An unusual species of *Wardomyces* (Hyphomycetes)*

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Wardomyces moseri, isolated from a dead petiole of *Mauritia minor* in Colombia, is described as a new species. It differs from other species of the genus by easily liberated conidia. Its classification is discussed.

Keywords: Hyphomycetes, taxonomy, *Wardomyces*, tropics.

Among isolates derived from plant material collected in Colombia in 1980 (see Veerkamp & Gams, 1983), a hyphomycete was preserved for several years in the CBS collection as *Wardomycopsis* sp. It was found in a Petri dish during the isolation of microfungi from plant litter collected by the author in a moist tropical biotope and it differed considerably from the type species of this genus, *W. inopinata* Udagawa & Furuya (1978), by the lack of annellides and noncatenate, though easily seceding slimy conidia. It strongly resembled *Ticogloea* G. Weber & al. (1994) but, upon closer examination, was found to differ in the mode of conidial germination. This fungus is therefore placed in *Wardomyces* though it differs from all known species by easily liberated conidia.

I dedicate this species to my esteemed teacher and former supervisor Meinhard Moser, Innsbruck, on the occasion of his 70th birthday.

The fungus was grown on 2% malt extract agar (MEA), oatmeal agar (OA), cornmeal agar (CMA), and potato-carrot agar (PCA). Cultures grown for 6 and 14 days on PCA were prepared for SEM analysis according to Samson & al. (1979).

Wardomyces moseri W Gams, sp. nov. - Figs 1, 2.

Coloniae 30-35 mm diam. post 10 dies 25 C, optime 21-27 C crescentes. Massa conidialis radiatim aggregata, partim superficialis, partim submersa. Cellulae conidiogenae ad hyphas radiantes dense aggregatae, sessiles seu ramulis singulis supportatae, $3.5-5.5 \mu$ m longae, e ventro 2-2.5(-3) μ m lato et uno vel compluribus

 $[\]ast$ This paper is dedicated to Professor M. Moser on the occasion of his seventieth birthday.

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Fig. 1. – Wardomyces moseri. – a. sporodochium-like fructifications. – b. conidia. – c. germinating conidia.

collulis 0.6–1.0 µm latis constantes. Conidia in massa mucida aggregata, ovoidea, ad hilum basilare truncata, levia, cito fuscescentia, nonnulla fissuram germinationis praebentia, 10–14 × 3–4.5 µm.

Holotypus colonia exsiccata, isolata e rachide *Mauritiae minoris* prope Villavicencio in Columbia, in COL, viva CBS 164.80.

Colonies rather fast-growing, reaching 30-35 mm diam in 10 days, daily radial increment appr. 1.6 mm at 25 C on MEA,



Fig. 2. – Wardomyces moseri. – a, b. Direct view of sporulation in undisturbed Petri dish culture, light micrograph at low power. – c, d. Sporulation in an 6-day-old PCA culture; polyblastic conidiogenous cells visible in d; SEM.

temperature optimum 21–27 C, minimum about 8 C, maximum about 35 C. Sporulation optimal on PCA, good on OA or CMA after about 5 days and poor on MEA, forming blackish, radially elongated, slimy conidial masses, partly superficial, partly submerged in the agar. – Conidiogenous apparatus often densely aggregated along radial hyphae, more or less hyaline, conidiogenous cells arising either directly from creeping vegetative hyphae or supported by short swollen stalk cells, $2-3 \times 1.5-2.5 \ \mu\text{m}$. – Conidiogenesis polyblastic, conidiogenous cells flask-shaped, about $3.5-5.5 \ \mu\text{m} \log$ (three) narrow conidiogenous necks, $0.6-1.0 \ \mu\text{m}$ wide. – Conidia forming irregular slimy masses, ovoid (flattened at the point of attachment), smooth-walled, soon becoming rather dark brown, some showing an inconspicuous longitudinal germ slit, $10-14 \times 3-4.5 \ \mu\text{m}$.

Material examined COLOMBIA: Dep. Meta, 35 km East of Villavicencio, on a river bank in the savannah area, isolated from dead petiole of the palm *Mauritia minor*, CBS 164.80.

Sporulation of this fungus is sporodochium-like, although the conidiogenous and stalk cells are not very firmly aggregated in several orders. Only more or less dense clusters of conidiophores arise laterally along trailing hyphae, either submerged in the agar or on its surface.

Conidiogenesis is polyblastic. Sympodial proliferation of the conidiogenous cells is observed quite frequently. Although the conidia are easily liberated and aggregate in slimy masses, no sign of multiple conidium formation through one opening could be observed in slide cultures grown on PCA in a device similar to that described by Cole & Kendrick (1968). No periclinal wall thickening characteristic of phialidic conidiogenesis is visible in light microscopy, although the openings appear rather wide in SEM (Fig. 2d). No trace of an annellation could be observed in SEM either. The conidiogenous cells are very irregular in SEM, apparently due to slime deposition.

Wardomycopsis Udagawa & Furuya is a genus analogous to Wardomyces Brooks & Hansford (incl. Gamsia Morelet) which has solitary pigmented blastoconidia provided with a germ slit and sometimes forming additional hyaline annelloconidia, which otherwise would fit Scopulariopsis Bain. Wardomycopsis, however, differs from the present fungus by the formation of pronounced annellides and catenate, dry, dark conidia with conspicuous germ slits.

The case of *Wardomycopsis* is somehow comparable to that of the *Chalara* anamorph of *Ceratocystis paradoxa* (Dade) C. Moreau, where

the pigmented phialoconidia also show a germ slit, while in some other species formerly classified in *Chalaropsis* Peyronel, but not in all, the germ slit occurs only on solitary blastoconidia (Nag Raj & Kendrick, 1976). Molecular studies by Blackwell & Spatafora (1992) and Spatafora & Blackwell (1994) have shown that the teleomorph genera *Ceratocystis* and *Microascus* are more closely related to each other than to other ascomycete genera with beaked perithecia such as *Ophiostoma*. Another genus having conidia with a germ slit is *Conioscypha* Höhnel, but this has a unique kind of percurrent conidiogenesis (Shearer, 1973; Shearer & Motta, 1973).

Ticogloea guttulata, isolated from roots of Ticodendron incognitum in Costa Rica and roots of Tilia platyphyllos in Hamburg, Germany, living cultures CBS 604.92 and 689.92, has a comparable sporodochium-like sporulation, but it differs from W. moseri by more sepia-brown colonies, and smaller conidia which are borne in greater numbers from a conidiogenous cell and are more broadly truncate at the base (Weber & al., 1994). After finding germ slits in W. moseri, the conidia of T. guttulata were reexamined and found to lack them altogether. Even upon germination, no trace of a shell-like splitting of the conidia was observed while this is quite evident in W. moseri (Fig. 1c). Therefore the new species cannot be placed in Ticogloea.

Although W. moseri differs from other species of Wardomyces (cf. Dickinson, 1964; Hennebert, 1968) by its narrowly attached and easily seceding conidia which tend to form slimy masses, I prefer to classify the new species in this genus rather than to describe another new genus, because of the aggregated production of one-celled pigmented conidia which are produced on polyblastic conidiogenous cells and are provided with a germ slit. For the Wardomyces anamorph of Microascus giganteus Malloch (1970), Wardomycopsis inopinata Udagawa & Furuya, and the Wardomycopsis anamorph of Microascus singularis (Sacc.) Malloch & Cain, the connections with Microascus teleomorphs have been proven. The features observed in W. moseri also suggest an affinity with the Microascaee, which cannot be ascertained for Ticogloea.

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References

Blackwell, M. & J. W. Spatafora (1992). Convergent evolution among arthropoddispersed perithecial ascomycetes (abstract). – Inoculum, Mycol. Soc. Amer. Newsletter 43: 26. Cole, G. J. & W B. Kendrick (1968). A thin culture chamber for time-lapse photomicrography of fungi at high magnifications. – Mycologia 60: 340–344.

Dickinson, C. H. (1964). The genus Wardomyces. Trans. Br. mycol. Soc. 47: 321-325.

- Hennebert, G. L. (1968). Echinobotryum, Wardomyces and Mammaria. Trans. Br. mycol. Soc. 51: 749-762.
- Malloch, D. (1970). New concepts in the Microascaceae illustrated by two new species. Mycologia 62: 727–740.
- Nag Raj, T. R. & W B. Kendrick (1976). A monograph of *Chalara* and allied genera. – Wilfrid Laurier University Press, Waterloo '1975'
- Samson, R. A., J. A. Stalpers, & W Verkerke (1979). A simplified technique to prepare fungal specimens for scanning electron microscopy. – Cytobios 24: 7-11.
- Shearer, C. A. (1973). Fungi of the Chesapeake Bay and its tributaries. II. The genus *Conioscypha.* – Mycologia 65: 128–136.

 & J. J. Motta (1973). Ultrastructure and conidiogenesis in Conioscypha (Hyphomycetes). - Can. J. Bot. 51: 1747-1751.

- Spatafora, J. W & M. Blackwell (1994). The polyphyletic origins of ophiostomatoid fungi.- Mycol. Res. 98: 1-9.
- Udagawa, S. & K. Furuya (1978). A new species of *Microascus* and its peculiar conidial state. – Mycotaxon 7: 91–96.
- Veerkamp, J. & W Gams (1983). Los hongos de Colombia VIII. Some new species of soil fungi from Colombia. – Caldesia 13: 709–717.
- Weber, G., F. Spaaij & W Gams (1994). Ticogloea, a new genus of Hyphomycetes from roots of Ticodendron incognitum from Costa Rica. Mycol. Res., 98: 660-664.

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