Pseudocercospora aspidospermatis - a new combination for Bactrodesmiella aspidospermatis

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Dianese, J. C. & M. P. S. Câmara (1994). Pseudocercospora aspidospermatis – a new combination for Bactrodesmiella aspidospermatis. – Sydowia 46(2): 225–232.

The holotype of *Bactrodesmiella aspidospermatis* on leaves of *Aspidosperma macrocarpon* and three other collections were studied. The species is shown to be incorrectly classified and is better accommodated in *Pseudocercospora* as *P. aspidospermatis* comb. nov.

Keywords: Deuteromycetes, taxonomy, new combination.

Aspidosperma macrocarpon Mart. (Apocynaceae) is an important component of the native flora of the cerrado around Brasília. Leaves recently collected were shown to be infected with a Pseudocercospora species which was previously classified by Batista & al. (1961) as Bactrodesmiella aspidospermatis Batista, Peres & Garnier. These collections and the holotype material of B. aspidospermatis were studied and the name of the species transferred to Pseudocercospora aspidospermatis (Batista, Peres & Garnier) Dianese & Câmara comb. nov. based on the concepts adopted by Braun (1992), Ellis (1959, 1971, 1976), Sutton & Pascoe (1989), and Crous & Braun (1994). A detailed description follows.

Pseudocercospora aspidospermatis (Batista, Peres & Garnier) J. C. Dianese et M. P. S. Câmara, comb. nov. – Figs. 1–13.

= Bactrodesmiella aspidospermatis Batista, Peres & Garnier In: Batista, Peres & Bezerra. Inst. Micol. Univ. Recife. Public. 343:9, 1961.

Lesions numerous, small, hypophyllous, dark brown with conspicuous cream-whitish center, containing masses of cream to pale brown conidia, circular to irregular, coalescing to cause extensive necrosis of the leaf blade (Fig. 1). The upper surface of the leaf becomes chlorotic, yellowish, before blight occurs. – Mycelium consisting of branched, septate, olivaceous to pale brown hyphae 2.5–3.5 µm wide, immersed, intercellular, infecting initially the lower epidermic cells and the palisade tissue and often colonizing the entire

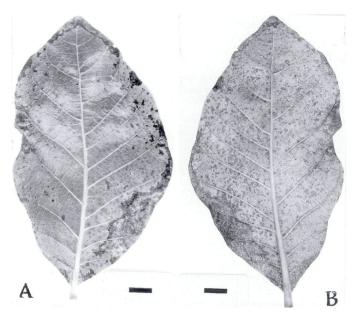
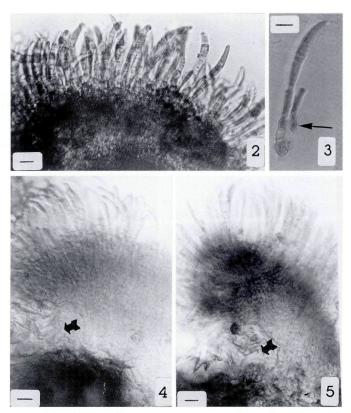


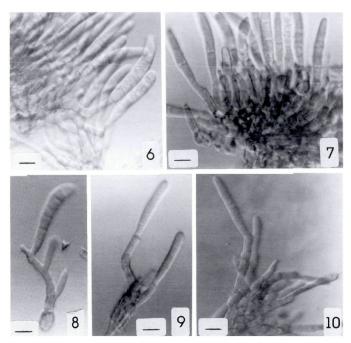
Fig. 1. Pseudocercospora aspidospermatis. – Lesions produced on the upper (A) and on the lower face (B) of a leaf of Aspidosperma macrocarpon. – Bar: 1 cm.

leaf parenchyma to reach the upper epidermis. - Conidiomata hypophyllous, sporodochial, dry, circular to irregular, subepidermal in origin, 36-160 µm diam. (Fig. 2), erumpent, 36-98 µm deep, containing an upper layer of ovoid to globoid cells which may be part of simple or branched conidiophores (Fig. 3); composed of pale brown thin-walled textura angularis 8 to 25-cells deep (Figs. 4 and 5). -Conidiophores short, 1-3 septate, light olivaceous, 15-20 µm long x 2.5-5 µm wide, simple or branched, terminating in 5-5.5 mm long, doliiform to ovoid cells, giving rise to the upper layer of the conidiomata which is comprised by the conidiogenous cells (Figs. 3, 6-11). - Conidiogenous cells integrated, terminal, erect, cylindrical, mostly straight, subhyaline to pale olivaceous, smooth, 7-15 µm long and 2.54 µm wide, with 1-3 enteroblastic percurrent and occasionally 1-2 holoblastic sympodial proliferations (Figs. 6-11). - Conidiogenous loci generally one to three per conidiogenous cell, flat, non-protuberant to slightly protuberant, unthickened



Figs. 2–5. Pseudocercospora aspidospermatis. – 2. Vertical median section of a sporodochium showing the general distribution of the conidia on the surface. – 3. A globoid cell from the surface of the sporodochium giving rise to a branched conidiophore with two terminal conidiogenous cells (arrow). – 4, 5. Two views of the erumpent textura angularis (arrows show host epidermis). – Bar: 10 μm.

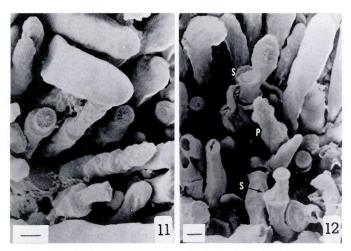
under the light microscope, 2–2.5 μm diam., terminal or lateral but revealing as slightly thickened when seen under the scanning electron microscope (Fig. 12). – Conidia 36–51 μm long and 4.5–8 μm wide, fusiform, slightly curved, smooth, pale olivaceous, formed holo-blastically, often at the apex of percurrently proliferated conidiogenous cells or being pushed aside in cells with sympodial



Figs. 6–10. Pseudocercospora aspidospermatis. – Samples of conidiogenous cells showing sympodial and percurrent proliferation as seen under the light microscope. – Bar: 10 μ m.

proliferation, (3-)5-6(-9)-euseptate, clearly guttulate when mature, with an obtuse apex; truncate unthickened base, 1.5-2.5 μ m (Fig. 13).

Specimens examined. – BRASIL: On leaves of Aspidosperma macrocarpon Mart., exsiccatum no. 15164, IMUR, Recife, Batista & Peres, coll. Heringer 15.9.1960, holotype; on leaves of A. macrocarpon, UB-3192 Coleção Micológica, Herbarium UB, Road to PADF, close to the Presídio da Papuda, Brasīlia, DF, J. C. Dianese, 18, II, 1993; on leaves of A. macrocarpon, UB-3334, Coleção Micológica, Herbarium UB, QI 4, Conj. 3 Lago Norte, Brasīlia, DF, J. C. Dianese, 7.3.1993; on leaves of A. macrocarpon, UB-3889, Coleção Micológica, Herbarium UB, Fazenda Roberto Ronald, Luziânia, Goiás, J. C. Dianese, 10.6.1993; on leaves of Aspidosperma dorycarpon A.D.C., Horto Florestal de Paraopeba, Minas Gerais, Leg. Dr. E. P. Heringer, 20, 6, 1960. Typus of Cercospora aspidospermatis Batista & Peres, 200078. IMUR.



Figs. 11–12. Pseudocercospora aspidospermatis. – 11. Conidiogenous cells showing percurrent proliferation as seen in the scanning electron microscope. – 12. Sympodially proliferated conidiogenous cells (S) showing unthickened scars (S and arrow) beside percurrently proliferated cells. – Bar: 1 µm.

Batista & al. (1961) described and illustrated Bactrodesmiella aspidospermatis Batista, Peres & Garnier as a new species of hyphomycete producing hyaline to pale brown 1-9-septate catenate or non-catenate conidia on a sporodochium topped by a layer of 60 μ m long conidiophores. According to our observations, the fungus does not form these long conidiophores but the conidiophores are at the most 20 μ m long on the surface of an erumpent, cream to light olivaceous stroma composed of a textura angularis. A portion of the stroma was apparently interpreted by Batista & al. (1961) as the continuation of the conidiophores.

Also the presence of catenate conidia, a characteristic of *Bactrodesmiella* M. B. Ellis, was never detected in the specimens examined and in the holotype. This fact associated with the presence of a large stroma covered by a layer of conidiogenous cells which proliferate mostly percurrently but also sympodially, does not allow for a classification of the fungus in *Bactrodesmiella* (Ellis, 1959; 1971). The fungus, however, could be placed in *Stigmina* Sacc. or in *Pseudocercospora* Speg.

Sutton & Pascoe (1989) discussed the complexity of the genus Stigmina and suggested its division in two groups: Stigmina stricto sensu would be applied to species associated with stomata and possessing "superficial and immersed mycelium, and percurrently proliferated verruculose, ragged conidiogenous cells". The other group would include among others those species with pulvinate sporodochia produced on leaves but retaining the percurrent proliferation as a characteristic of their smooth conidiogenous cells. Sympodial and percurrent proliferation of conidiogenous cells occurring simultaneously is not a main characteristic of Stigmina but it is recognized by Sutton & Pascoe (1989) as occurring in a few species. Also S. celastri (Kalchbr.) M. B. Ellis (Crous & Braun, 1994) cannot be placed in either of the groups mentioned. Observations under light microscope indicated percurrent proliferation to be frequent in B. aspidospermatis but simultaneous sympodial conidiogenesis occurred in most conidiogenous cells. The fungus is clearly not a member of Stigmina sensu Sutton & Pascoe (1989). which is a group segregating from a broad genus in need of a detailed revision which may eventually yield several new combinations into Pseudocercospora.

Deighton (1976, 1987) showed for the entire genus and recently Sutton (Dianese & al., 1993) for *Pseudocercospora punctata* (Wakefield) B. C. Sutton that the formation of a sporodochium containing conidiogenous cells that proliferate both percurrently and sympodially are major characteristics of *Pseudocercospora*. A similar interpretation was adopted by Sutton & al. (1987), and Sutton & Hodges (1990). In all cases it was shown that conidia formed by *Pseudocercospora* spp. always had unthickened basal scars, as seen in the light microscope and the same holds for scars on the conidiogenous cells. All these basic features related to conidiogenesis and morphology of conidia and conidiophores are present in *B. aspidospermatis* and support a new combination in *Pseudocercospora*. Therefore, the present species is transferred to *P. aspidospermatis*.

In the family Apocynaceae two *Pseudocercospora* species were previously reported: *Pseudocercospora apocynacearum* Gupta & al. (1987) described on *Iconocarpus frutescens* R. Br. (Apocynaceae) and *Cercospora aspidospermatis* Batista & Peres, on leaves of *Aspidosperma dorycarpon* A. D. C. (Batista & al., 1960). The first species differs from *P. aspidospermatis* by forming large infection spots (2 or more cm in diam.), not forming stromata, and producing longer and slender conidia (27-98 x 2.5-4.7 μ m) (Gupta & al., 1987).

The type material of C. aspidospermatis (pieces of leaves) was examined but it did not contain the fungus any longer. The leaf spots were larger, clearer, epiphyllous, and with better defined borders than those shown on leaves infected with P. aspidospermatis. Batista & al.

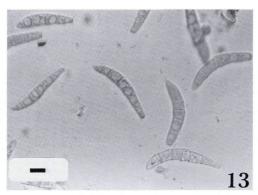


Fig. 13. Pseudocercospora aspidospermatis. – Multiseptate, guttulate conidia with unthickened truncate base. – Bar: 10 µm.

(1960) also showed C. aspidospermatis with 0-3 septate conidia completely different from the 3-9-septate conidia formed by P. aspidospermatis.

Acknowledgments

The authors acknowledge the assistance of Prof. Mariza Sanchez and Mr. Francisco Cesar Alves Santos. They also thank Dr. Leonor Maia, Curator, Herbário do Departamento de Micologia da Universidade Federal de Pernambuco. Dr. Pedro W. Crous (University of Stellenbosch, South Africa) is thanked for disclosing information contained in the the paper by Crous & Braun (1994) when still in press.

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(Manuscript accepted 11th April 1994)

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Digitale Literatur/Digital Literature

Zeitschrift/Journal: Sydowia

Jahr/Year: 1994

Band/Volume: 46

Autor(en)/Author(s): Dianese Jose C., Camara M. P. S.

Artikel/Article: Pseucdocercospora aspidospermatis - a new combination for

Bactrodesmiella aspidospermatis. 225-232