

On the Staphylinidae (Coleoptera) of Turkey IX. Five new species, a new synonymy, and additional records

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Abstract

A study of more than 3,700 specimens of Staphylinidae collected during three field trips to Turkey conducted in 2011 and 2012, as well as of additional material received from other sources yielded nearly 290 species. Five species are described and illustrated: *Atheta (Philhygra) scabens* **n. sp.** (NE-Turkey, N-Caucasus, Kyrgyzstan, Tajikistan); *Atheta brachati* **n. sp.** (Turkey: Bolu); *Cousya microdotoides* **n. sp.** (Turkey: Adana); *Meotica wunderlei* **n. sp.** (Turkey: Isparta); *Stenus (Hemistenus) bithynicus* **n. sp.** (Turkey: Kocaeli, Sakarya, Düzce). One synonymy is proposed: *Leptusa asiatica* Bernhauer, 1909 = *L. flagellulifera* Assing, 2009, **n. syn.** Ten species are recorded from Turkey for the first time; two new country records are reported for Greece and Iraq. The distributions of seven species are mapped. A recently published catalogue of the Staphylinidae of Asia Minor and Cyprus is reviewed. Most of the names synonymised in this catalogue are revalidated; the remaining synonymies are rectified.

Key words: Coleoptera, Staphylinidae, Turkey, Greece, Iraq, new species, new synonymy, revalidation, endemism, additional records, zoogeography, taxonomy, catalogue, review.

Zusammenfassung

Die Bearbeitung von mehr als 3700 Staphyliniden, die auf drei in den Jahren 2011 und 2012 in die Türkei unternommenen Exkursionen gesammelt wurden, sowie von weiterem Material, ergab insgesamt fast 290 Arten. Fünf Arten werden beschrieben und abgebildet: *Atheta (Philhygra) scabens* **n. sp.** (Nordosttürkei, Kirgisistan, Tadschikistan); *Atheta brachati* **n. sp.** (Türkei: Bolu); *Cousya microdotoides* **n. sp.** (Türkei: Adana); *Meotica wunderlei* **n. sp.** (Türkei: Isparta); *Stenus (Hemistenus) bithynicus* **n. sp.** (Türkei: Kocaeli, Sakarya, Düzce). *Leptusa flagellulifera* Assing, 2009, **n. syn.**, wird mit *L. asiatica* Bernhauer, 1909 synonymisiert. Zehn Arten werden erstmals aus der Türkei, je eine Art erstmals aus Griechenland bzw. dem Irak nachgewiesen. Die Verbreitungsgebiete von sieben Arten werden anhand von Karten illustriert. Ein kürzlich erschienener Katalog der Staphyliniden von Kleinasien und Zypern wird rezensiert. Fast alle in diesem Katalog synonymisierten Namen werden revalidiert; die restlichen Synonymien werden korrigiert.

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1 Introduction

The present paper is the ninth contribution to the Turkish staphylinid fauna providing descriptions and records of species from miscellaneous subfamilies and genera. Since the latest instalment (ASSING 2011a), additional material has become available primarily from two field trips to central southern and to northwestern Turkey conducted

by VOLKER BRACHAT (Geretsried) and HEINRICH MEYBOHM (Großhansdorf) in April 2011 and in April/May 2012, respectively, as well as one to southwestern Turkey conducted by PAUL WUNDERLE (Mönchengladbach) and the author in April 2011. Moreover, additional material was seen from several public and private collections. An examination of this material, in total nearly 290 species and approximately 3,700 specimens, yielded not only numerous

records of zoogeographic interest, among them ten new country records from Turkish territory, but also five species new to science.

Most of the material of genera such as *Sunius* Stephens, 1829, *Pella* Stephens, 1836, and *Geostiba* Thomson, 1858 has been, or will be, treated in separate revisionary contributions covering the West Palaearctic or the whole of the Palaearctic region (ASSING 2011b, 2011c, in prep.). As in previous contributions, the Scaphidiinae, Pselaphinae, and Scydmaeninae are not treated in the present paper; the latter two have been, or will be, studied by VOLKER BRACHAT and HEINRICH MEYBOHM, respectively.

In the recent past, a checklist (ANLAŞ 2009) and a catalogue (BORDONI 2010) of the Staphylinidae of Turkey were published. The former has been commented on explicitly or implicitly in earlier contributions (e. g., ASSING 2011a). The latter became available only after the previous contribution (ASSING 2011a) had been submitted and will be dealt with in section 4 of the present paper.

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I am indebted to VOLKER BRACHAT, Geretsried, and HEINRICH MEYBOHM, Großhansdorf, for the generous gift of their staphylinid by-catches collected during their field trips to Turkey. The following colleagues assisted with the identification of certain species or genera: JOHANNES FRISCH, Berlin (*Scopaeus* spp.), MIKHAIL GILDENKHOV, Smolensk (*Carpelimus atomus*), MICHAEL SCHÜLKE, Berlin (*Mycetoporus glaber*, *Tachyporus assingi*, *T. solutus*), and ADRIANO ZANETTI, Verona (*Dropephylla* spp., *Dialycera aspera*, *Eusphalerum caucasicum loebli*, *Omalium turcicum*). The criticism and comments of two reviewers, BENEDIKT FELDMANN, Münster, and MICHAEL SCHÜLKE, Berlin, are appreciated. VOLKER PUTHZ, Schlitz, ADRIANO ZANETTI, and ALEXEY SOLODOVNIKOV, Copenhagen, reviewed – and approved of – the comments on the catalogue by BORDONI (2010). VOLKER PUTHZ additionally confirmed the novelty of *Stenus bithynicus*.

2 Material and methods

The material studied in the present paper is deposited in the following public and private collections:

cAnl	private collection SINAN ANLAŞ, Turgutlu
cAss	author's private collection
cFel	private collection BENEDIKT FELDMANN, Münster
cSch	private collection MICHAEL SCHÜLKE, Berlin
cVav	private collection JIŘÍ VÁVRA, Ostrava-Krásné Pole
cWun	private collection PAUL WUNDERLE, Mönchengladbach
BMNH	The Natural History Museum, London (R. BOOTH)
EME	Entomology Museum, Erzurum (E. YILDIRIM)
MNHUB	Museum für Naturkunde der Humboldt-Universität, Berlin (J. FRISCH)
NHMW	Naturhistorisches Museum Wien (H. SCHILLHAMMER)
NMP	National Museum of Natural History, Praha (J. HÁJEK)

The morphological studies were conducted using a Stemi SV11 microscope (Zeiss Germany) and a Jenalab compound

microscope (Carl Zeiss Jena). A digital camera (Nikon Coolpix 995) was used for the photographs. The maps were created using MapCreator 2.0 (primap) software.

Head length was measured from the anterior margin of the clypeus (Aleocharinae) or from the anterior margin of the frons (Steninae) to the posterior margin of the head, elytral length at the suture from the apex of the scutellum to the posterior margin of the elytra, total length from the anterior margin of the mandibles (in resting position) to the apex of the abdomen, the length of the forebody from the anterior margin of the mandibles to the posterior margin of the elytra, and the length of the aedeagus from the apex of the ventral process to the base of the aedeagal capsule. The side of the aedeagus with the sperm duct opening is referred to as the ventral, the opposite side as the dorsal aspect.

The individual labels of type specimens are separated by slashes; they are cited in the original spelling and format, except that the following adaptations were made according to the general format requirements of the journal: names of persons (except authors of species) in small capitals, scientific names of genera and species in italics.

3 Results

3.1 General results

The field trips conducted in spring 2011 and in spring 2012 yielded a total of approximately 3,700 Staphylinidae (exclusive of Scaphidiinae, Pselaphinae, and Scydmaeninae) belonging to more than 275 species. The material seen from other sources comprised 56 specimens and yielded eleven additional species (see list below). The vast majority of specimens was identified to species level. Some, however, remain of doubtful identity either because they were represented only by females (e. g., *Mycetoporus nigricollis* group, *Astenus* spp., *Quedius* spp., *Gabrius* spp., *Xantholinus* spp.) or because they belong to species groups or genera that are currently in a state of taxonomic confusion and require revision (e. g., *Olophrum* sp., *Mycetoporus baudueri* group, *Sepedophilus* spp., *Anaulacaspis* spp., *Atheta (Microdota)* spp., *Atheta (Mocyta)* spp., *Cousya* spp., *Ocalea* spp., *Oxypoda (Baeoglana)* spp.).

The examined material not only included numerous new province records (not listed individually), new country records of ten species and the first primary record of an additional species from Turkey (see section 3.2), but also several species new to science. Some of them have already been described in the context of revisionary studies (ASSING 2011b, 2011c, 2012) or in other separate articles (SCHÜLKE 2013). The remainder is described in section 3.3. Two new country records are reported for Greece and Iraq.

A complete list of Staphylinidae (exclusive of Scaphidiinae, Pselaphinae, and Scydmaeninae) recorded from Turkey in 2011 and 2012 (sample numbers 1–95) and examined from other sources (sample numbers 96–130) is given below. In the localities column, the number of specimens is

given in parentheses behind the respective locality number. The chronological order of the subfamilies is arranged according to SMETANA (2004). The genera and species within the subfamilies are arranged in alphabetical order.

Localities and collection data corresponding to the sample numbers of the list below are as follows:

Staphylinidae collected in 2011–2012 (deposited in MNHUB, cAss, cFel, and cWun) (sample numbers 1–95):

Kocaeli (1–9; leg. BRACHAT & MEYBOHM): **1:** Kartepe, 40°39'N, 30°04'E, 1030 m, 24.IV.2012; **2:** Kartepe, 40°39'N, 30°05'E, 1040 m, 24.IV.2012; **3:** Suadiye, 40°41'N, 30°03'E, 180 m, 25.IV.2012; **4:** Suadiye, 40°40'N, 30°03'E, 540 m, 25.IV.2012; **5:** Pazarçayırı, 40°38'N, 30°04'E, 720 m, 25.IV.2012; **6:** same data, but 5.V.2012; **7:** Pazarçayırı, 40°39'N, 30°03'E, 880 m, 25.IV.2012; **8:** Kartepe, 40°39'N, 30°06'E, 1270 m, 5.V.2012; **9:** N Hereke, 40°49'N, 29°41'E, 460 m, 6.V.2012.

Sakarya (10–17; leg. BRACHAT & MEYBOHM): **10:** Geyve-Taraklı, 40°26'N, 30°24'E, 700 m, 26.IV.2012; **11:** Geyve-Taraklı, 40°26'N, 30°26'E, 710 m, 26.IV.2012; **12:** Geyve-Taraklı, 40°27'N, 30°28'E, 775 m, 26.IV.2012; **13:** 17 km N Hendek, 40°53'N, 30°46'E, 650 m, 3.V.2012; **14:** 14 km N Hendek, 40°54'N, 30°45'E, 700 m, 3.V.2012; **15:** 4 km S Dikmen, 40°40'N, 30°54'E, 1275 m, 4.V.2012; **16:** 2 km W Dikmen, 40°42'N, 30°53'E, 700 m, 4.V.2012; **17:** 14 km W Dikmen, 40°44'N, 30°50'E, 430 m, 4.V.2012.

Bolu (18–31; leg. BRACHAT & MEYBOHM): **18:** 20 km W Mudurnu, 40°31'N, 31°05'E, 620 m, 26.IV.2012; **19:** S Abant, 40°34'N, 31°16'E, 1010 m, 27.IV.2012; **20:** S Abant, 40°35'N, 31°16'E, 1180 m, 27.IV.2012; **21:** Abant, 40°36'N, 31°17'E, 1400 m, 27.IV.2012; **22:** 8 km NE Abant, 40°39'N, 31°22'E, 1010 m, 27.IV.2012; **23:** Yedigöller Milli Park, 40°53'N, 31°41'E, 1720 m, 28.IV.2012; **24:** Yedigöller Milli Park, 40°50'N, 31°40'E, 1120 m, 28.IV.2012; **25:** Yiğilca, 40°51'N, 31°39'E, 1060 m, 29.IV.2012; **26:** Bolu-Yiğilca, 40°51'N, 31°37'E, 1010 m, 29.IV.2012; **27:** Bolu-Yiğilca, 40°51'N, 31°37'E, 980 m, 29.IV.2012; **28:** 25 km S Bolu, 40°38'N, 31°37'E, 1580 m, 30.IV.2012; **29:** 23 km S Bolu, 40°38'N, 31°37'E, 1540 m, 30.IV.2012; **30:** 11 km S Bolu, 40°40'N, 31°38'E, 1080 m, 30.IV.2012; **31:** 10 km S Bolu, 40°40'N, 31°38'E, 1020 m, 30.IV.2012.

Düzce (32–36; leg. BRACHAT & MEYBOHM): **32:** 26 km S Düzce, 40°41'N, 31°09'E, 1200 m, 1.V.2012; **33:** 23 km S Düzce, 40°41'N, 31°08'E, 1020 m, 1.V.2012; **34:** 19 km S Düzce, 40°42'N, 31°10'E, 650 m, 1.V.2012; **35:** Kaplandede Dağı, 40°57'N, 31°04'E, 1100 m, 2.V.2012; **36:** Kaplandede Dağı, 40°55'N, 31°02'E, 710 m, 2.V.2012.

Afyon (37–41; leg. ASSING & WUNDERLE): **37:** Emir Dağları, 20 km S Emirdağ, 38°55'N, 31°07'E, 1170 m, pasture, under stones, 18.IV.2011; **38:** Emir Dağları, 20 km S Emirdağ, 38°53'N, 31°09'E, 1450 m, pasture, under stones, 18.IV.2011; **39:** Emir Dağları, 20 km S Emirdağ, 38°54'N, 31°08'E, 1230 m, N-slope with oak, sifted, 18.IV.2011; **40:** Sultan Dağları, 15 km SE Çay, 38°32'N, 31°11'E, 1430 m, oak forest, litter and bark sifted, 18.IV.2011; **41:** Sultan Dağları, 15 km SE Çay, 38°31'N, 31°09'E, 1810 m, cedar forest, sifted, 18.IV.2011.

Konya (42–53; leg. ASSING & WUNDERLE): **42:** 5 km W Yunak, Bayatolu Tepe, 38°49'N, 31°41'E, 1420 m, pasture, 17.IV.2011; **43:** 38 km SE Akşehir, 20 km N Hüyük, 38°04'N, 31°39'E, 1440 m, calcareous pasture, under stones, 19.IV.2011; **44:** 39 km SE Akşehir, NW Kayabeli geçidi, 38°02'N, 31°35'E, 1870 m, peak region, under stones, 19.IV.2011; **45:** same data,

but sifted, 20.IV.2011; **46:** 25 km N Beyşehir, Selki, 37°54'N, 31°44'E, 1320 m, moist calcareous road margin, under stones, 19.IV.2011; **47:** 30 km W Konya, S Kızılören, 37°51'N, 32°06'E, 1390 m, moist road margin, 19.IV.2011; **49:** 35 km ENE Beyşehir, E Hüyük, 37°57'N, 31°38'E, 1290 m, road margin, under stones, 20.IV.2011; **50:** 35 km ENE Beyşehir, E Hüyük, 37°57'N, 31°38'E, 1270 m, near reservoir, under stones, 20.IV.2011; **51:** 15 km NE Hüyük, Konakkale, 38°00'N, 31°41'E, 1490 m, field margin, under stones, 20.IV.2011; **52:** 20 km NE Hüyük, N Konakkale, 38°02'N, 31°40'E, 1520 m, near stream, litter and dead wood (*Salix*) sifted, 20.IV.2011; **53:** Sultan Dağları, NW Dereçin, 38°29'N, 31°15'E, 1320 m, oak forest, litter sifted, 21.IV.2011.

Isparta (54–70; leg. ASSING & WUNDERLE): **54:** Sultan Dağları, 15 km SSW Akşehir, Cankurtaran, 38°15'N, 31°24'E, 1860 m, under stones, 17.IV.2011; **55:** same data, but 22.IV.2011; **56:** 15 km SW Eğirdir, Davraz Tepe, 37°47'N, 30°45'E, 1680 m, stony calcareous slope, under stones, 23.IV.2011; **57:** same data, but 28.IV.2011; **58:** 5 km SW Eğirdir, Davraz Tepe, 37°47'N, 30°45'E, 1780 m, stony calcareous slope, under stones, 23.IV.2011; **59:** 15 km SW Eğirdir, Davraz Tepe, 37°47'N, 30°45'E, 1800 m, calcareous slope, pine litter sifted, 23.IV.2011; **60:** 24 km N Eğirdir, Barla Dağı, 38°05'N, 30°46'E, 1720 m, calcareous slope, under stones, 24.IV.2011; **61:** Barla Dağı, 38°06'N, 30°47'E, 1620 m, calcareous pasture, under stones, 24.IV.2011; **62:** Barla Dağı, 38°06'N, 30°47'E, 1680 m, litter and fungi under cedar sifted, 24.IV.2011; **63:** same data, but bark of dead pine sifted; **64:** Barla Dağı, 38°06'N, 30°48'E, 1590 m, calcareous slope, grass roots beneath juniper sifted, 24.IV.2011; **65:** 40 km ESE Eğirdir, Dedegöl Dağları, 8 km W Yenişarbademli, 37°44'N, 30°16'E, 1680 m, pasture, under stones, 25.IV.2011; **66:** 43 km ESE Eğirdir, Dedegöl Dağları, 5 km W Yenişarbademli, 37°43'N, 31°19'E, 1820 m, flooded pasture and calcareous slope, under stones, 25.IV.2011; **67:** 10 km SE Sütçüler, 37°25'N, 31°02'E, 1520 m, moist meadow, under stones, 26.IV.2011; **68:** same data, but grass and moss sifted; **69:** 10 km SE Sütçüler, 37°25'N, 31°02'E, 1520 m, calcareous slope, under stones, 26.IV.2011; **70:** 12 km N Sütçüler, 37°36'N, 30°59'E, 1100 m, oak forest, litter and grass, mostly between stones, sifted, 26.IV.2011.

Niğde (71–73; leg. BRACHAT & MEYBOHM): **71:** E Madenköy, 37°29'N, 34°42'E, 1200 m, 17.IV.2011; **72:** E Madenköy, 37°28'N, 34°40'E, 1330 m, 17.IV.2011; **73:** E Madenköy, 37°27'N, 34°39'E, 1615–1870 m, 17.IV.2011.

Mersin (74–75; leg. BRACHAT & MEYBOHM): **74:** E Çamlıyayla, Belcinar, 37°08'N, 34°42'E, 930 m, 25.IV.2011; **75:** E Çamlıyayla, Darıpınarı, 37°09'N, 34°43'E, 710 m, 25.IV.2011.

Adana (76–95; leg. BRACHAT & MEYBOHM): **76:** Belededik, 37°23'N, 34°55'E, 720 m, 15.IV.2011; **77:** Belededik, 37°21'N, 34°55'E, 710 m, 15.IV.2011; **78:** Belededik, 37°20'N, 34°55'E, 690 m, 16.IV.2011; **79:** E Pozanti, road to Armutoluğu, 37°26'N, 34°54'E, 1220 m, 16.IV.2011; **80:** E Pozanti, road to Armutoluğu, 37°26'N, 34°55'E, 1310 m, 16.IV.2011; **81:** E Pozanti, road to Armutoluğu, 37°26'N, 34°56'E, 1700 m, 16.IV.2011; **82:** Belededik, 37°21'N, 34°54'E, 740 m, 16.IV.2011; **83:** Kamışlı, 37°32'N, 34°54'E, 1400 m, 18.IV.2011; **84:** Kamışlı, 37°35'N, 34°53'E, 1350 m, 18.IV.2011; **85:** SW Hasandede geçidi, 37°30'N, 35°23'E, 1230 m, 19.IV.2011; **86:** 8 km S Feke, Akkaya, 37°46'N, 35°54'E, 760 m, 21.IV.2011; **87:** 11 km NE Feke, Cumhurlu, 37°53'N, 36°00'E, 710 m, 21.IV.2011; **88:** 14 km NE Feke, Cumhurlu, 37°54'N, 36°00'E, 720 m, 21.IV.2011; **89:** Feke, 21 km to Mansurlu, 37°51'N, 35°46'E, 965 m, 22.IV.2011; **90:** SW Aladağ, SW Kayadibi, 37°28'N, 35°21'E, 1080 m, with *Camponotus* sp.,

23.IV.2011; **91**: 3 km W Karaysali, 37°14'N, 35°01'E, 170 m, 24.IV.2011; **92**: 7 km E Kamışlı, 37°32'N, 35°00'E, 1330 m, 26.IV.2011; **93**: 5 km E Kamışlı, 37°32'N, 34°59'E, 1450 m, 26.IV.2011; **94**: 4 km E Kamışlı, 37°33'N, 34°59'E, 1385 m, 26.IV.2011; **95**: Belededik, 37°22'N, 34°55'E, 715 m, 27.IV.2011.

Material examined from other sources (sample numbers 96–130):

96: Antalya, 10 km N Akseki, 37°09'N, 31°48'E, 1250 m, mixed cypress and oak forest, litter sifted, 17.II.2011, leg. SCHÜLKE (cSch, cAss); **97**: Antalya, 10 km N Akseki, 37°08'N, 31°48'E, 1200 m, oak litter sifted, 14.II.2011, leg. SCHÜLKE (cSch, cAss); **98**: 8 km NNW Akseki, 37°07'N, 31°46'E, 1225 m, pine and fir litter sifted, 14.II.2011, leg. SCHÜLKE (cSch); **99**: Antalya, 10 km N Akseki, 37°09'N, 31°48'E, 1250 m, cypress and oak forest, litter sifted, 17.II.2011, leg. SCHÜLKE (cSch, cAss); **100**: 5 km S Ibradı, 37°04'N, 31°38'E, 810 m, oak litter and dead branches sifted, 17.II.2011, leg. SCHÜLKE (cSch); **101**: Antalya, N Bademli geçidi, 37°19'N, 31°44'E, 1400 m, mixed cypress and fir forest, litter sifted, 17.II.2011, leg. SCHÜLKE (cSch, cAss); **102**: 40 km SW Antalya, Göynük, 16.–20.IV.2004, leg. HULA (NMP, cAss); **103**: Antalya, Kumluca, 8.V.1998 (EME); **104**: Antalya, 40 km SW Antalya, Göynük, 16.–20.IV.2004, leg. HULA (NMP); **105**: Burdur, 21 km SE Burdur, W Çeltikçi geçidi, 37°34'N, 30°24'E, 1420 m, cypress and oak litter sifted, 16.II.2011, leg. SCHÜLKE (cSch); **106**: Muğla, N Fethiye, Çaliş, 36°40'N, 29°06'E, 10 m, bank of stream, I.X.2002, leg. ASSING (cAss); **107**: Muğla, NW Fethiye, Kargı, 36°43'N, 29°36'E, 10 m, gravel river bank, 31.III.2002, leg. ASSING (cAss); **108**: Muğla, Çetibeli, 36°58'N, 28°17'E, 10–30 m, floodplain forest, 30.IV.2001, leg. BRACHAT & MEYBOHM (cAss); **109**: Konya, Akşehir env., 19.IV.1960, leg. SCHUBERT (NHMW); **110**: Konya, Akşehir env., Sultan Dağları, 26.V.1960, leg. PETROWITZ & RESSL (NHMW); **111**: same data, but 30.V.1960 (cAss); **112**: Niğde, Orhaniye, Çamardı env., Ala Dağlar, 1800–2200 m, 19.V.2005, leg. VÁVRA (cVav, cAss); **113**: Niğde, Orhaniye, Çamardı env., Ala Dağlar, 1800–2200 m, 19.V.2005, leg. VÁVRA (cVav, cAss); **114**: Mersin, Çamlıyayla, 10.V.–3.VI.1963, leg. Schubert (NHMW, cAss); **115**: Mersin, Çamlıyayla, V.1967, leg. SCHUBERT (NHMW); **116**: Osmaniye, Osmaniye, 1200 m, VI.1967, leg. SCHUBERT (NHMW); **117**: Hatay, Kavalcık, Reynanlı, 26.IV.2007 (cAnl); **118**: Hatay, Hassa, Akbez, 5 km E Koruhöyük, 36°48'N, 36°38'E, 410 m, 23.IV.2008, leg. YAĞMUR (cAnl); **119**: Hatay, Yayladağı, 3 km N Leylekli, 35°59'N, 36°03'E, 670 m, 17.V.2008, leg. YAĞMUR (cAnl); **120**: Hatay, Belen geçidi, 2.5 km N Kıcı, 36°30'N, 36°14'E, 950 m, 9.V.2008, leg. YAĞMUR (cAss); **121**: İstanbul, Alem Dağı, leg. v. BODEMEYER (BMNH); **122**: Sakarya, Adapazari, Sapanogölü, 6.IV.1966 (NHMW, cAss); **123**: Samsun, 25 km S Samsun, NE Asarcık, 41°05'N, 36°16'E, 880 m, *Fagus-Quercus-Carpinus* forest, litter in dry stream sifted, 20.VII.2008, leg. SCHÜLKE (cAss); **124**: Ordu, road Ünye–Akkuş, 18 km NE Akkuş, 40°56'N, 37°07'E, 920 m, mixed deciduous forest (predominantly *Fagus*), litter sifted, 15.VII.2008, leg. SCHÜLKE (cSch); **125**: Sinop, Çangal Dağı, 8.–16.VII.1961, leg. SCHUBERT (NHMW); **126**: Sinop, Çangal Dağı, 16.–26.V.1957, leg. SCHUBERT (cAss); **127**: Artvin, Borçka, 1.–3.VI.1960, leg. SCHUBERT (NHMW); **128**: Erzurum, Dadaşköy, 1800 m, 7.VII.2005, leg. KESDEK (EME); **129**: Erzurum, Kombina, 5.IX.1988, leg. ÖZBEK (EME); **130**: Erzurum, Erzurum, 22.VI.1980, leg. ÖZBEK (EME); **131**: Rize, ca. 50 km SSE Rize, Ovitdağı geçidi, 40°38'N, 40°45'E, 2510 m, under stones, 25.VII.2006, leg. SCHÜLKE (cSch); **132**: Rize, 25 km

SSE Rize, 7 km E İkizdere, 40°47'N, 40°38'E, 1030 m, river bank, 31.VII.2006, leg. SCHÜLKE (cAss).

Omaliniinae

Acidota cruentata Mannerheim, 1830 – 96 (2)
Anthobium abantense (Fagel, 1968) – 23 (1)
Anthobium cf. *abantense* (Fagel, 1968) (dark morph) – 41 (1), 53 (1), 63 (1)
Anthobium anatolicum (Fagel, 1968) – 40 (6), 53 (4), 89 (1)
Anthobium fuscum (Erichson, 1839) – 59 (1), 62 (1), 92 (1)
Anthobium hamatum (Luze, 1905) – 8 (1), 35 (1)
Anthobium metallicum (Luze, 1905) – 62 (132), 92 (1)
Boreaphilus velox (Heer, 1839) – 88 (1)
Coryphodes anatolicus (Fagel, 1971) – 21 (1), 23 (1)
Coryphium angusticolle Stephens, 1834 – 23 (1)
Dialycera aspera (Eppelsheim, 1889) – 17 (1)
Dropephylla devillei (Bernhauer, 1902) – 63 (2)
Dropephylla ioptera (Stephens, 1834) – 2 (1), 22 (2), 25 (3), 26 (1), 32 (3)
Eusphalerum primulae Stephens, 1834 – 22 (2), 25 (1)
Eusphalerum caucasicum loebli Zanetti, 1993 – 13 (1), 22 (1)
Lesteva longoelytrata (Goeze, 1777) – 40 (1), 66 (5)
Mannerheimia brevipennis (Motschulsky, 1860) – 41 (2)
Omaliium assingi Zanetti, 2002 – 41 (2), 62 (2), 80 (2), 81 (1), 89 (1)
Omaliium cf. *cinnamomeum* Kraatz, 1857 (♀) – 9 (1)
Omaliium rivulare (Paykull, 1789) – 15 (1), 35 (2)
Omaliium rugatum Mulsant & Rey, 1880 – 62 (1), 78 (1), 80 (2)
Omaliium schuberti Zanetti, 2002 – 66 (1)
Omaliium turcicum Smetana, 1967 – 15 (1)
Olophrum cf. *piceum* (Gyllenhal, 1810) – 35 (1)

Proteininae

Megarthus depressus (Paykull, 1789) – 26 (1), 28 (1)
Metopsia assingi Zerche, 1998 – 26 (1), 28 (1)
Proteinus brachypterus (Fabricius, 1792) – 15 (1), 23 (4), 35 (1), 80 (2)
Proteinus longicornis Doderò, 1923 – 28 (1)
Proteinus ovalis Stephens, 1834 – 40 (18), 60 (1), 62 (80), 63 (1)
Proteinus utriarius Assing, 2004 – 21 (1), 29 (1), 41 (1), 62 (6)

Micropeplinae

Arrhenopeplus turcicus (Coiffait, 1958) – 78 (4)
Micropeplus fulvus Erichson, 1840 – 39 (27), 40 (28), 53 (31), 70 (194), 78 (1), 89 (5)
Micropeplus marietti Jacquelin du Val, 1857 – 11 (1)

Tachyporinae

Bryoporus multipunctus Hampe, 1867 – 73 (1), 84 (1)
Ischnosoma meyerbohmi Schülke, 2003 – 89 (1)
Ischnosoma splendidum (Gravenhorst, 1806) – 16 (2), 28 (5)
Lordithon exoletus (Erichson, 1839) – 40 (5), 53 (2), 76 (1), 83 (2)
Lordithon lunulatus (Linnaeus, 1760) – 6 (1), 22 (2), 30 (1), 32 (1)
Lordithon thoracicus thoracicus (Fabricius, 1777) – 6 (1), 22 (1), 30 (1), 40 (2), 52 (1), 53 (2)
Lordithon trinotatus (Erichson, 1839) – 62 (11)
Mycetoporus eppelsheimianus Fagel, 1968 – 41 (1)
Mycetoporus glaber glaber (Sperk, 1835) – 6 (1), 70 (1), 72 (1)
Mycetoporus ignidorsum Eppelsheim, 1880 (♀♀) – 26 (1), 57 (1)
Mycetoporus imperialis Bernhauer, 1902 – 39 (2), 93 (1)

- Mycetoporus punctus* (Gravenhorst, 1806) – 8 (1), 23 (2)
Mycetoporus reichei (Pandellé, 1869) – 1 (1), 6 (1), 70 (2), 71 (3), 72 (6), 73 (1), 89 (1)
Mycetoporus rufescens (Stephens, 1832) – 35 (1)
Mycetoporus spp. (*baudueri* group) – 10 (1), 20 (1), 25 (1), 40 (2), 53 (3), 60 (1), 71 (1), 77 (1), 83 (1), 93 (2)
Mycetoporus sp. (*nigricollis* group ♀♀) – 1 (1), 9 (1), 78 (1)
Parabolitobius inclinans (Gravenhorst, 1806) – 73 (1)
Parabolitobius ponticus (Fagel, 1968) – 32 (1)
Sepedophilus immaculatus (Stephens, 1832) – 10 (1), 11 (1), 20 (3), 24 (1), 25 (1), 70 (1), 92 (3)
Sepedophilus obtusus (Luze, 1902) – 39 (1), 43 (1), 52 (1), 53 (3), 72 (8), 73 (4), 77 (1), 78 (4), 80 (5), 81 (2), 83 (6), 84 (1), 87 (7), 89 (4), 93 (5)
Sepedophilus testaceus (Fabricius, 1792) – 25 (5), 29 (1), 39 (1), 40 (10), 52 (5), 53 (7), 70 (4), 71 (3), 72 (1), 78 (2), 82 (2)
Sepedophilus sp. – 38 (1)
Tachinus bonvouloiri Pandellé, 1869 – 41 (1), 55 (1), 59 (2)
Tachinus corticinus Gravenhorst, 1802 – 25 (1), 28 (2), 37 (2), 51 (1), 66 (1)
Tachinus discoideus Erichson, 1839 – 80 (1)
Tachinus fimetarius Gravenhorst, 1802 – 44 (1), 45 (1), 54 (1), 55 (4)
Tachinus lackneri Schülke, 2013 – 23 (1)
Tachinus rufipes (Linnaeus, 1758) – 39 (2)
Tachyporus assingi Schülke, 1997 – 11 (1)
Tachyporus cf. *atriceps* Stephens, 1832 (♀) – 89 (1)
Tachyporus caucasicus Kolenati, 1846 – 11 (1), 12 (1), 19 (3), 20 (1), 30 (2), 52 (1)
Tachyporus chrysomelinus (Linnaeus, 1758) – 43 (1)
Tachyporus hypnorum (Fabricius, 1775) – 60 (1)
Tachyporus nitidulus (Fabricius, 1781) – 53 (1), 62 (1), 64 (1), 68 (3), 72 (2), 83 (3)
Tachyporus pusillus Gravenhorst, 1806 – 43 (1), 51 (1), 66 (1)
Tachyporus solutus Erichson, 1839 – 12 (1)
- Habrocerinae
- Habrocerus pisidicus* Korge, 1971 – 10 (1), 11 (2), 40 (46), 53 (28), 70 (12), 76 (1), 78 (4), 95 (3)
Habrocerus simulans Assing & Wunderle, 1995 – 73 (2)
- Aleocharinae
- Acrotona piceorufa* (Mulsant & Rey, 1873) – 70 (45)
Aleochara grandeguttata Assing, 2009 – 52 (1), 68 (2), 93 (1)
Aleochara haematoptera Kraatz, 1858 – 113 (2)
Aleochara hamulata Assing, 2009 – 35 (1)
Aleochara lanuginosa Gravenhorst, 1802 – 124 (1)
Aleochara laticornis Kraatz, 1856 – 20 (1), 71 (1)
Aleochara subtumida (Hochhuth, 1849) – 6 (1), 25 (1), 32 (1)
Alevonota libanotica (Fagel, 1965) – 68 (1), 73 (1), 78 (4), 92 (1)
Alevonota rufotestacea (Kraatz, 1856) – 75 (1), 82 (5), 92 (1)
Aloconota gregaria (Erichson, 1839) – 49 (1), 50 (1), 51 (1), 67 (1)
Amarochara siculifera Assing, 2002 – 79 (1), 86 (1)
Amischa bifoveolata (Mannerheim, 1830) – 28 (3)
Amischa filum (Mulsant & Rey, 1870) – 68 (16)
Amischa forcipata Mulsant & Rey, 1873 – 122 (2)
Amischa nigrofusca (Stephens, 1832) – 68 (1)
Anaulacaspis cf. *nigra* (Gravenhorst, 1802) – 67 (1)
Atheta aegra (Heer, 1841) – 37 (1), 54 (1), 56 (2), 57 (1), 66 (1)
Atheta aeneicollis (Sharp, 1869) – 71 (1), 78 (2), 87 (1), 89 (4)
Atheta atramentaria (Gyllenhal, 1810) – 26 (1)
Atheta benickiella Brundin, 1948 – 4 (1), 6 (1), 15 (5), 21 (14), 22 (2), 23 (6), 25 (4), 28 (6), 30 (2), 53 (8)
Atheta brachati n. sp. – 28 (2)
Atheta britanniae Bernhauer & Scheerpeltz, 1926 – 1 (1), 23 (1)
Atheta castanoptera (Mannerheim, 1830) – 17 (1), 28 (1)
Atheta dadopora Thomson, 1867 – 35 (2)
Atheta laevigata (Hochhuth, 1849) – 50 (3)
Atheta palustris (Kiesenwetter, 1844) – 2 (1)
Atheta putrida (Kraatz, 1856) – 28 (1), 41 (5)
Atheta scabens n. sp. – 131 (1), 132 (1)
Atheta sodalis (Erichson, 1837) – 23 (4), 25 (3), 28 (3)
Atheta testaceipes (Heer, 1839) – 53 (1)
Atheta tibialis (Heer, 1839) – 28 (1)
Atheta trinotata (Kraatz, 1856) – 28 (1)
Atheta (Microdota) spp. – 13 (1), 52 (1), 55 (7), 56 (1), 58 (1), 60 (2), 78 (1), 79 (3), 82 (3), 85 (1), 89 (2), 92 (1)
Atheta (Mocytta) spp. – 11 (1), 30 (1), 41 (1), 76 (1), 78 (10), 86 (1), 87 (1), 89 (2), 92 (1)
Atheta sp. – 13 (2)
Bolitochara bella Märkel, 1844 – 16 (1), 36 (1)
Bolitochara laufferi Bernhauer, 1908 – 53 (6), 62 (2), 63 (6), 70 (1), 80 (1)
Callicerus rigidicornis (Erichson, 1839) – 35 (1)
Calodera aethiops (Gravenhorst, 1802) – 12 (1)
Caloderina hierosolymitana (Saulcy, 1865) – 86 (1), 114 (2), 115 (1), 116 (1)
Cordalia anatolica Assing, 2001 – 70 (10)
Cousya crocea Assing, 2004 – 82 (4), 89 (1)
Cousya microdotoides n. sp. – 79 (4), 81 (3)
Cousya spp. – 41 (1), 58 (3), 62 (1), 64 (2), 92 (1)
Cypha spathulata Assing, 2007 – 83 (1), 89 (1)
Cypha tarsalis (Luze, 1902) – 25 (1)
Cypha tenebricosa Assing, 2004 – 91 (1)
Drusilla limata Assing, 2005 – 56 (7), 57 (20), 59 (1)
Enalodroma hepatica (Erichson, 1839) – 53 (2)
Geostiba aequa Assing, 2011 – 73 (4)
Geostiba fodens Assing, 2011 – 56 (19), 57 (26), 58 (6)
Geostiba lucens (Benick, 1970) – 93 (2), 94 (1)
Geostiba oertzeni (Eppelsheim, 1888) – 4 (3), 5 (6), 6 (1), 20 (2), 23 (1), 24 (1), 35 (9)
Geostiba perfodens Assing, 2011 – 54 (6), 55 (186)
Geostiba pungens Assing, 2011 – 68 (6), 69 (25), 70 (1)
Geostiba rhodiensis Pace, 1983 – 85 (2), 89 (42)
Geostiba sultanica Assing, 2008 – 40 (1), 53 (1)
Geostiba uhligi Pace, 1983 – 1 (16), 8 (31)
Geostiba (Tropogastrosipalia) sp. (♀) – 23 (1)
Gyrophaena affinis Mannerheim, 1830 – 6 (1)
Gyrophaena bihamata Thomson, 1867 – 24 (1), 30 (2)
Gyrophaena gentilis Erichson, 1839 – 5 (1), 15 (1), 26 (7), 32 (1), 33 (1), 34 (1)
Gyrophaena joyioides Wüsthoff, 1937 – 22 (8), 25 (1), 26 (4), 30 (1)
Holobus flavicornis (Lacordaire, 1835) – 22 (1)
Homoeusa spp. – 70 (2), 75 (2), 76 (1), 82 (1), 84 (20)
Leptusa confinis Pace, 1982 – 6 (1), 17 (1)
Leptusa asiatica Bernhauer, 1909 – 6 (3), 8 (10), 13 (2), 14 (3), 15 (7), 16 (1), 17 (1), 21 (11), 22 (17), 23 (41), 24 (3), 25 (6), 26 (13), 27 (4), 28 (9), 29 (7), 30 (6), 31 (1), 32 (6), 33 (3), 34 (2), 35 (15), 36 (1)
Leptusa fuliginosa (Aubé, 1850) – 35 (2)

- Leptusa merkli* Bernhauer, 1900 – 1 (13), 2 (3), 3 (7), 5 (4), 6 (11), 7 (1), 9 (5), 18 (1), 20 (6)
Liogluta akiana Assing, 2004 – 55 (1), 56 (1), 68 (11)
Liogluta falcata Assing, 2010 – 23 (2)
Liogluta longiuscula (Gravenhorst, 1802) – 5 (1), 13 (1), 25 (1), 53 (1), 57 (10), 62 (16), 75 (1), 79 (1), 82 (1)
Maurachelia roubali (Lohse, 1970) – 67 (1)
Megalogastria cingulata (Eppelsheim, 1889) – 9 (2)
Meotica decolor Assing, 2004 – 78 (4)
Meotica subnigra Assing, 2006 – 108 (1)
Meotica truncata Assing, 2004 – 73 (1)
Meotica marchica Benick, 1953 – 4 (1)
Meotica wunderlei n. sp. – 67 (10)
Myrmecopora effeminata Assing, 2004 – 76 (1), 77 (4), 78 (6), 87 (2), 92 (1), 95 (4)
Myrmoecia perezii (Uhagón, 1876) – 2 (1)
Myrmoecia plicata (Erichson, 1837) – 83 (1)
Notothecta flavipes (Gravenhorst, 1806) – 40 (1)
Ocalea sp. aff. *ruficollis* Eppelsheim, 1888 – 80 (1)
Ocalea sp. – 6 (1), 7 (1), 8 (1), 12 (3), 15 (1), 16 (3), 22 (3), 23 (1), 25 (3), 41 (1)
Oligota pumilio Kiesenwetter, 1858 – 39 (1), 40 (1), 55 (1), 109 (2), 110 (1), 111 (1)
Oxypoda abdominalis (Mannerheim, 1830) – 112 (3)
Oxypoda acuminata (Stephens, 1832) – 22 (1)
Oxypoda antennata Bernhauer, 1902 – 28 (1), 40 (2), 41 (1), 45 (1), 62 (2)
Oxypoda brachati Assing, 2004 – 75 (1), 85 (2), 90 (1)
Oxypoda brevicornis (Stephens, 1832) – 1 (3), 4 (2)
Oxypoda obscuricollis Assing, 2007 – 62 (1)
Oxypoda cristata Assing, 2006 – 28 (2), 40 (1), 41 (2), 62 (3), 64 (2), 81 (1), 93 (2), 94 (1), 97 (3), 98 (1), 99 (2), 100 (1), 101 (8), 105 (1)
Oxypoda fissa Assing, 2006 – 41 (2), 54 (5), 55 (3), 64 (1)
Oxypoda flavicornis Kraatz, 1856 – 1 (4), 15 (5), 28 (23), 30 (1), 79 (2)
Oxypoda ignorata Zerche, 1996 – 62 (5)
Oxypoda libanotica Fagel, 1965 – 68 (4), 70 (1)
Oxypoda longipes Mulsant & Rey, 1861 – 62 (2)
Oxypoda lurida Wollaston, 1857 – 74 (1), 78 (2), 91 (1)
Oxypoda miricornis Assing, 2009 – 80 (1)
Oxypoda opaca (Gravenhorst, 1802) – 13 (6), 23 (1), 26 (2), 35 (4)
Oxypoda pungens Assing, 2012 – 23 (1)
Oxypoda scheerpeltziana (Fagel, 1968) – 80 (1), 81 (4), 83 (2), 88 (3), 92 (1), 93 (5)
Oxypoda subspectabilis Assing, 2007 – 123 (1)
Oxypoda vicina Kraatz, 1858 – 46 (1)
Oxypoda (Baeoglana) spp. – 9 (1), 13 (3), 23 (1), 25 (2), 28 (3), 29 (10), 40 (43), 41 (1), 52 (1), 53 (35), 71 (8), 78 (1), 84 (2)
Oxypoda spp. (♀♀) – 67 (1), 68 (2), 69 (1)
Pella erratica (Hagens, 1863) – 73 (1)
Pella similis (Märkel, 1844) – 12 (1)
Pella sp. aff. *humeralis* (Gravenhorst, 1802) – 2 (2), 13 (1), 17 (1), 20 (1), 23 (5), 24 (1), 25 (2), 26 (2), 30 (1), 32 (1)
Peltonia bodemeyeri (Bernhauer, 1936) – 95 (1)
Thiasophila angulata (Erichson, 1837) – 40 (14)
Tropimenelytron tuberiventre (Eppelsheim, 1880) – 125 (1), 126 (1), 127 (1)
- Oxytelinae
- Anotylus clypeonitens* (Pandellé, 1867) – 7 (1), 12 (1)
Anotylus inustus (Gravenhorst, 1806) – 5 (1), 74 (2), 87 (1), 92 (1)
- Anotylus pumilus* (Erichson, 1839) – 39 (1)
Anotylus sculpturatus (Gravenhorst, 1806) – 33 (1)
Aploderus lydicus Assing, 2007 – 53 (6)
Carpelimus atomus (Saulcy, 1864) – 106 (1), 107 (1)
Carpelimus corticinus (Gravenhorst, 1806) – 45 (1)
Platystethus brevipennis Baudi di Selve, 1857 – 51 (1), 60 (29), 67 (1)
Platystethus nitens (Sahlberg, 1832) – 11 (1), 12 (2), 68 (1)
- Steninae
- Stenus bithynicus* n. sp. – 1 (1), 7 (1), 8 (5), 15 (1), 33 (2)
Stenus capitulatus Assing, 1995 – 58 (1)
Stenus coarcticollis Eppelsheim, 1890 – 20 (1), 24 (1), 25 (1), 28 (4), 31 (1)
Stenus glacialis Heer, 1839 – 18 (1), 53 (5), 59 (3), 70 (3), 86 (2)
Stenus impressus Germar, 1824 – 11 (1)
Stenus ochropus Kiesenwetter, 1858 – 53 (5), 70 (12), 72 (2), 83 (5), 89 (4), 92 (1)
Stenus subaeneus Erichson, 1840 – 35 (4), 87 (1)
Stenus turbulentus Bondroit, 1912 – 10 (2), 11 (2), 12 (2), 70 (16), 75 (1), 76 (4), 77 (2), 78 (4), 82 (4), 87 (6), 88 (2), 89 (4), 95 (2)
- Paederinae
- Achenium anatolicum* Jarrige, 1952 – 66 (89), 67 (18), 68 (3)
Astenus lyonessius (Joy, 1908) – 69 (1)
Astenus procerus (Gravenhorst, 1806) – 74 (1)
Astenus sultanicus Assing, 2010 – 54 (6), 55 (86)
Astenus thoracicus (Baudi di Selve, 1857) – 42 (1), 43 (2), 49 (1), 54 (1), 55 (2), 56 (15), 57 (18), 58 (1), 61 (1), 65 (4), 66 (8), 67 (1)
Astenus sp. aff. *thoracicus* (Baudi di Selve, 1857) – 10 (1), 74 (11), 75 (1), 77 (1)
Astenus spp. (♀♀) – 49 (1), 56 (1), 67 (1)
Homaeotarsus chaudoirii (Hochhuth, 1851) – 87 (1), 121 (1)
Lathrobium bodemeyeri Bernhauer, 1903 – 10 (1), 13 (2), 14 (1), 16 (6), 26 (1), 32 (1), 36 (4)
Leptobium gracile (Gravenhorst, 1802) – 46 (3), 66 (7), 67 (67), 68 (27)
Leptobium syriacum (Saulcy, 1865) – 74 (3), 86 (1)
Luzea graeca (Kraatz, 1857) – 67 (1), 68 (3)
Medon brunneus (Erichson, 1839) – 9 (4)
Medon dilutus pythonissa (Saulcy, 1865) – 10 (1), 11 (2), 12 (5), 53 (1), 73 (2), 84 (1), 92 (1)
Medon ferrugineus (Erichson, 1840) – 11 (1), 12 (1), 54 (4)
Medon fuscus (Mannerheim, 1830) – 11 (2), 54 (1), 55 (1), 70 (1), 71 (2), 72 (2), 73 (2)
Medon lanugo Assing, 2004 – 70 (2), 71 (8), 72 (1), 73 (2), 78 (1), 87 (5)
Medon maronitus (Saulcy, 1865) – 70 (2), 78 (1)
Medon meyhohmi Assing, 2007 – 89 (4)
Medon semiobscurus (Fauvel, 1875) – 70 (3), 71 (6), 76 (1), 77 (1), 78 (5), 82 (5), 86 (1), 89 (2)
Micranops pilicornis (Baudi di Selve, 1870) – 50 (1), 78 (10)
Rugilus lesbius Assing, 2005 – 11 (1), 12 (2), 25 (1), 26 (1)
Rugilus rufipes Germar, 1836 – 4 (1), 16 (2)
Rugilus tauricus (Rougemont, 1988) – 40 (2), 70 (13), 83 (1 ♀)
Scopaeus bituberculatus Frisch, 2002 – 87 (1)
Scopaeus debilis Hochhuth, 1851 – 4 (1)
Scopaeus loebli Frisch, 1997 – 78 (1)
Scopaeus trifurcatus Frisch, 2002 – 55 (4)

Scymbalium anale (Nordmann, 1837) – 66 (2), 67 (4)
Sunius adanensis (Lokay, 1919) – 42 (3), 86 (1), 89 (2)
Sunius aequus Assing, 2011 – 68 (4), 69 (93)
Sunius melanocephalus (Fabricius, 1792) – 2 (1), 12 (1), 29 (2),
 41 (1), 55 (3)
Sunius sultanicus Assing, 2011 – 54 (13), 55 (221)
Sunius ulcerosus Assing, 2011 – 66 (17)

Staphylininae

Atrecus affinis (Paykull, 1789) – 15 (1), 21 (2)
Bisnius fimetarius (Gravenhorst, 1802) – 14 (1), 15 (1), 26 (2),
 32 (1), 34 (1)
Gabrius breviventer (Sperk, 1835) – 26 (1)
Gabrius spp. (♀♀) – 46 (1), 53 (1), 68 (1)
Gauropterus sanguinipennis (Kolenati, 1846) – 102 (6)
Gyrohypnus angustatus Stephens, 1833 – 23 (1), 40 (9), 128 (1),
 129 (1), 130 (1)
Gyrohypnus punctulatus (Paykull, 1789) – 33 (1)
Heterothops cf. *dissimilis* (Gravenhorst, 1802) – 40 (1), 55 (5),
 84 (1)
Heterothops cf. *minutus* Wollaston, 1860 – 57 (1)
Heterothops sp. – 45 (1)
Leptacinus nigerrimus Coiffait, 1971 – 68 (2)
Megalinus scutellaris (Fauvel, 1900) – 103 (1)
Nudobius lentus (Gravenhorst, 1806) – 26 (1)
Ocypus curtippennis Motschulsky, 1849 – 85 (1)
Ocypus fulvipennis Erichson, 1840 – 16 (1)
Ocypus mus (Brullé, 1832) – 51 (1), 84 (1)
Ocypus picipennis (Fabricius, 1792) – 42 (2), 54 (1), 58 (1),
 65 (1), 66 (12), 67 (13)
Othius laeviusculus Stephens, 1833 – 79 (1), 92 (1)
Othius lapidicola Märkel & Kiesenwetter, 1848 – 15 (1), 21 (2),
 28 (1), 39 (12), 40 (2), 41 (3), 53 (6), 73 (1)
Othius punctulatus (Goeze, 1777) – 32 (1), 39 (3), 40 (1)
Philonthus carbonarius (Gravenhorst, 1802) – 41 (1)
Philonthus concinnus (Gravenhorst, 1802) – 38 (1), 55 (1), 66 (1)
Philonthus debilis (Gravenhorst, 1802) – 47 (1)
Philonthus intermedius (Lacordaire, 1835) – 2 (1)
Philonthus parvicornis (Gravenhorst, 1802) – 68 (1)
Philonthus quisquiliarius (Gyllenhal, 1810) – 50 (1)
Platydracus chalcocephalus (Fabricius, 1801) – 1 (1)
Quedius boluensis Korge, 1971 – 25 (1), 30 (1)
Quedius boops (Gravenhorst, 1802) – 15 (1), 21 (1), 36 (1)
Quedius cinctus (Paykull, 1790) – 21 (2)
Quedius coloratus Fauvel, 1875 – 71 (1), 73 (1), 84 (1), 87 (1)
Quedius humeralis Stephens, 1832 – 35 (2)
Quedius lateralis (Gravenhorst, 1802) – 70 (1)
Quedius limbatus (Heer, 1839) – 8 (1), 20 (2), 23 (2), 30 (1), 36 (2)
Quedius cf. *molochinus* (Gravenhorst, 1806) (♀) – 67 (1)
Quedius nemoralis Baudi di Selve, 1848 – 1 (2), 9 (4), 20 (1),
 21 (1), 23 (1), 29 (1), 53 (10), 80 (1), 82 (1)
Quedius nitipennis (Stephens, 1833) – 9 (1)
Quedius cf. *ochropterus* Erichson, 1840 (♀♀) – 40 (3), 41 (1)
Quedius pseudonigriceps Reitter, 1909 – 9 (1), 10 (1), 72 (1),
 73 (2), 89 (3)
Quedius semiaeneus (Stephens, 1833) – 9 (1), 10 (1)
Quedius umbrinus Erichson, 1839 – 16 (3), 31 (1), 32 (2)
Quedius spp. (♀♀) – 35 (1), 39 (1), 40 (4), 53 (2), 62 (2), 70 (2),
 86 (1), 89 (1)
Stenistoderus cephalotes (Kraatz, 1858) – 46 (2), 67 (1)
Tasgius falcifer (Nordmann, 1837) – 38 (1)

Xantholinus audrasi Coiffait, 1956 – 7 (1), 16 (1), 84 (1)
Xantholinus ciliciae Bordoni, 1971 – 71 (1), 75 (1), 78 (1), 86 (1),
 95 (1)
Xantholinus laevigatus Jacobsen, 1849 – 28 (1), 32 (1)
Xantholinus rufipennis Erichson, 1839 – 43 (1), 104 (1), 117 (1),
 118 (1), 119 (1), 120 (1)
Xantholinus spp. (♀♀) – 38 (1), 43 (2), 46 (4), 60 (2), 61 (1),
 66 (4)

3.2 Remarks on some species

Acidota cruentata Mannerheim, 1830

This species has a trans-Palaeartic distribution (SME-TANA 2004), but was reported from Turkey (Kastamonu) for the first time only recently (ASSING 2011a). The specimens listed in section 3.1 represent the first record from southern Turkey. The aedeagus of the male from Antalya is somewhat smaller than that of material seen from other regions, but otherwise no evidence was found suggesting that the specimens should represent a distinct species. The elytra are distinctly longer than those of *Acidota brevis* Assing, 2004. Both specimens are micropterous.

Anthobium abantense (Fagel, 1968)

According to the original description, which is based on two males from the vicinity of the Abant Gölü (Bolu), *A. abantense* is distinguished from *L. atrocephalum* (Gyllenhal, 1827) by the darker coloration and by some minor differences in body proportions and other external characters (FAGEL 1968). Material matching the original description was examined from northwestern Turkey eastwards to Kastamonu, from western Anatolia (Izmir, Aydın), and from the Sultan Dağları in Konya. Some of these records were reported as *A. atrocephalum* by ASSING (2011a). Specimens of the typical coloration of *A. atrocephalum* have so far been seen only from northeastern Anatolia (Rize). Whether or not *A. abantense* and *A. atrocephalum* really represent different species requires clarification in the context of a revision on a larger scale. I have seen material with a similar coloration and other external characters also from various regions in Greece and from other Balkan countries.

Another morph of even darker coloration was recorded from southern Anatolia, from Afyon and Konya in the west to Adana and Adıyaman in the east. The records from the latter two provinces were reported as *A. atrocephalum* by ASSING (2004b, 2006a). At present, it is unclear if these populations represent a regional colour variation of *A. atrocephalum* or *A. abantense*, or a distinct species.

Anthobium metallicum (Luze, 1905) (Fig. 1)

Type material examined: Holotype (♀), “*melanoceph.* Akbes, Syria. STAUDINGER. / c. EPPLSH. STEIND. d. / Type. *metallicum* Luze / Typus” (NHMW).

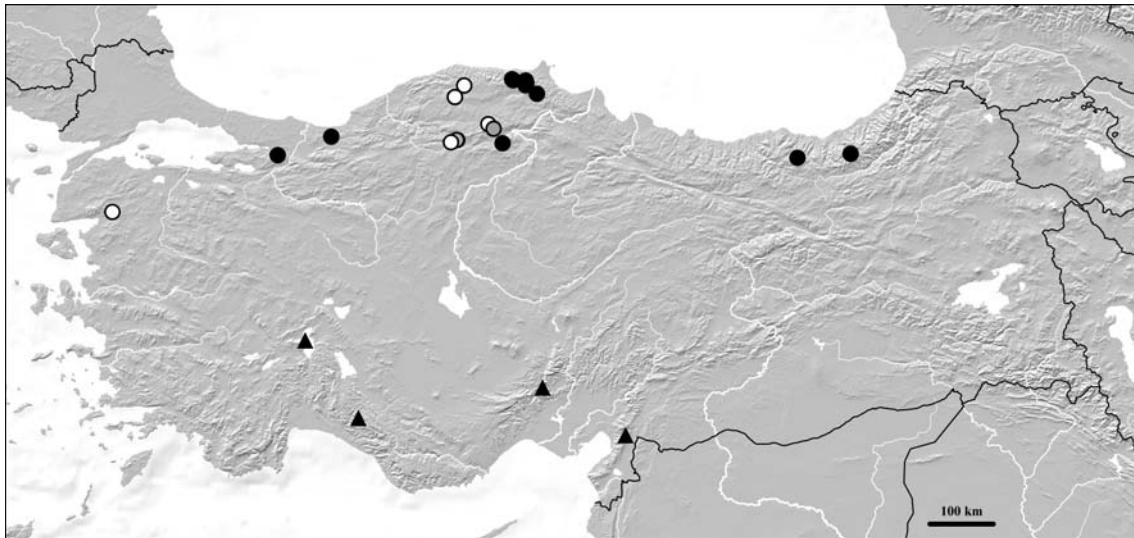


Fig. 1. Distribution of the species of the *Anthobium melanocephalum* group in Turkey: *A. melanocephalum* (white and grey circles); *A. hamatum* (black and grey circles); *A. metallicum* (triangles).

The original description is based on a single female from “Syrien (Akbes)” (LUZE 1905), which is probably Akbez in the Turkish province Hatay today. SCHEERPELTZ (1961) reported the species from Lebanon and Syria.

The species is distinguished from the similar and closely related *A. melanocephalum* (Illiger, 1794) particularly by the following characters: head posteriorly without transverse furrow connecting the ocelli, with less pronounced lateral furrows, and with flat median dorsal portion (*A. melanocephalum*: ocelli connected by more or less pronounced transverse furrows, lateral furrows pronounced; median dorsal portion of head convexly and often somewhat irregularly elevated); pronotum less broad and less transverse, with less strongly excavated anterior margin, and with less marked posterior angles; lateral margins of pronotum less strongly convex and – at most – very indistinctly sinuate posteriorly. According to LUZE (1905) and SCHEERPELTZ (1961), *A. metallicum* is characterised by a metallic hue on the head and pronotum, but such a hue is absent or indistinct both in the holotype and in the Turkish material examined.

The two specimens reported as *A. melanocephalum* from Antalya by ASSING (2004b) refer to *A. metallicum*. In Turkey, the distribution of *A. metallicum* is confined to the south (Fig. 2).

Anthobium melanocephalum (Illiger, 1794) (Fig. 1)

This species was recorded from Turkey by ASSING (2010a, 2011a). Since the previous record from Antalya is based on a misidentification, the distribution of *A. melanocephalum* in Turkey is confined to the northwest (Fig. 1).

Anthobium hamatum (Luze, 1905) (Fig. 1)

Anthobium hamatum had been known only from the Caucasus region (Georgia, Armenia) until recently when it was reported also from Turkey (ASSING 2010a, 2011a). In two localities it was found together with *A. melanocephalum*. In Turkey, its distribution is confined to the north (Fig. 1).

Coryphoides anatolicus (Fagel, 1971) (Fig. 2)

This rare and local species is endemic to northwestern Anatolia. Until recently, when it was reported from Sinop and Bolu (ASSING 2010a, 2011a), *C. anatolicus* had been recorded only from the type locality in the Uludağ (Bursa). The currently known distribution is illustrated in Fig. 2.

Coryphium angusticolle Stephens, 1834

The identification of the male specimen listed in section 3.1 is not absolutely certain. It has a somewhat larger and broader body than material of *C. angusticolle* that I have seen from other regions, but I have been unable to find any additional, convincing evidence that it should represent a distinct species. It is most unlikely to be conspecific with *C. turcicum* Zerche, 1993, whose description is based on a single teneral female from Artvin. According to ZERCHE (1993), *C. turcicum* has a forebody with pronounced microsculpture, which is absent in the specimen from Bolu. *Coryphium angusticolle* was previously unknown from Turkey.

Dropephylla ioptera (Stephens, 1834)

The previously known distribution of this widespread species ranged from the Balkans and Ukraine across East

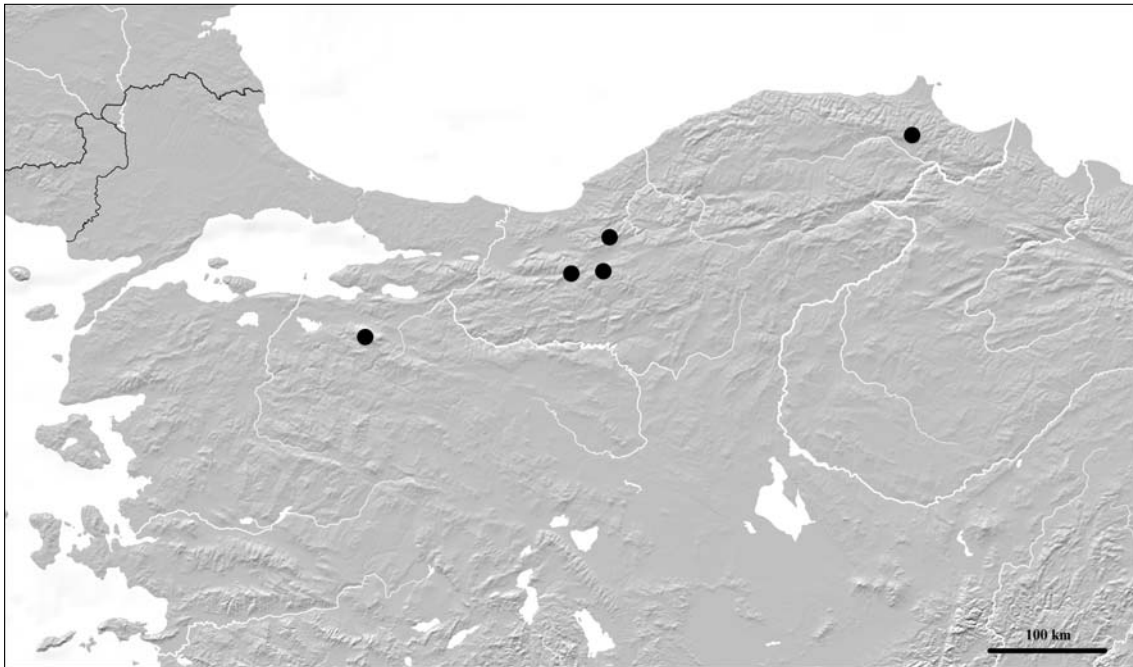


Fig. 2. Distribution of *Coryphoides anatolicus* in northwestern Turkey.

Europe, Central Europe, and Italy to France, the British Isles, and Scandinavia, with a record also from Algeria (SMETANA 2004). The specimens listed in section 3.1 expand the distribution to the southeast and represent the first records from Turkey.

Mannerheimia brevipennis (Motschulsky, 1860)

Additional material examined: Iraq: 1 ex., Rawanduz, 36°30'N, 44°36'E, 1200–1400 m, pitfall, XI.2007–III.2008, leg. REUTER (cFel).

The known distribution of this rare species ranges from Scandinavia to East Siberia. In Turkey, it had been reported from a few localities in Ankara, Kahramanmaraş, Adıyaman, Gümüşhane, and Kayseri provinces (ASSING 2007a). The material from Afyon (section 3.1) extends the distribution in Turkey further to the southwest. The above specimen from Iraq represents a new country record.

Proteinus longicornis Doderó, 1923

According to SMETANA (2004), the distribution of *P. longicornis* ranges from France across Central Europe and Italy to Ukraine. The specimen listed in section 3.1 represents the first record from Turkey and considerably expands the distribution towards the southeast.

Micropeplus marietti Jacquelin du Val, 1857

The distribution of *M. marietti* ranges from France to Turkmenistan. Very recently, it was reported from Turkey (Aksaray) for the first time (ASSING 2010a).

Parabolitobius ponticus (Fagel, 1968)

The original description of *P. ponticus* is based on five specimens from “Anatolie: Abant Dagħ” (FAGEL 1968). The male listed in section 3.1 represents the first record since the original description.

Tachyporus assingi Schülke, 1997

According to SCHÜLKE (2012), this species is distributed in the Balkans and Turkey, but primary records from Turkey were previously unknown.

Aleochara hamulata Assing, 2009

The distribution of *A. hamulata* includes Greece, Croatia, Iraq, and Anatolia, where it was previously known only from the southwest (ASSING 2009a). The specimen from Düzce (section 3.1) represents the first record from northwestern Turkey.

Aleochara subtumida (Hochhuth, 1849)

Aleochara subtumida, a rarely recorded species, is distributed in Turkey and the Caucasus region. For a map illustrating its distribution see ASSING (2009b).

Amarochara siculifera Assing, 2002

The distribution of *A. siculifera* is confined to central southern Anatolia, where it was previously known only from three localities in Mersin and Hatay (ASSING 2002, 2003a).

Calodera aethiops (Gravenhorst, 1802)

According to ASSING (1996), *C. aethiops* is widespread in the West Palaearctic region, but was previously unknown from Turkey. The male listed in section 3.1 represents a new country record. *Calodera aethiops* is the third species of the genus to be reported from Turkey.

Cypha tarsalis (Luze, 1902)

The previously known distribution of *C. tarsalis* ranges from West Europe across Italy, North, and Central Europe eastwards to the Russian Central European territory (SMETANA 2004). It had been reported neither from the Balkans nor from Turkey. The male listed in section 3.1 represents the first record from Turkey and, by far, the southeasternmost record of this species.

Drusilla limata Assing, 2005

Except for *D. canaliculata* (Fabricius, 1787), all the *Drusilla* species in southern Anatolia are local endemics. *Drusilla limata* was previously known from a few localities in Antalya and Karaman provinces (ASSING 2005, 2009c).

Leptusa merkli Bernhauer, 1900 (Fig. 3)

The distribution of *L. merkli* is confined to northwestern Turkey (ASSING 2009d) and adjacent to that of *L. asiatica* (Fig. 3).

Leptusa asiatica Bernhauer, 1909 (Fig. 3)

Leptusa flagellulifera ASSING, 2009d: 1290 ff.; **n. syn.**

Based on the shape and internal structures of the aedeagus, as well as for zoogeographic reasons, *L. flagellulifera*

was attributed to the subgenus *Stictopisalia* Scheerpeltz, 1966, and distinguished from its Turkish representatives. While examining the material listed in section 3.1, however, it was discovered that *L. flagellulifera* is conspecific with *L. asiatica*, which has been attributed to *Oncopisalia* Pace, 1982 (PACE 1989). Apart from *L. asiatica*, this subgenus previously included four species, three from Macedonia (Greece) and one from Albania. Based on both external and the male sexual characters, it seems somewhat doubtful that *L. asiatica* should be more closely related to *L. monachorum* Bernhauer, 1902, the type species of *Oncopisalia*, than to *L. merkli* and other Turkish species of *Stictopisalia*. Externally, *Leptusa asiatica* and *L. merkli* are practically indistinguishable. Moreover, no significant differences were found in the male secondary sexual characters and the general morphology of the aedeagus, except for the somewhat asymmetric structure of the internal sclerites. The apical internal structures are very similar. Future studies with a phylogenetic approach will have to show if *Oncopisalia* represents a distinct taxon or if it is nested within *Stictopisalia*.

Unlike other Turkish species of *Stictopisalia*, *L. asiatica* appears to be widespread in Turkey. The vast majority of records is from northwestern Anatolia. However, several specimens from the vicinity of the Van lake and from Osmaniye were found in the Schubert collection at the NHMW (ASSING 2003b). Since recent records from these regions are unknown, the possibility that the respective specimens are mislabelled cannot be ruled out completely. In northwestern Anatolia, the distribution of *L. asiatica* is adjacent to – and partly slightly overlaps with – that of *L. merkli* (Fig. 3).

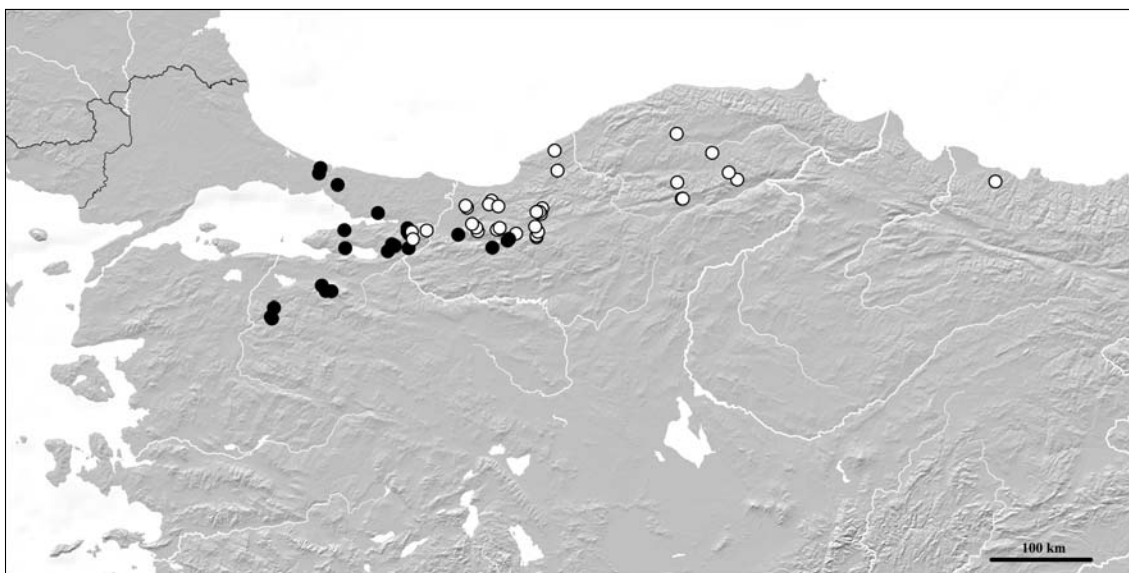


Fig. 3. Distributions of *Leptusa merkli* (black circles) and *L. asiatica* (white circles) in northwestern Turkey. Records of *L. asiatica* from other Turkish regions are omitted; they are mapped in ASSING (2003b).

Maurachelia roubali (Lohse, 1970)

Only few records of *M. roubali* have been reported from Austria, Slovakia, and Greece (ASSING 2004a). The specimen listed in section 3.1 represents the first record from Turkey.

Meotica marchica Benick, 1953 (Fig. 43)

The previously known distribution of *M. marchica* ranged from Central Europe to Greece (ASSING 2004a). The male listed in section 3.1 represents the first record from Turkey; its aedeagus is illustrated in Fig. 43.

Meotica subnigra Assing, 2006

Additional material examined. Greece: 1 ♂, Pelopónnisos, 10 km NE Githio, 36°24'N, 22°37'E, 23.III.1997, leg. ASSING (cAss).

This species was previously known only from the type locality in Kahramanmaraş (ASSING 2006b). The specimen from Muğla (section 3.1) represents the second record from Turkey, the above male from the Pelopónnisos the first record from Greece.

Myrmoecia perezii (Uhagón, 1876)

This extremely rare species was originally described based on two specimens found in association with "*Tapinoma erraticum* Latr." in Badajoz (Spain) (UHAGÓN 1876). It was subsequently reported also from Morocco, Slovakia, and the Czech Republic, always with the ant "*Tapinoma erraticum* (Latreille, 1789)" (BERNHAEUER 1940, DVOŘÁK 1965, ROUBAL 1932, SMETANA 2004, VÁVRA 2002). The male from Kocaeli (section 3.1), which was compared with a specimen from Slovakia (deposited in NHMW) and which too was found associated with *Tapinoma* sp., considerably expands the known distribution towards the southeast and represents the first record from Turkey.

Oxypoda obscuricollis Assing, 2007

Oxypoda obscuricollis was previously known only from two localities in Mersin and Kahramanmaraş provinces (ASSING 2007b, 2009e).

Oxypoda miricornis Assing, 2009

The specimen from Adana (section 3.1) represents the first record since the original description, which is based on two males from the type locality in Kahramanmaraş (ASSING 2009e).

Oxypoda pungens Assing, 2012

The distribution of this very recently described species includes both northern and southern Anatolia. The specimen from Bolu (section 3.1) represents a new province record.

Pella erratica (Hagens, 1863)

The distribution of *P. erratica* includes practically all of the Mediterranean and northwards extends into Central Europe, but records, particularly recent ones, are extremely scarce. Very recently, it was reported from Turkey (Sinop) for the first time (ASSING 2009c). The specimen from Niğde (section 3.1) represents the second record from Turkey and the first record from southern Anatolia.

Peltodonia bodemeyeri (Bernhauer, 1936)

Peltodonia bodemeyeri, the sole representative of the genus known from the West Palaearctic region, is widespread in Turkey, but rather rare. Its distribution is mapped by ASSING (2009c). The specimen from Adana represents a new province record.

Carpelimus atomus (Saulcy, 1864)

The distribution of *C. atomus* extends from Northwest Africa to the Middle East and Afghanistan. The specimens from Muğla (section 3.1) represent the first records from Turkey.

Stenus capitulatus Assing, 1995

This rare myrmecophilous species was originally described from southern Greece and subsequently reported also from southern Anatolia (Antalya) (ASSING 2004b). The specimen from Isparta (section 3.1) was found in a nest of harvester ants (*Messor* sp.) and represents the second record from Turkey.

Astenus sultanicus Assing, 2010

The original description of this very recently described, myrmecophilous species is based on a single male from the Sultan Dağları in Isparta (ASSING 2010a). The 92 specimens listed in section 3.1 were found at the type locality, all of them associated with *Tetramorium* sp. (Formicidae).

3.3 Descriptions of new species

3.3.1 Aleocharinae

3.3.1.1 Athetini

Atheta (Philhygra) scabens n. sp.

(Figs. 4–11)

Type material

Holotype ♂: "RU [29] – W-Caucasus, 13 km SW Teberda, 1450 m, gravel river bank, 43°20'00"N, 41°39'57"E, 28.VII.2011, V. ASSING / Holotypus ♂ *Atheta scabens* sp. n. det. V. ASSING 2012" (cAss).

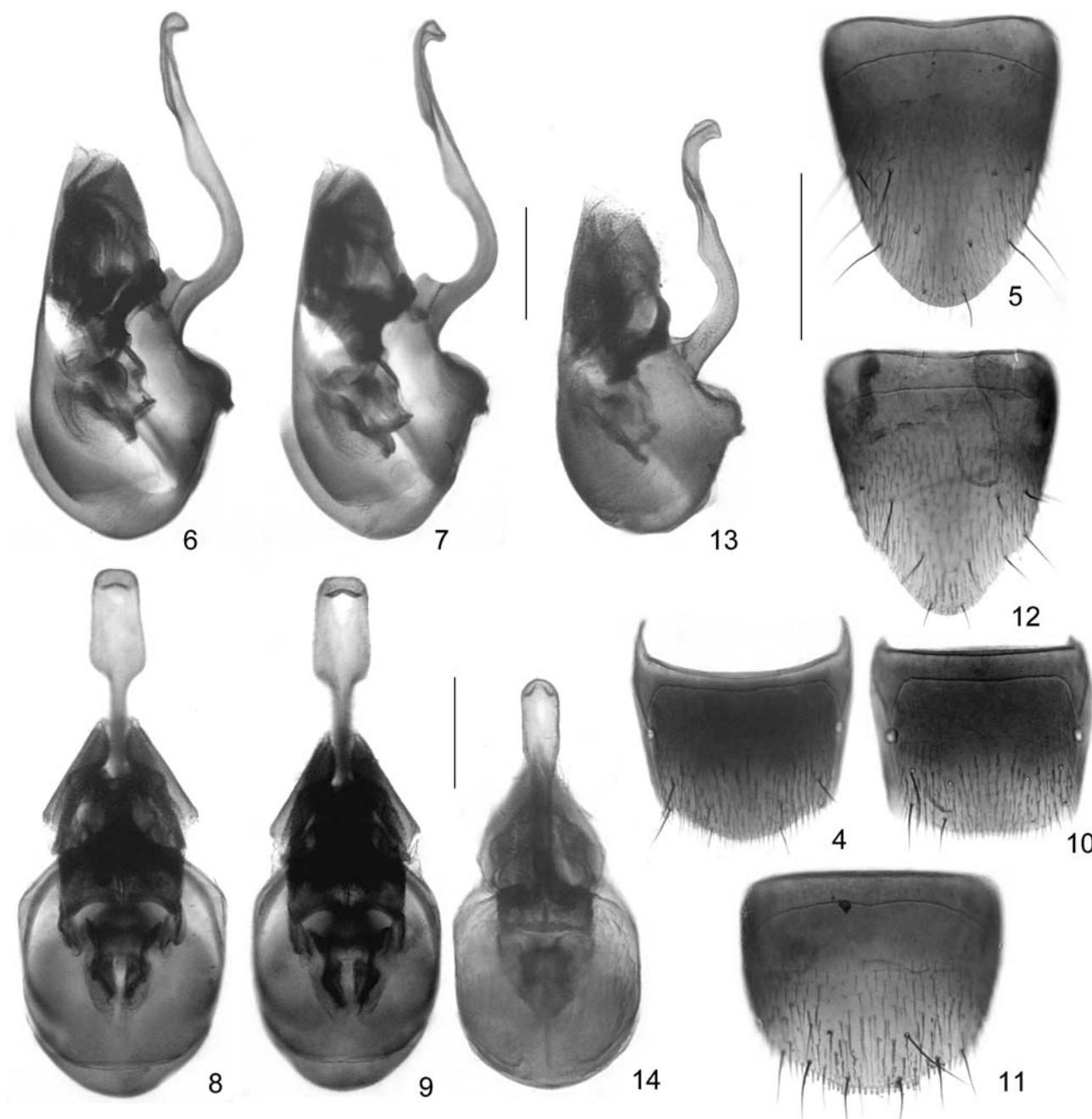
Paratypes: 3 ♂♂: same data as holotype; 1 ♂: "RU [19] – W-Caucasus, 4 km NNE Teberda, 1250 m, Teberda river bank, 43°29'20"N, 41°45'23"E, 24.VII.2011, V. ASSING" (cAss); 3 ♂♂, 4 ♀♀: "Kyrgyzstan/Talass, Tschytschkan-Valley, 15.–16.

VII.2003, 42°07'01N, 72°49'01E, 1700 m, leg. L. SCHMIDT" (cAss); 2 ♀♀: "Kyrgyzstan/Osch, NP Kara Shoro, Uzgenskij Chrebet, 20.VII.2003, 40°43'07N, 73°03'00E, 2900 m, leg. L. SCHMIDT" (cAss); 5 ♂♂, 1 ♀, 6 sex?: "Asia centr. Tadhikist., Pamir Alai, Hissar Mts., Adshuk-Cleft near Warsob, 1200 m, 1.-3.VII.1990, leg. SCHÜLKE & WRASE" (cAss, cSch); 1 ♂: "Tadshikistan, Hissar Alai, Warsob cleft, Rd. to Ansob-pass, km 55, 2000 m, Snowfieldedge [sic], 27.VI.1999, leg. M. SCHÜLKE" (cSch); 1 ♂: "TR [5] – Rize, ca. 50 km SSE Rize, Ovitdağı Geç.,

2510 m, under stones, 40°37'31N, 40°45'27E, 25.VII.2006, M. SCHÜLKE" (cSch); 1 ♂: "TR [24a] – Rize, 25 km SSE Rize, 7 km E İkizdere, 1030 m, river bank, 40°47'01N, 40°38'18E, 31.VII.2006, M. SCHÜLKE" (cAss).

Etymology

The specific epithet is the present participle of the Latin verb *scabere* and alludes to the shape of the ventral process of the aedeagus, which somewhat resembles a scraper.



Figs. 4–14. *Atheta scabens* n. sp. (4–11) and *A. hygrotopora* (12–14). – 4. Male tergite VIII. 5, 12. Male sternite VIII. 6–7, 13. Median lobe of aedeagus in lateral view. 8–9, 14. Median lobe of aedeagus in ventral view. 10. Female tergite VIII. 11. Female sternite VIII. – Scale bars: 0.5 mm (4–5, 10–12), 0.2 mm (6–9, 13–14).

Description

Body length 4.0–5.4 mm; length of forebody 1.9–2.2 mm. External characters as in *Atheta hygrotopora* (Kraatz, 1856); distinguished only by the primary and secondary sexual characters.

♂: tergite VIII broadly convex posteriorly (Fig. 4); sternite VIII oblong, distinctly longer than tergite VIII, posterior margin convexly produced (Fig. 5); median lobe of aedeagus 0.90–0.95 mm long and with ventral process of distinctive shape (Figs. 6–9).

♀: posterior margin of tergite VIII very weakly convex and in the middle weakly concave (Fig. 10); sternite VIII transverse, posterior margin broadly convex and with stout marginal setae (Fig. 11).

Comparative notes

Based on the similar external morphology, the similar modifications of the male and female terminalia, as well as on the similar morphology of the aedeagus, *A. scabens* is the adelphotaxon of *A. hygrotopora*, from which it is readily distinguished by the different shape of the male sternite VIII (*A. hygrotopora*: more narrowly produced posteriorly; Fig. 12) and by the morphology of the median lobe of the aedeagus (*A. hygrotopora*: smaller, 0.75–0.80 mm long; ventral process apically much less dilated and of different shape; Figs. 13, 14).

Distribution and natural history

Atheta scabens is currently known from northeastern Anatolia, the West Caucasus, Kyrgyzstan, and Tajikistan, which suggests that previous records of *A. hygrotopora* from Turkey, the Caucasus region, and Middle Asia probably refer to this species. Based on the similar habitat requirements and the evidently close relationship of both species, it seems likely that they have allopatric distributions. *Atheta hygrotopora* is widespread in Europe (material from Spain, Italy, Central Europe, and Bosnia-Herzegovina examined).

The type specimens of *A. scabens* were collected on the banks of rivers and streams, from under stones, and near the edge of a snowfield at altitudes of 1030–2900 m.

Atheta brachati n. sp.

(Figs. 15–24)

Type material

Holotype ♂: “TR – Bolu (20), Str. Bolu → Kibrıcık, ca. 25 km s. Bolu, 1580 m, N40°37'59, E31°36'49, 30.IV.2012, leg. MEYBOHM & BRACHAT / Holotypus ♂ *Atheta brachati* sp. n. det. V. ASSING 2012” (cAss).

Paratype ♂: same data as holotype (cAss).

Etymology

This species is dedicated to VOLKER BRACHAT, specialist of Pselaphinae, who collected the type specimens.

Description

Body length 3.2–3.5 mm; length of forebody 1.6–1.7 mm. Habitus as in Fig. 15. Coloration: body black with the posterior portion of the elytral disc diffusely blackish-brown; legs reddish-brown with darker femora; antennae black.

Head (Figs. 16, 17) transverse, approximately 1.1 times as wide as long, narrowed posteriorly immediately behind eyes; punctuation fine and rather sparse, somewhat indistinct in the pronounced microreticulation. Eyes larger, nearly twice as long as postocular region in dorsal view. Antennae slender, approximately 1.1 mm long; antennomere III approximately twice as long as broad and slightly shorter than II; IV approximately as long as broad; V–IX weakly transverse and gradually, but weakly increasing in width; X less than 1.5 times as broad as long; XI slightly longer than the combined length of IX and X.

Pronotum (Figs. 16, 18) strongly transverse, approximately 1.3 times as broad as long and 1.25 times as broad as head, maximal width approximately in the middle; punctuation fine, dense, and weakly granulose; interstices with pronounced microreticulation and almost matt; pubescence pattern: type II (i. e., pubescence directed caudad along midline and diagonally transversely laterad in lateral portions).

Elytra (Figs. 16, 19) approximately as long as, and distinctly broader than pronotum; punctuation dense, moderately fine, and somewhat granulose; interstices with pronounced microreticulation and almost matt. Hind wings fully developed.

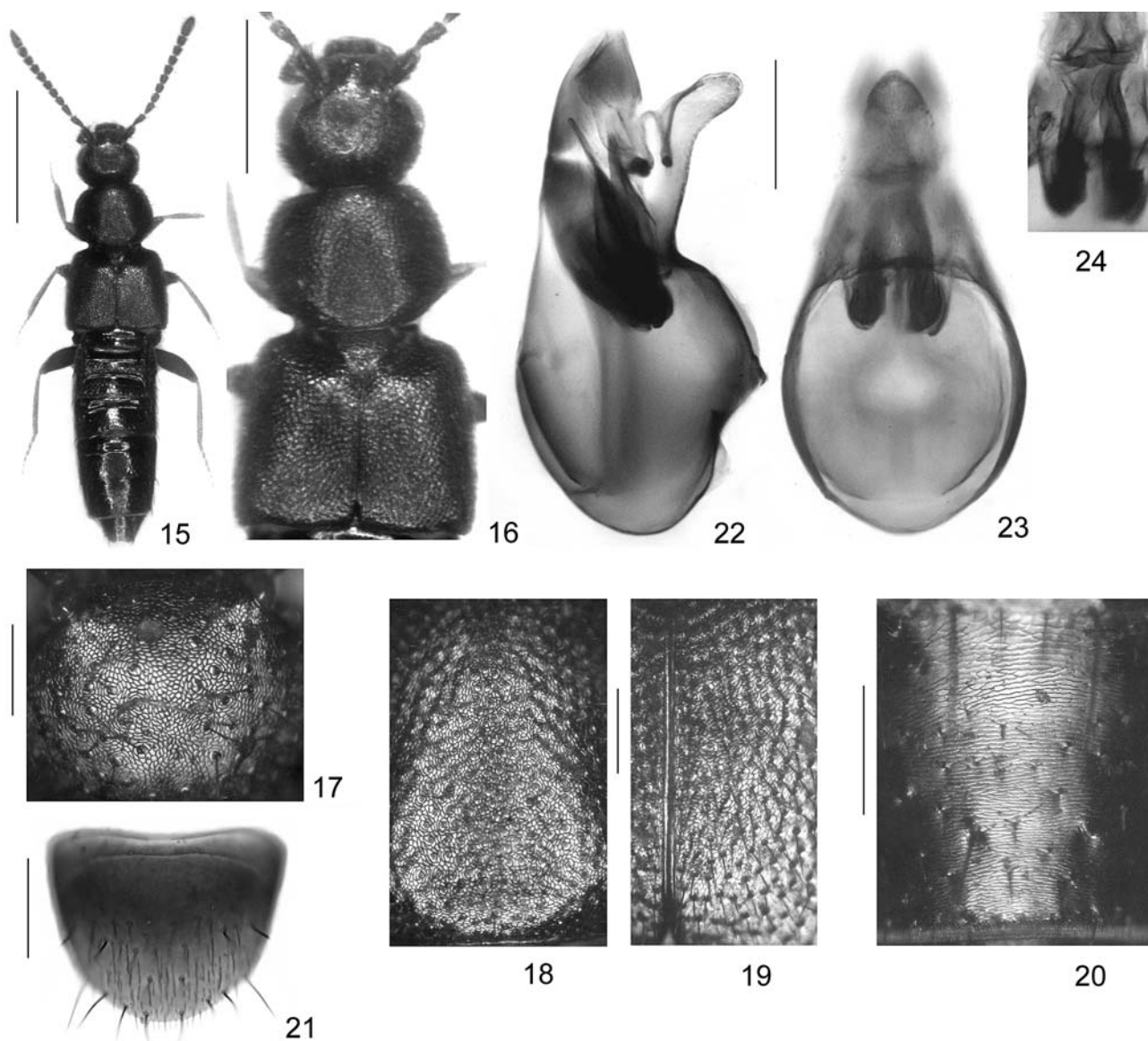
Abdomen distinctly narrower than elytra; punctuation fine and moderately dense on tergites III–V, very sparse and shallow on tergites VI–VIII; interstices with fine transverse microsculpture (Fig. 20); posterior margin of tergite VII with palisade fringe.

♂: posterior margin of tergite VIII weakly concave in the middle; sternite VIII transverse, convexly produced posteriorly (Fig. 21); median lobe of aedeagus approximately 0.35 mm long, with ventral process and internal structures of distinctive shape (Figs. 22–24).

♀: unknown.

Comparative notes

Based on the general morphology of the median lobe of the aedeagus, particularly the internal structures, *A. brachati* appears to be related to the species group allied to the widespread *A. vaga* (Heer, 1839) and *A. harwoodi* Williams, 1930. It is distinguished from these species by the more slender antennae, the different shape of the head, the absence of distinct impressions on the male head and



Figs. 15–24. *Atheta brachati* n. sp. – 15. Habitus. 16. Forebody. 17. Median dorsal portion of head. 18. Median portion of pronotum. 19. Sutural portion of elytra. 20. Median portion of male tergite VII. 21. Male sternite VIII. 22–23. Median lobe of aedeagus in lateral and in ventral view. 24. Internal structures of aedeagus in ventral view. – Scale bars: 1.0 mm (15), 0.5 mm (16), 0.2 mm (21), 0.1 mm (17–20, 22–24).

pronotum, the more pronounced and less dense punctation of the pronotum, the different pronotal pubescence pattern, the darker femora, and by the different male primary and secondary sexual characters.

Distribution and natural history

The type locality is situated to the south of Bolu, north-western Anatolia. The specimens were sifted in grassland near snow either from rotting wood or rotting grass (BRACHAT, pers. comm.) at an altitude of 1580 m.

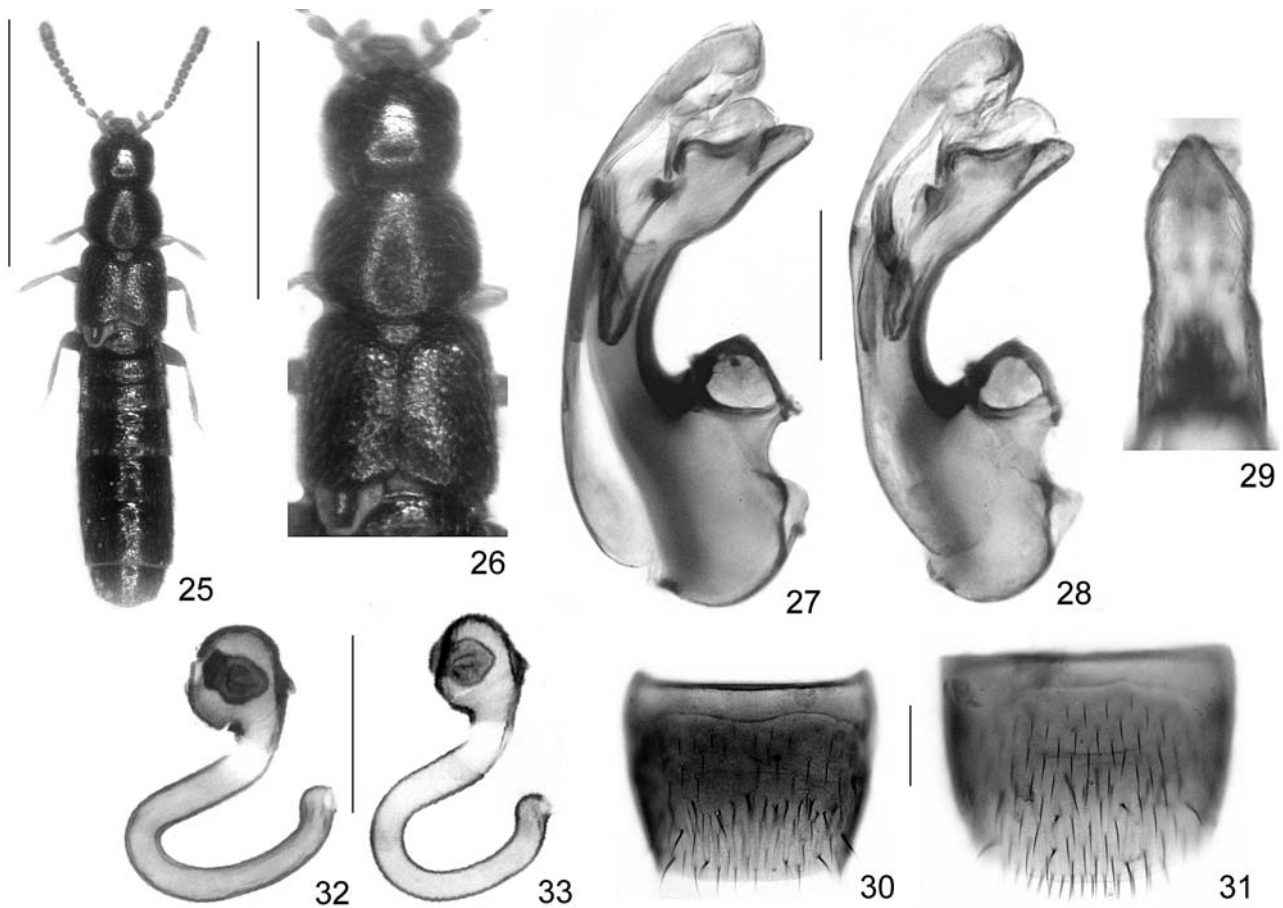
3.3.1.2 Oxypodini

Cousya microdotoides n. sp. (Figs. 25–33)

Type material

Holotype ♂: “N37°26'17 E034°56'09, TR Adana, 1700 m, 16.IV.2011, BRACHAT & MEYBOHM (7) / Holotypus ♂ *Cousya microdotoides* sp. n. det. V. ASSING 2012” (cAss).

Paratypes: 2 ♀♀: same data as holotype (cAss); 3 ♂♂, 1 ♀: “N37°26'08 E034°54'19, TR Adana Armutolugu, 1220 m, 16.IV.2011, BRACHAT & MEYBOHM (5)” (cAss).



Figs. 25–33. *Cousya microdotoides* n. sp. – 25. Habitus. 26. Forebody. 27–28. Median lobe of aedeagus in lateral view. 29. Ventral process of aedeagus in ventral view. 30. Female tergite VIII. 31. Female sternite VIII. 32–33. Spermatheca. – Scale bars: 1.0 mm (25), 0.5 mm (26), 0.1 mm (27–33).

Etymology

The specific epithet alludes to the external resemblance of this species with species of the subgenus *Microdota* Mulsant & Rey, 1873 (genus *Atheta* Thomson, 1858).

Description

Very small species; body length 1.6–2.2 mm; length of forebody 0.8–1.0 mm. Habitus as in Fig. 25. Coloration: forebody brown to blackish-brown, elytra sometimes slightly paler than head and pronotum; abdomen blackish with reddish-brown apex (posterior margin of segment VII; segments VIII–X); legs yellowish; antennae brown with slightly paler base (antennomeres I and II).

Head (Fig. 26) weakly oblong; punctuation shallow and moderately dense, barely noticeable in the pronounced microreticulation; dorsal surface with subdued shine. Eyes moderately convex and moderately large, slightly shorter than postocular region in dorsal view. Antennae short, approximately 0.5 mm long, and incrassate apically; anten-

nomere III much shorter than II, approximately as broad as long and of conical shape; IV–X strongly transverse, approximately twice as wide as long, and of gradually and distinctly increasing width; XI approximately as long as the combined length of IX and X.

Pronotum (Fig. 27) distinctly transverse, approximately 1.25 times as broad as long and 1.25 times as broad as head; posterior angles weakly marked; punctuation and microsculpture similar to those of head.

Elytra (Fig. 27) approximately 0.9 times as long as, and slightly broader than pronotum; punctuation fine, but more distinct than that of head and pronotum; interstices with pronounced microreticulation. Hind wings reduced to short stubs. Tarsi short; metatarsomere I approximately as long as the combined length of II and III.

Abdomen approximately as broad as elytra, widest at segments VI/VII; tergites III–V anteriorly with, tergite VI without transverse impression; punctuation very fine and relatively sparse, barely noticeable in the distinct micro-

sculpture; posterior margin of tergite VII with palisade fringe; posterior margin of tergite VIII broadly convex in both sexes.

♂: posterior margin of sternite VIII broadly convex, not produced in the middle; median lobe of aedeagus 0.32–0.33 mm long, with pronounced crista apicalis (Figs. 27–29).

♀: tergite VIII as in Fig. 30; sternite VIII broadly convex posteriorly, with stout and not particularly dense marginal setae (Fig. 31); spermatheca as in Figs. 32 and 33, of somewhat variable shape.

Comparative notes

In size and habitus, *C. microdotoides* somewhat resembles *C. deminuta* Assing, 2010 from Spain and *C. dimorpha* Assing, 2006 from Greece, but is distinguished from these species particularly by the morphology of the median lobe of the aedeagus (somewhat larger, with more pronounced crista apicalis and with ventral process of different shape). For illustrations of *C. deminuta* and *C. dimorpha* see ASSING (2006a, 2010b). All other described species represented in Turkey are separated from *C. microdotoides* by the different habitus (less transverse pronotum, longer elytra, etc.) and larger body size alone.

Distribution and natural history

The species was collected in two localities situated to the northeast of Pozanti in the northwest of Adana province, central southern Anatolia. The reduced hind wings and the absence of previous records from other localities suggest that the distribution of *C. microdotoides* is restricted. The specimens were sifted from leaf litter in mixed cedar and cypress forests (MEYBOHM, pers. comm.) at altitudes of 1220 and 1700 m.

Meotica wunderlei n. sp.

(Figs. 34–43)

Type material

Holotype ♂: “TR [26] – Isparta, 10 km SE Sütçüler, 1520 m, 37°24'55"N, 31°02'21"E, meadow, under stones, 26.IV.2011, leg. WUNDERLE / Holotypus ♂ *Meotica wunderlei* sp. n. det. V. ASSING 2012” (cAss).

Paratypes: 9 exs.: same data as holotype (cAss, cWun).

Etymology

The species is dedicated to my friend and field-trip companion PAUL WUNDERLE, who collected all the type specimens.

Description

Body length 2.0–2.3 mm; length of forebody 0.9–1.0 mm. Habitus as in Fig. 34. Coloration: head and pronotum reddish-brown; elytra yellowish-brown; abdomen brown to dark-brown with the apex (posterior portion of

segment VII, segments VIII–X) reddish; legs yellowish; antennae brown with yellowish antennomeres I–III.

Head (Fig. 35) 1.05–1.10 times as broad as long, somewhat wedge-shaped; punctation extremely fine, barely noticeable in the pronounced microreticulation; surface with weak, subdued shine. Eyes small, not projecting from lateral contours of head, little larger than antennomere I in cross-section, and composed of 10–15 ommatidia. Antenna distinctly incrassate apically, approximately 0.5 mm long; antennomere III much shorter than II, approximately as broad as long; IV–X increasingly transverse and of increasing width; X more than twice as broad as long; XI approximately as long as the combined length of IX and X.

Pronotum (Fig. 35) distinctly transverse, approximately 1.25 times as broad as long and 1.1 times as broad as head; posterior angles obtuse and moderately marked; punctation and microsculpture similar to those of head.

Elytra (Fig. 35) approximately as long as pronotum; punctation fine; microsculpture distinct, but less pronounced than that of head and pronotum; surface with subdued, but noticeable shine. Hind wings of reduced length (submacropterous).

Abdomen with pronounced microreticulation and with extremely fine punctation visible only at high magnification; tergite VII with palisade fringe.

♂: tergite VIII moderately transverse with weakly concave posterior margin (Fig. 36); sternite VIII strongly transverse, posterior margin rather broadly convex in the middle (Fig. 37); median lobe of aedeagus approximately 0.28 mm long, shape and internal structures distinctive (Figs. 38, 39); apical lobe of paramere black.

♀: tergite VIII approximately as long as broad and with weakly concave posterior margin (Fig. 40); sternite VIII approximately as long as broad, posterior margin broadly produced in the middle (Fig. 41); spermatheca shaped as in Fig. 42.

Comparative notes

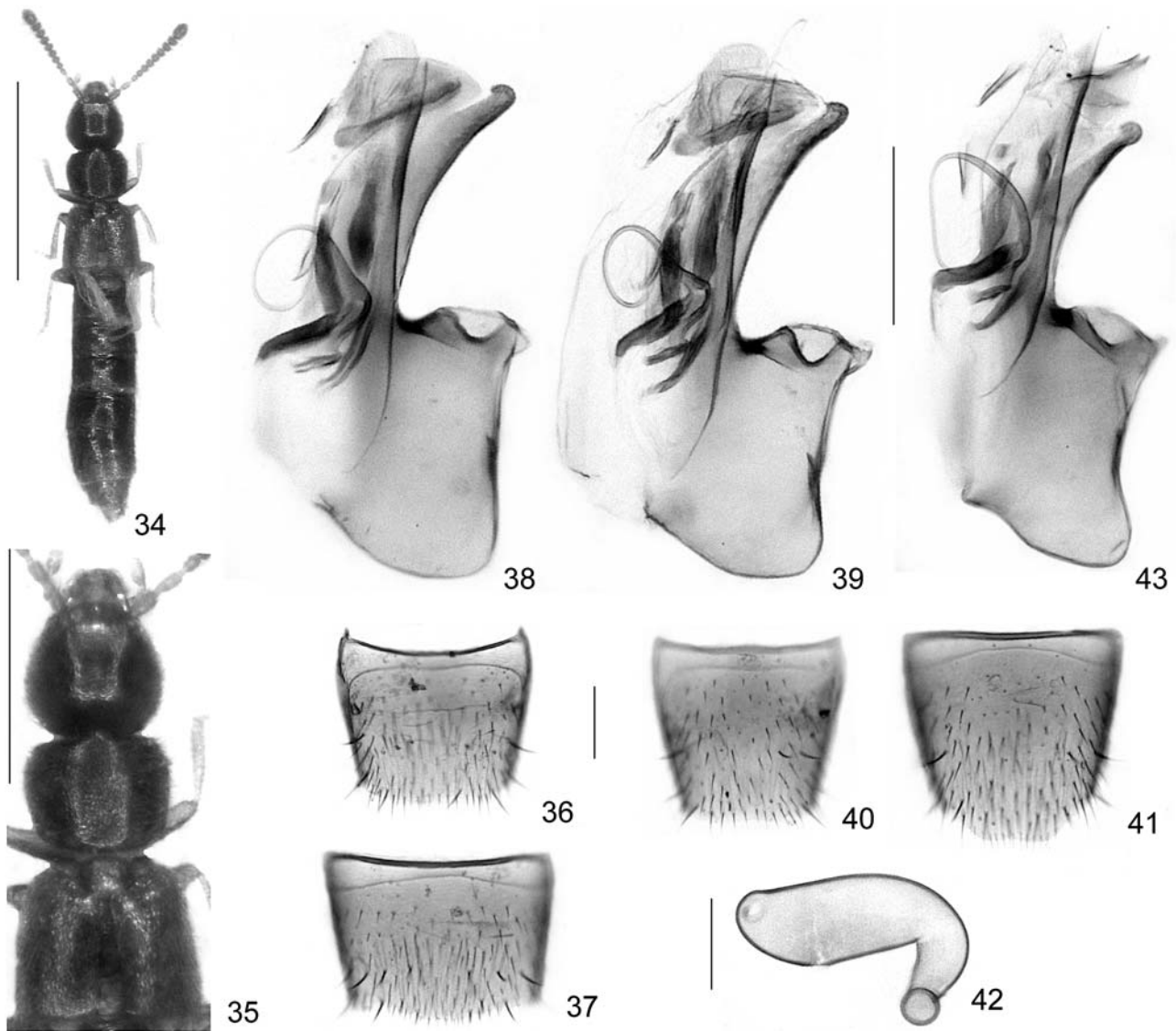
So far, five species of *Meotica* have been recorded from Turkey: *M. decolor* Assing, 2004, *M. hamata* Assing, 2011, *M. marchica* Benick, 1953, *M. subnigra* Assing, 2006, and *M. truncata* Assing, 2004. *Meotica wunderlei* is distinguished from all of them by the shape of the median lobe and of the internal structures of the aedeagus. In addition, it is separated from them as follows:

from *M. decolor* by less pronounced microsculpture on the whole body and by the black apical lobe of the paramere;

from *M. hamata* by paler coloration;

from *M. marchica* by the shape of the posterior margin of the male sternite VIII;

from *M. subnigra* by smaller eyes, paler coloration, the posteriorly more strongly tapering pronotum, and by the shorter elytra;



Figs. 34–43. *Meotica wunderlei* n. sp. (34–42) and *M. marchica* from Kocaeli (43). – 34. Habitus. 35. Forebody. 36. Male tergite VIII. 37. Male sternite VIII. 38–39, 43. Median lobe of aedeagus in lateral view. 40. Female tergite VIII. 41. Female sternite VIII. 42. Spermatheca. – Scale bars: 1.0 mm (34), 0.5 mm (35), 0.1 mm (36–41, 43), 0.05 mm (42).

from *M. truncata* by smaller eyes and the posteriorly convex posterior margin of the male sternite VIII (*M. truncata*: truncate or even weakly concave).

For illustrations of the species described from Turkey see ASSING (2004b, 2006b, 2011a); the aedeagus of *M. marchica* is illustrated in Fig. 43.

Distribution and natural history

The type locality is situated to the southeast of Sütçüler in Isparta province, southwestern Anatolia. The specimens were found under large, deeply embedded stones on a moist meadow on calcareous soil at an altitude of 1520 m.

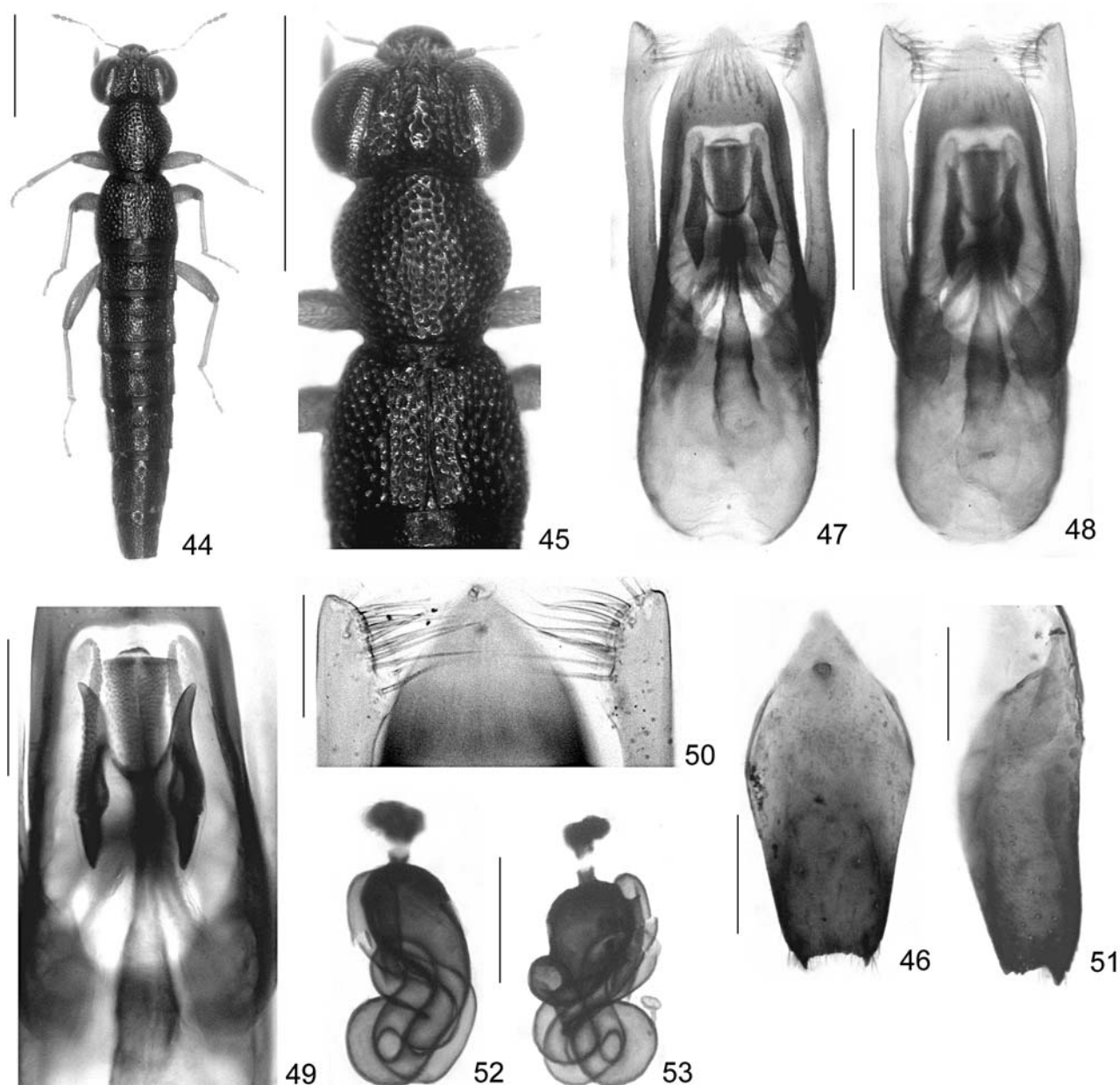
3.3.2 Steninae

Stenus (Hemistenus) bithynicus n. sp. (Figs. 44–54)

Type material

Holotype ♂: “N40°38'54 E30°05'32, TR Kocaeli 5.V.2012, Kartepe [= Goeck Dağı] 1270 m, BRACHAT & MEYBOHM (35) / Holotypus ♂ *Stenus bithynicus* sp. n. det. V. ASSING 2012” (cAss).

Paratypes: 2 ♀♀: same data as holotype (cAss); 2 ♀♀: “TR – Kocaeli (35), Kartepe [sic] Skizentrum 1270 m, N40°38'54, E30°5'32, 5.V.2012, leg. MEYBOHM & BRACHAT” (cAss); 1 ♀:



Figs. 44–53. *Stenus bithynicus* n. sp. – 44. Habitus. 45. Forebody. 46. Male sternite IX. 47–48. Aedeagus in ventral view. 49. Internal structures of aedeagus in ventral view. 50. Apices of parameres. 51. Female hemisternite IX. 52–53. Spermatheca. – Scale bars: 1.0 mm (44–45), 0.2 mm (46–48, 52–53), 0.1 mm (49–51).

“N40°38'47 E30°03'25, TR Kocaeli 25.IV.2012, Pazarcayiri 880 m, BRACHAT & MEYBOHM (6)” (cAss); 1 ♀: “TR – Kocaeli (1), SO Suadiye, Kaltepe [sic] 1030 m, N40°39'4, E30°3'42, 24.IV.2012, leg. MEYBOHM & BRACHAT” (cAss); 1 ♂: “N40°41'21 E31°08'18, TR Düzce 1.V.2012, Düzce 23 km S 1020 m, BRACHAT & MEYBOHM (25)” (cAss); 1 ♀: “TR – Düzce (25), 23 km s. Düzce, 1020 m, N40°41'21, E31°08'18, 1.V.2012, leg. MEYBOHM & BRACHAT” (cAss); 1 ♂: “N40°40'21 E30°54'22, TR Sakarya 4.V.2012, Dikmen 4 km S 1275 m, BRACHAT & MEYBOHM (32)” (cAss).

Etymology

The specific epithet is an adjective derived from Bithynia, the ancient name of the region where the localities are situated.

Description

Body length 4.0–5.0 mm; length of forebody 1.8–2.1 mm. Habitus as in Fig. 44. Coloration: body blackish, forebody sometimes with weak bronze hue; legs dark-yellowish, with the femoral apices usually weakly infusate;

antennae yellowish, with the 3–5 apical antennomeres somewhat infuscate.

Head (Fig. 45) dorsally with distinct, densely and coarsely punctured lateral furrows; median keel pronounced, less coarsely and less densely punctate, and with more shine.

Pronotum (Fig. 45) approximately 1.05 times as broad as long and approximately 0.85 times as broad as head across eyes; lateral margins strongly convex in dorsal view; maximal width slightly before middle; punctuation coarse and dense; interstices much narrower than diameter of punctures, with or without shallow microreticulation.

Elytra (Fig. 45) short, approximately 0.85 times as long as pronotum, posteriorly dilated; humeral angles almost obsolete; punctuation similar to that of pronotum, or coarser and denser; interstices usually with microreticulation. Hind wings completely reduced.

Abdomen with pronounced paratergites, those of segments III–V approximately as broad as maximal width of mesotibia; punctuation moderately dense and rather fine; interstices with very shallow to distinct microreticulation; posterior margin of tergite VII with or without indistinct rudiments of a palisade fringe.

♂: metatibia weakly curved in apical third; meso- and metatibia with fine subapical spine; sternite VIII with relatively deep V-shaped posterior excision; sternite IX as in Fig. 46; median lobe of aedeagus 0.63–0.66 mm long and shaped as in Figs. 47–49; parameres stout, apically dilated,

furnished with numerous long setae (Fig. 50), and extending slightly beyond apex of median lobe.

♀: meso- and metatibia unmodified; hemitergites IX shaped as in Fig. 51; spermatheca as in Figs. 52 and 53.

Comparative notes

Among the locally endemic and micropterous *Hemistenus* species known from Turkey, *S. bithynicus* is most similar to *S. cilicianus* Assing, 2003 from the Taurus range (region to the north of Alanya). It is distinguished from this species by the on average larger and broader body, the presence of microreticulation on the pronotum, the elytra, and the anterior abdominal tergites (*S. cilicianus*: very shallow microsculpture present only on tergites VII and VIII), the presence of subapical spines on the male meso- and metatibiae, the much deeper and larger posterior excision of the male sternite VIII, the slightly different shape of the aedeagus (parameres apically less strongly dilated), and by the much larger spermatheca. For illustrations of *S. cilicianus* see ASSING (2003a).

Distribution and natural history

The species represents the northwesternmost Turkish local endemic of the genus. It is known from several localities in Kocaeli, Sakarya, and Düzce provinces in northwestern Anatolia (Fig. 54). Its discovery came as quite a

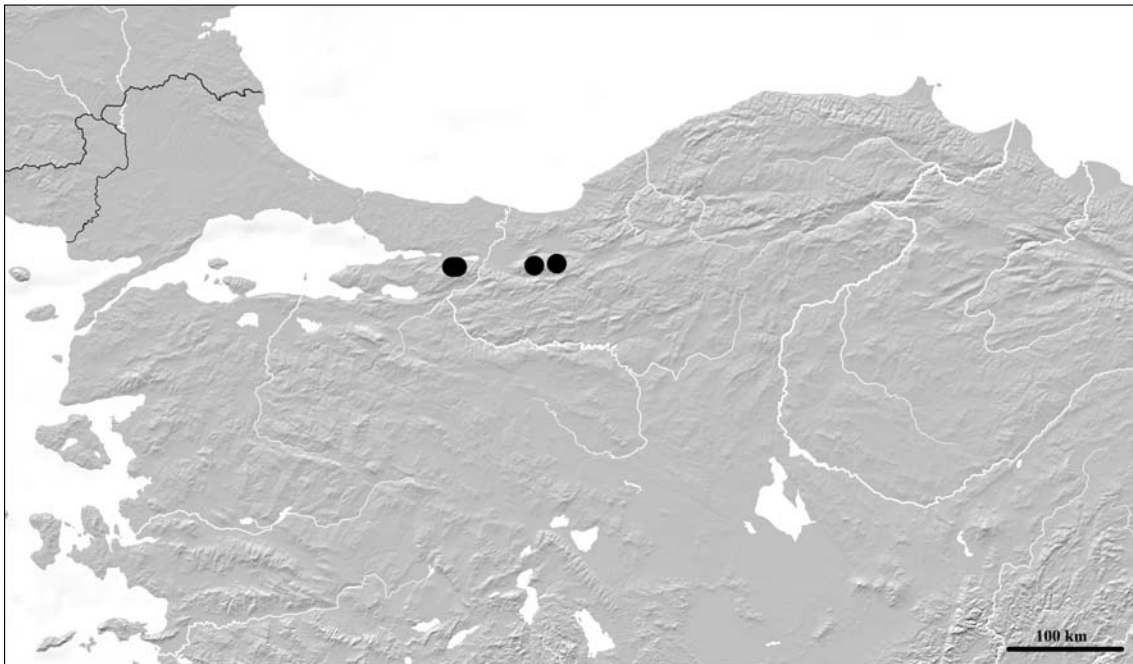


Fig. 54. Distribution of *Stenus bithynicus* n. sp. in northwestern Turkey.

surprise, since the geographically closest north Anatolian, micropterous, and locally endemic *Stenus* species are distributed in Ordu, since endemic Staphylinidae species are generally more widespread in northwestern Anatolia than elsewhere in Turkey, and finally since the region has repeatedly been visited by coleopterists before. The specimens were sifted from leaf litter in beech forests, partly with rhododendron undergrowth, in a hornbeam forest with rhododendron undergrowth, and in a mixed fir and beech forest at altitudes of 880–1270 m (MEYBOHM, pers. comm.).

4 Remarks on a recent catalogue of the Staphylinidae of Asia Minor and Cyprus

Catalogues can be most useful tools not only for taxonomists, but also for zoogeographic and ecological studies, since they compile information often scattered in the literature. Up until about a decade ago, the Turkish staphylinid fauna had been very poorly studied and addressed primarily in the context of species descriptions, of taxonomic revisions of certain taxa, or of identification keys and faunistic synopses not specifically dealing with the fauna of Turkey. In recent years, however, numerous articles addressing the Staphylinidae of Turkey have been published (see section 5), including descriptions of several hundred new species and even more new country records. Against this background, a comprehensive, critically reviewed, and up-to-date catalogue compiled by a taxonomist with a profound knowledge both of Turkey and the Staphylinidae could provide a state-of-the-art overview. A first attempt was made by ANLAŞ (2009), who published a checklist of all the Staphylinidae described and recorded from Turkish territory, indicated the respective provinces, and provided a nearly complete bibliography.

More recently, BORDONI (2010) published a catalogue of the Staphylinidae of Asia Minor and Cyprus “in the hope of contributing to better knowledge of Staphylinidae of one of the most interesting zoogeographical area of the Mediterranean” [sic]. The author has published numerous taxonomic articles, particularly on Xantholinini, as well as on some groups of West Palaearctic Paederinae, Staphylininae, and Omaliinae. It can be inferred from the material list, however, that he has never personally collected Staphylinidae in Turkey. The catalogue comprises more than 300 pages with a two-page introduction, a list of “acronyms” [sic], a list of taxa, an acknowledgements section, a summary, and a bibliography.

A reader hoping for a state-of-the-art synopsis of the Turkish staphylinid fauna, however, will be rather disappointed. A closer look raises considerable doubts regarding the scientific value of this paper. True, it is almost impossible to write a comprehensive work without mistakes and errors, but the shortcomings of the catalogue in ques-

tion are so numerous, so various, and so significant that it seems somewhat surprising that it ever got past an editor. Needless to say that it was evidently published without serious peer-reviewing.

The Scaphidiinae, Pselaphinae, Scydmaeninae, and Dasycerinae, formerly separate families, but now subfamilies of Staphylinidae are omitted. Nowhere in the catalogue does BORDONI mention that, or why, these subfamilies were not included. The data for the individual species are more or less copied from the primary sources. They are neither discussed nor evaluated, and hardly any new data are given. Instead of providing a complete bibliography, the reader is referred to the standard catalogues by HERMAN (2001) and SMETANA (2004) for part of the literature references.

Although the said catalogue was published as late as 2010, only data and references published before 2008 are considered, which rendered it hopelessly incomplete at the time of publication. Not only had at least 34 species been described and at least 40 additional species newly recorded from Turkey in 2008 and 2009 alone. Also, numerous new generic and subgeneric assignments, as well as new synonymies affecting the Turkish fauna had been established, and misidentifications had been rectified during this period. Moreover, it is particularly surprising that there is no reference whatsoever to the checklist of Turkish Staphylinidae published by ANLAŞ (2009), which BORDONI knew about and which, though not faultless, is significantly more correct, more comprehensive and complete (also regarding the bibliography), more detailed, more up-to-date, and more informative. Unlike BORDONI, ANLAŞ had sent parts of his checklist to various specialists for correction, critical review, and completion.

In the acknowledgements section, the author “wishes to thank for their kind cooperation [numerous] colleagues [sic]”, including myself. I am not aware, however, of having contributed anything to this book other than the publications listed in the bibliography section. The same is true of at least part of the other colleagues mentioned in this section. None of those that I have asked remembers any contribution.

The catalogue is subject to numerous formal shortcomings. It contains many hundreds, if not thousands, of linguistic errors (grammar, spelling, etc.), particularly in the introduction, but also in other parts of the catalogue. Turkish special characters are not given at all “for practical [sic] reasons” (what may these reasons be in an age where every text processor provides such characters?). Writing “Ardesen” rather than “Ardeşen” (example given in the introduction of the catalogue) would be equivalent to omitting accent marks in French or leaving out the “h” in macchia. Moreover, the omission of diacritic marks and the substitution of special characters with standard letters is not confined to Turkish localities, but also affects names of authors, as well as colleagues mentioned in the acknowl-

edgement section, e. g., “HLAVAC” rather than HLAVÁČ. The map of “Turkey” (p. 36) only depicts the Asian part of Turkey; the European part is omitted. The species are listed by subfamilies. However, the arrangement of genera within the subfamilies is partly somewhat confusing. For instance, the genus *Plataraea* is separated from other Athetini and appears only at the very end of the catalogue, right after the Aleocharini.

The distribution sections contain very little zoogeographic information. Some species recorded also from outside Turkey are categorised simply as “widespread”. Species recorded only from Turkey (or Cyprus) are given as “endemic” (endemic to what region?) without further specification. Turkey is zoogeographically highly diverse and there is not a single species that is distributed in all Turkish regions, but absent from any of the adjacent countries. Often, the general distribution sections are remarkably incomplete (see, e. g., *Medon rufiventris*; p. 124) or incorrect. In most cases, the zoogeographic data provided in the standard catalogues by HERMAN (2001) and SMETANA (2004) are significantly more informative. Turkish localities are often not specified, apparently because the primary records had not been found in the literature. They are neither standardised nor arranged systematically (e. g., by province), and they are subject to innumerable misspellings or other errors. In the case of *Atreucus parvioculatus* (p. 162), for instance, the collector is given as the locality.

Several species are erroneously listed for Turkey, based on evident misidentification, misinterpretation, or because relevant literature was either neglected or overlooked: e. g., *Diachus libanoticus*, *Mycetoporus niger*, *M. nigricollis*, *M. solidicornis*, *Tachinus flavolimbatus*, *Myrmecopora pygmaea*, and *Amarochara forticornis*. In addition, numerous doubtful records (most likely based on misidentifications) are reported; only some of them are classified as “to be confirmed”.

The catalogue is also subject to numerous taxonomic and nomenclatural errors, for instance, misspelled scientific names (e. g., *Omalium saulcy*, *Diachus atayus*, *Typhodes Microsauarus*, *Atheta chefssurica*, *A. cinammoptera*, *A. pittioni*), missing parentheses (see, e. g., the author of *Pella erratica*), omitted generic names (e. g., [*Myrmecopora*] *wunderlei*, [*Derocala*] *brachati*). Also, invalid genus-group names are used (e. g., *Ditroposipalia*, *Glossola*).

In several cases, one and the same species is erroneously listed twice (as valid) under different names, occasionally even in different genera or tribes, e. g. (valid name or combination given first) *Tachinus rufipes* and *T. signatus*; *Liogluta alpestris* and *L. nitidula*; *Geostiba oertzeni* (Athetini) and *Leptusa solitaria* (synonym of *G. oertzeni*; *Leptusa* belongs to Homalotini!); *Aloconota* (s. str.) *gregaria* and *Atheta* (*Glossola*, sic!) *gregaria*; *Nehemitropia lividipennis* and *Atheta sordida*; *Atheta clientula* and *Acrotone clientula*. Furthermore, some generic (e. g., *Lep-*

tusa solitaria, *Falagrioma pamphylica*, *Aleochara alata*, *A. cingulata*) and subgeneric assignments (e. g., *Geostiba attaleensis*, *G. bigibbera*, *G. kastamonuensis*, *G. occaecata*, *Atheta clientula*, *A. mucronata*, *Oxypoda cristata*, *O. flavocaudata*, *O. speculoclara*) are incorrect. Synonymies are given inconsistently, sometimes with, often without the original reference to the author of the respective synonymy.

One of the foremost objectives and principles of the Code (ICZN 1999) is the stability of nomenclature. In his catalogue, BORDONI proposes as many as thirteen new synonymies without presenting any arguments whatsoever. Instead he states that they “will be dealt with elsewhere” (p. 36). Remarkably, most of these synonymies are proposed in genera that the author has never published a single taxonomic article about and evidently has very little personal experience with (*Leptusa* Kraatz, 1856 and *Geostiba* Thomson, 1858). None of these synonymies is based on the study of type (or other) material of the synonymised names. What is more, in most cases the author probably has not even seen material of what he proposes as the senior name. If these synonymies are not based on research and study, what then are they based on? In any case, minimum scientific requirements and standards are not met, so that the synonymies would be unacceptable for these reasons alone.

Furthermore, the majority of the synonymies are rather bizarre, since the respective species pairs are not even particularly similar and even entomologists without much expertise in staphylinid taxonomy would realise at once that they represent distinct species if only they compared actual specimens or carefully looked at the descriptions and illustrations of sexual characters. Without exception, the below synonymies proposed by BORDONI are incorrect. The respective synonyms were all treated in the context of comprehensive taxonomic revisions (ASSING 2003c, 2004c, 2007a, 2007c, 2007d, 2009f). All of them represent distinct and valid species, so that they are here formally revalidated:

- Medon lanugo* Assing, 2004 [not synonym of *M. lydicus* Bordoni, 1980];
- Medon reliquus* Assing, 2007 [not synonym of *M. maronitus* (Saulcy, 1865)];
- Medon subquadratus* Assing, 2004 [not synonym of *M. seleucus* Bordoni, 1980];
- Xantholinus penicillatus* Assing, 2007 [not synonym of *X. puthzi* Bordoni, 1979];
- Leptusa longilobata* Assing, 2007 [not synonym of *L. fibula* Assing, 2003];
- Leptusa artviniensis* Pace, 1982 [not synonym of *L. ionopolitana* Pace, 1982];
- Leptusa amisensis* Pace, 1982 [not synonym of *L. ionopolitana* Pace, 1982];
- Leptusa diecki* Pace, 1983 [not synonym of *L. paphlagonica* Pace, 1982];

Leptusa spoliata Assing, 2002 [not synonym of *L. paphlagonica* Pace, 1982];
Geostiba lycaonica Pace, 2002 [not synonym of *G. uhligi* Pace, 1983].

Likewise, the three synonymies listed below are erroneous:

Leptusa confinis Pace, 1982 [of *L. paphlagonica* Pace, 1982];
Leptusa othmaniorum Pace, 1983 [of *L. paphlagonica* Pace, 1982];
Leptusa gurgentepensis Pace, 1989 [of *L. paphlagonica* Pace, 1982].

According to ASSING (2009d), the first revising author, the correct synonymies are as follows: *Leptusa confinis* Pace, 1982 = *L. paphlagonica* Pace, 1982 (syn.), = *L. othmaniorum* Pace, 1983 (syn.); *Leptusa diecki* Pace, 1983 = *L. gurgentepensis* Pace, 1989 (syn.).

According to BORDONI, “it believe that it can serve as a basis for further study of the Staphylinidae of Cyprus and Asia Minor, areas where in recent years has focused the attention of some scholars that researche continues [sic]”. In view of the significant shortcomings outlined above, however, it appears rather unlikely that this substandard catalogue will indeed prove to be a useful tool. If a synopsis of Turkish Staphylinidae is needed, the checklist by ANLAŞ (2009) will undoubtedly be the better choice.

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