18.III.; **3a**: same data, but north slope with sparse grass and herb vegetation, under stones; **3b**: same data as sample 3, but 24.III.; **4**: Oros Lepetimnos, 39°20'45"N, 26°16'56"E, 680 m, *Quercus ilex* forest, litter and herb roots sifted, 18.III.; **5**: SW Kalloni, 39°12'41"N, 26°11'42"E, 5 m, laguna, flood debris sifted, 19.III.; **5a**: same data, but 25.III.; **6**: SW Kalloni, 39°12'18"N, 26°10'39"E, 2 m, river bank, flood debris sifted and collected from soil (gravel), 19.III.; **6a**: same data, but 25.III.; **7**: N Kalloni, 39°15'52"N, 26°12'18"E, 220 m, pasture, under stones, 19.III.; **7a**: same data,

but 23.III.; **7b**: same data, but 24.III.; **7c**: same data, but 26.III.; **7d**: same data, but 27.III.; **7e**: same data, but 29.III.; **8**: 10 km SE Kalloni, 39°10'59"N, 26°17'54"E, 2 m, marshland, 19.III.; **9**: Olympos, 39°04'23"N, 26°21'15"E, 950 m, peak region, under stones, 20.III.; **10**: Olympos, 39°04'25"N, 26°21'17"E, 930 m, N-slope near peak, small stand of *Quercus*, *Juniperus*, and other deciduous trees, litter and moss sifted; **11**: track Olympos–Agiasos, 39°04'06"N, 26°21'40"E, 710 m, margin of track, under stones, 20.III.; **12**: track Olympos–Agiasos, 39°04'03"N, 26°22'20"E, 680 m, chestnut forest, litter sifted, 20.III.; **13**: track Olympos–Agiasos, 39°03'48"N, 26°23'39"E, 630 m, margin of pasture, under stones, 20.III.; **13a**: same data, but pine forest with undergrowth, litter, moss, and pine bark sifted; **14**: 14 km WNW Mytilini, S Lambou Mili, 39°07'51"N, 26°23'55"E, 50 m, stream bank, hand collected and floated from gravel, 20.III.; **14a**: same data, but stream bank, 22.III.; **14b**: same data, but 28.III.; **15**: Oros Lepetimnos, 39°20'41"N, 26°16'55"E, 700 m, pasture, under stones, 21.III.; **16**: Oros Lepetimnos, 39°20'47"N, 26°16'24"E, 790 m, grassy slope, under stones, 21.III.; **17**: Oros Lepetimnos, 39°21'02"N, 26°17'16"E, 610 m, grassy margin of track, under stones, 21.III.; **18**: Oros Lepetimnos, 39°21'16"N, 26°17'05"E, 570 m, grassy margin of track, under stone, 21.III.; **19**: Oros Lepetimnos, 39°21'03"N, 26°17'02"E, 580 m, litter and debris near temporary stream sifted, 21.III.; **20**: 3.5 km SSE Agiasos, 39°03'05"N, 26°23'02"E, 850 m, stony pasture with deciduous trees, under stones, 22.III.; **20a**: same data, but N-slope with rocks, *Quercus ilex*, and other deciduous trees, litter and roots of herbs and grass sifted; **20b**: same data, but stony pasture with deciduous trees, roots of grass and herbs sifted and floated; **20c**: same data, but stony pasture with deciduous trees, under stones, 28.III.; **20d**: same data, but roots of grass and herbs sifted and floated; **21**: 6 km S Agiasos, 39°01'51"N, 26°22'33"E, 770 m, pine forest, pine litter sifted, 22.III.; **21a**: same data, but forest margin, mushrooms sifted; **22**: 10 km W Kalloni, track Anemotia to Profitis Ilias, 39°13'52"N, 26°05'30"E, 550 m, margin of track, under stones, 23.III.; **23**: 9 km W Kalloni, track Anemotia to Profitis Ilias, 39°12'48"N, 26°06'32"E, 580 m, pasture, under stones, 23.III.; **24**: 9.5 km W Kalloni, Profitis Ilias, 39°12'29"N, 26°06'01"E, 790 m, peak, under stones, 23.III.; **24a**: same data, but bushes with scattered juniper and fern undergrowth, litter sifted; **24b**: same data, but pine forest and old pine tree on peak, pine litter sifted; **25**: 14 km E Kalloni, 39°13'29"N, 26°21'57"E, 240 m, margin of track, under stones, 24.III.; **26**: 15 km ENE Kalloni, 39°14'47"N, 26°22'28"E, 190 m, margin of track and pasture, under stones, 24.III.; **27**: 16 km ENE Kalloni, 39°15'02"N, 26°23'14"E, 120 m, stream bank, flood debris sifted, 24.III.; **27a**: same data, but gravel floated; **27b**: same data as sample 27, but 29.III.; **28**: Oros Lepetimnos, Prof. Ilias, 39°20'10"N, 26°14'54"E, 900 m, N-slope, under stones, 25.III.; **29**: Oros Lepetimnos, track to Prof. Ilias, 39°20'20"N, 26°15'06"E, 800 m, pasture, under stones, 25.III.; **29a**: same data, but litter beneath oak shrubs sifted; **29b**: same data, but horse dung sifted, 29.III.; **29c**: same data, but in cow dung, 29.III.; **30**: Oros Lepetimnos, track to Prof. Ilias, 39°20'16"N, 26°15'27"E, 730 m, pasture, under stones, 25.III.; **30a**: same data, but large rocks with *Quercus ilex*, litter sifted; **31**: N Kalloni, 39°15'41"N, 26°12'33"E, 190 m, pasture, under stones, 26.III.; **32**: S Mytilini, 4.4 km SSE Loutra, 39°01'08"N, 26°33'38"E, 1 m, beach, seaweed sifted, 26.III.; **33**: Olympos, 39°04'12"N, 26°20'59"E, 790 m, stream valley, *Platanus* litter sifted, 27.III.; **34**: Olympos, 39°04'08"N, 26°20'58"E, 800–820 m, N-slope, litter of *Platanus* and *Quercus ilex* and roots of grass and herbs sifted, 27.III.; **35**: Olympos, 39°04'11"N, 26°20'54"E, 810 m, grassy road margin, under stones, 27.III.; **36**: SW Olympos, track Ampeliko–Stavros, 39°02'44"N, 26°18'12"E, 310 m, olive grove with grass, under stones, 27.III.; **37**: 6.5 km ENE Vatera, Stavros, Vourkos river, 39°02'21"N, 26°16'09"E, 50 m, river bank with gravel, 27.III.; **38**: 2.7 km SSE Polihnitos, 39°03'30"N, 26°11'35"E, stream bank, 50 m, 28.III.

Sample data 2005 (only Pselaphinae and Scydmaeninae; all leg. Lompe & Meybohm): **39**: Skotino, 39°15'52"N, 26°12'25"E, 200 m, pasture on S-slope, near creek, 17.III.; **40**: Oros Lepetimnos, 39°21'11"N, 26°17'37"E, 570 m, pasture with *Quercus ilex*, 18.III.; **41**: Oros Lepetimnos, 39°20'47", 26°16'24"E, 800 m, 18.III.; **42**: Olympos, 39°04'11"N, 26°20'58"E, 790 m, *Platanus* leaf litter near stream, 29.III.; **42a**: same data, but 24.III.; **43**: Pigi, 39°10'58"N, 26°26'05"E, 90 m, litter under shrubs and bushes near stream, 21.III.; **44**: Oros Lepetimnos, 39°21'38"N, 26°16'43"E, 300 m, 21.III.; **44a**: same data, but 23.III.; **45**: W Skalohori, 39°15'35"N, 26°04'03"E, 360 m, oak leaf litter, 22.III.; **46**: W Argenos, 39°21'22"N, 26°15'20"E, 470 m, edge of stream with *Platanus*, 23.III.; **47**: Olympos, 39°04'29"N, 26°20'43"E, 720 m, meadow with *Quercus ilex*, in leaf litter and under stones, 24.III.; **48**: Lafionas, 39°17'34"N, 26°10'48"E, 300 m, arable land, sifted, 17.III.; **49**: Lambou Mili, 39°08'06"N, 26°23'18"E, 110 m, sifted, 19.III.; **50**: E Skalohori, 39°16'13"N, 26°05'37"E, 310 m, meadow, under stones, 22.III.

Doubtful records not included in the list: *Coproporus colchicus* KRAATZ, 1858, *Lordithon trimaculatus* (FABRICIUS, 1792), and *Quedius henroti*, all recorded from Lesbos by COIFFAIT (1970).

Previous records: articles are abbreviated as follows: A05a = ASSING (2005a); A05b = ASSING (2005b); A07e = ASSING (2007e); A08b = ASSING (2008b); A10 = ASSING (2010); A13d = ASSING (2013d); A13e = ASSING (2013e); App = ASSING (present paper); AW08 = ASSING & WUNDERLE (2008); BS12 = BESUCHET & SABELLA (2012); C70 = COIFFAIT (1970); F10 = FRISCH (2010); H65 = HORION (1965); M14 = MAKRANCZY (2014); Pa06 = PAŠNIK (2006); PC15 = Palaearctic Catalogue (SCHÜLKE & SMETANA 2015); Pu08 = PUTHZ (2008); S12 = SCHÜLKE (2012); SBK12 = SABELLA et al. (2012).

Footnotes: 1) cited as *Mycetoporus bosnicus* LUZE, 1901 (misidentification); 2) similar to an undescribed species from the environs of Istanbul, possibly conspecific; 3) listed as *Astenus (Astenochnathus)* sp.; 4) tentatively assigned; this species has been recorded also from Samos; 5) listed as *Pseudomedon* sp.; 6) listed as *Sunius anatolicus* ASSING, 1995 (synonym); 7) listed as *Sunius* sp.n.; 8) listed as *Ocypus olens* (MÜLLER, 1764) (misidentification); 9) listed as *Scopaenus* sp.; 10) listed as *Geostiba euboica* PACE, 1990 (synonym); 11) identification tentative; species requiring revision; 12) listed as *Anotylus tetracarinus* (BLOCK, 1799) (misidentification); 13) listed as *Ocypus integer* ABEILLE DE PERRIN, 1900 (synonym); 14) cited as *Quedius tristis* (GRAVENHORST, 1802) (unavailable name); 15) listed as synonym of *Atheta fungi* (GRAVENHORST, 1806) in SCHÜLKE & SMETANA (2015); 16) = *Atheta clientula* auct. (nec ERICHSON, 1839); 17) listed as *Alevonota rufotestacea* (KRAATZ, 1856) (misidentification).

Species	Sample numbers (specimen numbers)	Previous records
Omalinae		
<i>Omalium oxyacanthae</i> GRAVENHORST, 1806	3b(1)	
<i>Omalium rhodicum</i> ZANETTI & ASSING, 2013	1(1), 29b(1)	
<i>Omalium rugatum</i> MULSANT & REY, 1880	24b(1)	A05b
Proteininae		
<i>Metopsia assingi</i> ZERCHE, 1998		A05b
<i>Proteinus utrarius</i> ASSING, 2004	1(1), 1a(2), 1c(13), 3(6), 3b(7), 4(1), 10(145), 13a(1), 19(2), 20a(10), 20d(7), 21a(41), 24a(6), 24b(25), 29a(7), 33(2), 34(22)	A05b
Micropeplinae		
<i>Micropeplus fulvus</i> ERICHSON, 1840	34(2)	A05b
<i>Micropeplus staphylinoides</i> (MARSHAM, 1802)	20d(15), 24a(1), 24b(3)	
Pselaphinae		
<i>Brachygluta foveola foveola</i> (MOTSCHULSKY, 1840)	38(1)	
* <i>Bryaxis lesbius</i> sp.n.	42(1), 42a(1)	
<i>Bryaxis</i> sp. (♀)	1(1)	
* <i>Bythinus simplicialpis</i> sp.n.	42(1)	
<i>Bythinus</i> sp. (♀)	1(4), 1c(2), 46(1)	
<i>Faronus distinctus</i> BESUCHET, 1999	24a(2)	
* <i>Protamaurops assingi</i> sp.n.	24a(1)	
<i>Tribatus creticus</i> REITTER, 1884	50(1)	PC15
<i>Tychus apfelbecki</i> KARAMAN, 1955	19(4), 33(1), 42(1)	
<i>Tychus laminiger</i> BESUCHET, 1969	7c(1), 7d(2)	
* <i>Tychus lesbius</i> ASSING sp.n.	34(1), 35(1)	
* <i>Tychus moecha</i> KURBATOV, 2012	9(1), 39(2)	SBK12
* <i>Tychus torticornis</i> BESUCHET & SABELLA, 2012	1c(1), 7c(4), 7d(1), 7e(7), 39(1), 44(1), 44a(6), 46(1), 48(8)	BS12
* <i>Tychus triumphator</i> SABELLA, 2012	43(1), 49(4)	SBK12
Phloeocharinae		
<i>Phloeocharis longipennis</i> FAUVEL, 1875		A05b
Tachyporinae		
<i>Lordithon exoletus</i> (ERICHSON, 1839)		A05b
<i>Lordithon thoracicus</i> (FABRICIUS, 1777)	21a(1)	
<i>Lordithon trinotatus</i> (ERICHSON, 1839)	3b(1), 13a(1)	
<i>Mycetoporus ignidorsum</i> EPPELSHEIM, 1880	1c(1)	A05b
<i>Mycetoporus imperialis</i> BERNHAUER, 1902	27b(1)	A05b
<i>Mycetoporus reichei</i> (PANDELLÉ, 1869)	4(2), 10(1), 13a(1), 19(1), 24a(2), 30a(2), 33(1)	A05b
<i>Mycetoporus simillimus</i> FAGEL, 1965	1(1), 3(35), 3b(18), 4(2), 10(14), 13a(1), 19(2), 20a(4), 20d(6), 24a(3), 24b(1), 29a(1)	A05b ¹⁾
<i>Sepedophilus immaculatus</i> (STEPHENS, 1832)	13a(1), 20(1)	A05b

Species	Sample numbers (specimen numbers)	Previous records
<i>Sepedophilus obtusus</i> (LUZE, 1902)	6a(1), 7c(1)	A05b
<i>Sepedophilus testaceus</i> (FABRICIUS, 1792)	1c(1), 12(3), 13a(3)	A05b
<i>Tachinus corticinus</i> GRAVENHORST, 1802	16(1), 20(3), 20b(7), 20c(2), 20d(16), 30(1)	
<i>Tachyporus abner</i> SAULCY, 1865	7a(1), 31(1)	
<i>Tachyporus caucasicus</i> KOLENATI, 1846		A05b
<i>Tachyporus nitidulus</i> (FABRICIUS, 1781)	7b(2), 7e(1), 15(1), 28(1)	A05b
<i>Tachyporus pusillus</i> (GRAVENHORST, 1806)	7a(1)	
Habrocerinae		
<i>Habrocerus pisidicus</i> KORGE, 1971	1(21), 1c(10), 5(1), 12(3), 24a(2), 24b(1)	A05b
Aleocharinae		
<i>Acrotona muscorum</i> (BRISOUT, 1860)	1a(1), 4(1)	A05b
<i>Aleochara haematoptera</i> KRAATZ, 1858	37(1)	A05b
<i>Aleochara hamulata</i> ASSING, 2009	29c(1), 33	
<i>Aleochara lata</i> GRAVENHORST, 1802		A05b
<i>Aleochara laticornis</i> KRAATZ, 1856	1c(1)	
<i>Aleochara verna</i> SAY, 1833	34(1)	
<i>Alevonota libanotica</i> (FAGEL, 1965)		A05b ¹⁷⁾ , AW08
<i>Aloconota aegaea</i> sp.n.	14(4)	
<i>Aloconota cambrica</i> (WOLLASTON, 1855)	1b(1), 14(12)	A05b
<i>Aloconota gregaria</i> (ERICHSON, 1839)	33(2)	
<i>Aloconota lesbia</i> sp.n.	1b(1)	
<i>Amischa filum</i> (MULSANT & REY, 1870) ¹¹⁾	6(1)	A05b
<i>Anaulacaspis laevigata</i> (DUVIVIER, 1883)	14a(1)	A05b
<i>Anaulacaspis nigrina</i> (FAGEL, 1969)	14a(2), 14b(1)	
<i>Atheta aeneicollis</i> (SHARP, 1869)	1a(12), 12(1)	
<i>Atheta amicula</i> (STEPHENS, 1832)	6(1), 21a(1)	
<i>Atheta atramentaria</i> (GYLLENHAL, 1810)	29b(16), 29c(27)	
<i>Atheta atricolor</i> (SHARP, 1869)	29b(8)	
<i>Atheta cauta</i> (ERICHSON, 1837)	29b(3)	
<i>Atheta crassicornis</i> (FABRICIUS, 1792)	21a(2)	
<i>Atheta (Mocyta) cingulata</i> (HEER, 1839) ¹⁵⁾	10(2), 30a(1)	
<i>Atheta (Mocyta) clientula</i> (ERICHSON, 1839)	5a(1)	
<i>Atheta (Mocyta) pulchra</i> (KRAATZ, 1856) ¹⁶⁾	5(3), 5a(36), 6(4)	
<i>Atheta (Mocyta)</i> sp.	7e(1)	
<i>Atheta (Philhygra)</i> sp. (♀)	6(2), 6a(3), 38(2)	
<i>Brundinia meridionalis</i> (MULSANT & REY, 1853)	5a(16)	
<i>Cordalia anatolica</i> ASSING, 2001	12(4), 13a(1), 33(2)	A05b
<i>Cordalia obscura</i> (GRAVENHORST, 1802)	5(1)	
<i>Cypha longicornis</i> (PAYKULL, 1800)		A05b
<i>Cypha spathulata</i> ASSING, 2007	1c(1), 10(2), 20a(1)	
<i>Cypha tenebricosa</i> ASSING, 2004 (♀)	5(1)	
<i>Dinusa smyrnensis</i> ASSING, 2007	11(1), 13(9), 20(7), 20c(5)	
<i>Enalodroma hepatica</i> (ERICHSON, 1839)	30a(1)	
<i>Falagria sulcatula</i> (GRAVENHORST, 1806)	5a(3), 6a(2)	
<i>Geostiba maxiana</i> (TIKHOMIROVA, 1973)	3(3), 3b(2), 7a(2), 9(3), 10(3), 20b(2), 20d(11), 24a(1), 35(1)	A05b ¹⁰⁾
<i>Geostiba oertzeni</i> (EPPELSHEIM, 1888)	7a(1), 7b(2), 7c(2), 7d(2), 7e(1), 28(13)	A05b
<i>Gnypeta carbonaria</i> (MANNERHEIM, 1830)	38(1)	

Species	Sample numbers (specimen numbers)	Previous records
<i>Halobrecta algae</i> (HARDY, 1851)	32(5)	
<i>Halobrecta flavipes</i> THOMSON, 1861	5a(3)	
<i>Haploglossa villosula</i> (STEPHENS, 1832)		A05b
<i>Hydrosmecta</i> sp. 1	14(2), 37(1)	
<i>Hydrosmecta</i> sp. 2	27a(2)	
<i>Hydrosmecta</i> sp. 3	37(1)	
<i>Leptusa samia</i> ASSING, 2004	13a(1)	
<i>Liogluta longiuscula</i> (GRAVENHORST, 1802)	3(2), 3b(5), 4(1), 6a(1), 13(3), 20a(2), 20d(1), 21(1), 24a(4), 24b(8), 27b(2), 29a(1), 30a(1), 34(4)	
<i>Myllaena intermedia</i> ERICHSON, 1837	27b(1)	
<i>Myllaena lesbia</i> ASSING sp.n.	6(3), 6a(2), 27(1)	
<i>Myrmecopora convexula</i> ASSING, 1997	13(3), 20(8), 20c(1), 24(7)	A05b
<i>Myrmecopora sulcata</i> (KIESENWETTER, 1850)	32(5)	
<i>Myrmecopora uvida</i> (ERICHSON, 1840)	32(2)	
<i>Myrmoecia plicata</i> (ERICHSON, 1837)	3(1)	
<i>Notothecta pisidica</i> ASSING, 2004	29(1)	
<i>Ocalea</i> cf. <i>puncticollis</i> MULSANT & REY, 1875 (♀)	1(1)	
<i>Ocalea</i> cf. <i>rivularis</i> MILLER, 1852 (♀)	6a(1)	
<i>Oligota pumilio</i> KIESENWETTER, 1858		A05b
<i>Ousipalia caesula</i> (ERICHSON, 1839)	6a(1)	
<i>Oxypoda attenuata</i> (MULSANT & REY, 1853)	21(1)	
<i>Oxypoda carbonaria</i> (HEER, 1841)	29b(1)	
<i>Oxypoda ferruginea</i> ERICHSON, 1839	5a(1), 6(1)	
<i>Oxypoda haemorrhoea</i> (MANNERHEIM, 1830)	7c(1), 7e(1)	
<i>Oxypoda lesbia</i> ASSING, 2005	3(1), 20d(1)	A05b
<i>Oxypoda lurida</i> WOLLASTON, 1857	5(2)	
<i>Oxypoda</i> sp. (♀)	3b(1)	
<i>Pella cinctipennis</i> (EPPELSHEIM, 1884)	29a(1)	
<i>Piochardia reitteri</i> (WASMANN, 1894)	31(3)	
<i>Tachyusa nitella</i> FAUVEL, 1895		Pa06
<i>Tetralaucopora longitarsis</i> (ERICHSON, 1839)	6a(2), 14(3), 14b(1), 37(3), 38(5)	
<i>Zoosetha</i> sp. (♀)	3b(1)	A05b
O x y t e l i n a e		
<i>Anotylus clypeonitens</i> (PANDELLÉ, 1867)		A05b ¹²⁾ , S12
<i>Anotylus complanatus</i> (ERICHSON, 1839)	29b(2)	
<i>Anotylus inustus</i> (GRAVENHORST, 1806)	1a(1), 5(1), 6(2), 7b(1), 7e(1), 14(1), 20d(1), 21a(5), 25(4), 27b(1), 31(2), 33(1)	
<i>Anotylus pumilus</i> (ERICHSON, 1839)	29b(7), 29c(3)	
<i>Anotylus sculpturatus</i> (GRAVENHORST, 1806)	29b(5)	A05b
<i>Anotylus tetracarinatedus</i> (BLOCK, 1799)	29b(7)	
<i>Aploderus lydicus</i> ASSING, 2007	1(5)	
<i>Bledius frisius</i> LOHSE, 1978	8(1)	
<i>Bledius unicornis</i> (GERMAR, 1825)		A05b
<i>Carpelimus corticinus</i> (GRAVENHORST, 1806)	5a(1), 6(1), 37(1)	
<i>Carpelimus gracilis</i> (MANNERHEIM, 1830)	5a(1), 6(1), 37(1)	
<i>Carpelimus similis</i> (SMETANA, 1967)	1(1), 6a(3)	
<i>Ochtheophilus rosenhaueri</i> (KIESENWETTER, 1850)		M14
<i>Ochtheophilus andalusiacus</i> (FAGEL, 1957)	14(11)	
<i>Ochtheophilus lenkoranus</i> (SCHEERPELTZ, 1950)	14(44)	

Species	Sample numbers (specimen numbers)	Previous records
<i>Ochtheophilus venustus</i> (ROSENHAUER, 1856)	27(9), 27a(35), 27b(30)	
Steninae		
<i>Stenus aceris</i> STEPHENS, 1833	10(1), 20b(1), 24a(3)	A05b
<i>Stenus brunripes</i> STEPHENS, 1833	6(1), 6a(1), 27b(1)	
<i>Stenus glacialis</i> HEER, 1839	1c(1), 10(3), 20a(1)	A05b, Pu08
<i>Stenus hospes</i> ERICHSON, 1840	7a(2), 7e(3)	
<i>Stenus morio</i> GRAVENHORST, 1806		A05b
<i>Stenus picipes picipes</i> STEPHENS, 1833	2(1), 7c(1), 20c(1)	
<i>Stenus turbulentus</i> BONDROIT, 1912	12(5), 13a(2), 19(1), 20a(2), 24a(1), 24b(1)	A05b
Scydmaeninae		
* <i>Cephennium (Phemecium)</i> sp.n.	3(3)	
<i>Cephennodes</i> sp.n.	24b(1)	
* <i>Euconnus (Napochus)</i> sp. (♀)	19(1)	
<i>Leptomastax coquereli</i> (FAIRMAIRE, 1856)	45(2)	
* <i>Stenichnus lesbii</i> sp.n.	1(1), 17(1), 40(2), 41(2), 42a(1), 43(1), 44a(1), 45(1), 46(1)	
<i>Stenichnus pelliceus</i> (HOLDHAUS, 1908)	21(1), 39(1), 47(2)	
<i>Stenichnus</i> sp. 1 ²⁾	35(2)	
<i>Stenichnus</i> sp. 2 (♀)	20d(1)	
Paederinae		
<i>Achenium depressum</i> (GRAVENHORST, 1802)		A05b, A10
<i>Astenus lyonesis</i> (JOY, 1908)		A05b
<i>Astenus melanurus</i> (KÜSTER, 1853)	6(2), 7b(1), 11(2), 17(1),	A05b
<i>Astenus procerus</i> (GRAVENHORST, 1806)	18(1)	A05b ³⁾
<i>Astenus thoracicus</i> (BAUDI DI SELVE, 1857)	3a(3), 7a(4), 7b(2), 7c(2), 7d(2), 7e(8), 11(2), 16(4), 22(3), 23(1), 25(1), 30(1), 31(3), 36(1)	A05b
<i>Leptobium gracile</i> (GRAVENHORST, 1802)	5(20), 5a(5), 6a(2), 7(1), 7a(1), 7b(1), 7e(1),	A05b
<i>Leptobium illyricum</i> (ERICHSON, 1840)	16(7), 17(1), 20(2), 20c(1), 22(1), 30(1)	A05a, A05b
<i>Lobrathium rugipenne</i> (HOCHHUTH, 1851)	27(2), 37(1)	A05b, A07e, A13d
<i>Medon dilutus pythonissa</i> (SAULCY, 1865)	24a(1), 24b(5)	A05b
<i>Medon fusculus</i> (MANNERHEIM, 1830)	1(4), 1c(3), 2(1), 19(5)	
<i>Medon lydicus</i> BORDONI, 1980	1(26), 1c(11), 2(2), 12(7), 13a(4), 19(12), 20(1), 20a(11), 24a(10), 27(8), 27b(4), 30a(3), 33(5), 34(1)	A05b
<i>Medon maronitus</i> (SAULCY, 1865)	1(2), 19(5), 33(1)	A05b
<i>Medon rufiventris</i> (NORDMANN, 1837)		A05b
<i>Medon semiobscurus</i> (FAUVEL, 1875)	1(4), 1c(1), 13a(6)	A05b
<i>Micranops pilicornis</i> (BAUDI DI SELVE, 1870)		A05b
<i>Micrillus testaceus</i> (ERICHSON, 1840)	7(1)	
<i>Ochtheophilum brevipenne</i> (MULSANT & REY, 1861) (♀) ⁴⁾	5(1), 5a(3), 8(1)	
<i>Paederus littoralis</i> GRAVENHORST, 1802		A05b
<i>Pseudomedon dido</i> (SAULCY, 1865)	14(2)	A05b ³⁾ , A08b
<i>Pseudomedon obscurellus</i> (ERICHSON, 1840)	5a(1), 6(1)	
<i>Rugilus angustatus</i> (GEOFFROY, 1785)	27b(1)	A05b
<i>Rugilus lesbii</i> ASSING, 2005	12(4)	A05b

Species	Sample numbers (specimen numbers)	Previous records
<i>Rugilus similis</i> (ERICHSON, 1839)	20d(1)	
<i>Scopaeus cameroni</i> COIFFAIT, 1968	20c(1)	F10
<i>Scopaeus debilis</i> HOCHHUTH, 1851	5a(2)	
<i>Scopaeus gracilis</i> (SPERK, 1835)	37(2)	A05b ⁹⁾
<i>Scopaeus laevigatus</i> (GYLLENHAL, 1827)	6a(2)	
<i>Scopaeus</i> cf. <i>pusillus</i> KIESENWETTER, 1843 (♀)	35(1)	
<i>Scymbalium anale</i> (NORDMANN, 1837)		A05b, A13e
<i>Sunius melanocephalus</i> (FABRICIUS, 1792)	5a(1), 6(1)	A05b ⁶⁾
* <i>Sunius potti</i> ASSING sp.n.	7(2), 7c(1), 7e(1), 20(1), 20c(1), 20d(1)	A05b ⁷⁾
Staphylininae		
<i>Acylophorus glaberrimus</i> (HERBST, 1784)	27(2), 27a(1), 27b(6)	
<i>Astrapaeus ulmi</i> (ROSSI, 1790)	7c(1)	
<i>Creophilus maxillosus</i> (LINNAEUS, 1758)		A05b
<i>Erichsonius subopacus</i> (HOCHHUTH, 1851)	27(1), 27b(9)	
<i>Gabrius latro</i> JOY, 1913	5(6), 5a(1), 6(8), 6a(8), 27b(1)	
<i>Gabrius nigrutilus</i> (GRAVENHORST, 1802)	6a(1)	
<i>Gabronthus maritimus</i> (MOTSCHULSKY, 1858)	5a(1)	
<i>Gauropterus sanguinipennis</i> (KOLENATI, 1846)	37(2)	
<i>Gyrohypnus angustatus</i> STEPHENS, 1833	12(1), 33(3)	A05b
<i>Nudobius cypriacus</i> COIFFAIT, 1956		C70
<i>Ocyopus curtipennis</i> (MOTSCHULSKY, 1849)	24(1)	A05b ⁸⁾ , C70 ¹³⁾
<i>Ocyopus mus</i> (BRULLÉ, 1832)	12(1), 20(2), 20c(1)	A05b
<i>Ocyopus sericeicollis</i> (MÉNÉTRIÉS, 1832) (♀)	20(1), 24(1)	
<i>Othius laeviusculus</i> STEPHENS, 1833		A05b
<i>Othius lapidicola</i> MÄRKEL & KIESENWETTER, 1848	1(1)	A05b
<i>Philonthus micans</i> (GRAVENHORST, 1802)	5(1), 5a(1)	
<i>Philonthus nitidicollis</i> (LACORDAIRE, 1835)		A05b
<i>Philonthus rufimanus</i> ERICHSON, 1840	6(1), 6a(1), 14b(2), 37(85), 38(1)	
<i>Philonthus salinus</i> KIESENWETTER, 1844	5(1), 5a(2)	
<i>Rabigus pullus</i> (NORDMANN, 1837)	6(2)	
<i>Remus filum</i> (KIESENWETTER, 1849)	32(3)	
<i>Remus sericeus</i> HOLME, 1837	32(1)	
<i>Quedius coloratus</i> FAUVEL, 1875	10(1), 12(1)	
<i>Quedius fissus</i> GRIDELLI, 1938	13a(1), 19(1), 20a(1), 24b(1), 33(9)	A05b, C70
<i>Quedius humeralis</i> STEPHENS, 1832	4(1), 10(4), 13a(1), 19(2), 20a(3), 20d(1), 24b(2), 28(1), 30a(1), 33(2), 34(4), 35(1)	A05b
<i>Quedius job</i> COIFFAIT, 1963	12(1), 14b(1), 27(1), 27b(1)	
<i>Quedius levicollis</i> (BRULLÉ, 1832)		A05b, H65 ¹⁴⁾
<i>Quedius nemoralis</i> BAUDI DI SELVE, 1848	3(3), 3b(2), 4(1), 33(1)	
<i>Quedius semiaeneus</i> (STEPHENS, 1833)	20b(1), 31(2)	
<i>Xantholinus rufipennis</i> ERICHSON, 1839		App
<i>Xantholinus varnensis</i> COIFFAIT, 1972	5(2), 5a(2), 6a(1)	

Notes on some species

***Omalius rhodicum* ZANETTI & ASSING, 2013**

The original description of *O. rhodicum* is based on a unique male from Rhodos (ASSING 2013b). The species was subsequently reported also from Crete, again based on a unique male (ASSING 2015a). The records from Lesbos (two males) confirm that *O. rhodicum* is widespread in the Aegean region. So far, this species has been recorded only from islands.

***Metopsia assingi* ZERCHE, 1998**

This species was already recorded from Lesbos by ASSING (2005b), but is not listed for Greece in the Palaearctic Catalogue (SCHÜLKE & SMETANA 2015).

***Faronus distinctus* BESUCHET, 1999**

The original description is based on material from southwestern and western Anatolia (Antalya, Muğla, Izmir) and from Rhodos (BESUCHET 1999), from where it was reported again recently (ASSING 2013b). The species was subsequently recorded also from Samos (ASSING 2015c). The specimens listed in Tab. 1, a male and a female, represent the first record from Lesbos.

***Aleochara verna* SAY, 1833**

According to SCHÜLKE & SMETANA (2015), *A. verna* is widespread in the Holarctic Region, but was previously unknown from Greece.

***Anaulacaspis nigrina* (FAGEL, 1969)**

This species was previously known only from the type locality near Alanya (Turkey: Antalya) (FAGEL 1969). The males listed in Tab. 1 and two of the specimens erroneously listed as *A. laevigata* in ASSING (2005b) represent the first records from Greece.

***Atheta atricolor* (SHARP, 1869)**

Atheta atricolor had been recorded from several countries in West and Central Europe, Turkey, and West Siberia (SCHÜLKE & SMETANA 2015). The material listed in Tab. 1 represents the first record from Greece.

***Cypha spathulata* ASSING, 2007**

Though widespread in the Mediterranean, this species was only recently recorded from Greece (Karpathos) for the first time (ASSING 2016a).

***Cypha tenebricosa* ASSING, 2004**

The previously known distribution of *C. tenebricosa* was confined to Turkey and the Greek islands Chios and Karpathos (ASSING 2015b, 2016a).

***Dinusa smyrnensis* ASSING, 2007**

Previously, only the unique female holotype of this species from Izmir was known (ASSING 2007b). *Dinusa* species are associated with ants of the genus *Messor* FOREL, 1890 and generally found on very rare occasions; several species are currently known only from their respective type localities and/or represented only by unique holotypes. Therefore, the 22 specimens from Lesbos, the first records of *D. smyrnensis* from Greece, represent an exceptional strike of luck. Never before has such a number of *Dinusa* specimens been collected in one region and in one period. Remarkably, the sex ratio was somewhat biased. All nine specimens collected from a single *Messor* nest in sample number 13 are males and all seven specimens from sample number 20 are males, too. In all, only four females were found (sample numbers 11 and 20c).

***Geostiba maxiana* (TIKHOMIROVA, 1973)**

This widespread species is wing-dimorphic (ASSING 2015b). All 28 specimens collected in Lesbos are micropterous.

***Geostiba oertzeni* (EPPELSHEIM, 1888)**

Like *G. maxiana*, *G. oertzeni* is widespread and wing-dimorphic. The previous observation that the material from Lesbos is of exceptionally dark coloration (ASSING 2005b) is confirmed. Interestingly, all eight specimens from samples 7a–e are macropterous, whereas all 13 specimens from sample 28 are micropterous.

***Leptusa samia* ASSING, 2004**

This species was originally described from Samos (ASSING 2004a) and subsequently recorded also from one locality in Manisa province in western Anatolia (ASSING 2007a).

***Notothecta pisidica* ASSING, 2004**

Unlike other West Palaearctic *Notothecta* species, *N. pisidica* is not associated with formicine ants, but with myrmicine ants of the genus *Messor* (ASSING 2007b). The species was originally described from Konya (ASSING 2004b) and subsequently recorded also from Aydın and Antalya provinces in Turkey (ASSING 2007b). The male listed in Tab. 1 was found in a nest of *Messor* sp. and represents the first record from Greece and Europe.

***Oxypoda lesbia* ASSING, 2005**

This species was originally described based on a unique male from Lesbos (ASSING 2005b) and subsequently recorded also from the Greek mainland (ASSING 2006b), Turkey (Aydın, Izmir, and Denizli provinces) (ASSING 2007c), and the Aegean islands Rhodos (ASSING 2013b), Crete (ASSING 2015a), and Chios (ASSING 2015b).

***Pella cinctipennis* (EPPELSHEIM, 1884)**

Pella cinctipennis is a rare species with only few records known from Middle Asia (Tajikistan, Uzbekistan), Azerbaijan, one from Muğla in southwestern Anatolia, and one from Lebanon (ASSING 2008c, 2016b, MARUYAMA 2006). The female listed in Tab. 1 represents the first record from Greece.

***Piochardia reitteri* (WASMANN, 1894)**

This rare myrmecophilous species is associated with *Cataglyphis nodus* (BRULLÉ, 1833). Its distribution ranges from the Balkans to the Caucasus region and the Middle East. In Greece, it was previously known only from one locality in Thessalía (ASSING 1999). The specimens listed in Tab. 1 were collected from two nests of *Cataglyphis nodus* after a cold night with rainfall. Subsequent attempts at collecting more specimens were unsuccessful.

***Zoosetha* sp.**

Unfortunately, both specimens that are currently available, one of them collected in 2005 and one in 2016, are females. They probably represent an undescribed species. Males would be required for an adequate description.

***Anotylus complanatus* (ERICHSON, 1839)**

According to SCHÜLKE & SMETANA (2015), *A. complanatus* is widespread in the Palaearctic Region and has also been reported from the Australian and Neotropical Regions, but was previously unknown from Greece.

***Aploderus lydicus* ASSING, 2007**

The previously known distribution of *A. lydicus* was confined to the West Anatolian provinces Aydın, Izmir, and Konya (ASSING 2007b, d, 2013c). The specimens from Lesbos represent the first record from Greece.

***Stenus hospes* ERICHSON, 1840**

All five specimens listed in Tab. 1 are females and were found in the same locality; two of them were collected from nests of *Messor* spp. For comments on the taxonomy and myrmecophily of this species see ASSING (2015c).

***Leptomastax coquereli* (FAIRMAIRE, 1856)**

According to H. Meybohm (e-mail 14 April, 2016), the specimens from Lesbos are distinguished from material from northern Turkey by distinctly longer antennae. However, no significant differences were observed in the male sexual characters, and the length of the antennae and antennomeres is generally considered as subject to pronounced intraspecific variation in this species.

***Pseudomedon dido* (SAULCY, 1865)**

The distribution of *P. dido* ranges from the eastern Aegean islands across Turkey to Iran and Cyprus. In Greece it has been recorded only from Lesbos and Karpathos (ASSING 2009).

***Othius lapidicola* MÄRKEL & KIESENWETTER, 1848**

Othius lapidicola is widespread and common in the East Mediterranean. In the Aegean region, it is usually one of the dominant species in leaf litter of all kinds. Surprisingly, only a single specimen was collected in Lesbos in 2016. This specimen is infested with Laboulbeniales.

***Quedius fissus* GRIDELLI, 1938**

The known distribution of this species is confined to Turkey and the Greek islands Rhodos and Lesbos (ASSING 2005b, 2013b).

***Quedius job* COIFFAIT, 1963**

Quedius job had been known only from Turkey and Lebanon until it was recently recorded from Greece (Karpathos) for the first time (ASSING 2016a).

***Xantholinus rufipennis* ERICHSON, 1839**

MATERIAL EXAMINED: **LESBOS**: 1 ♂, Ahladeri env., 39°09'N, 26°17'E, 5 m, sea shore, 21.–22.VII.2007, leg. Hájek (cAss).

This species is widespread in the Mediterranean. It was not recorded during the two field trips in 2005 and 2016.

***Xantholinus varnensis* COIFFAIT, 1972**

The distribution of *X. varnensis* ranges from Bulgaria and northeastern Greece across West Turkey and Rhodos southwards to Karpathos (ASSING 2008b). The male-based records from Lesbos confirm that the females from Samos (ASSING 2015c) most likely belong to *X. varnensis* (and not to the sibling species *X. graecus* KRAATZ, 1858).

Descriptions of new species***Myllaena lesbia* ASSING sp.n. (Figs. 2–13)**

TYPE MATERIAL: **Holotype** ♂: “GREECE: Lesbos [6a], SW Kalloni, river bank, 39°12'18"N, 26°10'39"E, 2 m, 25.III.2016, V. Assing & A. Hetzel / Holotypus ♂ *Myllaena lesbia* sp. n. det. V. Assing 2016” (cAss). **Paratypes**: 1 ♂, 2 ♀♀: same data as holotype; 1 ♂, 1 ♀: same data as holotype, but “[6] ... 19.III.2016” (cAss); 1 ♀: “GREECE: Lesbos [27] 16 km ENE Kalloni, 39°15'02"N, 26°23'14"E, 120 m, 24.III.2016, V. Assing & A. Hetzel” (cAss).

ETYMOLOGY: The specific epithet (Latin, adjective) alludes to the fact that this species is currently known only from Lesbos.

DESCRIPTION: Body length 2.8–3.4 mm; length of forebody 1.3–1.5 mm. Coloration: body blackish-brown to black; legs dark-yellowish; antennae dark-brown, with the apical 1–3 antennomeres often paler.

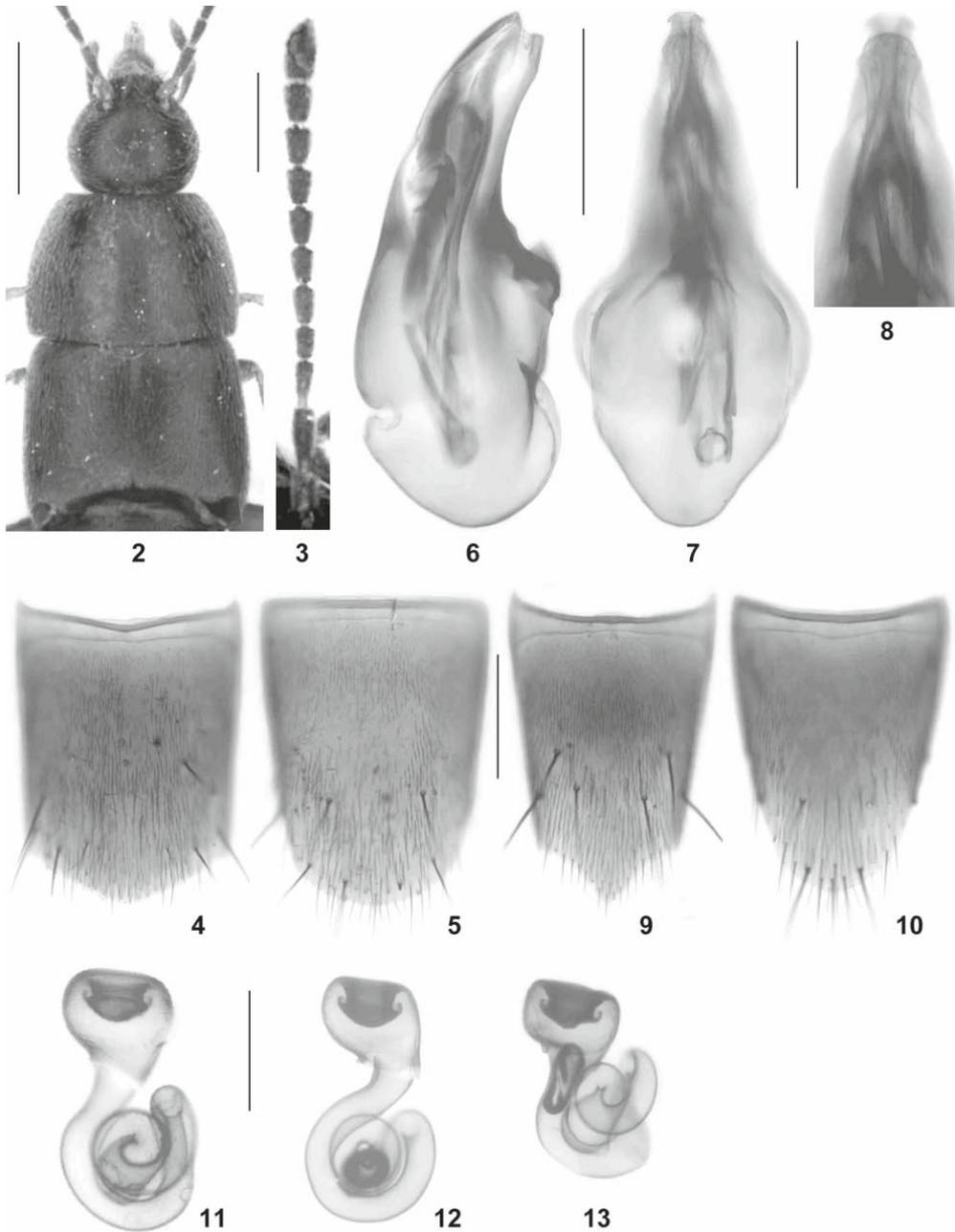
Head (Fig. 2) approximately 1.1 times as broad as long, broadest behind eyes, wedge-shaped. Eyes slightly shorter than postocular region in dorsal view. Antenna approximately 1.0 mm long; all antennomeres distinctly oblong (Fig. 3).

Pronotum (Fig. 2) approximately 1.3 times as broad as long and 1.5 times as broad as head; posterior margin weakly sinuate.

Elytra (Fig. 2) 0.80–0.85 times as long as pronotum; punctation extremely fine and dense, slightly more distinct than that of head and pronotum.

Abdomen: tergite VIII and sternite VIII with pronounced sexual dimorphism.

♂: posterior margins of tergite VIII (Fig. 4) and sternite VIII (Fig. 5) convex; median lobe of aedeagus (Figs. 6–8) approximately 0.5 mm long; ventral process and internal structures of distinctive shapes.



Figs. 2–13: *Myllaena lesbia*: 2) forebody; 3) antenna; 4) male tergite VIII; 5) male sternite VIII; 6–7) median lobe of aedeagus in lateral and in ventral view; 8) apical portion of median lobe in ventral view; 9) female tergite VIII; 10) female sternite VIII; 11–13) spermatheca. Scale bars: 2) 0.5 mm; 3–7, 9–10: 0.2 mm; 8, 11–13: 0.1 mm.

♀: posterior margin of tergite VIII (Fig. 9) acutely produced in the middle; posterior margin of sternite VIII (Fig. 10) strongly convex (much more so than in male); spermatheca as in Figs. 11–13.

COMPARATIVE NOTES: This species is characterized particularly by the sexual dimorphism of tergite VIII and sternite VIII, as well as by the distinctive morphology of the aedeagus. In external characters, it is nearly indistinguishable from the common, widespread, and syntopic *M. intermedia*, except for the slightly less transverse pronotum. However, in *M. intermedia*, tergite VIII and sternite VIII are acutely produced in both sexes and the median lobe of the aedeagus is of completely different morphology (0.6 mm long; crista apically strongly developed and of distinctive shape; ventral process apically very acute, also in ventral view, and slightly curved ventrad in lateral view; internal structures of different shapes). The spermathecae of both species are nearly identical. For illustrations of the aedeagus of *M. intermedia* see STRAND (1967).

DISTRIBUTION AND NATURAL HISTORY: *Myllaena lesbia* is currently known from two localities in the northeast and the centre of Lesbos. Since *Myllaena* species are generally widespread, it seems likely that *M. lesbia* will eventually be recorded also from Turkey and other Aegean islands. The species may previously have been confounded with *M. intermedia*.

The specimens were floated from fine gravel, hand-collected from gravel, and sifted from flood debris on stream banks at altitudes of 50 and 120 m, in one locality together with *M. intermedia*.

Aloconota (Aloconota) aegaea ASSING sp.n. (Figs. 14–22)

TYPE MATERIAL: **Holotype** ♂: "GREECE: Lesbos [14], 14 km WNW Mytilini, 39°07'51"N, 26°23'55"E, 50 m, 20.III.2016, V. Assing & A. Hetzel / Holotypus ♂ *Aloconota aegaea* sp. n. det. V. Assing 2016" (cAss). **Paratypes**: 3 ♀♀: same data as holotype (cAss); 2 ♀♀: "Greece: Samos [34+1], S Karlovasi, river bank, 37°46'34"N, 26°42'17"E, 50 m, debris & gravel 5.IV.2014, leg. Forcke" (cAss).

ETYMOLOGY: The specific epithet (Latin, adjective: Aegean) alludes to the fact that the species is currently known only from two Aegean islands.

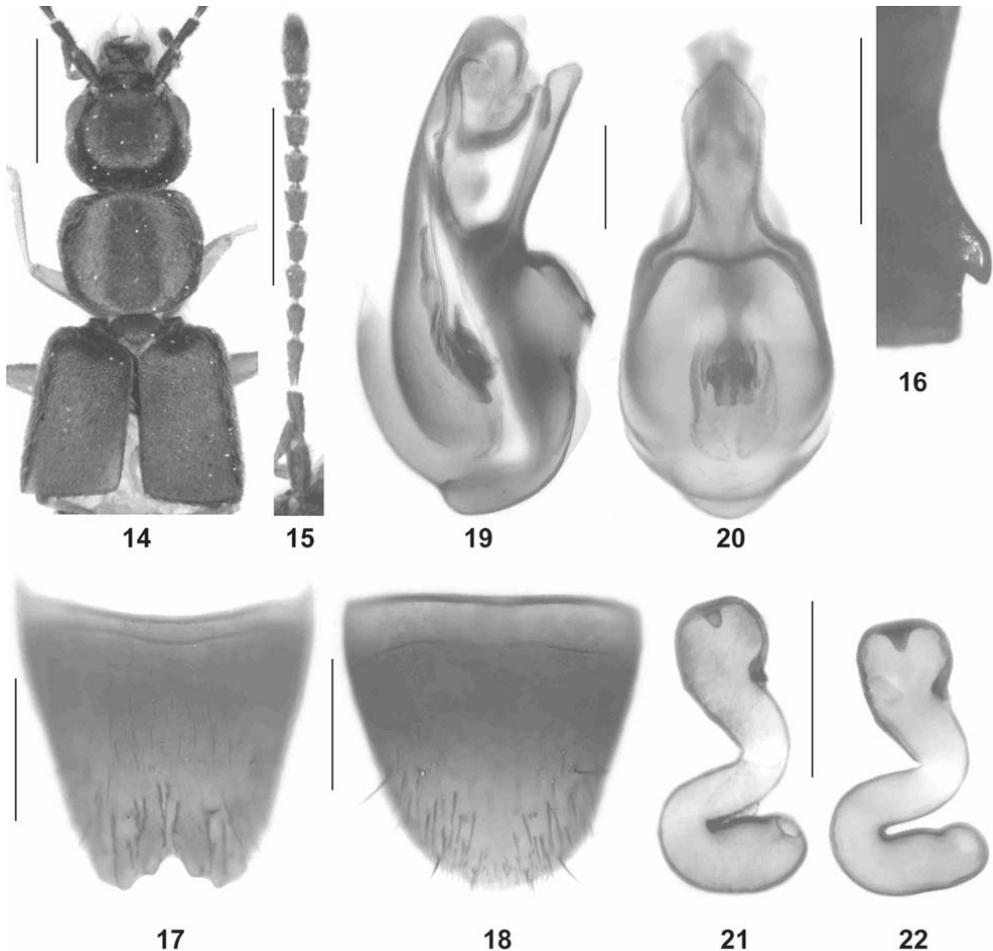
DESCRIPTION: Body length 3.8–4.5 mm; length of forebody 1.7–1.9 mm. Coloration: body black, sometimes with the elytra blackish-brown; legs yellowish; antennae black.

Head (Fig. 14) weakly transverse, of subquadrate shape or weakly dilated posteriorly; punctuation dense and extremely fine; interstices with distinct microreticulation and subdued shine. Eyes large, longer than postocular region in lateral view; genal carina distinct posteriorly and indistinct anteriorly (lateral view). Antenna (Fig. 15) 1.3–1.4 mm long, slender; all antennomeres longer than broad; antennomere XI slightly longer than the combined length of antennomeres IX and X.

Pronotum (Fig. 14) flat, depressed or shallowly impressed in postero-median portion, 1.15–1.20 times as broad as long and approximately 1.15–1.20 times as broad as head, broadest in anterior half; pubescence of midline directed anteriorly; punctuation very fine and dense; interstices with microreticulation and subdued shine.

Elytra (Fig. 14) approximately 1.1 times as long as pronotum; punctuation extremely dense; microsculpture distinct; surface nearly matt. Hind wings fully developed. Metatarsomere I approximately as long as the combined length of metatarsomeres II and III.

Abdomen narrower than pronotum; punctuation dense and distinct on tergites III–VI, sparse and fine on tergites VII–VIII; tergites VII and VIII with pronounced sexual dimorphism; posterior margin of tergite VII with palisade fringe.



Figs. 14–22: *Aloconota aegaea*: 14) forebody; 15) antenna; 16) male tergite VII in lateral view; 17) male tergite VIII; 18) male sternite VIII; 19–20) median lobe of aedeagus in lateral and in ventral view; 21–22) spermatheca of females from Lesbos (21) and Samos (22). Scale bars: 14–15: 0.5 mm; 16–18: 0.2 mm; 19–22: 0.1 mm.

♂: tergite VII (Fig. 16) with pronounced postero-median tubercle directed obliquely posteriad in lateral view; tergite VIII (Fig. 17) with posterior excision of distinctive shape; sternite VIII (Fig. 18) with convex posterior margin; median lobe of aedeagus 0.45 mm long and shaped as in Figs. 19–20.

♀: tergites VII and VIII without modifications; sternite VIII with weakly convex posterior margin; spermatheca shaped as in Figs. 21–22.

COMPARATIVE NOTES: *Aloconota aegaea* is distinguished from the similar *A. debilicornis* (ERICHSON, 1839), with which it shares a similar habitus, body size, depressed pronotum, and the slender antennae, by distinctly denser and coarser punctation of the elytra, completely different modifications of the male tergite VIII, and the shapes of the median lobe of the aedeagus and of

the spermatheca. For illustrations of *A. debilicornis* and numerous other species distributed in the Mediterranean see TRONQUET (2014).

DISTRIBUTION AND NATURAL HISTORY: The distribution of this species is currently confined to one locality in Lesbos and one in Samos. The distributions of other ripicolous *Aloconota* species, the habitat, and the low altitude suggest that *A. aegaea* is probably more widespread at least in the Aegean islands and in western Turkey. The specimens from Lesbos were floated from fine gravel of a stream bank at an altitude of 50 m, those from Samos were hand-collected from debris and gravel of a river bank at an altitude of 50 m.

Aloconota (Aloconota) lesbia ASSING sp.n. (Figs. 23–29)

TYPE MATERIAL: **Holotype** ♂: "GREECE: Lesbos [1b], 7 km ESE Mithimna, 39°21'23"N, 26°15'18"E, 440 m, 18.III.2016, V. Assing & A. Hetzel / Holotypus ♂ *Aloconota lesbia* sp. n. det. V. Assing 2016" (cAss).

ETYMOLOGY: The specific epithet (Latin, adjective) alludes to the fact that this species is currently known only from Lesbos.

DESCRIPTION: Body length 2.8 mm; length of forebody 1.3 mm. Coloration: body black, with the elytra indistinctly paler; legs dark-yellowish; antennae blackish-brown.

Head (Fig. 23) weakly transverse, of subquadrate shape; punctuation extremely fine, barely noticeable; interstices with distinct microreticulation composed of isodiametric meshes and subdued shine. Eyes large, slightly longer than postocular region in lateral view; genal carina visible only in posterior portion of head (lateral view). Antenna (Fig. 24) 0.95 mm long, not particularly slender; antennomeres IV and V weakly oblong, VI approximately as broad as long, and VII–X weakly transverse; antennomere XI approximately as long as the combined length of antennomeres IX and X.

Pronotum (Fig. 23) moderately convex in cross-section, approximately 1.15 as broad as long and 1.12 times as broad as head, broadest in anterior half; pubescence of midline probably directed anteriorly (somewhat disturbed in the holotype); punctuation extremely fine, visible only at high magnification, and moderately dense; interstices with distinct microreticulation composed of isodiametric meshes and with subdued shine.

Elytra (Fig. 23) approximately 1.05 times as long as pronotum; punctuation extremely dense, fine, but much more distinct than that of head and pronotum; microsculpture pronounced. Hind wings fully developed. Metatarsomere I slightly shorter than the combined length of metatarsomeres II and III.

Abdomen narrower than pronotum; punctuation dense and distinct on tergites III–VI, sparse and fine on tergites VII–VIII; microsculpture of tergites III–VI composed of transverse meshes, that of tergite VII of isodiametric and short transverse meshes; tergites VII and VIII with pronounced sexual dimorphism; posterior margin of tergite VII with palisade fringe.

♂: tergite VII (Fig. 25) with pronounced postero-median tubercle directed obliquely posteriorly in lateral view; tergite VIII (Figs. 26–27) with a pair of oblong elevations posteriorly, these elevations extending into blunt tooth-like projections, posterior margin additionally with a sharp tooth on either side; sternite VIII transverse and with strongly convex posterior margin; median lobe of aedeagus 0.32 mm long and shaped as in Figs. 28–29; ventral process slightly bent dorsad in lateral view.

♀: unknown.

COMPARATIVE NOTES: This species is distinguished from the similar widespread common, and syntopic *A. cambrica* by slightly smaller body size, a more transverse pronotum, slightly

darker coloration (particularly of the legs), different modifications of the male tergite VIII (*A. cambrica*: median projections of posterior margin closer together; lateral teeth shorter), and the shape of the median lobe of the aedeagus (*A. cambrica*: median lobe nearly 0.4 mm long; ventral process not bent dorsad in lateral view). For characters distinguishing it from the similar *A. samia* sp.n. see the comparative notes in the appendix at the end of this article.

For illustrations of *A. cambrica* and numerous other species distributed in the Mediterranean see TRONQUET (2014).

DISTRIBUTION AND NATURAL HISTORY: The type locality is situated near the northern coast of Lesbos. The presence of a similar different species in Samos suggests that the distribution of *A. lesbia* may be restricted. The holotype was floated from fine gravel at the bank of a temporary stream at an altitude of 440 m, together with a specimen of *A. cambrica*.

Sunius potti ASSING sp.n. (Figs. 36–43)

TYPE MATERIAL: **Holotype** ♂: “GREECE: Lesbos [7e], N Kalloni, 39°15'52"N, 26°12'18"E, 220 m, pasture, 29.III.2016, V. Assing & A. Hetzel / Holotypus ♂ *Sunius potti* sp. n. det. V. Assing 2016” (cAss). **Paratypes:** 1 ♂, 1 ♀: same data as holotype, but “[7] ... 19.III.2016” (cAss); 1 ♀: same data as holotype, but “[7c] ... 26.III.2016” (cAss); 1 ♀: “GREECE: Lesbos [20], 3.5 km SSE Agiasos, 39°03'05"N, 26°23'02"E 850 m, 22.III.2016, V. Assing & A. Hetzel” (cAss); 1 ♀: same data, but “[20c] ... 28.III.2016” (cAss); 1 ♀: same data, but “[20d] ... 28.III.2016” (cAss); 1 ♀: “N39°15'52 E026°12'25, GR Lesbos 17.3.2005, Skotino 200 m (2), Lompe & Meybohm” (cAss).

ETYMOLOGY: This species is dedicated to Jürgen Pott (Hannover), a most enthusiastic biologist, dear friend, and teaching colleague soon to be retired from the Käthe-Kollwitz-Schule Hannover, also in appreciation of his own repeated, albeit hopelessly futile efforts at collecting an undescribed species and having it named after him.

DESCRIPTION: Body length 2.7–3.2 mm; length of forebody 1.35–1.50 mm. Habitus as in Fig. 36. Coloration: forebody yellowish-red to pale reddish-brown; abdomen blackish-brown to black; legs and antennae yellowish. Pronotum with very dense punctation; interstices much narrower than diameter of punctures. Other external characters as in other species of the *S. seminiger* group.

♂: sternite VII with very weakly concave posterior margin, otherwise unmodified; sternite VIII (Figs. 37–38) approximately as long as broad, with a pronounced, in lateral view fin-shaped, tubercle and with a broadly V-shaped posterior excision; aedeagus 0.36–0.37 mm long and shaped as in Figs. 39–42; ventral process in lateral view with conspicuously straight ventral side in apical half.

COMPARATIVE NOTES: The similar external characters (dense punctation of the pronotum; body distinctly bicolored) and the similar modifications of the male sternite VIII (with fin-shaped median tubercle) and the aedeagus (shapes of ventral process and internal structures) suggest that *S. potti* is closely allied to *S. plasoni* (EPPELSHEIM, 1875) (Turkey: Manisa: Karadağ) and *S. pinniger* ASSING, 2007 (Turkey: Izmir: Karaburun), together with which *S. potti* would key out in the key to the *Sunius* species of the West Palaearctic Region in ASSING (2008a). The new species is distinguished from both of them only by the male sexual characters.

In *S. plasoni*, the ventral process of the aedeagus is more slender, more acute, and ventrally distinctly concave in the apical half and more deeply concave in the basal half (lateral view), and the median tubercle of the male sternite VIII is broader, posteriorly more broadly convex (ventral view), and more pointed (lateral view).

In *S. pinniger*, the ventral process of the aedeagus is curved and ventrally distinctly concave in the apical half, rendering the median angle acute, and more broadly concave in the basal half

(lateral view), and the median tubercle of the male sternite VIII is posteriorly more acute (ventral view) and more broadly truncate (lateral view).

For illustrations of *S. pinniger* and *S. plasoni* see ASSING (2006a: figs. 12–24, 49–62).

COMMENT: The three females from the locality near Agiasos are tentatively included in the type series; a male from this locality would be required to confirm that they are conspecific with the material from the type locality. However, the low altitude of the type locality suggests that *S. potti* is widespread in Lesbos.

DISTRIBUTION AND NATURAL HISTORY: As can be inferred from the restricted distributions of other species of the *S. seminiger* group, *Sunius potti* is most likely endemic to Lesbos. The specimens from the type locality were collected by turning stones at the margin of a pasture at an altitude of only 220 m (Fig. 43), together with an undescribed species of Anillina (Carabidae). Despite repeated efforts (in all six visits to this locality), only four specimens were collected. The females from the sample plot to the southeast of Agiasos were collected from the margin of a track with grass and herbs, two of them by turning stones, and one by washing soil.

***Bryaxis lesbius* BRACHAT sp.n. (Figs. 44–45)**

TYPE MATERIAL: **Holotype** ♂: “N39°04'11 E026°20'58, GR Lesbos, 24.03.2005, Olymbos, 790 m (10a), Lompe & Meybohm / Holotypus ♂ *Bryaxis lesbius* sp. n. det. V. Brachat 2016” (cBra).

ETYMOLOGY: The specific epithet is an adjective (Latin) derived from Lesbos.

DESCRIPTION: Body length 1.88 mm. Body reddish-brown, with long, sparse, and more or less erect yellowish pubescence.

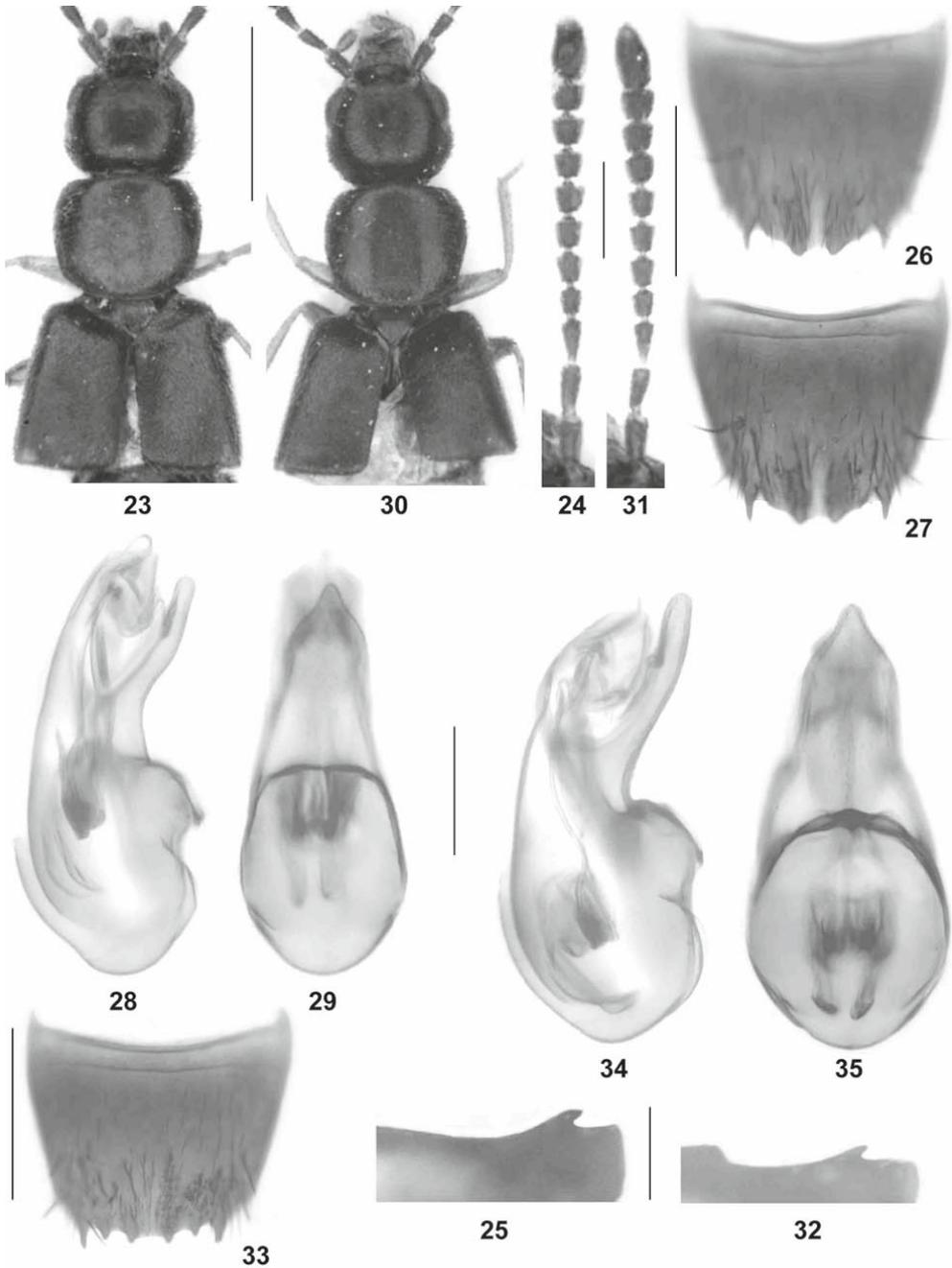
Head weakly transverse, 0.34 mm long and 0.36 mm broad; vertex with long median keel anteriorly extending beyond intra-ocular pits; postocular region weakly rounded in dorsal view; gular pit distinct, anteriorly with a small coniform projection; microsculpture absent. Eyes small, projecting from lateral contours of head, and composed of 13 ommatidia. Antenna 0.8 mm long; antennomere I (Fig. 44) stout, 1.4 times as long as broad, distally with a coniform process directed obliquely upwards; antennomere II (Fig. 44) much smaller, weakly oblong; antennomeres III distinctly oblong, IV and V weakly oblong, VI–VIII approximately as broad as long, IX–XI of increasing width and forming a club, XI elongate and approximately as long as the combined length of VII–X. Maxillary palpomeres II and III with scattered tubercles on inner side, IV slightly more than twice as long as broad, broadest in basal third, with short and dense pubescence.

Pronotum glossy, indistinctly transverse. Elytra (combined) transverse, 0.80 mm broad and 0.68 mm long, and with coarse punctation; humeral angles rounded; sub-humeral pit pronounced. Legs long and with strongly dilated femora; protibia with a deep and basally dentate excision in apical third; mesotibia unmodified; metatibia flattened, dilated, and with pronounced apical spine.

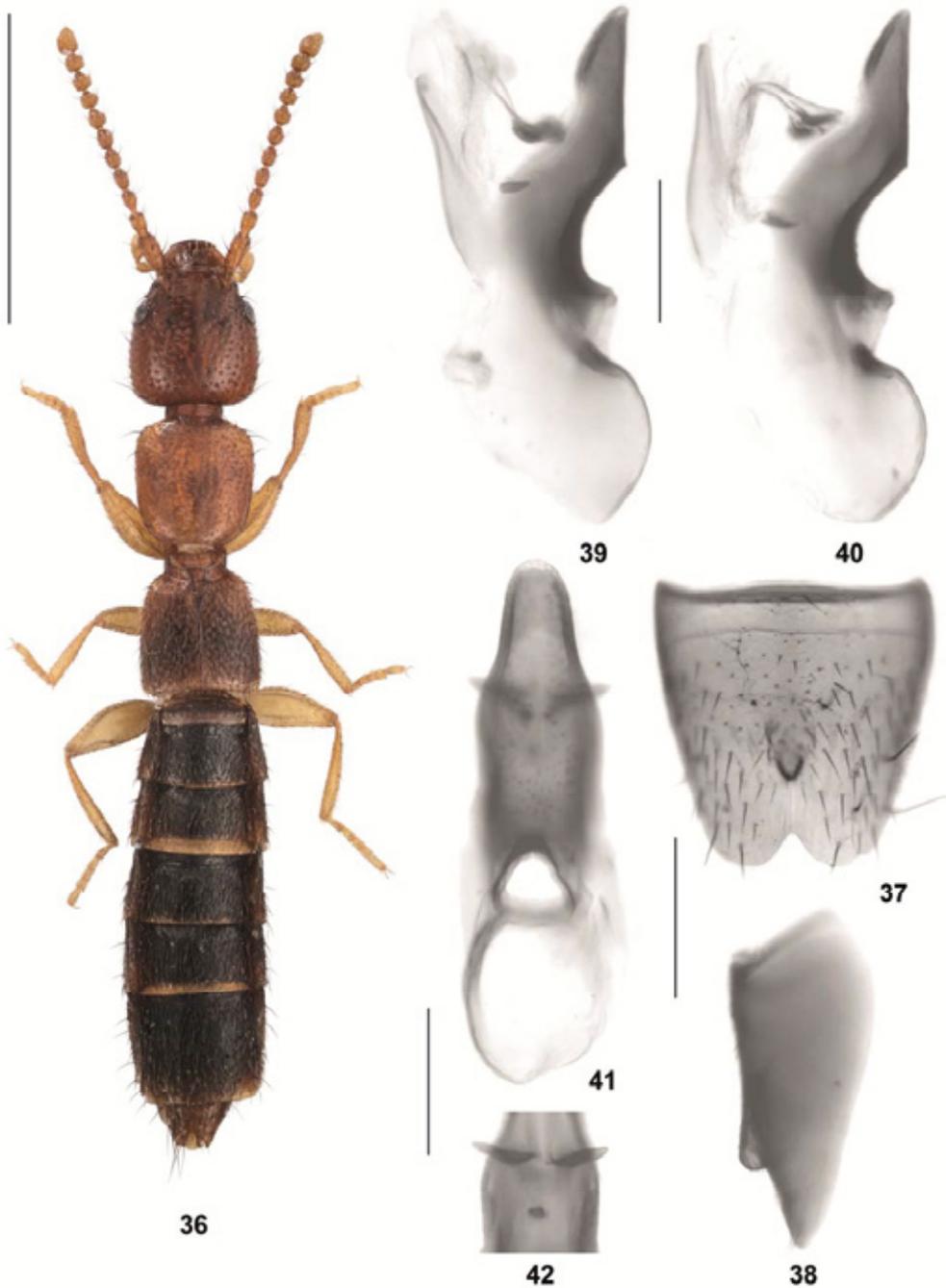
♂: antennomeres I–II and legs distinctly modified (see above); aedeagus (Fig. 45) 0.55 mm long (total length); internal sac with five apically curved sclerotized spines.

♀: unknown.

COMPARATIVE NOTES: *Bryaxis lesbius* is distinguished from the similar *B. theanus* (REITTER, 1894) (Macedonia, northwestern Turkey) and *B. mohamedis* (REITTER, 1903) (northwestern Turkey) by larger body size (*B. theanus* and *B. mohamedis*: 1.5–1.7 mm), more distinctly oblong antennomeres I and II, and by the internal structures of the aedeagus.



Figs. 23–35: *Aloconota lesbia* (23–29) and *A. samia* (30–35): 23, 30) forebody; 24, 31) antenna; 25, 32) male tergite VII in lateral view; 26–27, 33) male tergite VIII; 28–29, 34–35) median lobe of aedeagus in lateral and in ventral view. Scale bars: 23, 30: 0.5 mm; 24, 26–27, 31, 33: 0.2 mm; 25, 28–29, 32, 34–35: 0.1 mm.



Figs. 36–42: *Sunius potti*: 36) habitus; 37) male sternite VIII in ventral view; 38) male sternite VIII in lateral view; 39–41) aedeagus in lateral and in ventral view; 42) internal structures of aedeagus in ventral view. Scale bars: 36: 1.0 mm; 37–38: 0.2 mm; 39–42: 0.1 mm. Habitus photo: H. Schillhammer.



Fig. 43: Type locality of *Sunius potti* (sample number 7). An undescribed species of *Anillina* (Carabidae) was collected in this locality too.

DISTRIBUTION AND NATURAL HISTORY: The type locality is situated in Oros Olympos in the south of Lesbos, where the species is most likely endemic. The holotype was sifted from litter of *Platanus* and *Rubus* at an altitude of 790 m.

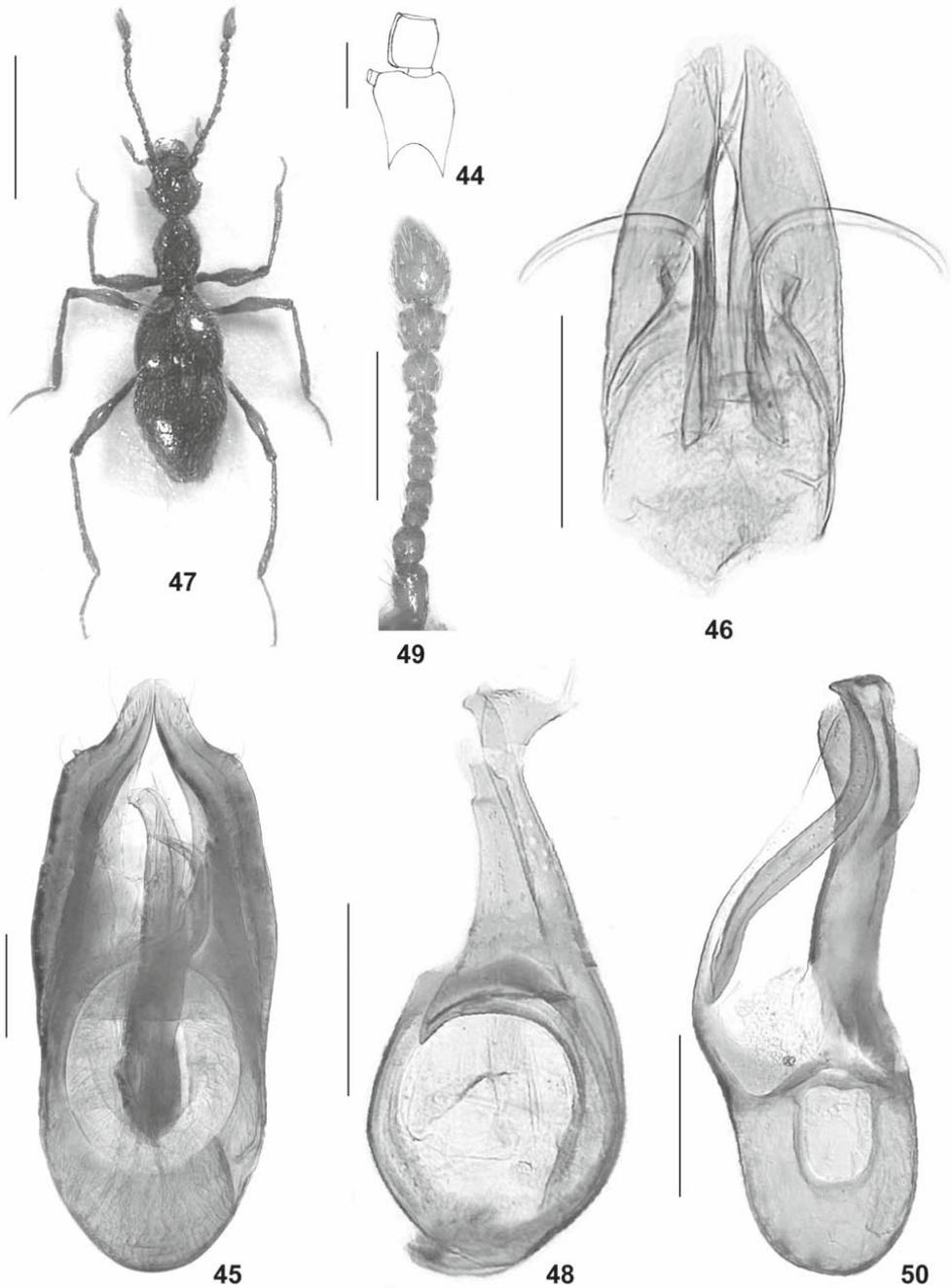
***Bythinus simplicipalpis* BRACHAT sp.n. (Fig. 46)**

TYPE MATERIAL: **Holotype** ♂: “N39°04'11 E26°20'58, GR Lesbos, 20.3.2005, Olympos 790m (10), Lompe & Meybohm / Holotypus ♂ *Bythinus simplicipalpis* sp. n. det. V. Brachat 2016” (cBra).

ETYMOLOGY: The specific epithet (adjective) alludes to the unmodified maxillary palpomere IV.

DESCRIPTION: Body length 1.43 mm. Body dark reddish-brown, with long and suberect yellowish pubescence; legs, antennae, and maxillary palpi pale-brown.

Head indistinctly transverse, 0.28 mm long and 0.29 mm broad; vertex with median keel anteriorly extending to an imaginary line between the anterior margins of the eyes; gular pits deep, anterior margin interrupted in the middle. Eyes large, distinctly projecting from lateral contours of head, and composed of about 28 ommatidia. Antennae 0.55 mm long; antennomeres I and II unmodified, I 0.09 mm long and 0.06 mm broad, II 0.07 mm long and 0.05 mm broad, III–VIII approximately as long as broad, IX–XI forming a club and gradually increasing in width, XI elongate and approximately as long as the combined length of VI–X. Maxillary palpomere III with scattered tubercles, palpomere IV large, 0.19 mm long and 0.08 mm broad, without distinct dorsal tubercle (unique in the genus), and with short erect pubescence.



Figs. 44–50: *Bryaxis lesbius* (44–45), *Bythinus simplicipalpis* (46), *Protamaurops assingi* (47–48), and *Tychus lesbius* (49–50): 44) male antennomeres I–II; 45–46) aedeagus in dorsal view; 47) habitus; 48, 50) aedeagus in ventral view; 49) male antenna. Scale bars: 47: 1.0 mm; 49: 0.25 mm; 44–46, 48, 50: 0.1 mm.

Pronotum broader than head, 0.33 mm long and 0.35 mm broad, without microsculpture.

Elytra 0.55 mm long and 0.64 mm broad (combined), with coarse punctation; humeral angles rounded. Hind wings present. Legs long; protibia weakly indented and with small tooth in apical fourth; metatibia with apical spine.

♂: pro- and metatibiae modified (see above); gular pits pronounced; aedeagus (Fig. 46) 0.26 mm long (total length); internal structures symmetrical, on either side with two basally connected branches on either side, with the inner branches directed towards, and nearly extending to, the apices of the parameres, the lateral branches abruptly bent laterad and extending far beyond lateral margins of parameres.

♀: unknown.

COMPARATIVE NOTES: *Bythinus simplicipalpis* is distinguished from the similar *B. icariensis* BESUCHET, 1964 (Greece: Ikaria) and *B. parvipenis* MEGGIOLARO, 1966 (Turkey, Cyprus) by the absence of a distinct dorsal tubercle on the maxillary palpomere IV and the internal structures of the aedeagus. For illustrations of the aedeagus of *B. icariensis* and *B. parvipenis* see BESUCHET (1964) and MEGGIOLARO (1966), respectively.

DISTRIBUTION AND NATURAL HISTORY: The holotype was collected together with that of *Bryaxis lesbius* (see above).

***Protamaurops assingi* BRACHAT sp.n. (Figs. 47–48, 51)**

TYPE MATERIAL: **Holotype** ♂: "GREECE: Lesbos [24a], W Kalloni, Prof. Ilias, 39°12'29"N, 26°06'01"E, 790 m, 23.III.2016, V. Assing & A. Hetzel / Holotypus ♂ *Protamaurops assingi* sp. n. det. V. Brachat 2016" (cBra).

ETYMOLOGY: The specific epithet is dedicated to the collector of the holotype.

DESCRIPTION: Body length 2.40 mm. Habitus as in Fig. 47. Body glossy, pale reddish-brown, with long, suberect (forebody) or depressed (abdomen) yellowish pubescence.

Head oblong, 0.50 mm long and 0.40 mm broad (width not including lateral spines); dorsal surface with distinct scattered punctation; antennal insertions prominent, extending into a sharp longitudinal keel; median keel on vertex anteriorly extending to an imaginary line connecting the two lateral spines. Eyes completely reduced, with distinct lateral spine. Antennae 1.32 mm long; all antennomeres distinctly oblong; antennomeres II–VIII approximately twice as long as broad, IX–XI forming a club; IX 0.05 mm long and 0.03 mm broad, X 0.45 mm long and 0.03 mm broad, and XI 0.12 mm long and 0.05 mm broad.

Pronotum 0.48 mm long and 0.38 mm broad, with scattered fine punctation; dorsal keels pronounced, near base of pronotum with an acute tooth; area between the keels strongly impressed, near the basal teeth with pit-like impression; near posterior and anterior margins with a short median keel.

Elytra strongly convex in cross-section, strongly tapering towards base, 0.62 mm long and 0.70 mm broad (combined), with scattered and extremely fine punctation; at base with two broad pits and a short humeral fold; humeral angles obsolete. Hind wings completely reduced. Legs very long; pro- and metatibiae weakly curved.

Abdomen: tergite IV large, longer than the combined length of the remaining tergites, at base with three densely pubescent impressions and with two parallel keels separated from each other by approximately half the width of tergite and extending for approximately one-fourth of the length of tergite; lateral keels of tergite IV parallel to lateral margins and extending along anterior three-fourths of tergite.

♂: metatrochanter with a minute tooth; sternite VIII with pronounced impression; aedeagus (Fig. 48) 0.3 mm long (total length); right paramere (ventral view) pronounced, apically dilated, and with a bidentate process.

♀: unknown.

COMPARATIVE NOTES: *Protamaurops assingi* is distinguished from the similar *P. macedonicus* (MÜLLER, 1934) (northern Greece) and *P. dentatithorax* (PIC, 1900) (Turkey: Bursa) by the morphology of the aedeagus, from *P. macedonicus* additionally by the presence of a small tooth at the base of the dorsal keels of the pronotum (*P. macedonicus*: only an angular elevation present) and from *P. dentatithorax* by much longer antennae (*P. dentatithorax*: antennae 0.42–0.44 mm long). For an illustration of the aedeagus of *P. dentatithorax* (as *Schweigeria uludagensis* LÖBL, 1967) see LÖBL (1967).

DISTRIBUTION AND NATURAL HISTORY: This species is most likely endemic to Lesbos. The holotype was collected at the peak of Profitis Ilias in the Ordimnos range in the west of the island by sifting litter in a stand of shrubs with scattered juniper and fern undergrowth at an altitude of 790 m (Fig. 51).



Fig. 51: Type locality of *Protamaurops assingi* (sample number 24a).

Tychus lesbius BRACHAT sp.n. (Figs. 49–50)

TYPE MATERIAL: **Holotype** ♂: “GREECE: Lesbos [35], Olympos, 39°04'11"N, 26°20'54"E, 810 m, road margin, 27.III.2016, V. Assing & A. Hetzel / Holotypus ♂ *Tychus lesbius* sp. n. det. V. Brachat 2016” (cBra). **Paratype** ♀: “GREECE: Lesbos [34], Olympos, 39°04'08"N, 26°20'58"E, 800–820 m, N-slope, 27.III.2016, V. Assing & A. Hetzel” (cBra).

ETYMOLOGY: The specific epithet is an adjective (Latin) derived from Lesbos.

DESCRIPTION: Body length 1.34–1.40 mm. Body pale-brown and glossy with sparse pubescence and scattered longer erect setae.

Head weakly oblong, 0.26 mm long and 0.24 mm broad; antennal insertions separated by narrow furrow; laterally with a small tooth anterior to eyes; frons between eyes with a pair of small pits; postocular region rounded and with dense and short pubescence. Eyes sexually dimorphic. Antenna (Fig. 49) 0.70 mm long; antennomeres I twice as long as broad, II distinctly oblong, III–VIII narrower than II and approximately as long as broad, IX–XI forming a club and increasing in width, IX and XI approximately as long as broad, and XI slightly longer than the combined length of IX and X. Maxillary palpomere IV very short, less than twice as long as broad (length 0.175 mm; width 0.100 mm).

Pronotum 0.28 mm long and 0.30–0.32 mm broad, distinctly broader than head, at posterior margin with a large median impression, two large lateral impressions, and a series of indistinct puncture-like impressions.

Elytra 0.36 mm long and 0.52–0.54 mm broad, with scattered fine punctation; each elytron with two basal pits; discal striae extending to middle of elytra; humeral angles weakly pronounced. Hind wings completely reduced. Legs long; tibia unmodified; mesotrochanter anteriorly with a distinct spine in both sexes.

Abdomen: tergite IV much longer than the following tergites, with two very short medio-basal keels separated from each other by one-fourth of the width of tergite, area between these keels weakly impressed and with short pubescence; sternites not distinctive.

♂: eyes larger and composed of 16 ommatidia; tergite VII broadly and weakly impressed, at base with a small median cluster of diverging setae partly concealed by the posterior margin of tergite VI; aedeagus (Fig. 50) 0.375 mm long (total length).

♀: eyes smaller and composed of five ommatidia; metatibia with a long and thin apical spine.

COMPARATIVE NOTES: *Tychus lesbius* is characterized by several unique characters (very short maxillary palpomere IV; very short elytra; modifications of the male abdominal tergites) at the same time obscuring possible affiliations to any of the previously established species groups. The species may be related to *T. brevipennis* (SAHLBERG, 1908) from Lebanon and numerous allied unnamed species distributed in the southeastern Mediterranean, which partly have modified male abdominal tergites. For an illustration of the aedeagus of *T. brevipennis* see BESUCHET (1960).

DISTRIBUTION AND NATURAL HISTORY: The type specimens were collected in two close localities situated in Oros Olympos at an altitude of approximately 800 m, the holotype from under a stone on a grassy road margin and the paratype by sifting roots of grass and herbs, as well as litter of *Platanus* and *Quercus*.

Stenichnus (Stenichnus) lesbius MEYBOHM sp.n. (Figs. 52–55)

TYPE MATERIAL: **Holotype** ♂: “GREECE: Lesbos [17], Oros Lepetimnos, 39°21'02"N, 26°17'16"E, 610 m, 21.III.2016, V. Assing & A. Hetzel / *Stenichnus lesbius* m. Meybohm 2016 det / Holotypus” (cMey). **Paratypes:** 2 ♀ ♀: “GR Lesbos Lepetimnos 570 m (3) N39°21'11 E26°17'37 18.3.2005 Lompe & Meybohm” (cMey); 2 ♂ ♂: “GR Lesbos Lepetimnos 800 m (4), N39°20'47 E26°16'24 18.3.2005 Lompe & Meybohm” (cMey); 1 ♀: “GR Lesbos Olympos 790 m (10a) N39°04'11 E26°20'58 24.3.2005 Lompe & Meybohm” (cMey); 1 ♂: “GR Lesbos Pigi 90 m (13) N39°10'58 E26°26'05 21.3.2005 Lompe & Meybohm” (cAss); 1 ♀: “GR Lesbos Lepetimnos 300 m (14a) N39°21'38 E26°16'43 23.3.2005 Lompe & Meybohm” (cMey); 1 ♂: “GR Lesbos Skalohori 360 m (17) N39°15'35 E26°04'03 22.3.2005 Lompe & Meybohm” (cMey); 1 ♂: “GR Lesbos Argenos 470 m (19) N39°10'58 E26°26'05 23.3.2005 Lompe & Meybohm” (cMey); 1 ♀: “GR Lesbos Lepetimnos 300 m (14a) N39°21'38 E26°16'43

23.3.2005 Lompe & Meybohm" (cMey); 1 ♀: "GREECE: Lesbos [1], 7 km ESE Mithimna, 39°21'23"N, 26°15'18"E, 440 m, 18.III.2016, V. Assing & A. Hetzel" (cMey).

ETYMOLOGY: The specific epithet is an adjective (Latin) derived from Lesbos.

DESCRIPTION: Male habitus as in Fig. 52. Body length 1.35–1.45 mm. Body weakly bicolored; head brown to blackish brown; pronotum brown to blackish-brown or black; elytra black; legs, antennae, and palpi pale-brown.

Head (Fig. 53) broadest across the large and moderately projecting eyes; width of head including eyes 0.27–0.29 mm; length of head measured from front of frons to occipital constriction 0.19–0.21 mm; strongly transverse (including eyes), 1.4 times as broad as long and 0.8 times as broad as pronotum. Eyes composed of approximately 20 ommatidia. Antenna slender, 0.75 mm long, antennomeres I–VII oblong, up to twice as long as broad, VIII about as long as broad, IX and X moderately transverse, XI distinctly shorter than the combined length of IX and X.

Female. Postocular region in dorsal view distinctly tapering behind eyes, as long as diameter of the eyes; vertex flattened, transverse, anteriorly confluent with frons; frons between eyes flattened, supra-antennal tubercles weakly pronounced. Punctures on vertex and frons very fine, interstices glossy; vertex and frons between eyes with long and curved suberect setae, which are as long as antennomere II and directed anteriorly; a longer erect seta near posterior margin on either side; setae in lateral portions of head directed upwards; anterior subtriangular part of frons without punctures or setae.

Male (Fig. 53). Vertex behind eyes and anterior part of frons bumpy, without punctures and setae, glossy; vertex and frons between the bumps with punctation as distinct as that of elytra, with setae thicker, less curved, and less erect than in female.

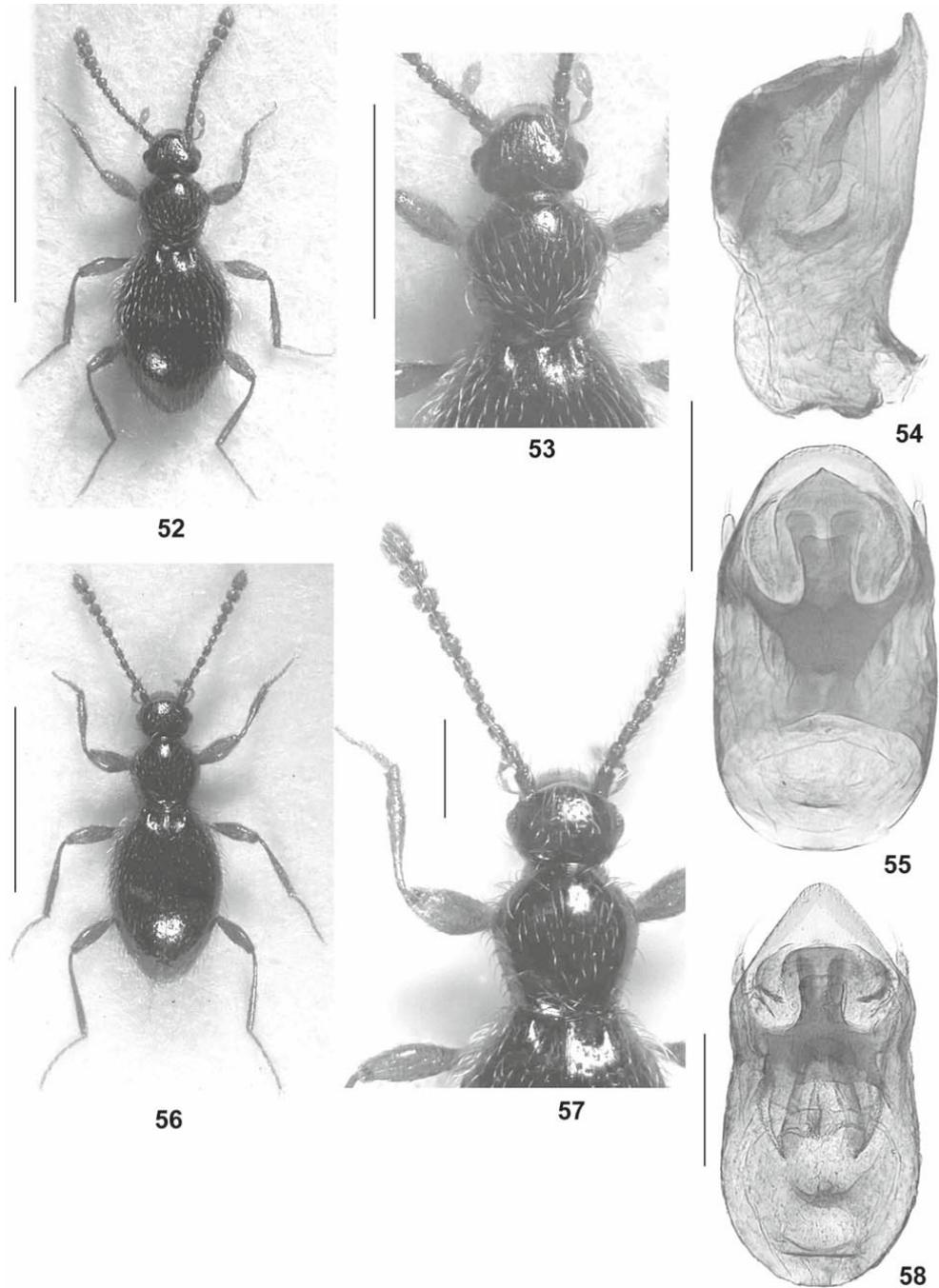
Pronotum of sub-trapezoid shape in dorsal view, weakly oblong, 0.35–0.37 mm long and 0.34–0.36 mm broad, broadest at anterior third; anterior margin weakly rounded, lateral margins strongly rounded in anterior third and straight in posterior half; posterior angles broadly rounded; punctures very fine; pubescence long and suberect; setae at anterior margin directed towards the middle of anterior margin, in lateral portions of posterior half directed medially, otherwise directed posteriorly; basally only with two more or less minute pits separated from each other by the same distance as from the posterior margin.

Elytra broadly oval, dorsally flattened, 0.84–0.87 mm long and 0.55–0.59 mm broad, broadest at anterior third; height approximately one-third of elytral length (lateral view); humeral angle or fold absent; at base of each elytron with a large and deep inner and a small outer impression separated by a longitudinal ridge; pubescence long and suberect, longest setae approximately as long as the combined length of antennomeres III and IV; punctation of elytra distinct, moderately fine. Elytral apices truncate.

Legs relatively long and slender, with moderately dilated femora; profemora without sexual dimorphism, only slightly more dilated than meso- and metafemora.

Aedeagus (Figs. 54–55) 0.24 mm long and 0.13 mm broad; sclerotized portion of dorsal aspect relatively long; dorsal plate longer than broad, apically emarginate; ventral plate much broader than dorsal plate, apically obtusely pointed; ventral lobe with broadly rounded apex, parameres each with two apical setae.

COMPARATIVE NOTES: This species is distinguished from similar and closely related congeners by the chaetotaxy of the head (setae long and directed anteriorly on vertex and frons), the male modifications of the head, and the morphology of the aedeagus. *Stenichnus* species with a sexually dimorphic head have been recorded from Chios (see Appendix) and Karpathos (MEYBOHM 2016), but are unknown from western Anatolia.



Figs. 52–58: *Stenichnus lesbius* (52–55) and *Stenichnus chius* (56–58): 52, 56) male habitus; 53, 57) male head and pronotum; 54) aedeagus in lateral view; 55, 58) aedeagus in dorsal view. Scale bars: 52, 56: 1.0 mm; 53: 0.5 mm; 57: 0.25 mm; 54–55, 58: 0.1 mm.

DISTRIBUTION AND NATURAL HISTORY: *Stenichnus lesbius* is endemic to Lesbos. The specimens were collected from under stones and by sifting litter in a pasture and on the grassy margin of a track at altitudes of 90–800 m.

Appendix

Ocalea brachyptera FAGEL, 1971

The two males from Chios listed as *Ocalea* sp. in ASSING (2015b) belong to *O. brachyptera* FAGEL, 1971, a species previously known only from the type locality (Turkey: Bursa: Ulu Dağ). Thus, these specimens represent the first record from Greece.

Aloconota (Aloconota) samia ASSING sp.n. (Figs. 30–35)

TYPE MATERIAL: **Holotype** ♂: “Greece: Samos [2a+1], SW Karlovasi, Oros Kerkis: NE-slope, 670 m, 37°44'14"N, 26°39'04"E, 27.III.2014, V. Assing / Holotypus ♂ *Aloconota samia* sp. n. det. V. Assing 2016” (cAss).

ETYMOLOGY: The specific epithet (Latin, adjective) alludes to the fact that this species is currently known only from Samos.

DESCRIPTION: Body length 2.8 mm; length of forebody 1.35 mm. Coloration: body black, with the elytra indistinctly paler; legs dark-yellowish; antennae dark-brown.

Head (Fig. 30) weakly transverse, 1.05 times as broad as long, of subquadrate shape; punctation extremely fine, barely noticeable; interstices with distinct microreticulation composed of isodiametric meshes and subdued shine. Eyes large, slightly longer than postocular region in lateral view; genal carina visible only in posterior portion of head (lateral view). Antenna (Fig. 31) 0.95 mm long, not particularly slender; antennomeres IV and V weakly oblong, VI approximately as broad as long, and VII–X weakly transverse; antennomere XI approximately as long as the combined length of antennomeres IX and X.

Pronotum (Fig. 30) moderately convex in cross-section, approximately 1.12 as broad as long and 1.12 times as broad as head, broadest in anterior half; pubescence of midline probably directed anteriad (somewhat disturbed in the holotype); punctation extremely fine, visible only at high magnification, and moderately dense; interstices with distinct microreticulation composed of isodiametric meshes and with subdued shine.

Elytra (Fig. 30) approximately 1.05 times as long as pronotum; punctation extremely dense, fine, but more distinct than that of head and pronotum; microsculpture pronounced; surface nearly matt. Hind wings fully developed. Metatarsomere I slightly shorter than the combined length of metatarsomeres II and III.

Abdomen narrower than pronotum; punctation dense and distinct on tergites III–VI, sparse and fine on tergites VII–VIII; microsculpture of tergites III–VI composed of transverse meshes, that of tergite VII of isodiametric and very short transverse meshes; tergites VII and VIII with pronounced sexual dimorphism; posterior margin of tergite VII with palisade fringe.

♂: tergite VII (Fig. 32) with small postero-medial tubercle; tergite VIII (Fig. 33) with six distinct teeth at posterior margin, the external ones longer than the four median ones; sternite VIII transverse and with strongly convex posterior margin; median lobe of aedeagus 0.36 mm long and shaped as in Figs. 34–35; ventral process slightly bent dorsad in lateral view.

♀: unknown.

COMPARATIVE NOTES: In external characters, this species is practically identical to *A. lesbia*, from which it is distinguished by the smaller tubercle on the male tergite VII, the

completely different shape of the male tergite VIII (less convex and with six teeth), and by the larger median lobe of the aedeagus with a ventral process of slightly different shape (lateral view).

For illustrations of numerous other *Aloconota* species distributed in the Mediterranean see TRONQUET (2014).

DISTRIBUTION AND NATURAL HISTORY: The type locality is situated on the northeastern slope of Oros Kerkis. The holotype was collected from gravel at the bank of a temporary stream at an altitude of 670 m.

***Stenichnus (Stenichnus) chius* MEYBOHM sp.n.** (Figs. 56–58)

TYPE MATERIAL: **Holotype** ♂: “GREECE – Chios [2], Oros Pelinnee: W-slope 38°33'16"N, 25°59'31"E, 700 m, sifted, 22.XII.2014, V. Assing / *Stenichnus chius* m. Meybohm 2016 det / Holotypus” (cMey).

ETYMOLOGY: The specific epithet is an adjective (Latin) derived from Chios.

DESCRIPTION: Male habitus as in Fig. 56. Body length 1.53 mm. Body weakly bicolored; head brown, pronotum blackish-brown, elytra black; legs, antennae, and palpi light brown.

Head broadest across the large and moderately projecting eyes; width of head including eyes 0.30 mm, length of head (measured from front of frons to occipital constriction) 0.23 mm, strongly transverse (width measured across eyes), 1.4 times as broad as long, 0.8 times as broad as pronotum. Eyes composed of approximately 20 ommatidia. Antenna slender, 0.80 mm long; antennomeres I–VII elongate, up to twice as long as broad, VIII about as long as broad, IX and X moderately transverse, XI distinctly shorter than the combined length IX and X.

Head (Fig. 57): postocular region distinctly tapering behind eyes in dorsal view, as long as diameter of the eyes; vertex flattened, transverse, anteriorly confluent with frons; frons between eyes flattened, supra-antennal tubercles nearly obsolete; punctures on vertex distinct and as large as those on elytra; punctures on frons distinct, but finer than those on elytra; cuticle glossy; vertex with long curved and suberect setae, which are as long as setae on pronotum and directed anteriorly; frons with short suberect straight setae directed anteriorly; setae near the posterior margin directed towards the middle of posterior margin; setae in lateral portions of head directed upwards; anterior subtriangular portion of frons without punctures or setae.

Pronotum subtrapezoidal in dorsal view, weakly oblong, 0.39 mm long and 0.38 mm broad, broadest at anterior third; anterior margin weakly rounded, lateral margins in anterior third strongly rounded, in posterior half straight, posterior angles broadly rounded; punctures very fine; pubescence long and suberect; setae directed medially in lateral portion of posterior half, otherwise directed posteriorly; basally only with two minute pits separated from each other by the same distance as from posterior margin.

Elytra broadly oval, dorsally flattened, 0.91 mm long and 0.60 mm broad, broadest at anterior third, height approximately one-third of elytral length (lateral view); humeral angle and humeral fold absent; at base of each elytron with a large deep inner and a small outer impression separated by a longitudinal ridge; pubescence long and suberect, longest setae approximately as long as the combined length of antennomeres III–IV; punctuation on elytra distinct and moderately fine; elytral apices truncate.

Legs relatively long and slender, with moderately dilated femora; profemora only slightly more dilated than meso- and metafemora.

Aedeagus (Fig. 58) 0.32 mm long and 0.16 mm broad; sclerotized portion of dorsal aspect relatively short; dorsal plate as long as broad, apically emarginate; ventral plate much broader

than dorsal plate, apically weakly convex; ventral lobe with straight sides, only apex rounded; parameres each with two apical setae.

♀: unknown.

COMPARATIVE NOTES: *Stenichnus chius* is most similar to *S. lesbius*. Both species share a similar chaetotaxy of the head (setae on vertex and frons long and directed anteriorly). *Stenichnus chius* is distinguished from *S. lesbius* by greater body size, the modifications of the male head, and by the morphology of the aedeagus. The shape of the male head in *S. chius* is similar to that of the female head in *S. lesbius*, while the setae on vertex and frons are longer and more erect than in both sexes of *S. lesbius*.

DISTRIBUTION AND NATURAL HISTORY: *Stenichnus chius* is most likely endemic to Chios. The holotype was collected in Oros Pelinneeon in the north of the island by sifting litter and grass roots on a rocky slope with scattered oak trees at an altitude of 700 m.

Acknowledgements

Several colleagues identified taxa or confirmed identifications: Volker Brachat (Pselaphinae), Heinrich Meybohm (Scydmaeninae), Michael Schülke, Berlin (*Anotylus complanatus*, *A. tetracarinatus*, *A. pumilus*, *Mycetoporus ignidorsum*, *M. imperialis*, *Tachyporus abner*, *T. pusillus*), Volker Puthz, Schlitz (*Stenus hospes*), Johannes Frisch, Berlin (*Scopaeus* spp.), Heinrich Terlutter, Münster (*Ocalea* spp.), Vladimir Gusarov, Oslo (*Mocyta* spp.) and György Makranczy, Budapest (small samples of *Ochtheophilus* spp.). Jürgen Vogel (Görlitz) reviewed the descriptions of *Aloconota* spp. Volker Brachat and Heinrich Meybohm communicated additional data on Pselaphinae and Scydmaeninae, respectively, from a field trip to Lesbos in 2005 and provided additional information on the distributions of the individual species of these subfamilies. Harald Schillhammer, Wien, provided the habitus photo of *Sunius potti*. Benedikt Feldmann (Münster) proof-read the manuscript.

Zusammenfassung

Bei Freilanduntersuchungen auf der griechischen Insel Lesbos im Frühjahr 2016 wurden insgesamt 1640 Individuen aus 169 Staphyliniden-Arten nachgewiesen. Neun Arten werden beschrieben und abgebildet: *Myllaena lesbia* ASSING sp.n., *Aloconota (Aloconota) aegaea* ASSING sp.n. (auch auf Samos verbreitet) und *A. (A.) lesbia* ASSING sp.n. aus der Unterfamilie Aleocharinae, *Sunius potti* ASSING sp.n. (Paederinae), *Bryaxis lesbius* BRACHAT sp.n., *Bythinus simplicipalpis* BRACHAT sp.n., *Protamaurops assingi* BRACHAT sp.n. und *Tychus lesbius* BRACHAT sp.n. aus der Unterfamilie Pselaphinae sowie *Stenichnus (Stenichnus) lesbius* MEYBOHM sp.n. (Scydmaeninae). Zumindest die neuen Arten der Paederinae, Pselaphinae und Scydmaeninae sind mit hoher Wahrscheinlichkeit auf Lesbos endemisch. Der bisher als Endemit von Lesbos betrachtete *Quedius henroti* COIFFAIT, 1970 kommt nach derzeitigem Kenntnisstand auf Lesbos nicht vor, sondern ist auf der Peloponnes verbreitet. Eine Gesamtliste der derzeit von Lesbos bekannten beschriebenen und unbeschriebenen Arten wird erstellt. Zahlreiche beschriebene Arten werden erstmals von Lesbos gemeldet, darunter auch acht Erstnachweise für Griechenland. Die derzeit bekannte Staphylinidenfauna der Insel umfasst 199 Arten, 184 benannte und 15 unbeschriebene oder nicht sicher identifizierte. Bei neun (5 %) der beschriebenen Arten und mindestens zwei der unbeschriebenen Scydmaeninae handelt es sich höchstwahrscheinlich um Inselendemiten. Trotz insgesamt höherer Diversität und allgemein ähnlicher Zusammensetzung enthält die Fauna von Lesbos weniger Inselendemiten als die der nahen und deutlich kleineren Insel Samos. Erwartungsgemäß sind deutliche Beziehungen der Fauna von Lesbos zum benachbarten türkischen Festland erkennbar. Einige Arten erreichen auf Lesbos

(und teilweise auch auf Chios und Samos) ihre westliche Verbreitungsgrenze. Insgesamt sechs myrmekophile Arten wurden von Lesbos nachgewiesen, fünf davon bei *Messor* spp. und eine bei *Cataglyphis nodus* (BRULLÉ, 1833). In einem Appendix werden *Aloconota* (*Aloconota*) *samia* ASSING sp.n. (Aleocharinae) von Samos und *Stenichnus* (*Stenichnus*) *chius* MEYBOHM sp.n. (Scydmaeninae) von Chios beschrieben. *Ocalea brachyptera* FAGEL, 1971 wird von Chios und somit zum ersten Mal aus Griechenland gemeldet.

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Dr. Volker ASSING

Gabelsbergerstr. 2, D – 30163 Hannover, Germany (vassing.hann@t-online.de)

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