New species and records of Staphylinidae from Greece, with two new synonymies (Insecta: Coleoptera)

V. Assing

Abstract: Based on a study of mostly recently collected material from Greece, ten species of Staphylinidae are (re-)described and illustrated: *Aphaenostemmus rhodicus* sp.n., *Xantholinus* (*Helicophallus*) *moreanus* sp.n., *X.* (*Xantholinus?*) *creticus* sp.n., *Tectusa recta* sp.n., *Cousya dimorpha* sp.n., *Oxypoda* (*Bessopora*) *cretica* sp.n., *O. (B.) afimbriata* sp.n., *O. (B.) praecisa* sp.n., *O. (B.) longicollis* Bernhauer, and *Aleochara* (*Ceranota*) *mortuisimilis* sp.n. In addition, the previously unknown male sexual characters of *Lathrobium vitsiense* Assing and those of the holotype of *L. anophthalmum* Fauvel are described and illustrated. *Gyrohypnus liber* Assing is formally revalidated. More than 150 species are listed, among them at least 82 first records from Greece, in addition to the new species described in this paper. 15 species reported from Greece before 2002, but omitted from the recent Palaearctic catalogue are listed. The following synonymies are established: *Oxypoda bimaculata* Baudidel 1870 = *O. corcyrica* Scheerpelz 1958, syn.n.; *Haploglossa villosula* (Stephens 1832) = *Aleochara hellenica* Pace 2005, syn.n. A lectotype is designated for *Oxypoda longicollis* Bernhauer, a species to be deleted from the list of Greek Staphylinidae. The distributions of five species are mapped.

Keywords: Coleoptera, Staphylinidae, *Aphaenostemmus*, *Xantholinus*, *Gyrohypnus*, *Tectusa*, *Cousya*, *Lathrobium*, *Oxypoda*, *Haploglossa*, Palaearctic region, Greece, Ukraine, taxonomy, new species, new records, new synonymy.

1. Introduction

In the recent past, the Greek staphylinid fauna has been addressed in several taxonomic and faunistic articles (e.g. Assing 1999, 2002, 2004a, 2005a, 2005b, 2005g, Assing & Schülke 2000, Assing & Wunderle 1999, 2001, Zerche 2002). However, the continuous discovery of undescribed species and of species that had previously been unknown from the region indicates that the currently known species inventory is still far from complete. Thus, it was not surprising that material collected during a short-term field trip to several mountain ranges in northern Greece in May 2005 and during several previous field trips yielded various new species and numerous records of zoogeographic interest.

The second part of the paper deals with new species and records based on an examination of material collected on previous occasions or seen from other collections. The recently published Palaearctic catalogue (Löbl & Smetana 2004), though somewhat incomplete
regarding faunistic data, nevertheless represents a useful tool for compiling an inventory of Greek Staphylinidae. A comparison of the unpublished results of several field trips to Greece with the respective entries in the catalogue revealed that apparently even some widespread and common species had not been reported from Greek territory before.

2. Material and measurements

The material referred to in this study is deposited in the following public institutions and private collections:

- DEI.................. Deutsches Entomologisches Institut, Müncheberg (L. Behne, L. Zerche)
- FMNH.............. Field Museum of Natural History, Chicago (J. Boone)
- MNHNP........... Muséum national d’Histoire naturelle, Paris (A. Taghavian)
- MNHUB........... Museum für Naturkunde der Humboldt-Universität Berlin (J. Frisch)
- NHMW ............ Naturhistorisches Museum Wien (H. Schillhammer)
- OÖLL............... Oberösterreichisches Landesmuseum Linz
- SMNS............... Staatliches Museum für Naturkunde (W. Schwaller, K. Wolf-Schwenninger)
- cAss.............. author’s private collection
- cFel .............. private collection B. Feldmann, Münster
- cSch.............. private collection M. Schülke, Berlin
- cWun............... private collection P. Wunderle, Mönchengladbach

Head length and elytral length were measured from the anterior margin of the clypeus to the posterior margin of the head and from the apex of the scutellum to the elytral hind margin, respectively.

The maps were generated using the online generic mapping tool (GMT) of the Geomar website at www.aquarius.geomar.de/omc.

3. Results of a field to northern Greece in spring 2005

The Staphylinidae collected by Paul Wunderle and the author at montane and alpine elevations in the Oros Vitsi, Oros Varnous, Oros Timfi, and the Oros Gramos range in spring 2005 altogether amounted to more than 1700 specimens, the vast majority of which were identified at species level (Tab. 1). In some cases, however, a sufficiently reliable identification was not possible, either because the respective genera or species groups are in a state of taxonomic confusion and require revision (e. g. some subgenera and species groups of *Atheta*), or because only females were available (e. g. *Eusphalerum* sp., *Xantholinus* sp.). Surprisingly, more than 20 species – some of them rather common in many parts of Europe – are not listed for Greece by Löbl & Smetana (2004) and apparently represent first records from Greek territory (Tab. 1). Those records that deserve special attention are referred to in more detail below. Also, if available, additional material seen from other parts of Greece and collected on other occasions is listed. The description of *Geostiba varnousica* and a discussion of the records of other *Geostiba* species have been dealt with elsewhere (Assing 2005h).
**Eusphalerum limbatum limbatum** *(ERICHSON 1840)*

In the Balkans, this subspecies was previously known only from Bosnia-Herzegovina and Yugoslavia (LÖBL & SMETANA 2004); it is here recorded from Greece for the first time.

**Eusphalerum zerchei** ZANETTI 1992

The species was described from Macedonia and is here reported from Greece for the first time.

**Phyllodrepa melanocephala** *(FABRICIUS 1787)*

According to LÖBL & SMETANA (2004), *P. melanocephala* was previously unknown from Greece.

**Omalium excavatum** STEPHENS 1834

LÖBL & SMETANA (2004) do not list this species for Greece, but it was reported from there by ASSING & WUNDERLE (2001).

**Coryphium atratum** BREIT 1911

According to ZERCHE (1990), the species has a trans-Adriatic distribution. For the Balkans, he lists records only from Croatia and Bosnia-Herzegovina. Hence, the specimen from the Oros Arénes, which was compared with male type specimens (NHMW), represents the first record from Greece. The locality where the Greek specimen was collected is illustrated in Fig. 32.

**Scopaeus illyricus** FRISCH 1997

Previously, only the three type specimens from Albania and Corfu had become known (FRISCH 1997).

**Lathrobium vitsiense** ASSING 2004 *(Figs 1-7)*

The specimen listed in Tab. 1 was collected at the type locality. When I described this species based on a single female – despite the fact that most *Lathrobium* species can be separated only by the male sexual characters – I did so, because *L. vitsiense* can be distinguished from all other subanophthalmous congeners occurring in Greece by the completely reduced eyes alone and because I did not expect additional material to become available in the near future. In contrast to the female holotype, which was found under a stone in a beech forest, the male listed in Tab. 1 was collected by sifting grass roots and moss near rocks. The male primary and secondary sexual characters are as follows:

♀: sternite VII with weakly concave posterior margin and with weakly delimited patches of dark modified setae (Fig. 6); sternite VIII as in Fig. 7; median lobe of aedeagus highly distinctive, without large sclerotised spines in internal sac, apex of ventral process almost spear-shaped (Figs 1-5).

For comparison, the – completely different – aedeagus of the holotype of *Lathrobium
anophthalmum FAUVEL 1885 from Serbia, another Balkans species with completely reduced eyes, is illustrated in Figs 8-9.

Figs 1-9: Lathrobium vitsiense ASSING (1-7) and L. anophthalmum FAUVEL, holotype (8-9). (1, 8) - aedeagus in lateral view; (2) aedeagus in ventral view; (3, 9) aedeagus in dorsal view; (4) internal structures of aedeagus (dorsal view); (5) apical part of ventral process of aedeagus; (6) ♂ sternite VII; (7) ♀ sternite VIII. Scale bars: 1-4, 6-9: 0.5 mm; 5: 0.2 mm.

Gyrohypnus liber ASSING 2003

The species was described only recently (ASSING 2003a) and is currently known only
from Greece. Without even seeing any material of this species, let alone types, and without presenting any further evidence, BORDONI (2005) expresses the opinion that *G. liber* is conspecific with *G. atratus* (HEER 1839). This approach does not meet basic requirements in scientific taxonomy (and, unfortunately, also contributes to nomenclatural instability), so that *G. liber* is here formally revalidated.

**Philonthus cognatus** STEPHENS 1832

Additional material examined: 1 ex., Thessalia, Ossa Oros, NE Spilia, near refuge, 39°48N, 22°41E, 1450 m, N-slope with scattered *Abies*, grass and ferns, 22.VII.2004, leg. Assing (cAss); 11 exs., Makhedonia, Olympos, above Refuge A, 40°05N, 22°22E, 2460 m, grassy cirque, under stones, 25.VII.2004, leg. Assing (cAss).

**Heterothops dissimilis** (GRAVENHORST 1802)

Additional material examined: 1 ex., Makhedonia, Vermion, above Seli, 1500 m, mixed forest, sifted, 11.IV.1998, leg. Assing (cAss); 8 exs., Makedhonía, Kavála, Pangéo, peak region, 1900 m, 28.V.1999, leg. Assing (cAss).

**Quedius bernhaueri** RAMBOUSEK 1915

This remarkable endogean *Quedius* species was recently recorded from Greece for the first time (ASSING & WUNDERLE 2001) and again by SOLODOVNIKOV (2005). Some of the specimens listed in Tab. 1 are slightly teneral.

**Quedius persimilis** MULSANT & REY 1876

Additional material examined: 1 ex., Fthiotis, ca. 20 km SSE Lamia, Oros Kallidromo, 38°44N, 22°32E, 1250 m, N-slope, under stones on meadow, 7.IV.2001, leg. Assing (cAss); 1 ex., NW-Evvoia, Oros Lichas, 38°51N, 22°54E, 525 m, 8.IV.2001, leg. Assing (cAss); 3 exs., Evvoia, Oros Dirfs, N Kato Steni, SE peak, 38°37N, 23°52E, 1120 m, 8.IV.2001, leg. Assing (cAss).

**Quedius pseudopyrenaeus** COIFFAIT 1967

LÖBL & SMETANA (2004) do not list the species for Greece, but it was reported from there by ASSING & WUNDERLE (2001).

**Leptusa jelineki** PACE 1983

*Leptusa jelineki* was only recently recorded from Greece for the first time by ASSING & WUNDERLE (2001), who reported the species from the Oros Vermio and the Oros Vitsi. Subsequently, additional records from northern Greece (Verno, Smolikas, Piéria) were published by ZERCHE (2002), suggesting that the species is rather common and widespread in montane beech forests of the region.

**Leptusa winneguthiana** PACE 1983

The only previous Greek record of this species is from the same mountain range (Varvous), but from a different locality (ASSING & WUNDERLE 2001).
**Leptusa meschniggi** Bernhauer 1935

The type locality is situated in the "Perister", a synonym of the Oros Varnous, which crosses the border between Macedonia and Greece; so, it is unknown if the types were collected in the Macedonian or the Greek part of the mountain range. The specimens listed in Tab. 1 were compared with the holotype (NHMW) and represent the first confirmation of the presence of the species in Greece. The holotype has a pronounced median carina on the abdominal tergite VII; this carina, however, is missing in the illustration of the holotype in PACE (1989: fig. 452). The only other published locality of *L. meschniggi* is "Sandanaki" in western Bulgaria (PACE 1989).

**Leptusa peristerica** PACE 1989

The species was for the first time recorded from Greece by ASSING & WUNDERLE (2001), who collected it in the Oros Varnous. The specimens listed in Tab. 1 were found in the same mountain range, but in a different locality. ZERCHE (2002) reported it from the Vérno, not far from the southernmost parts of the Oros Varnous.

**Atheta orosana** SCHEERPFLTZ 1931

The species was previously known only from the Greek island Levkas (SCHEERPFLTZ 1931). The specimen listed in Tab. 1 was identified by J. Vogel (Görlitz).

**Atheta laevicauda** SAHLBERG 1876

The species is not uncommon in montane and alpine habitats, especially in the Alps, but according to LÖBL & SMETANA (2004), it had not been recorded from Greek territory.

**Pella funesta** (GRAVENHORST 1806)

Additional material examined: 4 exs., Makedhonía, Kavála, Pangéo, road from Akrovouni to ski resort, oak forest, 500 m, with *Lasius fuliginosus*, 28.V.1999, leg. Assing (cAss).

**Pella humeralis** (GRAVENHORST 1802)

Additional material examined: 1 ex., Makedhonía, Sérrai, Vrontous, above Lailias, 1800 m, 25.V.1999, leg. Assing (cAss); 3 exs., same data, but 1750 m, beech forest (cAss).

**Cousya dimorpha** sp. n. (Figs 10-21)

Holotype, ♀: GR. [1] Flórina, 20 km SSW Flórina, Oros Vitsi, N-slope, 1850-1900 m, 40°38'32N, 21°22'46E, 22.V.2005, V. Assing / Holotypus ♀ *Cousya dimorpha* sp. n. det. V. Assing 2005 (cAss). Paratypes: 7♂ ♀, 10♂♀: same data as holotype (OÖLL, cAss); 4♂ 4♀: same data, but leg. P. Wunderle (cWun).

**Description:** Very small species, 1.9-2.4 mm (abdomen fully extended). Coloration: forebody uniformly dark brown to blackish; abdomen blackish with the apex slightly paler; legs brown; antennae dark brown, with the basal two antennomeres often slightly paler. Habitus as in Figs 10-11.

Head approximately as wide as long; eyes moderately large and protruding from lateral outline of head, slightly shorter than postocular region in dorsal view; microsculpture distinct; punctuation very fine and sparse, barely noticeable. Mouthparts as in Fig. 12.

Antenna with antennomeres I and II of subequal length and about twice as long as wide;
III much shorter than II and only slightly longer than wide; IV-X of increasing width and increasingly transverse; IV almost 1.5 times as wide as long; X approximately twice as wide as long; XI about 1.5 times as long as wide and slightly longer than the combined length of IX and X.

Pronotum transverse, approximately 1.25 times as wide as long and 1.25 times as wide as head; maximal width approximately in the middle; posterior angles weakly marked; microsculpture and punctuation similar to those of head; pubescence along midline directed posteriad.

Elytra with microsculpture and punctuation more pronounced than those of head and pronotum, surface matt. Width and length of elytra polymorphic. Macropterous specimens (Fig. 10); elytra 1.3-1.4 times as wide and at suture approximately 1.1 times as long as pronotum; hind wings fully developed. Brachypterous specimens (Fig. 11); elytra about 1.2-1.3 times as wide and at suture 1.0-1.1 times as long as pronotum. Hind wing development dimorphic, either fully developed or of reduced length and only slightly longer than elytra.

Abdomen subparallel, in macropterous specimens distinctly (0.8-0.9 x) narrower than elytra, in brachypterous specimens approximately as wide as elytra; tergites III-V with shallow anterior transverse impression; tergite VI without such impression; microsculpture distinct; punctuation fine and sparse; posterior margin of tergite VII with palisade fringe in both morphs; tergite VIII without appreciable sexual dimorphism, its posterior margin weakly convex in both sexes.

♂: posterior margin of sternite VIII strongly convex and with long thin marginal setae, especially in the middle (Fig. 13); median lobe of aedeagus and apical lobe of paramere as in Figs 14-17.

♀: posterior margin of sternite VIII broadly convex, sometimes with shallow concavity in the middle, and with row of moderately stout modified marginal setae (Fig. 18); spermatheca as in Figs 19-21.

Etymology: The name (adjective) refers to the dimorphism of the wings.

Comparative notes and systematics: Cousya dimorpha is readily distinguished from its Western Palearctic congeners by its small size, the distinctly transverse pronotum, the wing dimorphism, the relatively short antennae and maxillary palpi, as well as by the morphology of the genitalia. In body shape, as well as in the morphology of the mouthparts, the antennae, and the genitalia, C. dimorpha resembles species of the genus Zoosetha Mulsant & Rey, from which it is distinguished by the much finer punctuation of head and pronotum. However, the character constellation of this species raises the question if Cousya Mulsant & Rey and Zoosetha should really be considered distinct genera.

Distribution and bionomics: The species is known only from the peak of the Oros Vitsi, a mountain inhabited by several species with restricted distributions or even local endemics: Pareudectus vitsiensis Assing & Wunderle, an undescribed species of Coryphiodes Bernhauer, Lathrobium vitsiense, Quedius bernhaueri, Geostiba torisuturalis, and an undescribed species of Tectusa Bernhauer. The fact that C. dimorpha was not found on any of the neighbouring mountains suggests that the species may have a restricted distribution, too. The types were collected by sifting grass roots and moss on a north slope at an altitude of approximately 1900 m. Seven type specimens (1♂, 6♀) are macropterous, 22 (14♂, 8♀) are brachypterous.
Figs 10-21: Cousya dimorpha sp.n. (10) forebody of macropterous specimen; (11) forebody of brachypterous specimen; (12) mouthparts; (13) ♂ sternite VIII; (14-16) median lobe of aedeagus of three ♂ in lateral and in ventral view; (17) apical lobe of paramere; (18) ♀ sternite VIII; (19-21) spermathecae of three ♀. Scale bars: 10-11: 0.5 mm; 12-21: 0.1 mm.
**Tectusa recta** sp.n. (Figs 22-32)

Holotype ♂: GR. [12] N-Pindos, ca. 40 km NNE Konitsa, Oros Arênes, 1900 m, 40°17'39N, 20°55'22E, 27.V.2005, V. Assing / Holotypus ♂ *Tectusa recta* sp. n. det. V. Assing 2005 (cAss).

Paratypes: 2♂ 2♀: same data as holotype (cAss); 2♀ 2♀: same data, but leg. P. Wunderle (cWun).

**Description:** 2.5-3.0 mm (abdomen fully extended); habitus as in Fig. 22. Coloration: forebody uniformly blackish brown, with the abdominal apex slightly paler; legs reddish brown, with the femora slightly darker; antennae reddish brown, usually gradually darkened towards apex.

Head approximately as wide as long; eyes small, composed of less than 20 ommatidia, and weakly protruding from lateral outline of head, less than half the length of postocular region in dorsal view; microreticulation of integument distinct; puncturation very fine and sparse, barely noticeable (Fig. 23). Antenna with antennomeres I and II of subequal length and about twice as long as wide; III somewhat shorter than II and about 1.5 times as long as wide; IV weakly transverse; IV-X of increasing width; IX almost twice as wide as long; X less than IX and about 1.5 times as wide as long; XI of ovoid shape and shorter than the combined length of IX and X.

Pronotum distinctly convex in cross-section, transverse, approximately 1.1 times as wide as long, and about 1.15 times as wide as head; maximal width approximately in the middle; posterior angles weakly marked; midline without impression; microsculpture and puncturation similar to those of head; pubescence along midline directed posteriad, in anterior 1/6 of midline sometimes anteriad, and on either side of midline laterad (Fig. 23).

Elytra about as wide and at suture about 0.6 times as long as pronotum; microsculpture pronounced; puncturation fine, but somewhat more distinct than that of head and pronotum; pubescence short and more or less depressed (Fig. 23). Hind wings reduced.

Abdomen slightly wider than elytra, widest at segments V/VI (Fig. 22). Anterior impressions of tergites III-V moderately shallow, that of tergite VI very shallow; microsculpture distinct; puncturation fine and sparse; posterior margin of tergite VII without palpisde fringe; tergite VIII without appreciable sexual dimorphism.

♂: posterior margin of sternite VIII strongly convex and with relatively short thin marginal setae (Fig. 24); median lobe of aedeagus at base of ventral process with pronounced folds (see arrows in Figs 25-27), ventral process in lateral view straight, not curved (Figs 25-28).

♀: posterior margin of sternite VIII broadly and weakly convex, marginal setae not distinctly modified (Fig. 29); spermatheca as in Figs 30-31.

**Etymology:** The name (Lat., adjective) refers to the straight ventral process of the median lobe of the aedeagus.

**Comparative notes:** Greece is inhabited by numerous endemic species of *Tectusa*, most of which, however, are undescribed. Among the described species, the geographically closest congeners are *T. vrontousensis* Assing & Wunderle from the Vrontous range near Séres and *T. meschniggi* (Bernhauer) from the Olympos range. From all known species of *Tectusa*, the new species is separated especially by the distinctive shape of the median lobe of the aedeagus. From *T. vrontousensis*, it is additionally distinguished by its distinctly smaller eyes (in *T. vrontousensis* with more than 30 ommatidia), the lighter coloration of the legs and antennae, the much finer and more...
Figs 22-31: *Tectusa recta* sp.n.: (22) habitus (holotype); (23) forebody (paratype); (24) ♀ sternite VIII; (25-27) median lobe of aedeagus in lateral and in ventral view (26-27: holotype); (28) apical part of median lobe of aedeagus in lateral view; (29) ♂ sternite VIII; (30-31) spermatheca. Scale bars: 22: 1.0 mm; 23: 0.5 mm; 24, 29: 0.2 mm; 25-28, 30-31: 0.1 mm.
indistinct puncturation of the forebody, the absence of a median impression on the pronotum, the shallower anterior impression of the abdominal tergite VI, as well as by the much longer duct of the spermatheca. From *T. meschniggi*, it is also separated by the finer puncturation and the darker coloration of the forebody, by the darker femora, the more convex pronotum, by the posteriorly more strongly convex male sternite VIII, and by the much longer spermathecal duct. For illustrations of the genitalia of *T. vrontousensis* and *T. meschniggi* see Assing & Wunderle (2001) and Assing (2004a), respectively.

**Distribution and bionomics:** The type locality is situated in the Oros Arénes, which is part of the Oros Grámos range, very close to the Albanian border. The specimens were sifted from grass roots, moss, and debris close to the edge of snowfields at an altitude of 1900 m (Fig. 32).

---

**Oxypoda vittata Märkel 1842**

Additional material examined: 22 exs., Makedhónia, Kavála, Pangéo, road from Akrovouni to ski resort, oak forest, 500 m, with *Lasius fuliginosus*, 28.V.1999, leg. Assing (cAss); 1 ex., Fthiotis, ca. 20 km SSE Lamia, Oros Kalidromo, 38°45N, 22°28E, 940 m, 2.IV.2001, leg. Assing (cAss); 1 ex., Pelopónnisos, Agios Petros, Vourvouni, 920 m, *Quercus ilex* forest, 22.III.1997, leg. Assing (cAss).

**Oxypoda (Bessopora) moreatica Scheerpeltz 1962**

Additional material examined: Mainland Greece: 8 exs., Fthiotis, Oros Íti, W-slope, 38°49N, 22°14E, 1450 m, *Abies* forest, 10.IV.2001, leg. Assing, Wunderle (cAss, cWun); 1 ex., Fthiotis, SSE Lamia, Oros Kalidromo, 38°44N, 22°31E, 1250 m, 7.IV.2001, leg. Assing (cAss); 1 ex., Fthiotis, Parnassos Oros, ski resort, 38°33N, 22°35E, 1760 m, *Abies* wood, 15.IV.2000, leg. Assing (cAss); 1 ex., Parnassos, Lilea, 20.V.1997, leg. Wolf (cSch). Pelopónnisos: 3 exs., Panahaiko, above Ano Kastritsi, 38°15N, 21°52E, 1500 m, 28.III.1997, leg. Assing (cAss); 5 exs., 10 km E Kalavrita,
Aroania range, 38°01N, 22°10E, 1450 m, 29.III.1997, leg. Assing, Wunderle (cAss, cWun); 1 ex., Killini range, S Trikalon, 37°59N, 22°27E, 1300 m, 31.III.1997, leg. Assing (cAss); 2 exs., Killini, Ano Trikala, 1450 m, pine litter, 7.VI.1996, leg. Wunderle (cWun); 2 exs., Erimanthos, above Kalendzi, 37°57N, 21°47E, 1500 m, 27.III.1997, leg. Assing (cAss); 1 ex., Erimanthos, above Kalendzi, 37°57N, 21°47E, 1200 m, 27.III.1997, leg. Assing (cAss); 1 ex., Erimanthos, Kalendzi, 1350 m, Abies forest, 27.III.1997, leg. Wunderle (cWun); 5 exs., 20 km S Derveni, Sarandapaho, 38°02N, 22°23E, 1250 m, 3.IV.1997, leg. Assing (cAss); 1 ex., Parnon range, 1100-1450 m, Pinus, Abies, 13.VI.1996, leg. Wunderle (cWun).

Comments: The species had not been recorded since its original description, which is based on type material from the Pelopónnisos.

Aleochara maculata Brisout de Barneville 1863

In the Balkans, A. maculata was previously recorded from Croatia, Bosnia-Herzegovina, and Bulgaria (Löbl & Smetana 2004). The specimens from the Oros Vitsi (Tab. 1) represent the first record from Greece.

Aleochara gridellii Bernhauer 1936

The species was previously known only from Macedonia and Albania. It is here recorded from Greece for the first time. The specimen listed in Tab. 1 was compared with type material from the Bernhauer collection.

Tab. 1: Results of the 2005 field-trip to northern Greece (leg. P. Wunderle & V. Assing). r: species marked with a "*" are listed for Greece neither in the Palaearctic catalogue (Löbl & Smetana 2004), nor by Assing & Wunderle (2001), and probably first records from Greece. Sample data:

<table>
<thead>
<tr>
<th>Species</th>
<th>sample no.</th>
<th>r</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
<th>11</th>
<th>12</th>
</tr>
</thead>
<tbody>
<tr>
<td>Proteinus brachypterus (Fabricius 1792)</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Eusphalerum sp.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eusphalerum limbatum limbatum (Erichson 1840)</td>
<td></td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Eusphalerum zerchei Zanetti 1992</td>
<td></td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Phyllodrepa ioptera (Stephens 1834)</td>
<td></td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
</tr>
</tbody>
</table>

Species marked with a "*" are listed for Greece neither in the Palaearctic catalogue (Löbl & Smetana 2004), nor by Assing & Wunderle (2001), and probably first records from Greece.
<table>
<thead>
<tr>
<th>Species \ sample no.</th>
<th>r</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
<th>11</th>
<th>12</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phyllodrepa melanocephala (FABRICIUS 1787)</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Omalium excavatum STEPHENS 1834</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>5</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Omalium cinnamonum KRAATZ 1857</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Omalium caesum GRAVENHORST 1806</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anthobium atrocephalum (GYLENHAL 1827)</td>
<td></td>
<td></td>
<td>6</td>
<td>7</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anthobium melanosecephalum (ILLIGER 1794)</td>
<td></td>
<td></td>
<td>2</td>
<td>1</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amphichromus canaliculatum (ERICHSON 1840)</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lesteva longoelytrata (GOEZE 1777)</td>
<td>4</td>
<td>100</td>
<td>127</td>
<td>37</td>
<td>13</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Coryphium atratum BREIT 1911</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stenus glacialis HEER 1839</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>4</td>
<td>5</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stenus cordatoides PUTZ 1972</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stenus subsaeneus ERICHSON 1840</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stenus ochropus KIESENWETTER 1858</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stenus coarcticollis drepanensis PUTZ 1980</td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>5</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stenus aceris STEPHENS 1833</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Paederus schoenherri CZWALINA 1889</td>
<td>1</td>
<td></td>
<td></td>
<td>3</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Astenus gracilis (PAYKULL 1789)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Astenus procerus GRAVENHORST 1806</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Scopaeus illyricus FRISCH 1997</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Scopaeus sp.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Medon brunneus (ERICHSON 1839)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>11</td>
<td>10</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lathrobium vitiense ASSING 2004</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gyrohypnus liber ASSING 2003</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Xantholinus laevigatus JACOBSEN, 1849</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Xantholinus cf. linearis (OLIVIER 1795)</td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Xantholinus sp.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Othius punctulatus (GOEZE 1777)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Othius lapidicola MARKEL &amp; KIESENWETTER 1848</td>
<td>8</td>
<td>1</td>
<td>2</td>
<td>11</td>
<td>10</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Philonthus decorus (GRAVENHORST 1802)</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Philonthus mannerheimi FAUVEL 1869</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Philonthus carbonarius GRAVENHORST 1802</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gabrias sp.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Heterothops dissimilis GRAVENHORST 1802</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius mesomelius MARSHAM 1802</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Species \ sample no.</td>
<td>r</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
</tr>
<tr>
<td>----------------------</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>----</td>
<td>----</td>
<td>----</td>
</tr>
<tr>
<td>Quedius bernhaueri Rambousek, 1915</td>
<td></td>
<td>1</td>
<td>6</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius cinctus (Paykull 1790)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius paradisianus (Heer 1839)</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>9</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius nemoralis Baudi 1848</td>
<td></td>
<td>7</td>
<td>8</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius pseudonigriceps Reitter 1909</td>
<td></td>
<td>2</td>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius limbatis (Heer 1839)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius pseudopyrenaeus Coiffait 1967</td>
<td></td>
<td>4</td>
<td>11</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quedius persimilis Mulsant &amp; Rey 1876</td>
<td>+ 16</td>
<td>1</td>
<td>2</td>
<td>6</td>
<td>1</td>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Habrocerus pisidicus Korge 1971</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>5</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Mycetoporus punctipennis Scriba 1868</td>
<td></td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mycetoporus dispersus Schülke &amp; Kocian 2000</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Mycetoporus rufescens Stephens 1832</td>
<td>+</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mycetoporus puncatus Gravenhorst 1806</td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mycetoporus himaculatus Lacordaire 1835</td>
<td>+</td>
<td></td>
<td></td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mycetoporus bosniacus Luze 1901</td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mycetoporus erichsonanus Fagel 1965</td>
<td></td>
<td>1</td>
<td>3</td>
<td>1</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tachysoma splendidum (Gravenhorst 1806)</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sepedophilus immaculatus (Stephens 1832)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>1</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tachyphorus nitidulus (Fabricius 1781)</td>
<td></td>
<td>2</td>
<td></td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tachyphorus scitulus Erichson 1839</td>
<td></td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tachinus bonvouloiri Pandelle 1869</td>
<td></td>
<td>9</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Holobus flavicornis (Lacordaire 1835)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Leptusa aspatica Eppelesheim 1888</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>2</td>
<td>1</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Leptusa ruficollis (Erichson 1839)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>6</td>
<td>1</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Leptusa jelinekii Pace 1983</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td>15</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Leptusa winneguthiana Pace 1983</td>
<td></td>
<td>9</td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Leptusa meschniggi Bernhauer 1935</td>
<td>+?</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>9</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Leptusa peristerica Pace 1989</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>41</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Callicerus rigidicornis Erichson 1839</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aloconota gregaria (Erichson 1839)</td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aloconota sulcifrons (Stephens 1832)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amischa analis (Gravenhorst 1802)</td>
<td></td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Geostiba armata (Eppelesheim 1878)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>101</td>
<td>278</td>
<td></td>
</tr>
<tr>
<td>Geostiba torisuturalis Assing 2000</td>
<td>100</td>
<td></td>
<td>28</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Species \ sample no.</td>
<td>r</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>5</td>
<td>6</td>
<td>7</td>
<td>8</td>
<td>9</td>
<td>10</td>
<td>11</td>
<td>12</td>
</tr>
<tr>
<td>---------------------</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>---</td>
<td>----</td>
<td>----</td>
<td>----</td>
</tr>
<tr>
<td>Geostiba varnousica</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>62</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Geostiba euboica</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>12</td>
<td>9</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Liogluta alpestris</td>
<td>14</td>
<td>1</td>
<td>3</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Liogluta longianulc (GRAVENHORST 1802)</td>
<td>1</td>
<td>6</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta (Microdota) sp.</td>
<td>9</td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta orosana SCHRERPELTZ 1931</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta (Paralpinia) sp.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta benickiella BRUNDIN 1948</td>
<td>16</td>
<td></td>
<td></td>
<td>4</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta bosonica GANGLBAUER 1895</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta paetida KRAATZ 1856</td>
<td>2</td>
<td>7</td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta laevicuda SAHLBERG 1876</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atheta (Mocyta) spp.</td>
<td>2</td>
<td></td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acrotonaetroglodytes MOTSCILSKY 1858</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alevonota egregia (RYE 1876)</td>
<td>+</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alevonota gracilenta (ERICHSON 1839)</td>
<td>+</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pella funesta (GRAVENHORST 1806)</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pella humeralis (GRAVENHORST 1802)</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tectusa recta sp.n.</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cousy dimorpha sp.n.</td>
<td>+</td>
<td>30</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cousy sp.</td>
<td></td>
<td>1</td>
<td>1</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxylea spp.</td>
<td>21</td>
<td>36</td>
<td>2</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxypoda viitata MÄRKL 1842</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxypoda brevicornis (STEPHENS 1832)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxypoda recondita KRAATZ 1856</td>
<td>8</td>
<td>4</td>
<td>3</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxypoda (Baeoglena) sp.</td>
<td>31</td>
<td>101</td>
<td>8</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxypoda moreatica SCHRERPELTZ 1962</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxypoda annularis MANNERHEIM 1830</td>
<td>2</td>
<td>2</td>
<td>1</td>
<td>8</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Homoessa sp.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aleochara maculata BRISOUT 1863</td>
<td>+</td>
<td>17</td>
<td>17</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aleochara laevigata GYLLENHAL 1810</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aleochara gridellii BERNHAUER 1936</td>
<td>+</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aleochara verna SAY 1833</td>
<td>+</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aleochara bipustulata (LINNAEUS 1760)</td>
<td>2</td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

© Biologiezentrum Linz/Austria; download unter www.biologiezentrum.at
4. Additional new records, new species, omissions in the Palaearctic catalogue, and a new synonymy

The records given below are based on material collected during five field trips to Greece in 1997, 1998, 1999, 2001, and 2004, as well as on material seen from other collections. A species list of a field trip in 2000 and part of the records of the 1999 and the 2004 excursions are given by Assing & Wunderle (2001), Assing & Wunderle (1999), and Assing (2004a), respectively. For records of species belonging to recently revised genera such as Medon Stephens, Leptothion Casey, Drusilla Leach, Geostiba Thomson, etc. see the respective revisions (e. g. Assing 1999, 2004b, 2005c, 2005d). Some groups (especially of the Tachyporinae and the Aleocharinae) will be dealt with in the course of future revisions. If not indicated otherwise, all the species listed below are not indicated for Greece by Löbl & Smetana (2004) and probably first records.

15 species had previously been recorded from Greece by Assing (2003b) and Assing & Wunderle (2001), but are not listed for this country in the Palaearctic catalogue: Micropeplus ripicola Kerstens 1964, Omalium excavatum Stephens 1834, Carpelimus subtilis (Erichson 1839), Ochthephilus angustior (Bernhauer 1943), O. rosenhaueri (Kiesenwetter 1850), O. venustulus (Rosenhauer 1856), Gabrius astutoides (Strand 1946), Quedius microps Gravenhorst 1847, Q. pseudopyrenaeus Coiffait 1967, Q. xanthopus Erichson 1839, Mycetoporus brucki (Pandelle 1869), M. forticornis Fauvel 1875, Sepedophilus binotatus (Gravenhorst 1802), Tachinus discoideus Erichson 1839, Poronnius procidua (Erichson 1837).

Proteinus crenulatus Pandelle 1867

Material examined: 2 exs., Makedonía, Pieria range, above Skotina, 900-1000 m, mixed forest, sifted, 9.IV.1998, leg. Assing (cAss); 1 ex., Pelopónnisos, Agios Petros, 1100m, oak forest with Erica arborea, 22.III.1997, leg. Assing (cAss).

Omalium riparium Mulsant & Rey 1861


Hypopycna rufula (Erichson 1840)

Material examined: 1 ex., Pelopónnisos, 7 km NE Lálas, 700 m, Quercus, 4.X.2004, leg. Schawaller (SMNS).

Xylostiba monilicornis (Gyllenhal 1810)

Material examined: 3 exs., Thessalia, Kato Olympos, E Kallipefki, 1300 m, under bark of Abies, 6.IV.1998, leg. Assing (cAss).

Hapalaraea pygmaea (Paykull 1800)

Material examined: 1 ex. (det. Wunderle), Corfu, Zigos env., 400 m, Quercus ilex forest with moss, sifted, 24.IX.1994, leg. Wunderle (cWun).
Anthobium unicolor (MARSHAM 1802)


Olophrum piceum (GYLLENHAL 1810)

Material examined: 14 exs., Thessalia, Stómio, ca. 10 m, flood plain forest, 5.IV.1998, leg. Assing (cAss).

Xylodromus affinis (GERHARDT 1877)

Material examined: 1 ex., Makhedonia, Kavála, Pangéo, near ski resort, 1700 m, beech forest, 24.V.1999, leg. Assing (cAss).

Lesteva albanica BERNHAUER 1936


Comment: The species was previously known from Albania, Bulgaria, and Cyprus, but had not been recorded from Greece.

Aphaenostemmus rhodicus sp.n. (Figs 60-63)


Description: 3.8-4.0 mm; habitus as in Fig. 60. Coloration: head and pronotum castaneous; elytra yellowish brown; abdomen dark brown to blackish brown; legs and antennae brown.

Head weakly transverse, approximately 1.05 times as wide as long; eyes large and bulging, about twice as long as postocular region in dorsal view; punctuation moderately coarse and dense, interstices in central dorsal area on average as wide as diameter of punctures, in lateral areas narrower; microsculpture absent (Fig. 62). Antenna as in Fig. 63.

Pronotum approximately 1.10 times as wide as head and 1.20 times as wide as long, maximal width a short distance anterior to middle; posterior angles obtusely marked; punctuation coarser and denser than that of head, interstices distinctly narrower than diameter of punctures; microsculpture absent. Hind wings fully developed.

Abdomen slightly wider than elytra; punctuation very fine, barely noticeable; integument with pronounced microreticulation rendering the surface almost completely matt; posterior margin of tergite VII with palisade fringe.

♂: unknown.

Etymology: The name (Lat.: adj.) is derived from Rhódos, where the type locality is situated.
Comparative notes: In the Palaearctic region sensu Löbl & Smetana (2004), the genus *Aphaenostemmus* Peyerimhoff 1914 previously included only five species (Herman 2001). Two species had become known from the Western Palaearctic: *A. bordei* Peyerimhoff 1914 (Algeria, Tunisia) and *A. testaceus* (Koch 1936) (Egypt, Saudi Arabia, Oman). From the former, *A. rhodicus* is separated by the lighter coloration of the head, pronotum, legs, and antennae, by slightly smaller size, by the relatively narrower and smaller pronotum, by the shorter and more slender antennae, and by the much denser puncturation of the head and pronotum. The habitus of *A. bordei* is illustrated in Assing (2005f). Based on the details specified by Koch (1936), the new species is separated from *A. testaceus*, of which I have seen no material, by the bicoloured body (in *A. testaceus* uniformly rufotestaceous), larger size, and by the denser puncturation of the forebody.

Distribution and biornomics: The species is known only from one locality situated in the Greek island Rhódos, off the southwest Anatolian coast. The types were apparently collected on a river bank; further bionomic data are not available.

*Thinodromus arcuatus* (Stephens 1834)

*Thinodromus distinctus* Fairmaire & Laboulbène 1856

*Carpelimus erichsoni* (Sharp 1871)

*Scopaeus portai* Luze 1910

*Platydomene springeri* (Koch 1987)
Material examined: 2 exs., Fthiotis, ca. 30 km SW Lamia, bank of Inachos river near Perivoli, 38°50N, 22°05E, 470 m, 5.IV.2001, leg. Assing (cAss).

*Stenistoderus turcicus* (Coiffait 1956)
Material examined: 1 ex., Pelopónnisos, N Kalávrita, bank of Vouraikos river, 680 m, 30.III.1997, leg. Assing (cAss).

*Nudobius cypriacus* Coiffait 1956

Comment: The species had been reported from Greece before, but is apparently very rare.
Xantholinus (Helicophallus) moreanus sp. n. (Figs 47-54, Map 1)

Holotype δ: GR. Pelopónnisos, 40 km SE Tripoli, SW Ag. Petros, 1160 m, 37°18'51N, 22°31'10E, 22.III.1997, V. Assing / Holotypus δ Xantholinus moreanus sp. n. det. V. Assing 2005 (cAss).


Description: Species of intermediate size, 8.0-9.2 mm (abdomen fully extended). Coloration variable, ranging uniformly rufous to distinctly bicoloured, with the head and abdomen blackish brown to blackish, and pronotum and elytra rufous; legs rufous; antennae with the basal three antennomeres rufous, antennomeres IV-X reddish brown, and XI apically testaceous.

Habitus of forebody as in Fig. 47-48. Head approximately 1.25 times as long as wide, weakly dilated posteriad; eyes very small, approximately 1/5 the length of postocular region; puncturation moderately sparse and relatively fine; microsculpture absent. Pronotum 0.9 times as wide as head and approximately 1.45 times as long as wide; dorsal rows of punctures composed of about 10 punctures; microsculpture absent. Elytra short, approximately 0.55 times as long as pronotum; puncturation relatively dense, rather coarse, and well-defined, interstices on either side of suture much narrower than diameter of punctures; microsculpture absent. Hind wings reduced; when fully unfolded, apices just protruding from under the elytra. Abdomen with fine and moderately sparse punctuation; microsculpture distinct, transverse everywhere; posterior margin of tergite VII without palisade fringe.

δ: posterior segments without conspicuous modifications. Aedeagus basally with two series of distinctly sclerotised spines, one with ca. 15 long spines, and one with more numerous smaller spines (Figs 49-53); long spines present also in anterior part of aedeagus (Fig. 54).

Etymology: The name (Lat., adj.) is derived from Morea, the Latin name of what is the Pelopónnisos today.

Comparative notes: The geographically closest species of the subgenus Helicophallus is X. bulgaricus COIFFAIT from the Bulgarian Stara Planina, which, based on the original description, as well as on an examination of the holotype from the Coiffait collection (MNHNP) and a non-type male from the Stara Planina, is distinguished from the new species by the distinctly microsculptured elytra with almost indistinct punctuation, by the less slender (shorter and relatively broader) head and pronotum, by the smaller aedeagus (0.75-0.80 times the length of that of X. moreanus), and by the shorter and less massive proximal spines in the aedeagus. For an illustration of the aedeagus of X. bulgaricus see figure 98A in COIFFAIT (1972).

Comment: KORGE (1973) reports X. bulgaricus also from the Parnon range in the
Pelopónnisos, stating he had found only negligible differences between the material from the Parnon and the original description of *X. bulgaricus*. As emphasised above, however, there are differences both in external morphology and in the internal structures of the aedeagus. In addition, there are other arguments suggesting that the populations from the Stara Planina and from the Pelopónnisos represent distinct species. First, aedeagal character divergence is generally low in the subgenus, so that striking differences like those in many other species groups of *Xantholinus* are not to be expected. Second, both *X. bulgaricus* and *X. moreanus* are micropterous and have small eyes, from which one can infer that dispersal power is low and the distributions are very likely to be restricted. Finally, despite much increased collecting activity in various parts of Greece in the past decade, no further records of *Helicophallus* have become known. Korge’s material was not examined, but it may hypothesised that it is conspecific with *X. moreanus*.

It was not possible to examine the aedeagus of the – evidently slightly teneral – holotype of *X. bulgaricus* ("Bulg. St.planina, Midzur, 2150-2168 m, 26.10.1968, P. Beron leg. / Préparation microscopique / Holotype / Xantholinus (Helicophallus) bulgaricus H. Coiffait 1971"), since it is apparently kept separately (see labels). The transverse microsculpture of the head emphasised in the original description is visible only in the lateral parts; the dorsal median area is free of microsculpture. Fortunately, a male from the Stara Planina in the collections of the DEI was available for examination, which confirmed that *X. bulgaricus* and *X. moreanus* are indeed different species. The collection data of this specimen are: Stara Planina, Etropolska Planina, Murgana peak, 42°42N, 24°03E, 1600-1630 m, 11.V.2001, leg. Zerche & Behne.

**Distribution and bionomics:** The species is known only from several localities in the Pelopónnisos (Map 1), where it is probably endemic, as can be inferred from the absence of records from other Greek localities, as well as from the reduced eyes and wings. It was collected in oak and mixed forests (oak, fir, maple, pine) at altitudes of 1100-1300 m.

**Map 1:** Distribution of *Xantholinus moreanus* sp.n. in southern Greece.
Xantholinus (Xantholinus?) creticus sp.n. (Figs 55-59)


Description: Relatively small species, 6.8 mm (abdomen fully extended). Coloration: head blackish; pronotum dark brown, with the anterior and posterior margins diffusely paler; elytra brown; abdomen dark brown with the tergal margins reddish; legs and antennae rufous.

Forebody as in Fig. 55. Head almost 1.3 times as long as wide, with subparallel lateral margins; eyes less than one third the length of postocular region; punctuation sparse and relatively fine; microsculpture absent. Pronotum 0.9 times as wide as head and approximately 1.45 times as long as wide; dorsal rows of punctures composed of 9-10 punctures; microsculpture absent. Elytra approximately 0.7 times as long as pronotum; punctuation relatively sparse and well-defined; microsculpture absent. Hind wings present, but possibly not fully developed. Abdomen with very fine and sparse punctuation; microsculpture distinct, on tergites III-VII transverse, on tergite VIII predominantly isodiametric; posterior margin of tergite VII with palisade fringe.

♂: posterior segments without conspicuous modifications. Aedeagus with membranous tube, two large black spines, and two series of rather long brownish spines (Figs 56-59).

Etymology: The name (Lat., adj.) refers to Crete, where the species may be endemic.

Comparative notes: Xantholinus creticus is distinguished from all other species of the genus by the internal structures of the aedeagus. In X. linearis (OLIVIER), which too has two long black spines in the internal sac, these spines are distinctly angled, the additional spines in the aedeagus are much shorter, and the head and pronotum have distinct transverse microsculpture.

Distribution and bionomics: The type locality, the Samaria valley, is located in the south of western Crete. Bionomic data are not available.

Xantholinus pantokratoris BORDONI 1975 (Map 2)

Material examined: Mainland: 1 ♂, Thessalia, Pilion, 5 km NE Vólós, Stagiate, 500-700 m, 15.V.1996, leg. Schulz & Vock (cAss); 2 ♀♂, Voiotia, Oros Elíkonas, Kiriaki, 38°23N, 22°50E, 900 m, 4.IV.2001, leg. Assing (cAss). Evvoia: 1 ♂, Oros Líchas, 38°52N, 22°53E, 620 m, 9.IV.2001, leg. Assing (cAss). Kérkira (Corfu): 3 ♂♂, Kérkira, 11.&.13.X.1987 (cAss); 1 ♂, Paleokastrítra, ruderal meadow, 14.X.1987 (cAss). Pelopónnisos: 1 ♂, Agios Mammás, 36°37N, 22°56E, 3.X.1980, leg. Malicky (cAss); 1 ♂, Olympia, 1.IV.1986, leg. Assing (cAss); 1 ♂, pass 25 km S Kalávrita, 1100 m, 5.X.2004, leg. Schawaller (cAss); 1 ♂, Sparta, stream bank, 2.IV.1986, leg. Assing (cAss); 1 ♂, Parnon, 4.5 km W Karies, 37°17N, 22°28E, 930 m, oak litter sifted, 19.IV.1999, leg. Behne (DEI).

Comment: The species was previously considered to be endemic to the Greek island Kérkira (=Corfu) (BORDONI 1975, 1990), but is apparently widespread in southern and western Greece (Map 2).
Neobisnius villosulus (Stephens 1833)

Material examined: 1 ex., Peloponnisos, N Kalavrita, bank of Vouraikos river, 680m, 30.III.1997, leg. Assing (cAss).

Cafius xantholoma (Gravenhorst 1806)

Material examined: 3 exs., Levkas, Sivota, beach debris, 27.IX.1993, leg. Assing (cAss); 8 exs., Peloponnisos, 10 km NE Githio, beach debris, 23.&26.III.1997, leg. Assing (cAss).

Remus filum (Kiesewetter 1849)


Remus sericeus Holme 1837

Material examined: 2 exs., Levkas, Sivota, beach debris, 27.IX.1993, leg. Assing (cAss); 3 exs., Peloponnisos, 10 km NE Githio, beach debris, 23.&26.III.1997, leg. Assing (cAss).
Gabrius exspectatus SMETANA 1952

Additional material examined: 2 exs., Fthiotis, ca. 20 km SSE Lamia, Oros Kallidromo, 38°45N, 22°32E, 1250 m, N-slope, under bark of *Abies*, 7.IV.2001, leg. Assing (cAss); 22 exs., Pelopónnisos, Aroania, E Káliávrita, 1450 m, *Abies* forest, 29.III.1997, leg. Assing (cAss); 6 exs., Pelopónnisos, E Chelmós, 2 km S Zarákhla, *Abies*, *Pinus*, 1100 m, 23.IX.2004, leg. Schawaller (SMNS, cAss); 4 exs., Chelmós, ski resort, 1800 m, leg. Bense (SMNS).

Comment: The species is not listed for Greece by LöBL & SMETANA (2004), but was reported from there by Assing & Wunderle (2001).

Quedius brevis ERICHSON 1840


Quedius nigrocaeruleus FAUVEL 1876

Material examined: 3 exs., Pelopónnisos, Chélmos, ski resort, *Abies* forest, 1600 m, 22.IX.2004, leg. Schawaller (SMNS); 6 exs., same data, but 1800 m, leg. Bense (SMNS, cAss); 5 exs., Pelopónnisos, Taygetos, Neohóri, 1100 m, *Platanus*, *Abies*, 30.IX.2004, leg. Bense (SMNS, cAss).

Comment: The species was previously known neither from Greece nor from adjacent countries (LÖBL & SMETANA 2004).

Quedius umbrinus ERICHSON 1839

Material examined: 1 ex., Flórina, 40 km NE Flórina, Oros Voras, 40°53N, 21°48E, 1650 m, 22.IV.2000, leg. Assing (cAss); 1 ex., Pella, 40 km NE Flórina, Oros Voras, 40°54N, 21°50E, 1950 m, 22.IV.2000, leg. Assing (cAss); 1 ex., Thessalia, Pílon Oros, 4 km SW Zagora, 39°25N, 23°05E, 450 m, 3.IV.1998, leg. Assing (cAss); 1 ex., Fthiótis, 40 km W Lamia, bank of Sperchios river, 38°56N, 21°55E, 16.IV.2000, leg. Assing (cAss); 11 exs., Pelopónnisos, Taygetos, Manganári source, 1000 m, *Abies*, *Pinus*, 29.IX.2004, leg. Schawaller (SMNS, cAss).

Comment: *Quedius umbrinus* had been reported from Greece before. Interestingly, the material from the Pelopónnisos is characterised by distinctly longer and larger elytra, as well as by darker coloration than material seen from other European regions.

Tachinus discoideus Erichson 1839

Additional material examined: 1 ex., Fthiotis, Oros Oxia, SW Paleochori, 38°50N, 21°56E, 1500 m, N-slope, 11.IV.2001, leg. Assing (cAss).

Comment: The species is not listed for Greece by LöBL & SMETANA (2004), but was reported from there by Assing & Wunderle (2001).

Gyrophaena joyioides WÜSTHOFF 1937

Material examined: 2 exs., Thessalia, Sóstio, ca. 10 m, flood plain forest, 5.IV.1998, leg. Assing (cAss).

Gyrophaena gentilis ERICHSON 1839

Material examined: 1 ex., Makedhonía, Pieria range, above Skotina, 900-1000 m, mixed forest, sifted, 9.IV.1998, leg. Assing (cAss).
Placusa complanata Erichson 1839

Material examined: 73 exs., Thessalia, Kato Olympos, E Kallipefki, 1300 m, under bark of Abies, 6.IV.1998, leg. Assing (cAss); 1 ex., Peloponnisos, E Chelmós, 2 km S Zarúkhla, 1100 m, 23.IX.2004, leg. Bense (SMNS).

Placusa tachyoporoides (Waltl 1838)

Material examined: 5 exs., Thessalia, Kato Olympos, E Kallipefki, 1300 m, under bark of Abies, 6.IV.1998, leg. Assing (cAss).

Tachysida gracilis (Erichson 1837)

Material examined: 6 exs., Makedhonía, Olympos, NW Stavros refuge, 1450 m, under pine bark, 7.IV.1998, leg. Assing (cAss).

Megaloscapa punctipennis (Kraatz 1856)

Material examined: 1 ex., Peloponnisos, Erimanthos, above Kalendzi, 1500 m, grass, moss, and leaf litter sifted, 27.III.1997, leg. Assing (cAss).

Phytosus balticus Kraatz 1859


Gnypeta rubrior Tottenham 1939

Material examined: 1 ex., Fthiotis, ca. 30 km SW Lamia, bank of Inachos river near Perivoli, 38°50N, 22°05E, 470 m, 5.& 10.X.1991, leg. Wunderle (cWun).

Trichiusa immigrata Lohse 1984

Material examined: 1 ex., Peloponnisos, N Kalávrita, bank of Vouraikos river, 680 m, 30.III.1997, leg. Assing (cAss).

Aloconota planifrons (Waterhouse 1863)

Material examined: 1 ex., Peloponnisos, N Kalávrita, bank of Vouraikos river, 680 m, 30.III.1997, leg. Assing (cAss).

Hydrosmecta longula (Heer 1839)


Amischa forcipata Mulsant & Rey 1873

Material examined: 1 ex., Peloponnisos, N Kalávrita, bank of Vouraikos river, 680 m, 30.III.1997, leg. Assing (cAss).
**Brundinia meridionalis** (Mulsant & Rey 1853)

**Material examined:** 18 exs., Thessalia, Stómio, coast laguna, 5.IV.1998, leg. Assing (cAss); 6 exs., Makedhonia, Kavála, 60 km W Kavála, beach of Ophrynia, laguna, 30.V.1999, leg. Assing (cAss); 1 ex., Thessalia, Stómio, ca. 10 m, flood plain forest, 5.IV.1998, leg. Assing (cAss); 1 ex., Pelopónnisos, 10 km NE Gíthio, beach debris, 23.&26.III.1997, leg. Assing (cAss).

**Liochlata granigera** (Kiesenwetter 1850)

**Material examined:** 1 ex., Makedhonia, NW Kavála, Pangró, road to ski resort, 1200 m, beech forest, 28.V.1999, leg. Wunderle (cWun); 1 ex., Makedhonia, N Drama, Falakró, road to ski resort, 1000 m, N-slope, 26.V.1999, leg. Wunderle (cWun).

**Enalodroma hepatica** (Erichson 1839)

**Material examined:** 1 ex., Makedhonia, Kavála, Pangró, peak region, 1900 m, 24.V.1999, leg. Assing (cAss); 2 exs., Pangró, near ski resort, 1700 m, beech forest, 24.V.1999, leg. Assing (cAss); 2 exs., same data, but 1650 m, 24.&28.V.1999, leg. Assing (cAss); 1 ex., Pangró, road from Akrovoúni to ski resort, beech forest, 1350 m, 24.V.1999, leg. Assing (cAss).

**Atheta cadaverina** (Brisout de Barneville 1860)

**Material examined:** 1 ex., Thessalia, Ossa Oros, NE Spilia, 39°49N, 22°40E, 1130 m, mixed Abies and Fagus forest, 22.VII.2004, leg. Assing (cAss).

**Atheta harwoodi** Williams 1930

**Material examined:** 1 ex., Pelopónnisos, northern Taygetos, E Perivolia, 1300 m, creek valley, 24.III.1997, leg. Assing (cAss); 1 ex., Pelopónnisos, SE Zákhiro, 6 km E Néa Figalia, 700 m, 2.X.2004, leg. Bense (SMNS).

**Atheta ischnocera** Thomson 1870

**Material examined:** 1 ex., Makedhonia, Dráma, Falakró, above ski resort, 1750-1850 m, 26.V.1999, leg. Assing (cAss).

**Atheta laticollis** Stephens 1832

**Material examined:** 1 ex., Pelopónnisos, Stímfalia, 800 m, lakeshore, 24.IX.2004, leg. Schawaller (SMNS).

**Atheta nigra** Kraatz 1856

**Material examined:** 1 ex., Pelopónnisos, 10 km NE Gíthio, beach debris, 23.&26.III.1997, leg. Assing (cAss); 1 ex. (det. J. Vogel), Crete, Lefka Ori, Mt. Psári, 1700 m, VI.2002, leg. Feldmann (cFel).

**Atheta nigritula** (Gravenhorst 1802)

**Material examined:** 1 ex., Thessalia, Ossa Oros, 6 km W Karitsa, 750 m, beech forest, 4.IV.1998, leg. Assing (cAss); 1 ex., Thessalia, Ossa Oros, E-slope, 1150-1200 m, beech forest, 8.IV.1998, leg. Assing (cAss).
**Atheta paracrassicornis** BUNDIN 1954

*Material examined:* 1 ex., Thessalia, Kato Olympos, E Kallipeftki, 1450-1500 m, beech and fir forest, sifted, 6.IV.1998, leg. Assing (cAss).

**Atheta episcopalis** BERNHAUERI 1910

*Material examined:* 1 ex. (det. J. Vogel), Ipiros, Peristeri range near Athohori, 1700-2200 m, 4.VI.2000, leg. Marggi (cFd).

**Atheta ravilla** (ERICHSON 1839)

*Material examined:* 1 ex., Makhedonía, Vermion, above Seli, 1500 m, mixed forest, sifted, 11.IV.1998, leg. Assing (cAss).

**Acrotona aterrima** (GRAVENHORST 1802)

*Material examined:* 1 ex., Makedhonía, Kavála, Pangéo, road from Akrovouni to ski resort, beech forest, 1350 m, 24.V.1999, leg. Assing (cAss).

**Acrotona benicki** ALLEN 1940

*Material examined:* 1 ex., Pelopónnisos, Erimanthos, above Kalendzi, 1200 m, Abies forest, 27.III.1997, leg. Assing (cAss).

**Acrotona obfuscata** (GRAVENHORST 1802)

*Material examined:* 1 ex., Fthiotis, ca. 30 km SW Lamia, bank of Inachos river near Perivoli, 38°50N, 22°05E, 470 m, 5.IV.2001, leg. Assing (cAss).

**Acrotona piceorufa** (MULSANT & REY 1873)

*Material examined:* 5 exs., Pelopónnisos, 30 km NE Pirgos, Koumani, oak forest, 630 m, 1.IV.1997, leg. Assing (cAss).

**Pella cognata** (MÄRKEL 1842)

*Material examined:* 2 exs., Thessalia, Sórmio, ca. 10 m, flood plain forest, 5.IV.1998, leg. Assing (cAss); 3 exs., Fokís, Oros Elikonas, road from Kiriaki to Elikonas, 38°23N, 22°49E, 900 m, N-slope, sifted Abies litter and under stones, 4.IV.2001, leg. Assing (cAss).

**Pella laticollis** (MÄRKEL 1845)

*Material examined:* 11 exs., Makedhonía, Kavála, Pangéo, road from Akrovouni to ski resort, oak forest, 500 m, with Lasius fuliginosus, 28.V.1999, leg. Assing (cAss).

**Dinarda dentata** (GRAVENHORST 1806)

*Material examined:* 3 exs., Fthiotis, Oros Iti, 38°49N, 22°14E, 1400 m, N-slope, subalpine meadow, 10.IV.2001, leg. Assing (cAss).
Phloeopora corticalis (GRAVENHORST 1802)

Material examined: 1 ex., Peloponnisos, 7 km SE Kalavrita, 1200 m, Abies, 21.IX.2004, leg. Bense (cAss); 1 ex., Peloponnisos, Chelmnos, ski resort, 1800 m, Abies, 22.IX.2004, leg. Bense (SMNS).

Apimela mulsanti (GANGLBAUER 1895)

Material examined: 1 ex., Fthiotis, ca. 30 km SW Lamia, bank of Inachos river near Perivoli, 38°50N, 22°05E, 470 m, 5.IV.2001, leg. Assing (cAss).

Oxypoda (Thliboptera) micans KRAATZ 1855

Material examined: Greece: 14 exs., Levkas, Vouno peak, 1050 m, 25.IX.1993, leg. Assing (cAss, cWun); 1 ex., Fthiotis, 30 km W Lamia, W Kalithea, 38°53N, 22°06E, 800 m, oak forest, 16.IV.2000, leg. Assing (cAss); 4 exs., N Larissa, Kato Olympos, above Goni, 39°54N, 22°27E, 550 m, roadside, nest of Messor sp., 6.IV.1998, leg. Assing (cAss); 7 exs., N Larissa, Kato Olympos, N Larissa, Kato Olympos, E Kallipefki, 39°58N, 22°29E, 1500-1580 m, 6.&12.IV.1998, leg. Assing (cAss, cWun); 1♀, Makedhonia, NW Kavála, Pangéo, near ski resort, 1650 m, 24.V.1999, leg. Assing (cAss); 1 ex., Peloponnisos, Ahaia, Agios Nikolaos, 4.IV.1999, leg. Wachtel (cAss); 2 exs., Ikaría, Néas, 37°37N, 26°03E, 10-100 m, sifted from grass under Mastix shrubs, 26.IV.2003, leg. Brachat & Meybohm (cAss).

Comments: The species was originally described from "Griechenland" (KRAATZ 1855), the locality is not specified. Recent records have not become known. Oxypoda micans is somewhat similar to, but not conspecific with O. referens MULSAN & REY from Corsica. The latter is of larger average body size, slightly paler coloration, and has shorter elytra. Most importantly, however, the median lobe of the aedeagus is of different morphology (shape of internal structures, smaller crista apicalis). These differences are not very pronounced, but constant.

The record from the Pangéo is somewhat doubtful, since it is based on a single female; males are required to confirm the presence of the species in northeastern Greece.

Oxypoda (Deropoda) bimaculata BAUDI DI SELVE 1870

Oxypoda bimaculata BAUDI DI SELVE 1870: 377.
Oxypoda (Oxypoda) corcyrica SCHEERPETZ 1958: 425 ff.; syn.n.

Type material examined: Holotype ♀: Ipsos, 16.IV.1932 / Insel Korfu, Dr. M. Beier leg. / Typus Oxypoda corcyrica O. Scheerpeltz / Oxypoda bimaculata Baudi det. V. Assing 2005 (NHMW).


Comments: The original description of Oxypoda corcyrica is explicitly based on a female holotype from "Ipsos auf der Insel Korfu" (SCHEERPETZ 1958), not from Levkas, as indicated by LöHL & SCHEERPETZ (2004); that of O. bimaculata is based on two syntypes from Cyprus (BAUDI DI SELVE 1870). An examination of the holotype of O. corcyrica revealed that it is conspecific with O. bimaculata, which is widespread in the Eastern Mediterranean. Material of this species was seen from Cyprus, Greece (see above), and from Turkey (ASSING 2004c, in press).
Oxypoda (Bessopora) cretica sp.n. (Figs 64-70)

Holotype ♂: N35°11', E25°31', GR Kreta Lassithi, Selia Afhin 1000 m, Meybohm 10.3.2001 / Holotypus ♂ Oxypoda cretica sp. n. det. V. Assing 2005 (cAss).

Description: 3.5 mm. Coloration: whole body pale reddish brown, with the anterior areas of abdominal tergites V-VI infuscated; legs and antennae yellowish brown. Habitus as in Fig. 64.

Head 1.2 times as wide as long; punctuation dense and distinct, interstices narrower than diameter of punctures and without microsculpture (Fig. 65); eyes moderately large (Fig. 66), slightly shorter than postocular region in dorsal view. Antennae relatively long and massive (Fig. 67), antennomeres 4-10 distinctly coniform, antennomere X approximately 1.5 times as wide as long, XI rather large. Preapical joint of maxillary palpus slightly more than twice as long as wide.

Pronotum more than 1.4 times as wide as head and more than 1.3 times as wide as long (Fig. 65); maximal width approximately in the middle; punctuation very dense, denser and less well-defined than that of head; microsculpture absent.

Elytra approximately as wide as pronotum and at suture 0.65 times as long as pronotum; near lateral margins shallowly impressed (Fig. 65); punctuation coarser than that of pronotum, very dense, asperate, and ill-defined; microsculpture absent. Hind wings reduced. Legs moderately slender; metatarsus almost as long as metatibia; metatarsomere I approximately as long as the combined length of metatarsomeres II-IV.

Abdomen approximately as wide as elytra, widest at segments III/IV, moderately tapering posteriorly (Fig. 64); punctuation fine and rather dense on tergites III-VI, sparser on tergites VII-VIII; microsculpture absent; posterior margin of tergite VII without palisade fringe.

♂: posterior margin of sternite VIII obtusely angled in the middle; median lobe of aedeagus shaped as in Figs 68-69, ventral process apically bifid; apical lobe of paramere very long, with two long basal setae, short apical seta, and subapical seta of intermediate length (Fig. 70).

Etymology: The name (Lat., adj.) is derived from Creta, the name of the island where the type locality is situated.

Comparative notes: Based on external and the male sexual characters, O. cretica is closely related to O. cingulum Bernhauer from the Middle East and to O. hatayana Assing from central southern Anatolia. It is distinguished from the former, whose male sexual characters are unknown, by the uniformly pale coloration of the forebody (in O. cingulum the head is darker than the pronotum) and from the latter by larger body size and by the shape of the aedeagus (apically straight ventral process in lateral view). From both species, it is additionally separated by the coarser punctuation of the forebody, the absence of microsculpture, the less transverse pronotum, the lateral impressions on the elytra, the relatively longer metatarsus, and the longer first metatarsomere. For illustrations of the habitus and the sexual characters of O. hatayana see Assing (2004c, in press).

Distribution and bionomics: The species is known only from one locality in eastern Crete. The reduced hind wings and palisade fringe at the posterior margin of tergite VII suggest that it is endemic to the island.
Oxypoda (Bessopora) lesbia ASSING 2005 (Figs 72-79, Map 3)

Material examined: 4 exs., Kato Olympos, E Kallipefki, 39°58N, 22°29E, 1500-1580 m, 6.IV.1998, leg. Assing, Wunderle (cAss, cWun); 2 exs., Evritania, W Lamia, N Karpenisi, Timfristos, ski resort, 38°57, 21°48E, 1900 m, 14.IV.2000, leg. Assing, Wunderle (cAss, cWun); 1 ex., Fokis, 45 km SSW Lamia, Oros Giona, 38°40N, 22°20E, 1700 m, 17.IV.2000, leg. Assing (cAss).

Map 3: Distribution of Oxypoda lesbia ASSING (filled circles), O. praecisa sp.n. (open circles), and O. afimbriata sp.n. (square) in Greece.

Comments: The species was only recently described from the Greek island Lesbos (ASSING 2005g). A comparative study of the external and sexual characters of the above material and of the holotype male of O. lesbia revealed some differences between populations (microsculpture, punctuation, shape of median lobe of aedeagus and of spermatheca), but no convincing evidence that they should represent distinct species. The intraspecific variation of the aedeagus and of the – previously unknown – spermatheca are illustrated in Figs 72-74 and Figs 77-79. The species is closely related to O. brachyptera (STEPHENS), as can be inferred from the highly similar morphology of the aedeagus (shape and internal structures of median lobe, apically bifid ventral process of the median lobe, shape and chaetotaxy of the apical lobe of the paramere, shape of the spermatheca). For comparison, the median lobe of the aedeagus of O. brachyptera is illustrated in Fig. 71. All the specimens seen so far have hind wings of reduced length (approximately 1.5 times as long as elytra and distinctly projecting from under the posterior elytra margins when unfolded). However, the presence of a palisade fringe at the posterior margin of the abdominal tergite VII, as well as the wide distribution suggest that O. lesbia may be wing-dimorphic.
**Oxypoda (Bessopora) afimbriata** sp.n. (Figs 80-82, 84-90, Map 3)

*Holotype* ♂: GR-Peloponnes, Erymanthos, Kalendzi, Pass, 1500 m, Rasen, Tanne, 27.5.97, P. Wunderle / Holotypus ♂ Oxypoda afimbriata sp. n. det. V. Assing 2005 (cAss). *Paratypes*: 2♀ ♂: GR. Peloponnisos, Erimanthos, oberh. Kalendzi, Sattel, 1500 m, 37°56'38N; 21°46'30E, 27.III.1997, V. Assing (cAss, cWun).

**Description:** 2.5-2.7 mm. Coloration: whole body reddish yellow, with the central part of tergite VI and the anterior half of tergite VII infuscated; legs yellow. Habitus as in Fig. 84.

Head indistinctly transverse; punctuation dense, but very shallow and somewhat ill-defined; microsculpture very shallow (Fig. 85); eyes relatively small (Fig. 86), weakly projecting from lateral outline of head, distinctly shorter than postocular region in dorsal view. Antenna as in Fig. 87, antennomere X approximately 1.5 times as wide as long. Preapical joint of maxillary palpus somewhat dilated, slightly less than twice as long as wide.

Pronotum 1.20-1.25 times as wide as head and 1.25-1.30 times as wide as long (Fig. 85); maximal width approximately in or a short distance anterior to middle; punctuation very dense, ill-defined, and asperate; surface almost matt.

Elytra approximately as wide as pronotum and at suture 0.85-0.90 times as long as pronotum (Fig. 85); surface somewhat flattened; punctuation coarser than that of pronotum, very dense, rasp-like, and ill-defined; microsculpture present; surface almost matt. Hind wings reduced, slightly projecting from under elytral hind margins when unfolded (Fig. 85). Legs relatively short; metatarsomere I longer than the combined length of the two following metatarsomeres, approximately as long as the combined length of metatarsomeres II-IV or nearly so.

Abdomen approximately as wide as elytra, widest at segments V/VI, segments VII-VIII weakly tapering (Fig. 84); punctuation fine and very dense on tergite III, gradually decreasing in density from tergites IV to VIII; microsculpture absent; posterior margin of tergite VII without palisade fringe.

♂: posterior margin of sternite VIII obtusely angled in the middle (Fig. 80); median lobe of aedeagus shaped as in Figs 88-89, ventral process apically bifid; apical lobe of paramere of similar shape and chaetotaxy as in other species of the *O. brachyptera* group, very long, with two long basal and two short apical setae (Fig. 90).

♀: posterior margin of sternite VIII weakly convex (Fig. 81); spermatheca as in Fig. 82.

**Etymology:** The name (Lat., adj.: not fimbriate) refers to the absence of a palisade fringe at the posterior margin of tergite VII.

**Comparative notes:** *Oxypoda afimbriata* belongs to the *O. brachyptera* group, as is suggested by the similar external characters and especially by the morphology of the median lobe of the aedeagus (general shape, internal structures, apically bifid ventral process), the shape and chaetotaxy of the apical lobe of the paramere, and by the morphology of the spermatheca. The new species is distinguished from the widespread *O. brachyptera* and *O. lesbia* by the paler coloration, which somewhat resembles that of some species of the subgenus *Deropoda*, by the much coarser and more asperate punctuation of the pronotum and the elytra, by the absence of a palisade fringe at the posterior...
margin of tergite VII, by the shape of the median lobe of the aedeagus, and by the shape of the spermathecal duct, which is longer and more slender than in *O. lesbia* (see Figs 77-79, 82).

**Distribution and bionomics:** The species is known only from the Erimanthos range in the northwestern Peloponnisos, Greece (Map 3). The reduced hind wings and palisade fringe at the posterior margin of tergite VII suggest that it has a restricted distribution.

**Oxypoda (Bessopora) praecissa** sp. n. (Figs 83, 91-107, Map 3)


**Description:** 2.5-3.0 mm. Coloration variable: body of almost uniformly dark colour, with the head and pronotum dark brown, the elytra slightly paler brown, and the abdomen blackish with paler tergal margins and paler apex, or bicoloured, with the pronotum and elytra reddish, the head slightly darker, and the abdomen reddish with tergite VI and anterior half of tergite VII infuscated; legs yellowish to yellowish brown; antennae brown, with antennomeres I-II yellowish to reddish yellow. Habitus as in Fig. 91.

Head approximately 1.1 times as wide as long; punctuation dense, but usually very shallow, ill-defined, and indistinct; microsculpture shallow (Fig. 92); eyes relatively small (Fig. 93), not projecting from lateral outline of head, distinctly shorter than postocular region in dorsal view. Antenna as in Fig. 94, antennomere X approximately 1.5 times as wide as long. Preapical joint of maxillary palpus approximately twice as long as wide.

Pronotum approximately 1.25 times as wide as head and 1.30 times as wide as long (Fig. 92); maximal width approximately in the middle; punctuation very dense, much coarser than that of head, and somewhat asperate; microsculpture very shallow; surface with subdued shine.

Elytra approximately as wide as pronotum and at suture 0.70 times as long as pronotum.
(Fig. 92); puncturation coarser than that of pronotum, very dense, and rasp-like; microsculpture indistinct; surface with some shine. Hind wings reduced, not projecting from under elytral hind margins when unfolded (Fig. 85). Legs relatively short; metatarsomere I longer than the combined length of the two following metatarsomeres, but shorter than the combined length of metatarsomeres II-IV.

Abdomen approximately as wide as elytra, widest at segments VI/VII, apically weakly tapering (Fig. 92); puncturation distinct and very dense on tergite III, decreasing in density from tergites IV to VIII; microsculpture absent, interstices with distinct shine; posterior margin of tergite VII without palisade fringe.

♂: sternite VIII obtusely angled in the middle (Fig. 100); median lobe of aedeagus as in Figs 95-98, with pronounced crista apicalis, ventral process apically not bifid; apical lobe of paramere relatively short, with one very short apical and three long basal and subapical setae (Fig. 99).

♀: posterior margin of sternite VIII weakly convex, similar to that of O. afimbriata (cf. Fig. 81); spermatheca of highly distinctive, but somewhat variable shape (Figs 83, 101-107).

E t y m o l o g y : The name (Lat., adj.: shortened) refers to the short duct of the spermatheca.

I n t r a s p e c i f i c  v a r i a t i o n : A comparison of specimens from different mountain ranges revealed some variability of the spermatheca. In the material from the Panahaiko and the Chelmos, the spermathecal duct tends to be shorter, whereas in females from the Killini and the Melanon it is somewhat longer and proximally more or less smoothly curved (Figs 83, 101-107). However, since convincing constant differences were found neither in the morphology of the aedeagus nor in external characters and, furthermore, since the shape of the spermatheca is subject to some variation even within populations, these differences are attributed to intra- rather than interspecific variation.

C o m p a r a t i v e  n o t e s : In external morphology, Oxypoda praecisa somewhat resembles species of the O. brachyptera group. However, the morphology of the apically unincised median lobe of the aedeagus, the shape and chaetotaxy of the apical lobe of the paramere, as well as the shape of the spermatheca suggest that it belongs to a different species group. From O. brachyptera and O. lesbia, the new species is readily distinguished by the coarser puncturation of the pronotum and elytra, the shorter elytra, the more pronounced shine especially of the abdomen, by the absence of a palisade fringe at the posterior margin of tergite VII, as well as by the completely different shape of the spermatheca. From O. afimbriata, it is separated by the darker coloration, the less reduced shine of the pronotum and elytra, the less dense puncturation of the pronotum, the shorter elytra, the more pronounced puncturation of the abdomen, and by the different shape of the spermatheca.

D i s t r i b u t i o n  a n d  b i o n o m i c s : The species inhabits several mountain ranges (Panahaiko, Chelmos, Killini, Melanon) in the northern Peloponnisos, Greece (Map 3). The reduced hind wings and palisade fringe at the posterior margin of tergite VII suggest that it has a restricted distribution. The type specimens were sifted from moss, grass, litter of coniferous trees and shrubs, often near snow, and the roots of grass and herbs at altitudes of 1450-2010 m.
Oxypoda (Bessopora) longicollis BERNAUER 1902 (Figs 108-116)

Oxypoda longicollis BERNAUER 1902: 209.


**Additional material examined:** 1 ♀, Ukraine, Krim (FMNH).

**Comments:** The original description of this species is based on syntypes from the Krim peninsula, Ukraine (one male and two females), and from the Parnass range in Greece (one female). The type specimens from both localities are not conspecific, so that a lectotype designation is mandatory and the male from the Krim peninsula is here designated as the lectotype. The female from the Parnass belongs to an unidentified – possibly undescribed – species; its general appearance is similar to that of *O. brachyptera*, but the spermathecal duct is shorter and of different shape.

The similar morphology of the median lobe of the aedeagus, the shape and chaetotaxy of the apical lobe of the paramere, as well as the morphology of the spermatheca suggest that *O. longicollis* is not affiliated with species of the *O. brachyptera* group, but rather closely related to *O. praecisa*.

**Redescription:** Small species, 2.3-2.7 mm; habitus of forebody as in Fig. 108.

**Coloration:** Reddish brown, with abdominal segment VI and the anterior half of segment VII somewhat infuscate; legs testaceous; antennae pale reddish.

**Head:** As wide as long or weakly transverse, relatively large in relation to pronotum (Fig. 108); puncturation fine and indistinct; microsculpture shallow, but distinct; eyes relatively small (Fig. 110), little more than half the length of postocular region in dorsal view. Antennae as in Fig. 109. Penultimate joint of maxillary palpus of moderate length, about 2.5-3.0 times as long as wide.

**Pronotum:** 1.20-1.25 times as wide as long and only 1.15-1.20 times as wide as head; maximal width in anterior half; puncturation shallow and dense, somewhat ill-defined, but more distinct than that of head; microsculpture shallow (Fig. 108).

**Elytra:** Slightly wider than pronotum and at suture about 0.75 times as long as pronotum; puncturation dense and distinct, much more so than that of head and pronotum (Fig. 108). Hind wings reduced. Metatarsomere I slightly longer than the combined length of II-III.

**Abdomen:** About as wide as elytra; puncturation on anterior tergites dense and distinct, gradually decreasing in density towards apex of abdomen; microsculpture absent or indistinct; posterior margin of tergite VII without palisade fringe.

**♂:** Posterior margin of sternite VIII strongly convex, but not angled in the middle (Fig. 112); aedeagus with apically unincised and, in lateral view, somewhat angled ventral process (Figs 113-114); apical lobe of paramere as in Fig. 111.

**♀♀:** Posterior margin of sternite VIII weakly concave in the middle (Fig. 115); spermatheca with rather short duct (Fig. 116).
Distribution: The species is known only from the Krim peninsula, Ukraine.

Oxypoda (Bessopora) moczarskii Bernhauser 1906

Oxypoda moczarskii Bernhauser 1906: 127 f.


Comments: The original description is explicitly based on two syntypes from the Pelopónnisos and Corfu (Bernhauser 1906). Since both type specimens are females, a lectotype designation is refrained from until males become available from the vicinity of either of the two type localities.

Haploglossa gentilis (Märkel 1845)

Material examined: 2 exs., Makedhonía, Kavála, Pangéo, road from Akrovouni to ski resort, oak forest, 500 m, with Lasius fuliginosus, 28.V.1999, leg. Assing (cAss).

Haploglossa villosula (Stephens 1832)

Aleochara (Aleochara) hellenica Pace 2005: 278; syn.n.


Comments: The description of Aleochara hellenica is based on a female holotype from "Etol.-Akar., O. Váltou, 2° polje NW di Perdikáki" deposited in the collection of Mauro Giachino (Torino). The details indicated in the original description, especially the illustration of the distinctive spermatheca, gave rise to the suspicion that A. hellenica was in fact a synonym of Haploglossa villosula. A subsequent examination of the holotype confirmed that it is doubtlessly conspecific with H. villosula; hence the synonymy proposed above.

Haploglossa villosula has been recorded from Greece before (Assing 2005g, Löbl & Smetana 2004). The species is widespread and common in Europe; a recent record from China (Assing 2005e) shows that it is present also in the Eastern Palaeartic region.

Aleochara inconspicua Aubé 1850

Material examined: 1 ex., Makedhonía, Dráma, Falakró, above ski resort, 1750-1850 m, 26.V.1999, leg. Assing (cAss); 1 ex., Falakró, below ski resort, 1500 m, pine forest, 26.V.1999, leg. Assing (cAss); 1 ex., Makedhonía, Kavála, Pangéo, peak region, 1900 m, 28.V.1999, leg. Assing (cAss); 1 exs., Pangéo, near ski resort, 1650 m, beech forest, 28.V.1999, leg. Assing (cAss).

Aleochara (Ceranota) purkynei Roubal 1937

Material examined: 1 ♀, Pelopónnisos, Killini range, Mosia, 4.IV.1992, leg. Frisch (cWun).

Comments: The species was originally described based on a male and a female (Likovsky 1973) – not two males, as indicated in the original description – from the Pelopónnisos, but has not been recorded since.
**Aleochara (Ceranota) mortuisimilis** sp. n. (Figs 33-46)

**Holotype ♂:** GR. N Larissa, No. 11, Kato Olympos, 1500-1580 m, E Kallipefki, 39°57'34"N, 22°29'15", 6.IV.1998, V. Assing / Holotypus ♂ *Aleochara mortuisimilis* sp. n. det. V. Assing 2005 (cAss). **Paratypes:** 4♂, 3♀: same data as holotype (cAss); 3♂, 1♀: same data, but leg. P. Wunderle (cWun).

**Description:** Relatively small and slender species, 4.3-5.3 mm (abdomen fully extended). Coloration variable. Light-coloured specimens: head brown; pronotum and elytra light brown to reddish brown; abdomen reddish brown with the central tergal areas infuscate; legs testaceous; antennae brown, with the basal 2-3 antennomeres paler. Dark-coloured specimens: head blackish; pronotum and elytra castaneous, with the elytra at least slightly paler than the pronotum; abdomen dark brown to blackish brown, with the tergal margins slightly paler; legs rufotestaceous; antennae brown, with the basal 2-3 antennomeres paler. Habitus as in Fig. 33.

Head approximately as wide as long; eyes moderately large and protruding from lateral outline of head, postocular region approximately 1.5 times the length of eyes in dorsal view (Fig. 34); integument with pronounced and dense microreticulation, matt; puncturation fine, shallow, and relatively sparse; pubescence long and suberect. Antenna with antennomeres I-III approximately 2.5 times as long as wide and of subequal length, IV weakly transverse; IV-X of gradually increasing width and increasingly transverse; X approximately 1.5 times as wide as long; XI approximately as long as the combined length of IX and X (Fig. 35).

Pronotum transverse, approximately 1.25 times as wide as long and 1.40-1.50 times as wide as head; maximal width approximately in the middle; posterior angles weakly marked, almost obsolete; microsculpture almost as pronounced as that of head; puncturation more distinct and somewhat denser than that of head; pubescence more or less depressed, along midline directed posteriorly, on either side of midline laterad.

Elytra approximately 1.1 (bacthyaerous morph) to 1.18 (macropterous morph) times as wide and at suture 0.80-0.85 times as long as pronotum; microreticulation similar to that of head and pronotum; puncturation of similar density as that of pronotum, but somewhat granulose; pubescence directed diagonally latero-caudad. Hind wing length dimorphic, either almost completely reduced or fully developed.

Abdomen with segments III-V of subequal width, posterior segments (from segment VI) slightly tapering; tergites III-V with moderately deep anterior impressions, tergite VI with shallow impression; microsculpture very shallow, integument more shiny than that of forebody; puncturation relatively fine and sparse, in anterior impressions of tergites III-V very fine and very sparse; posterior margin of tergite VII with palisade fringe in both morphs; tergites III and VII with sexual dimorphism.

♂: tergite III with weakly elevated median elevation (Fig. 35); tergite VII in the middle of posterior margin with broad, smoothly rounded (antero-dorsal view), and not very strongly elevated tubercle (Figs 36-38); posterior margin of tergite VIII convex and with very sparse setae posteriorly (Fig. 39); posterior margin of sternite VIII pointed and with moderately long marginal setae (Fig. 40); median lobe of aedeagus as in Figs 41-43.

♀: posterior margin of tergite VIII broadly convex and with more numerous short setae posteriorly (Fig. 44); posterior margin of sternite VIII broadly convex and with row of
short, moderately stout, modified marginal setae (Fig. 45); spermatheca as in Fig. 46.

**Intraspecific variation:** The species appears to be subject to lower intraspecific variation than most of its congeners; this conclusion, however, is based only on twelve specimens from one locality. As indicated above, coloration is rather variable, whereas variation in size is moderate. The male secondary sexual characters were present in all eight males, but especially the tubercle on tergite VII may be of reduced size.

**Etymology:** The name (Lat., adj.: seemingly dead) refers to the observation that the species displays thanatosis when moved or touched.

**Comparative notes:** *Aleochara mortuisimilis* is readily separated from all other species of the subgenus by the following character combination: relatively small size; pronotum less than 1.3 times as wide as long; the pronounced microsculpture especially on the head, but also on the pronotum and elytra; the relatively short elytra; the dimorphism of the hind wings; the male secondary sexual characters, as well as the shapes of the aedeagus and the spermatheca.

Two species of *Ceranota* have been recorded from Greece: *Aleochara erythroptera* GRAVENHORST and *A. purkynei* ROUBAL. From both, *A. mortuisimilis* is distinguished by distinctly smaller size, the pronounced microsculpture of the forebody, the shorter and less slender antennae, the less smaller and less transverse pronotum, the much sparser and more indistinct puncturation of the forebody, the fine and sparse punctuation of the anterior impressions of the abdominal tergites, the differently shaped male secondary sexual characters (especially the shape of the tubercle on the abdominal tergite VII), and by the shape of the genitalia. From *A. erythroptera* it is additionally separated by the different and more uniform coloration (in *A. erythroptera* the elytra are ferrugineous and distinctly contrasting with the dark pronotum) and from *A. purkynei* by the unmodified male tergite IV. For illustrations of the habitus and the male secondary sexual characters of *A. purkynei* see ROUBAL (1937).

**Distribution and bionomics:** The species is known only from the peak of the Kato Olympos, northeastern Thessalia. The types were collected by sifting moss and litter of beech and fir, partly near snow fields at an altitude of 1500-1580 m. Interestingly, the specimens displayed thanatosis when moved or touched. *Aleochara mortuisimilis* is apparently the first example of wing dimorphism to become known in the subgenus: two females are macropterous, the remaining eight males and two females are brachypterous.

**Tinotus morion** (GRAVENHORST 1802)

**Material examined:** 1 ex., Makedonía, Olympos, NW Stavros refuge, 1500 m, under pine bark, 7.IV.1998, leg. Assing (cAss).
Figs 33-46: *Aleochara mortuisimilis* sp.n. (34-40, 43: holotype). (33) habitus (brachypterous specimen); (34) head in lateral view; (35) abdominal tergite III; (36) abdominal tergite VII in dorsal view; (37) abdominal segment VII in lateral view; (38) abdominal tergite VII in antero-dorsal view; (39) tergite VIII; (40) sternite VIII; (41-43) median lobe of aedeagus in lateral and in ventral view; (44) tergite VIII; (45) posterior part of sternite VIII; (46) spermatheca.

Scale bars: 33-38: 1.0 mm; 39-40, 44: 0.5 mm; 41-43, 45-46: 0.2 mm.
Figs 47-53: *Xantholinus moreanus* sp.n. (47-48) forebody; (49-52) aedeagus of 4♂♂; (53) spine series in internal sac (proximal part of aedeagus). Scale bars: 47-48: 1.0 mm; 49-53: 0.5 mm.
Figs 54-59. Xantholinus moreanus sp.n. (54) and X. creticus sp.n. (55-59). (54) internal tube of aedeagus, anterior part; (55) forebody; (56) aedeagus; (57-58) posterior part of aedeagus; (59) membranous tube of aedeagus. Scale bars: 55: 1.0 mm; 56-57: 0.5 mm; 54, 58-59: 0.25 mm.
Figs 60-70: Aphaenostemmus rhodicus sp.n. (60-63) and Oxypoda cretica sp.n. (64-70). (60, 64) habitus; (61, 65) forebody; (62) head and pronotum; (63, 67) antenna; (66) head in lateral view; (68) median lobe of aedeagus in lateral view; (69) apex of median lobe of aedeagus in ventral view; (70) apical lobe of paramere. Scale bars: 60, 64: 1.0 mm; 61-63, 65-67: 0.5 mm; 68-70: 0.1 mm.
Figs 71-83: *Oxypoda brachyptera* (STEPHENS) (71), *O. lesbia* ASSING (72-79), *O. a fibrata* sp.n. (80-82), and *O. praecisa* sp.n. (83). (71-74) median lobe of aedeagus in lateral and in ventral view (72, 74: Timfristos; 73: Kato Olympos); (75) apical lobe of paramere; (77-79, 82-83) spermatheca (77: Timfristos; 78: Oros Giona; 79: Kato Olympos; 83: Panahako); (76, 81) ♀ sternite VIII; (80) ♂ sternite VIII. Scale bars: 76, 80-81: 0.2 mm; 71-75, 77-79, 82-83: 0.1 mm.
Figs 84-94: *Oxyopa afimbriata* sp.n. (84-90) and *O. praecisa* sp.n. (91-94). (84, 91) habitus; (85, 92) forebody; (86, 93) head in lateral view; (87, 94) antenna; (88-89) median lobe of aedagus in lateral and in ventral view; (90) apical lobe of paramere. Scale bars: 84, 91: 1.0 mm; 85, 92: 0.5 mm; 86-87, 93-94: 0.2 mm; 88-90: 0.1 mm.
Figs 95-111: Oxypoda praccisa sp.n. (95-107) and O. longicollis BernHauer (108-111). (95-98) median lobe of aedeagus in lateral and in ventral view; (99, 111) apical lobe of paramere; (100) ♀ sternite VIII; (101-107) spermathecae of ♀♀ from Panahako (101), Chelmos (102-103), Killini (104-105), and Melanon (106-107); (108) forebody; (109) antenna; (110) head in lateral view. Scale bars: 96-99, 101-107, 111: 0.1 mm; 10, 108-110: 0.2 mm.
Fig. 112-116: Oxypoda longicollis BERNHAUER (112-114: lectotype). (112) ♂ sternite VIII; (113-114) median lobe of aedeagus in lateral view; (115) posterior margin of ♀ sternite VIII; (116) spermatheca. Scale bars: 112: 0.2 mm; 113-116: 0.1 mm.

Acknowledgements

I am indebted to the colleagues listed in the material section for the loan of material from the collections under their care, as well as to Jean-Claude Lecoq for his assistance with the loan of some Xantholinus types from the Coiffait collection (MNHNP). Paul Wunderle provided the habitat photograph (Fig. 32) and additional records. The following colleagues assisted in the identification of some of the species: Michael Schülke (some Tachyporinae), Jürgen Vogel, Görlitz (some Atheta), Adriano Zanetti, Verona (Eusphalerum, Lesteva), and Lothar Zerche (Oxypoda moreatica, O. micans). Benedikt Feldmann proof-read the manuscript.

Zusammenfassung

References


Author’s address: Dr. Volker ASSING
Gabelsbergerstr. 2
D-30163 Hannover, Germany
E-mail: vassing.hann@t-online.de
New species and records of Staphylinidae from Greece, with two new synonymies (Insecta: Coleoptera) 333-379